

Supplementary material

Feedback-regulated 3D-printed hydrogel micropatch for integrated therapy of fungal keratitis

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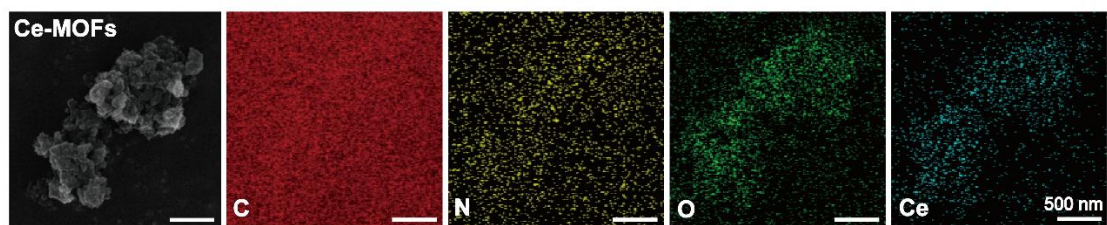
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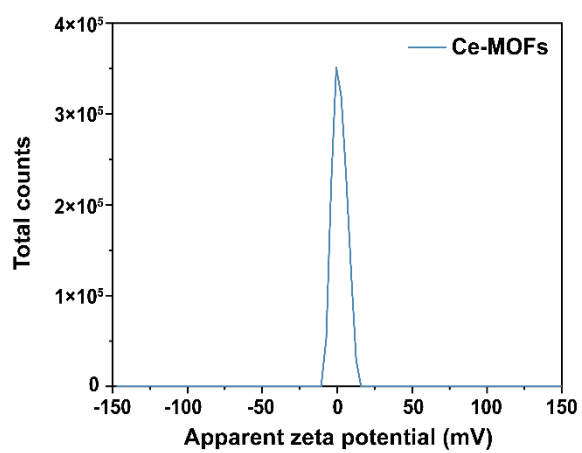
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Figure S1. Scanning electron microscopy (SEM) image and corresponding energy-dispersive spectroscopy (EDS) elemental mapping images of C, N, O, and Ce of cerium-based metal-organic frameworks (Ce-MOFs).



24 Figure S2. Zeta potential distribution map of Ce-MOFs.

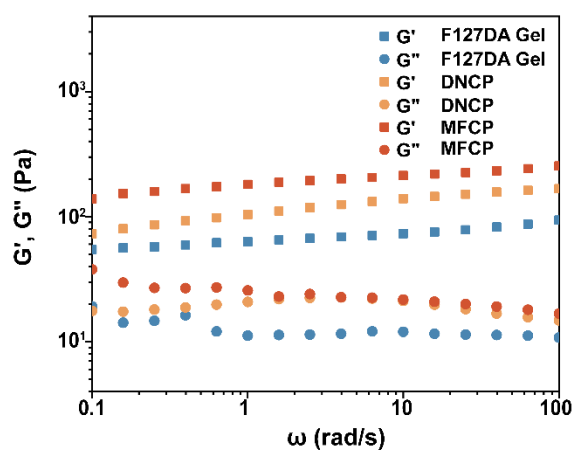
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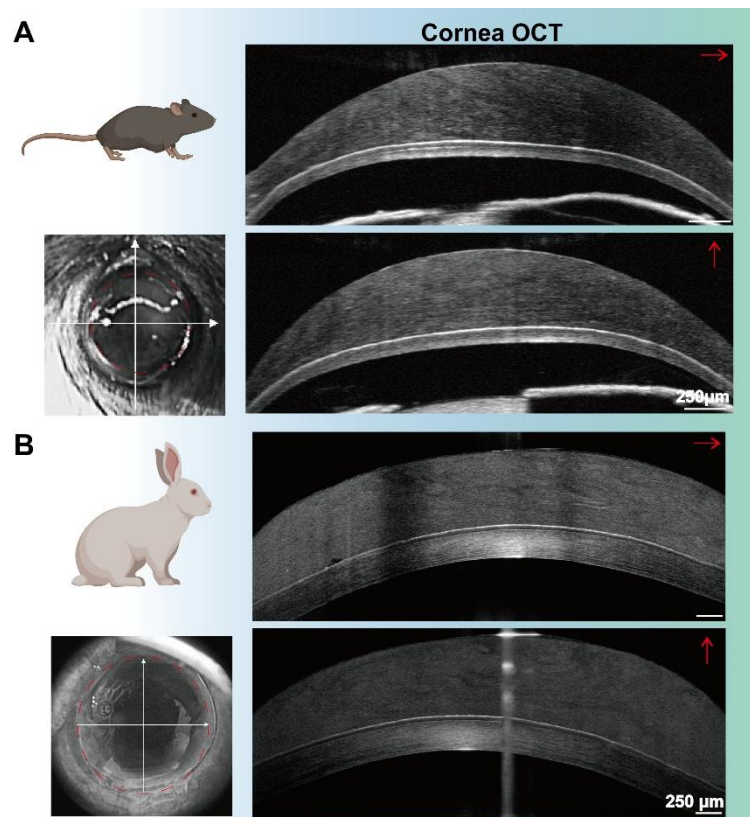
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28 Figure S3. The storage moduli (G') and loss moduli (G'') of F127DA-Gel, DNCP and MFCP.
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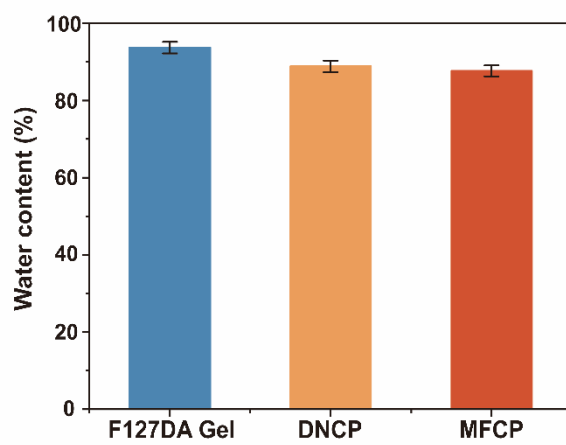


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Figure S4. Application of MFCP based on corneal curvature via 3D printing in different animal corneas. (A) Application on the C57BL/6J mouse cornea. (B) Application on the New Zealand white rabbit cornea.



38 Figure S5. The moisture content of F127DA Gel, DNCP and MFCP.
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Figure S6. Quantitative analysis of the live/dead staining fluorescence on HCECs and HCFs was performed using ImageJ software.

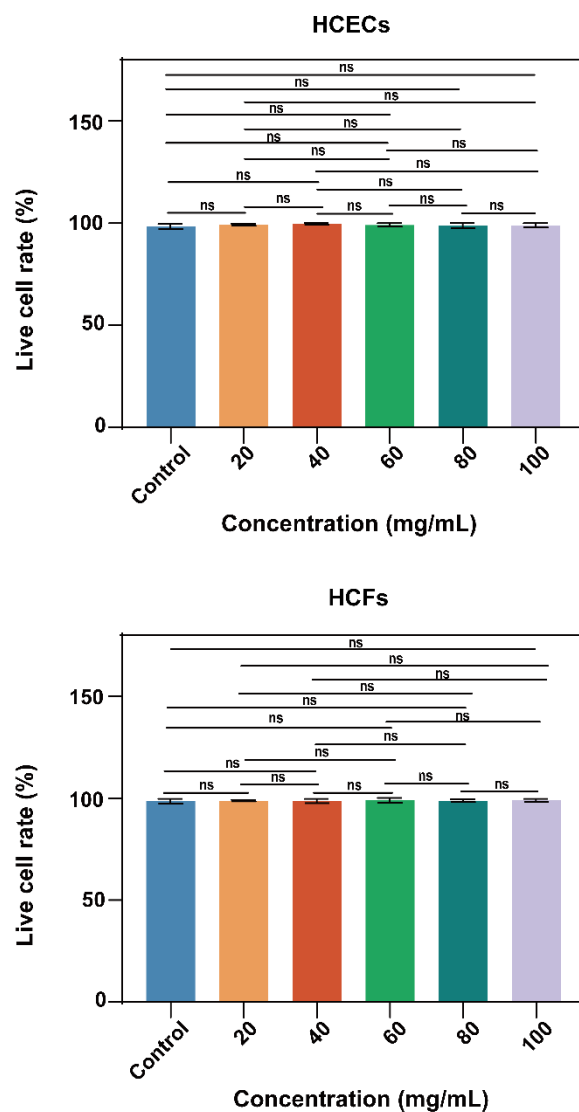


Figure S7. Quantitative real-time PCR was used to quantitatively analyze the mRNA expression levels of (A) α -SMA and (B) TGF- β_1 .

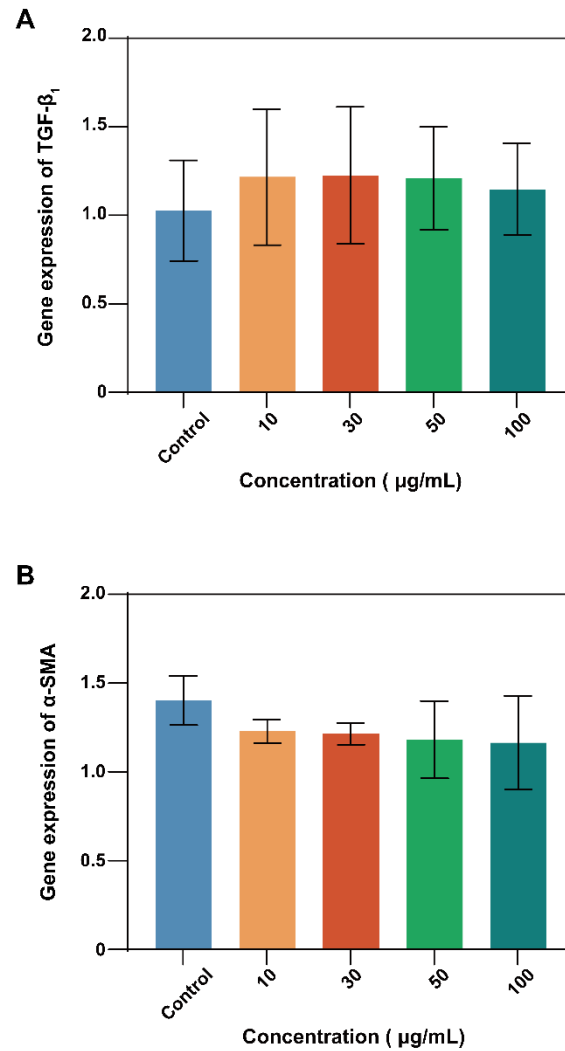


Figure S8. The bright-field image of the mouse cornea at 0, 24, and 48 h after treatment with PBS, 1% VCZ, GAT-MOFs, DNCP, or MFCP (n = 3).

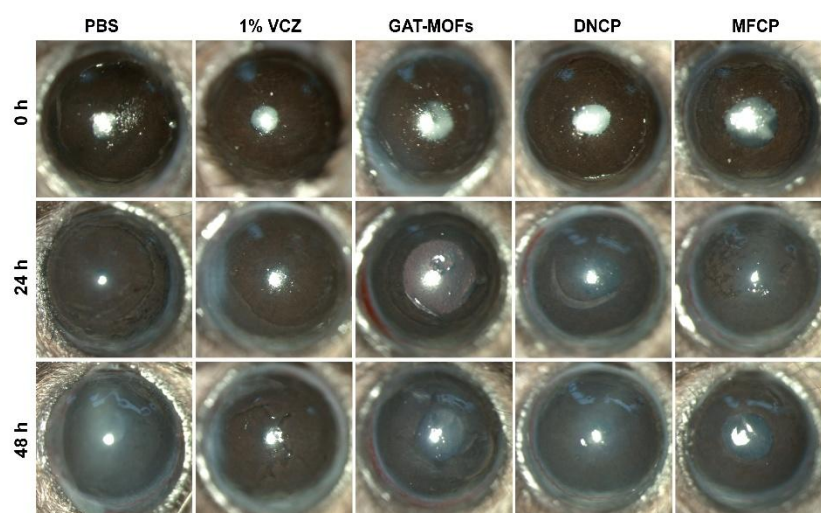


Figure S9. Quantitative analysis of ROS clearance fluorescence in HCECs and HCFs was conducted using ImageJ software.

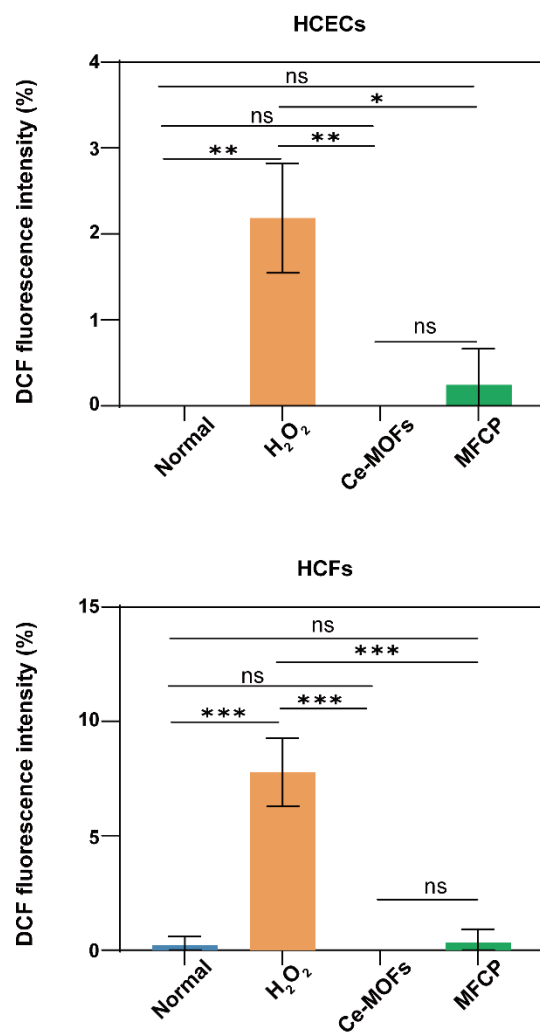


Figure S10. (A) Inhibition zone diameters against *S. epidermidis* and *S. aureus* for free GAT solution, (B) quantitative analysis of inhibition zone diameters between GAT and GAT-MOFs.

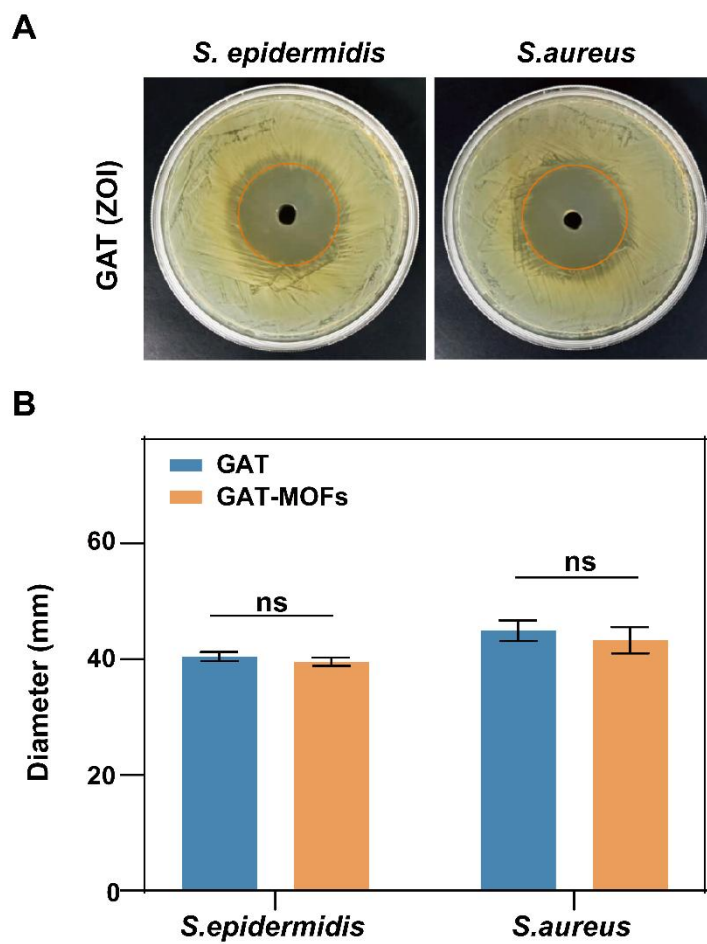


Figure S11. Fungal burden kinetics in MFCP-treated mouse corneas. Corneas were collected on days 1, 3, 5, and 7 post-infection (n = 3). (A) Images showing fungal colonies from the colony counting assay, accompanied by (B) statistical analysis of CFUs.

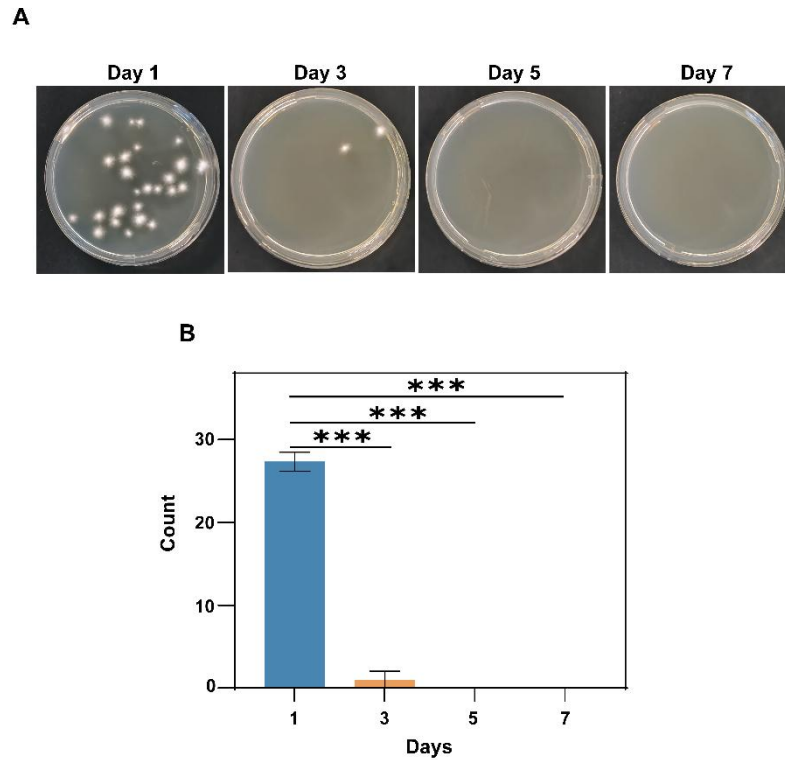


Figure S12. Fungal hyphal penetration depth in corneal stroma on day 3 post-infection.

