

## Supplementary Figure Legends

**Supplementary Figure 1.** Size distribution and Zeta potential of  $\beta$ -glucan and haemolytic tests for different concentrations of G-PL. **(A)** Process of G-PL synthesis. **(B)** Characteristic peaks corresponding to  $\beta$ -glucan and poly-L-lysine. **(C)**  $n = 3$ , biologically independent replicates. Data are presented as mean  $\pm$  SD. **(D)** A series of concentrations of G-PL was added to the erythrocytes. Haemolysis was assessed 1 h later. Haemolysis was observed when the G-PL concentration was higher than 1 mg/mL.

**Supplementary Figure 2.** **(A)** and **(B)** Activation of RAW264.7 by G-PL. G-PL-coated GL261 cells were co-cultured with RAW264.7 for 48 h, and the CD80 and CD86 expressed on macrophages were analysed by flow cytometry. **(C)** Co-culture of the splenocytes, DCs, and GL261 cells. A total of  $1 \times 10^5$  GL261 cells or PBS was co-cultured with  $1 \times 10^5$  DCs; 48 h later,  $1 \times 10^6$  splenocytes were added for further co-culture to monitor the development and cytotoxicity of GL261-specific T cells according to the apoptosis of GL261 and proliferation of T cells. In the PBS group, GL261 cells were not added to the co-culture system, and PBS solution was used instead. In the glucan group, 500  $\mu$ g/mL  $\beta$ -glucan was added to the co-culture supernatant. In the G-PL group, the GL261 cells were pretreated with 500  $\mu$ g/mL G-PL solution and then added to the co-culture system. Representatives of tumour cells that were alive are marked with arrows. Scale bar, 200  $\mu$ m.

**Supplementary Figure 3.** **(A)** The ratio of terminal fragment versus full length of Dectin-1 and the T cell subtypes after tumour-DC - T cells co-culture. DCs co-cultured with GL261 cells were collected at different time points after co-culture. GL261 cells were pretreated with 500  $\mu$ g/mL G-PL solution before co-culture in G-PL group. In the glucan group, the GL261 cells were not pre-coated with G-PL, but 500  $\mu$ g/mL  $\beta$ -glucan was added to the supernatant of the co-culture medium instead. **(B)** T cells were separated from the Tumour - DC - splenocytes co-culture system by using anti-CD3 magnetic beads. For the T cell subtype analysis, IFN- $\gamma$  and IL-17 expressed by CD4<sup>+</sup> T cells were used for Th1 and Th17 detection during flow cytometry analysis. **(C)** The average of three replicates was calculated. **(D)** and **(E)** WT and Dectin-1 KO mice were independently subcutaneously inoculated with  $1 \times 10^5$  ICC@G-PL, and the Th17 ratio in splenocytes was analysed 7 days later.

**Supplementary Figure 4.** **(A)** Xenograft mice were inoculated with 500  $\mu$ g G-PL or PBS (control) twice on days 0 and 7 following GL261 inoculation. No survival benefits were found. **(B)** Inguinal lymph nodes were extracted 48 h later to measure the DC maturation level based on CD86 expression. **(C)** Splenocytes were extracted on day 20 after inoculation and co-cultured with GL261 at a ratio of 10:1 for 3 days; the expression of IFN- $\gamma$  was further measured. **(D)**, **(E)**, and **(F)** Apoptosis of GL261 cells induced by gradient irradiation. GL261 cells were cultured and irradiated by a medical accelerator. Cell apoptosis was analysed to find the appropriate dose of

irradiation for ICC@G-PL construction by using Western Blot detection of the apoptosis marker, cleaved Caspase-3, and recording the growth of GL261 cells after gradient radiation exposure *in vitro* and *in vivo* based on cell counting and IVIS radiance, respectively. **(G)** and **(H)** Local inflammation induced by ICC@G-PL during footpad injection. ICC@G-PL cells ( $1 \times 10^6$  in 50  $\mu$ L of PBS) were inoculated into the footpad of mice, and the injection sites and draining lymph nodes were monitored 48 h later. Significant redness and swelling were observed at the injection site (footpad) after injection of ICC@G-PL, compared with the control group. Enlarged draining lymph nodes were observed. Left to right: PBS, ICC, ICC+ $\beta$ -glucan, ICC@G-PL.

**Supplementary Figure 5.** Example of the CBA analysis. Serum samples from the different groups of mice were diluted at a ratio of 1:100 and then analysed by CBA. The bead-conjugated antibodies of IL-6, IL-10, MCP-1, IFN- $\gamma$ , TNF- $\alpha$ , and IL-12 were labelled with PE fluorochrome at different intensities.

**Supplementary Figure 6.** Humoral immunity induced by ICC@G-PL vaccination. Serum samples from different groups of C57BL/6 mice were collected after an 8-week-long consecutive vaccination, and the titres of tumour-specific IgG1 and IgG2a were measured by ELISA. The endpoint titres were determined as the titre range at which the absorbance was twice the absorbance of the blank. \*:  $p < 0.05$ , \*\*:  $p < 0.01$ , \*\*\*:  $p < 0.0001$ . Data are represented as mean  $\pm$  SEM. Statistical significance was calculated by one-way ANOVA with Tukey's post-hoc test for multiple comparison correction. **(A)** Illustration of the experiment design. **(B)** Titration of IgG1 by serial dilution. **(C)** Titration of IgG2a by serial dilution.

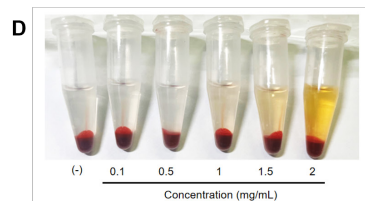
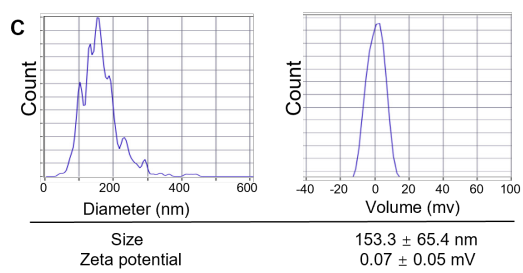
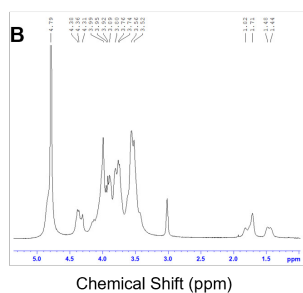
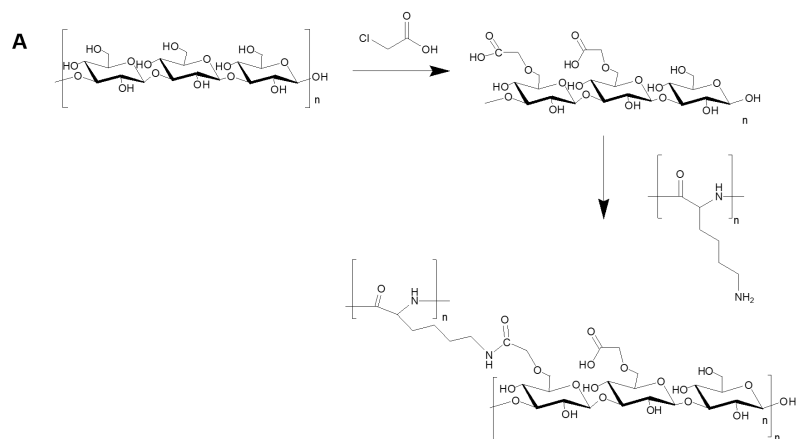
**Supplementary Figure 7.** Major organ damage screening. **(A)** H&E staining of the heart, kidney, spleen, lung, and liver. The organs of the vaccinated mice were collected for H&E staining 10 weeks after the first vaccination. No significant organ damage was observed after the ICC@G-PL vaccination. Scale bar, 200  $\mu$ m. **(B)** Serum levels of alanine aminotransferase (ALT), aspartate aminotransferase (AST), triglycerides, total cholesterol, uric acid, blood urea nitrogen (BUN), glucose, and amylase were measured.  $n = 3$  for each group.

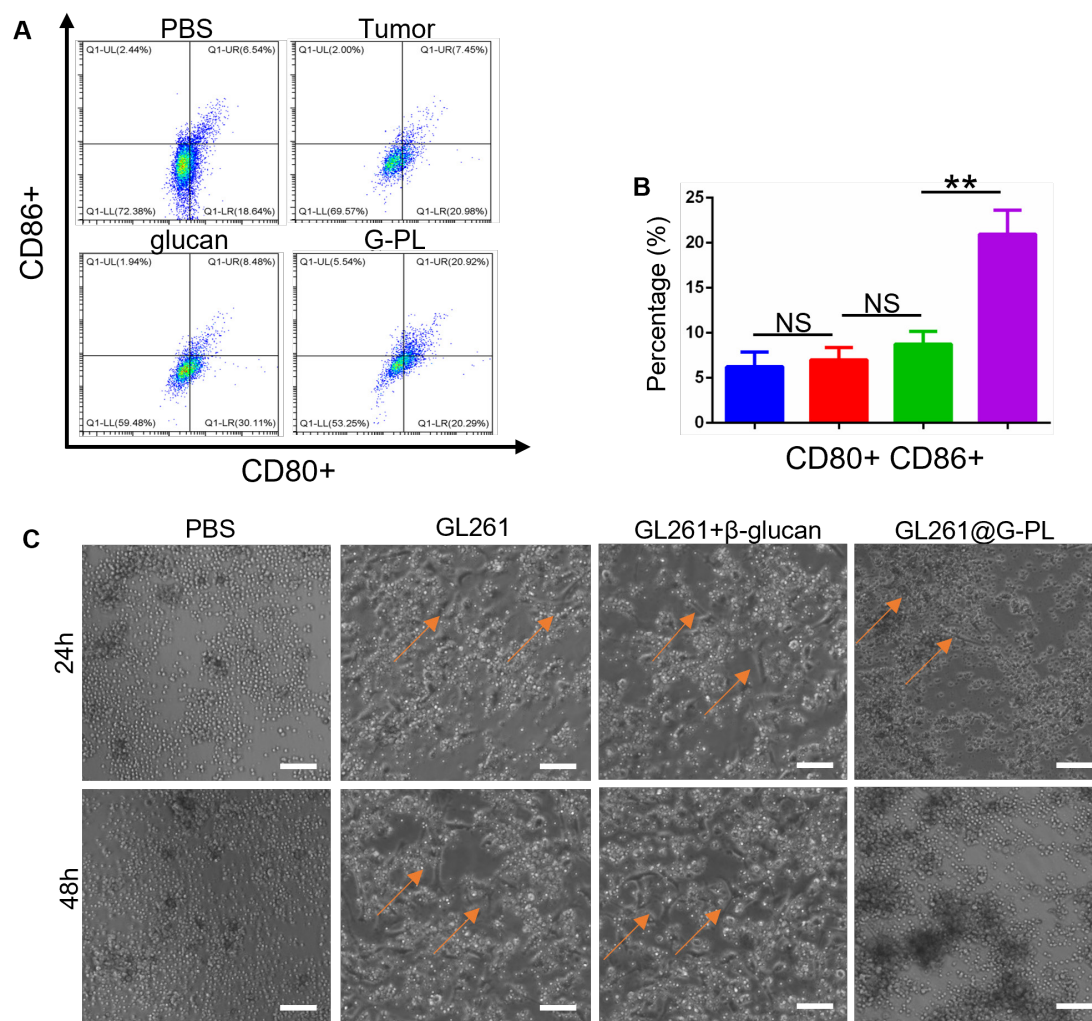
**Supplementary Figure 8.** Rechallenge of the treated mouse with GL261 cells. The mouse that underwent complete tumour relief in the therapeutic vaccine model was rechallenged by intracranial inoculation of  $1 \times 10^4$  GL261 cells. IVIS was conducted every 48 h starting on day 7, and the radiance was recorded. Control group,  $n = 3$ .

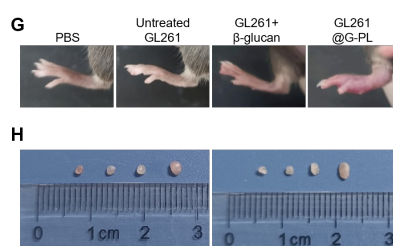
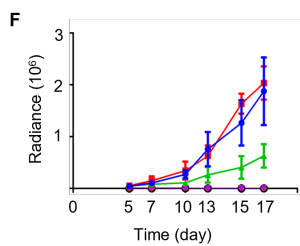
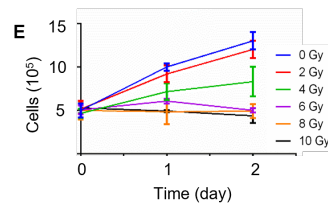
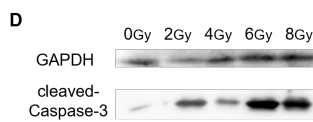
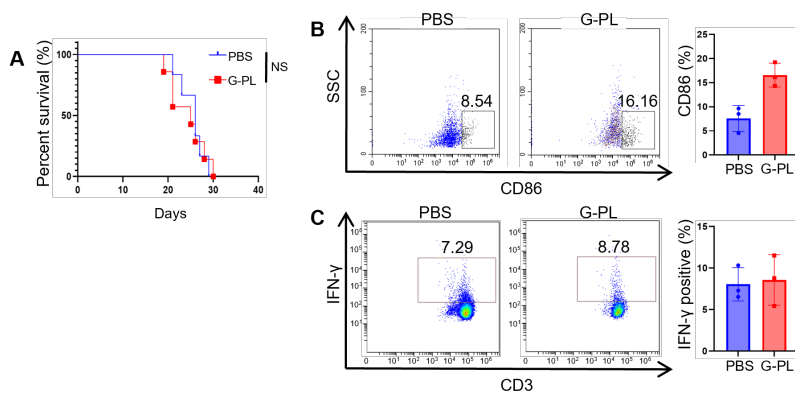
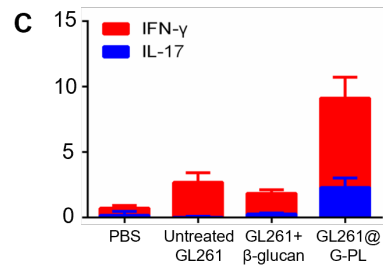
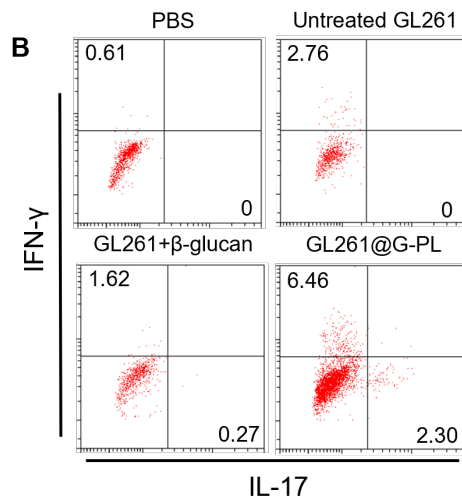
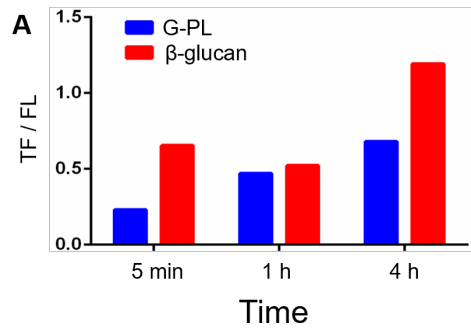
**Supplementary Figure 9.** **(A)** Flow cytometry gate strategy for tumour microenvironment analysis. The tumour specimens of the xenograft mice were collected and dissociated into single-cell suspensions. The live immune cells in the tumour were recognised by FVS620 $^-$  and CD45 $^+$ , and the cell subtypes such as T cells, macrophages, and neutrophils were further differentiated by characterised markers. **(B)**

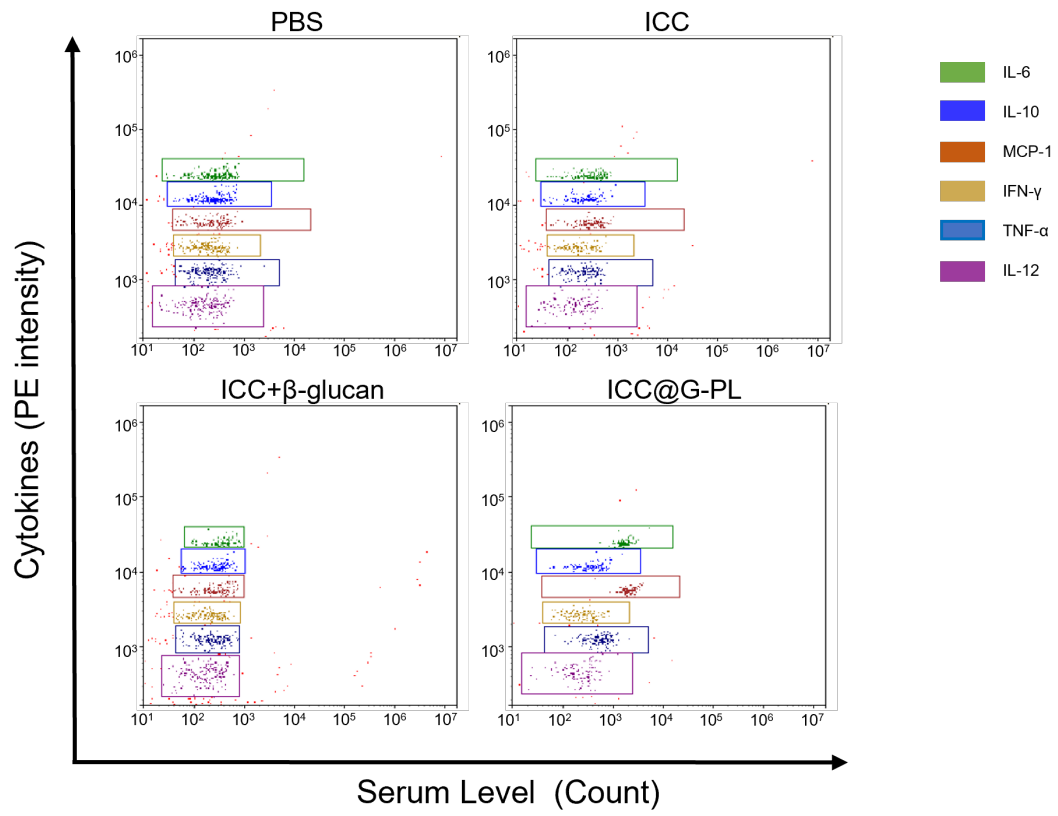
and (C) After therapeutic inoculation with ICC@G-PL, the GBM xenograft mice were intraperitoneally injected with 5 mg/kg anti-IL-17A every other day for 2 weeks, starting from the day of GL261 inoculation. For CD4<sup>+</sup> and CD8<sup>+</sup> T cell depletion, the mice were intraperitoneally injected with four doses of 200  $\mu$ g per mouse, twice a week. The infiltration of neutrophils and T cells was measured using immunofluorescence on day 20. The survival of the mice was observed in a parallel study.



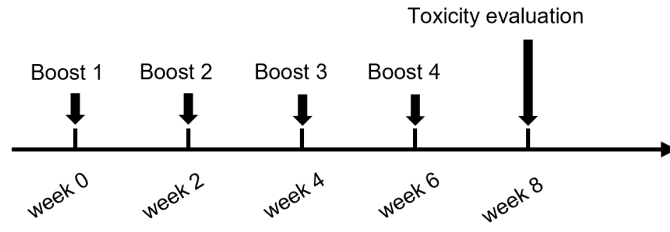
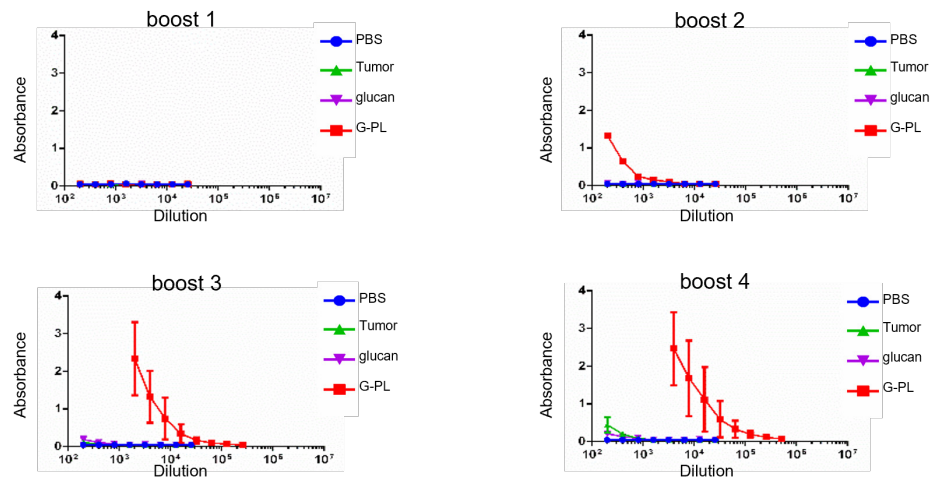










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