

Research Paper

CXCR4-directed endoradiotherapy with [¹⁷⁷Lu]Pentixather added to total body irradiation for myeloablative conditioning in patients with relapsed/refractory acute myeloid leukemia

Krischan Braitsch¹, Theo Lorenzini^{2,3}, Maike Hefter¹, Katrin Koch¹, Katharina Nickel¹, Jan C Peeken⁴, Katharina S Götze¹, Wolfgang Weber^{2,3,5,6}, Anne Allmann², Calogero D'Alessandria², Julia Brosch-Lenz², Florian Bassermann^{1,3,5,6}, Martina Rudelius⁷, Mareike Verbeek¹, Matthias Eiber^{2,3,6,#,✉}, Peter Herhaus^{1,#,✉}

1. Department of Internal Medicine III, School of Medicine and Health, Technical University of Munich, Munich, Germany.
2. Department of Nuclear Medicine, School of Medicine and Health, Technical University Munich, Munich, Germany.
3. German Cancer Consortium (DKTK), partner site Munich, a partnership between DKFZ and University Hospital rechts der Isar (MRI), Germany.
4. Department of Radiation Oncology, School of Medicine and Health, Technical University Munich, Munich, Germany.
5. Center for Translational Cancer Research (TranslaTUM), Technical University of Munich, Munich, Germany.
6. Bavarian Cancer Research Center (BZKF), Munich, Germany.
7. Ludwig-Maximilians-Universität München, Institute of Pathology, Munich, Germany.

#equal contribution

✉ Corresponding author: Peter Herhaus, MD, Department of Internal Medicine III, School of Medicine and Health, Technical University of Munich, Ismaninger Str. 22, 81675 Munich, Germany, peter.herhaus@tum.de.

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Abstract

Rationale: Despite recent advances in the targeted therapy of AML, the disease continues to have a poor prognosis. Allogeneic hematopoietic stem cell transplantation (alloSCT) remains to be the curative therapy option for fit patients with high-risk disease. Especially patients with relapsed or refractory (r/r) AML continue to have poor outcomes. Myeloablative total body irradiation (TBI) based conditioning can be used in AML patients refractory to multiple lines of standard therapy, but the optimal conditioning regimen remains unclear for patients considered to be chemotherapy- refractory. Feasibility of C-X-C-motif chemokine receptor 4 (CXCR4)-directed endoradiotherapy (ERT) has previously been demonstrated in AML patients with CXCR4 expression on leukemic blasts.

Methods: Here, we report on a small cohort of seven AML patients refractory to multiple lines (range 3-7) of therapy, who received CXCR4-directed ERT with [¹⁷⁷Lu]Pentixather in combination with TBI and chemotherapy prior to alloSCT. We report outcomes with a focus on toxicity, engraftment, the impact on the bone marrow (BM) niche and efficacy.

Results: In this intensively pre-treated group of patients, promising response (6 out of 7 patients) and engraftment (6 out of 7 patients) rates were observed. Histopathological analysis showed that niche compartments are spared and allow for engraftment to occur despite the combined ERT and TBI conditioning.

Conclusion: To the best of our knowledge, we report on the first seven patients who received CXCR4-directed ERT in sequential combination with TBI and chemotherapy, providing an effective, individualized conditioning regimen for intensively pre-treated r/r AML patients.

Keywords: acute myeloid leukemia, CXCR4, allogeneic stem cell transplantation, endoradiotherapy, conditioning regimens

Introduction

In recent years, therapeutic advances for acute myeloid leukemia (AML) were observed [1-4], but allogeneic hematopoietic stem cell transplantation (alloSCT) remains standard of care for fit patients

with high-risk disease [5]. However, relapsed or refractory (r/r) AML patients undergoing alloSCT have a poor prognosis, and relapse remains the main cause of AML treatment failure [6]. Although a second alloSCT after relapse may achieve long-term remissions in selected AML patients, treatment options for patients with relapsed and uncontrolled AML are extremely limited [7]. The increased risk of relapse after transplant is in part attributed to disease biology but also to incomplete eradication of malignant cells by induction, salvage and conditioning therapy [8, 9]. Myeloablative conditioning (MAC) using total body irradiation (TBI) based protocols can be used in AML patients refractory to multiple lines of standard chemotherapy or in patients with AML relapse after first alloSCT who are considered chemotherapy-refractory. However, relapse rates are similar to those of other MAC regimens [10].

C-X-C-motif chemokine receptor 4 (CXCR4) is a G-protein coupled transmembrane receptor that regulates pivotal processes in hematopoiesis, immune response, embryogenesis and stem cell homing, together with its natural ligand CXCL12 [11-14]. Physiologically, CXCR4 is abundantly expressed in the hematopoietic system, especially in hematopoietic stem cells [15]. In the context of malignancy, CXCR4 overexpression is commonly found in various types of cancer promoting proliferation and metastasis and mediating resistance to chemotherapy [16-20]. The bone marrow (BM) niche plays an essential role in protecting healthy hematopoietic stem cells as well as leukemic blasts from cytotoxic stress. The CXCR4/CXCL12 axis is believed to mediate a protective microenvironment [17, 21]. The protective niche is thought to be an important factor in disease relapse occurrence and mediation of drug resistance [22].

Radiolabeled peptides targeting CXCR4, such as [⁶⁸Ga]Pentixafor and [¹⁷⁷Lu]Pentixather were developed for diagnostic positron emission tomography (PET) and endoradiotherapy (ERT), respectively [23, 24]. [⁶⁸Ga]Pentixafor enables *in vivo* molecular CXCR4 imaging in a subset of AML patients [25]. Pentixather is coupled with beta minus emitting radionuclides, like ¹⁷⁷Lutetium or ⁹⁰Yttrium which cause localized radiation damage within a few millimeters around the binding site. ERT of the CXCR4/CXCL12 axis is especially promising in refractory AML patients undergoing alloSCT, as it can destroy malignant cells directly but also dislocate them from their protective niche, rendering them more vulnerable to sequential or combined therapy approaches [26]. The feasibility of CXCR4-directed ERT has previously been demonstrated in AML patients with CXCR4 expression on leukemic blasts

[27]. Naturally, hematological toxicity is a major concern of CXCR4-directed ERT, as it directly targets hematopoietic stem cells in the BM and the supporting niche by the “cross-fire effect”. In the context of alloSCT, the hematological toxicity itself is part of MAC and is bypassed by the stem-cell rescue, but damaging the hematopoietic niche may lead to engraftment failure or prolonged cytopenia [28].

Methods

Patient selection

Seven patients with r/r AML were treated at our center between February 2019 and October 2023. ERT with [¹⁷⁷Lu]Pentixather was offered as compassionate use and in compliance with §37 of the Declaration of Helsinki. Written informed consent was obtained from all patients prior to treatment.

[⁶⁸Ga]Pentixafor PET/CT imaging

All patients underwent [⁶⁸Ga]Pentixafor PET/computed tomography (CT) imaging to confirm *in vivo* CXCR4 expression on leukemic blasts. Increased tracer uptake of AML lesions as compared to normal tissue was visually determined by a board-certified nuclear medicine physician. Only patients with significantly visually increased tracer retention were considered eligible for ERT. Representative CXCR4 PET images of each patient are shown in Figure S1.

Pre- and post-therapy dosimetry

Radiolabeling procedures of [¹⁷⁷Lu]Pentixather are described in the supplemental material. All patients underwent pre-therapeutic dosimetry as follows: after intravenous injection of standard amounts of activity of 200 MBq (mean 198 ± 9.7 MBq, range 182-213 MBq) of [¹⁷⁷Lu]Pentixather, whole-body scintigraphy or quantitative single photon emission (SPECT)/CT imaging was performed at 1 hour, 4 hours, 24 hours, 48 hours, and 7 days (d) post injection (p.i.). Post-therapeutic imaging was performed at the same time points following intravenous injection of the therapeutic activity of [¹⁷⁷Lu]Pentixather. For whole-body scintigraphy, each scan was obtained at a speed of 12 cm/min on a dual-headed SYMBIA T6 (Siemens Medical Solutions, Erlangen, Germany) equipped with 9.5 mm NaI(Tl) crystals and medium-energy low-penetration collimators. A 12% and a 20% energy window were placed around the 208 keV and 113 keV photo-peaks of ¹⁷⁷Lu, respectively. SPECT imaging was performed with 32 steps per head with 40 s acquisition duration per step using the 208 keV photo peak (12% width with upper and lower scatter windows with 15% and 10% width,

respectively). A low dose CT was simultaneously acquired for attenuation and scatter correction. SPECT images were reconstructed in a 128x128 matrix using Flash 3D with 8 iterations and 4 subsets using CT-based attenuation and scatter correction. A camera- and isotope-specific calibration factor was applied to convert the images from counts to units of Bq.

For planar dosimetry, an extracorporeal background region of interest (ROI) was drawn for the liver and its content was scaled according to the size of the individual organ and subtracted. For the kidneys, the ROI was placed in the thigh. For the BM, the absorbed dose was determined by delineating the L2-L4 lumbar vertebrae following the guideline of the European Association of Nuclear Medicine (EANM) and the values were scaled accordingly [29, 30]. The ROI for background subtraction was positioned adjacent to the BM. Following the administration of [¹⁷⁷Lu]Pentixather and immediately before each planar whole-body scintigraphy, a probe counter measurement was conducted to normalize the counts to the residual activity in the patient's body at each time point. Count rates for each ROI were extracted using the open-source DICOM software OsiriX (version 5.1, 64-bit, Pixmeo, Geneva, Switzerland), and the geometric mean of the anterior and posterior counts for each ROI was calculated. The conversion of counts to activity was achieved through normalization using the probe measurements.

For dosimetry calculation with SPECT/CT, segmentations of volumes of interest (VOIs) were performed in MIM v7.2.8 software using their artificial intelligence (AI)-based segmentation model for CT images. Kidneys and liver were segmented using the AI-segmentation method and VOIs were copied to the co-registered SPECT images across imaging time points. For BM, three spherical VOIs were placed in the lumbar vertebrae L2 to L4 and total red BM was calculated above following the EANM guidelines [30].

Total organ activities were extracted and time-activity curves were fitted using either a mono- or bi-exponential fit function in MATLAB. Organ absorbed doses were calculated using phantom organ values from OLINDA/EXM v1.0 for whole-body scintigraphy and IDACDose 2.1 from the International Commission of Radiation Protection for SPECT/CT dosimetry using the organ time-integrated activities and patient-individual organ masses [31].

The results of the pre-therapeutic dosimetry were used to personalize the therapeutic amount of radioactivity of [¹⁷⁷Lu]Pentixather based on scaling the absorbed dose per unit of administered activity

(Gy/MBq) to organs at risk (maximum of 23 Gy to the kidneys and/or a maximum of 1 Gy to BM at the time of transplantation), yielding the amount of activity to be administered during therapy.

Post-therapeutic imaging was performed using the same protocols following intravenous injection of the therapeutic activity of [¹⁷⁷Lu]Pentixather, using either whole-body scintigraphy or quantitative SPECT/CT imaging, followed by dosimetry calculations using the same approaches as described above.

Treatment regime

Conditioning prior to alloSCT consisted of [¹⁷⁷Lu]Pentixather (mean activity of 12.4 ± 2.7 GBq, range 7.6-16.1 GBq) ERT on d-15 or d-14; TBI 8-10 Gy, depending on patient fitness and age, on d-9 to d-7 and chemotherapy, based on donor type (Figure 1).

ERT was performed in the department of nuclear medicine according to German Radiation Protection regulations. In addition, a 1000 mL saline solution containing 2% L-arginine and L-lysine was intravenously administered starting 30 minutes before ERT and continued for a total duration of 4 hours, as is commonly performed in patients with neuroendocrine tumors undergoing peptide receptor radionuclide therapy and has been adapted in prior studies with [¹⁷⁷Lu]Pentixather [32, 33].

TBI was performed with twice-daily fractions of 2 Gy up to a desired dose of 8 (n=3) to 10 Gy (n=4). Patients were treated with a linear accelerator positioned supine and treated with lateral fields using 6-MV photons or with a tomotherapy system using intensity-modulated radiotherapy. The lungs were attenuated in all patients to a median dose of 8 Gy using the arms and brass compensators.

Chemotherapy regimens were as follows: fludarabine 30 mg/m² d-5 to d-2 or cyclophosphamide 60mg/m² d-5 and d-4 for matched unrelated donors (MUD) and matched related donors (MRD); fludarabine 30 mg/m² d-6 to d-2 and cyclophosphamide 14.5 mg/m² d-6 to d-5 for haploidentical donors (HID) or mismatched unrelated donors (MMUD). For patients with matched donors, immunosuppression consisted of rabbit antithymocyte globulin (5 mg/kg for MRD and 10 mg/kg for MUD) d-3 to d-1, mycophenolatmofetil and a calcineurin inhibitor. For HID and MMUD, post-transplant cyclophosphamide 50mg/kg on days 3 and 4, mycophenolatmofetil and a calcineurin inhibitor were used according to standard of care.

Response assessment

AML response assessment was done by BM biopsy during routine follow-up according to

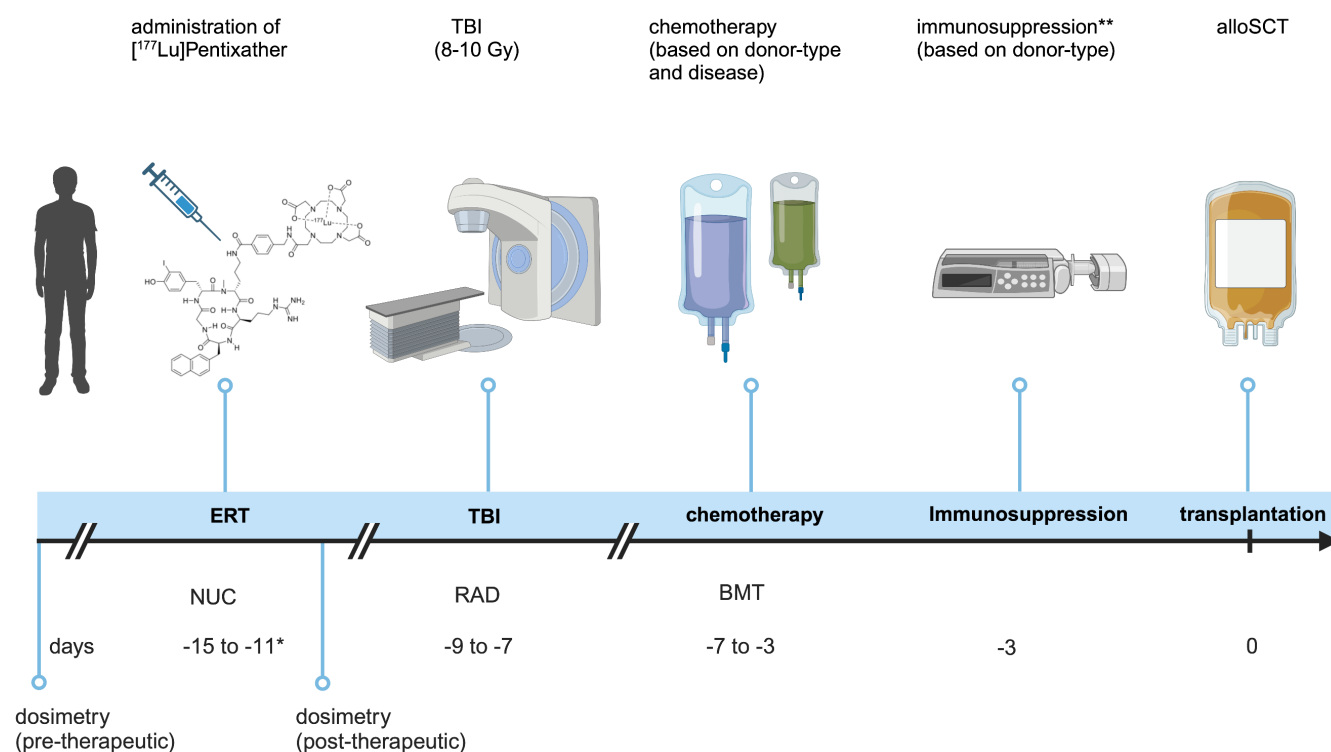


Figure 1. Treatment schedule. Schematic overview of the conditioning regimen. *Duration of in-patient stay varied based on German Radiation Protection Rules; **For patients with HID/MMUD, immunosuppression was started on day +3 instead. TBI = total body irradiation; alloSCT = allogeneic hematopoietic stem cell transplantation; NUC=nuclear medicine; RAD = radiation oncology; BMT=bone marrow transplantation. (Created in BioRender).

standard of care as described in the European LeukemiaNet (ELN) recommendations [5]. Patients with extramedullary disease were either staged by biopsy or by fluorodeoxyglucose (FDG)-PET-CT. Chimerism analysis was done by digital droplet polymerase chain reaction from the PB.

Multispectral imaging

After antigen retrieval, immunofluorescence was performed using an Opal 6-Plex Detection Kit (Akoya Biosciences, Marlborough, MA, USA) according to the manufacturer's protocol. The following antibodies were used MPO (Agilent DAKO, Santa Clara, CA, USA), CD34 (Agilent, DAKO), E-cadherin (Cell Marque, Darmstadt, Germany), CD42b (Agilent, DAKO), CD3 (Agilent, DAKO) and CD20 (Cell Marque). A PhenoImager (Akoya Bioscience) was used for multispectral imaging.

Results

Patient characteristics

Median patient age was 46 (range 42-57) years. Five patients had de novo AML and two had secondary (s)AML. Patients had previously received a median of four (3-7) lines of intensive therapy, including alloSCT in four patients (Table S1). Two patients were primary refractory and five patients relapsed after achieving a complete remission (CR).

Prior to conditioning, one patient achieved morphologic leukemia-free state (MLFS) after salvage chemotherapy, all other patients were refractory and had active AML at the beginning of conditioning. Two patients had relapsed with extramedullary disease. Median hematopoietic cell Transplantation-Comorbidity Index (HCT-CI) Score was 1 (0-5) and at the start of conditioning, five patients had severe neutropenia with an absolute neutrophil count of $<500/\mu\text{l}$ and four patients required broad-spectrum antibiotics at initiation of conditioning for uncontrolled infection. Median baseline creatinine was 0.8 mg/dl (0.4-1.1), and baseline bilirubin was 0.8 mg/dl (0.3-1.1). All patients received a peripheral blood (PB) stem cell graft with a median of 5.39×10^6 (4.5-10.3) CD34+ cells/kg. Four patients received grafts from matched donors (3MUD and 1MRD = 4 matched), two from HIDs and one from a MMUD (Table 1).

CXCR4 PET imaging

In vivo CXCR4 expression was confirmed by [^{68}Ga]Pentixafor PET/CT in BM or extramedullary lesions (Figure 2) and patients with significantly visually increased tracer retention were considered eligible for ERT. Maximum standardized uptake value (SUVmax) was increased in skeletal system and was measured in the representative regions as

follows: skull base 7.64 (range 4.61-12.47, not available for 3 patients), fourth cervical vertebra (C4) 5.19 (1.13-10.72), fifth thoracic vertebra (T5) 6.55 (1.78-14.42); fifth lumbar vertebra (L5) 6.93 (1.77-15.78) and left and right ilium 6.25 (2.31-10.74) and 6.67 (2.50-12.78), respectively. Of note, two patients (patient 5 and patient 7) had primary extramedullary disease, with a mean SUVmax of 1.90 (1.13-2.50) and 5.08 (2.93-6.09) across all measured sites.

Table 1. Patient Characteristics. Table showing the baseline characteristics of the seven included patients. *Treatment lines were defined when a regimen change occurred due to relapse or progress of underlying disease. sAML = secondary acute myeloid leukemia; ELN = European LeukemiaNet; MUD = Matched unrelated donor; MRD = Matched related donor; MMUD = Mismatched unrelated donor; HID = Haploidentical Donor; MLFS = Morphologic leukemia-free state; RD = refractory disease.

Parameter	Value
Age, median (range)	46 (42-57)
Sex, n (%)	
male	4
female	3
AML type, n	
denovo	5
sAML	2
ELN 2022 risk group, n	
favorable	2
intermediate	3
adverse	2
Previous treatment lines*, median (range)	4 (3-7)
Previous alloSCT, n	4
Donor type, n	
MUD	3
MMUD	1
MRD	1
HID	2
Prior remission, n	
MLFS	1
RD	6
HCT-CI Score, median (range)	1 (0-5)

Dosimetry and conditioning

Radiolabeling details and methods for ¹⁷⁷Lu]Pentixather can be found in Supplemental Material (Supplemental Methods and Table S2).

Pre-ERT dosimetry was available for all patients. The median kidney absorbed dose across patients was 1.44 ± 0.70 Gy/GBq (range 0.73-2.97), the median liver absorbed dose was 0.40 ± 0.25 Gy/GBq (0.16-0.92), and the median BM absorbed dose was 0.42 ± 0.12 Gy/GBq (0.01-0.57). Post-therapy dosimetry was only available for 4 out of 7 patients. The median kidney absorbed dose across patients was 0.78 ± 0.22 Gy/GBq (range 0.68-1.21), the median liver absorbed dose was 0.34 ± 0.07 Gy/GBq (0.20-0.36), and the BM absorbed dose was 0.22 ± 0.12 Gy/GBq (0.10-0.41) (Table 2).

Consequently, the potential therapeutic activity of ¹⁷⁷Lu]Pentixather was limited either to a maximum of 23 Gy renal dose and/or a maximum of 1 Gy to BM at the time of transplantation (Figure 3). The mean injected therapeutic activity of ¹⁷⁷Lu]Pentixather was 12.4 ± 2.7 GBq (range 7.6-16.1). No immediate reactions or adverse effects were observed during the obligatory 48-hour stay in the nuclear medicine ward post ERT as required by German Radiation Protection Rules. Patients were subsequently transferred to the SCT unit to receive TBI and chemotherapy.

Table 2. Pre- and post-therapeutic dosimetry. Estimated doses from pre- and post-therapeutic dosimetry delivered to indicated organs and the final injected activity. Post-therapeutic dosimetry was only available for 4/7 patients. Indicated doses are for ERT and should be considered as adjunctive to the sequential TBI. n.a.=not available, BM = bone marrow. *derived from SPECT-imaging

Patient	Kidney-Dose [Gy/GBq] (pre / post)	Liver-Dose [Gy/GBq] (pre / post)	BM-Dose [Gy/GBq] (pre / post)	Injected activity [GBq]
#1	1.59 / 0.87	0.29 / 0.20	0.42 / 0.14	12.0
#2	1.44 / 1.21	0.40 / 0.36	0.57 / 0.41	14.4
#3	1.73 / n.a.	0.47 / n.a.	0.47 / n.a.	13.0
#4	1.18 / 0.68	0.66 / 0.32	0.44 / 0.30	11.4
#5	0.73 / n.a.	0.16 / n.a.	0.11 / n.a.	12.2
#6	1.30 / 0.68	0.38 / 0.36	0.16 / 0.10	16.1
#7	2.97* / n.a.	0.92* / n.a.	0.01* / n.a.	7.6

Toxicity

ERT, TBI and chemotherapy were generally well tolerated. No tumor lysis syndrome was observed during the conditioning period and no unexpected adverse events occurred. All patients developed fever and received broad-spectrum antibiotic and antifungal treatments during their hospitalization. Blood-stream infections were common and occurred in all seven patients and CT findings of pneumonia were seen in six patients. Mucositis II°-III° occurred in all patients. Four patients required intensive care unit (ICU) treatment due to infectious complications with hypotension or respiratory failure, two of these four patients required invasive ventilation during ICU stay. ICU admission occurred after a median 19 days (15-24) from the start of conditioning. An increase in serum creatinine of ≥ 1.5 folds was observed in three patients after conditioning therapy, in all three cases creatinine elevation was attributed to infectious complications. Among those patients, two required hemodialysis during their ICU stay. Hepatotoxicity was common, five patients developed I/II° and two patients III° hyperbilirubinaemia.

Three patients died during hospital stay after a median of 41 (30-83) days following conditioning. Among these patients, two died in the ICU due to

septic shock (n=1) and acute respiratory distress syndrome (n=1) and one patient had refractory AML and died with palliative support. All deaths were considered to be either transplant-related or disease-related but not specific to ERT.

Response and engraftment

One patient died prior to formal response assessment but achieved MLFS at day 15 after alloSCT. Five patients achieved complete remission with incomplete count recovery (CRi) and one patient was refractory and regenerated with 11% blasts in the PB at day eleven. Two patients with primarily extramedullary disease were either staged by FDG-PET-CT (for extramedullary disease) or by biopsy (for known chloroma of the skin). Both patients achieved complete remission. The recipient chimerism at or after day 28 was <1% (0.18% (0%-0.89%)) in all responding patients. Time to leukocyte recovery in the responding patients was 23 (12-28) days and platelet engraftment occurred at day

35 (12-55) in median, one patient had received a stem cell boost from the same donor after initial graft failure. Patients that were discharged had been hospitalized for a median of 41 days (35-89) for the transplantation period.

To further assess whether engraftment was impaired by the intensified conditioning regimen, multispectral imaging of exemplary BM sites biopsied before, during and after therapy was performed for patients with available samples. Before conditioning, a compact infiltrate of CD34-positive myeloblasts in a BM biopsy from a patient with AML with reduced trilineage hematopoiesis and few intermingled lymphocytes was seen. Following CXCR4-directed ERT, TBI and chemotherapy, hematopoiesis was markedly reduced and no residual blasts were detected. After engraftment, recovery of the BM microenvironment was observed with a normal distribution of hematopoiesis and the hematopoietic niche (Figure 4).

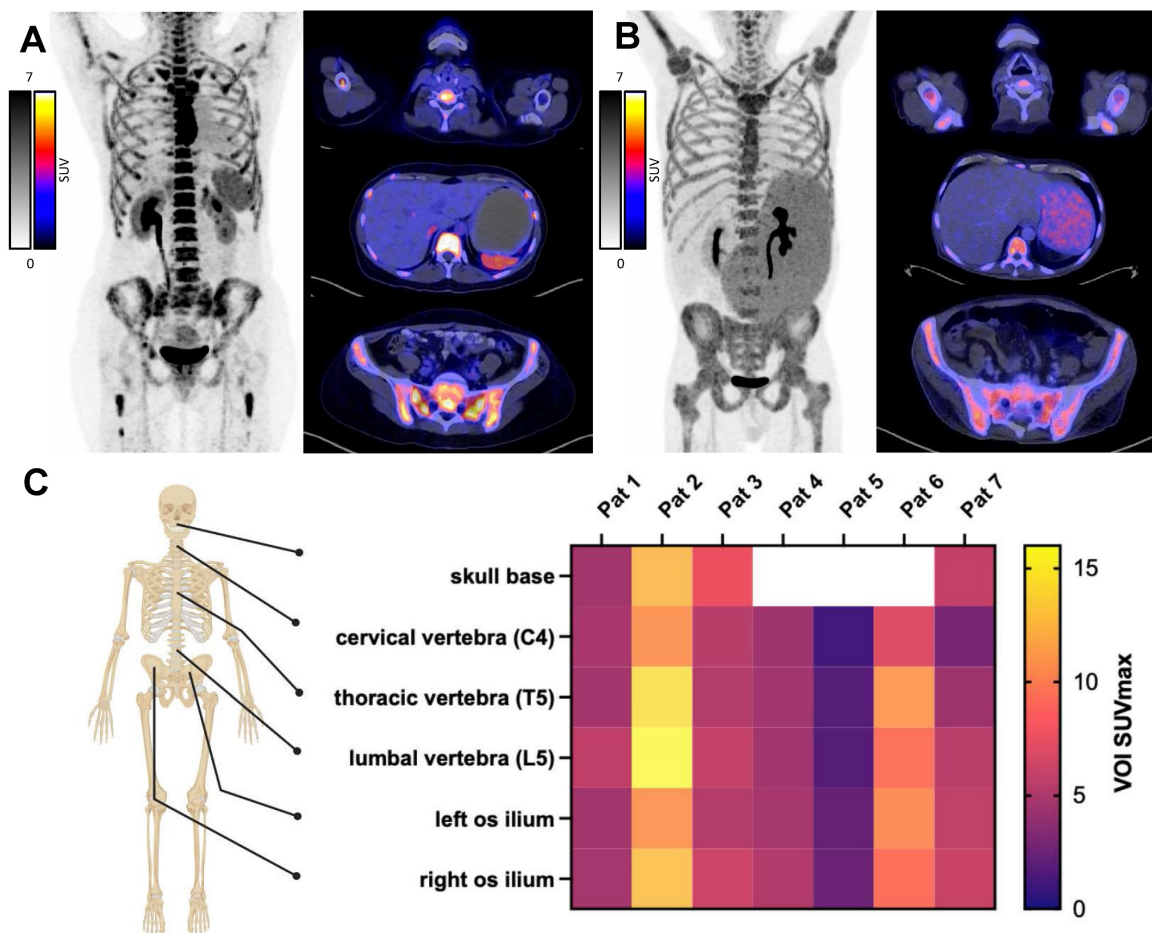


Figure 2. In vivo CXCR4 PET Imaging. Pre-therapeutic PET/CT with the CXCR4 ligand [⁶⁸Ga]Pentixafor to confirm *in vivo* CXCR4 expression. Patient examples with high (A) and moderate (B) CXCR4 expression on PET-imaging. C) SUVmax from pre-therapeutic CXCR4 imaging, measured at indicated sites. Patients 5 and 7 exhibited CXCR4 positive extramedullary disease. (Created in BioRender).

Follow-up

Both patients with extramedullary only disease at the time of alloSCT had relapsed after three months. One patient had hematologic relapse, the other had isolated central nervous system (CNS) relapse. Both patients went to palliative care and died shortly after.

Two years after alloSCT, two patients were alive. One had developed CNS relapse twelve months after alloSCT but was successfully treated with donor lymphocyte infusions (DLI), 5-Azacitidine and radiotherapy. More than two years after alloSCT, the same patient developed pneumocystis jirovecii pneumonia and died in the ICU. One patient is in ongoing molecular CR more than four years after alloSCT. In the two patients with available follow-up, no long-term hepatic or renal toxicity was observed,

as evidenced by normal creatinine and bilirubin levels in routine follow-ups (Table S3).

Discussion

This retrospective analysis demonstrates that incorporating CXCR4-directed ERT with TBI and chemotherapy is feasible and results in promising response rates for patients with AML refractory to multiple lines of therapy.

TBI-based conditioning prior to alloSCT has proven highly effective in controlling AML, as evidenced by low relapse-associated mortality rates in various studies [34]. However, this does not translate into improved OS, particularly for older r/r AML patients, due to substantial treatment-related mortality [35].

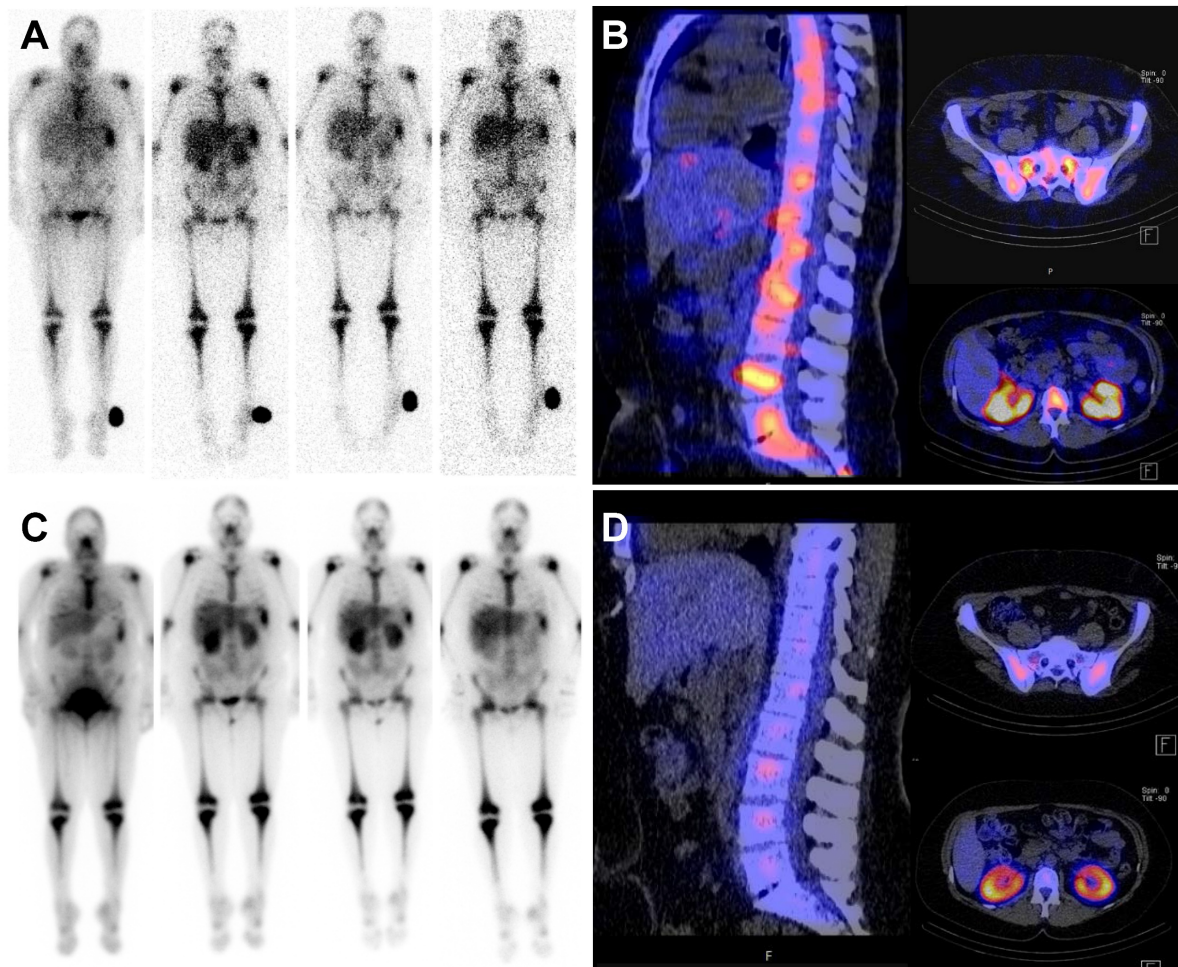


Figure 3. Pre- and post-therapeutic dosimetry. Pre- and post-therapeutic scintigraphy and SPECT/CT imaging of a 47 year old patient with refractory AML. Pre-therapeutic whole-body scintigraphy at 1 hour post injection (p.i.); 22h p.i.; 48h p.i. and 6 days p.i. (A, from left to right) for dosimetry including fused SPECT/CT (B). Post-therapeutic whole-body scintigraphy at 2 h, 24 h, 48 h and 6 d after administration of 14.4 Gbq [¹⁷⁷Lu]Pentixather (C) and fused SPECT/CT (D).

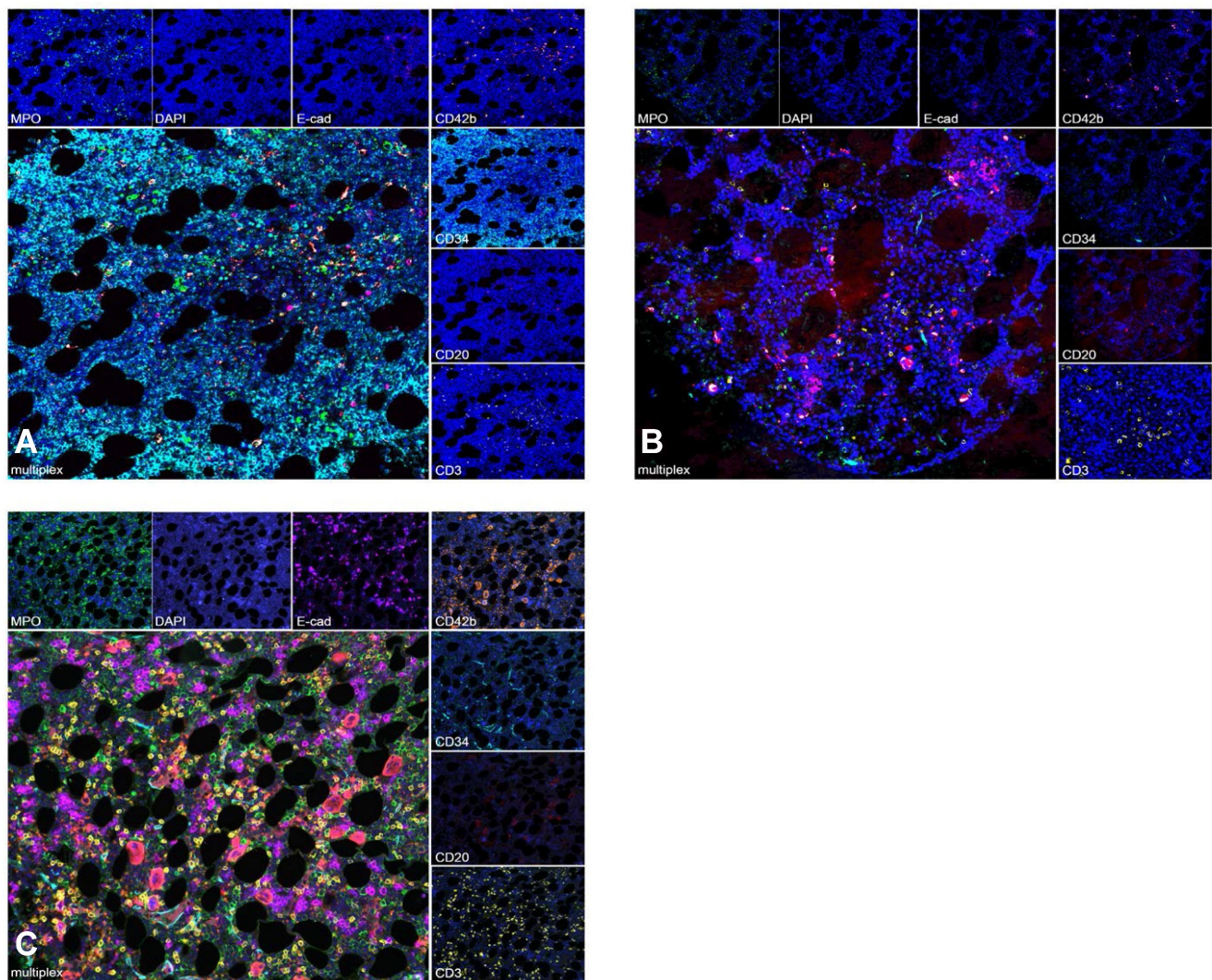


Figure 4. Multispectral Imaging. Multispectral image of an exemplary BM biopsy before treatment shows a compact infiltrate of CD34 positive AML blasts with reduced trilineage hematopoiesis (A). During aplasia, BM with clearance of AML blasts and markedly reduced trilineage hematopoiesis is detected (B). After engraftment, recovery of the BM microenvironment with normal distribution of hematopoiesis and the hematopoietic niche is shown (C). Granulopoiesis is detected by MPO, erythropoiesis by E-cadherin and megakaryopoiesis by CD42b. CD20 highlights B- lymphocytes and CD3 T- lymphocytes. Myeloblasts are marked by CD34.

In AML, the CXCR4/CXCL12 axis is thought to play an important role in the BM-mediated resistance by providing a local protective environment. CXCR4-targeted ERT causes radiation-absorbed dose in AML-infiltrated organs, particularly the BM [36]. Therefore, targeting CXCR4 in a sequential therapy approach and bypassing the hematologic toxicity by SCT is an intriguing concept for disease control in high-risk patients unable to achieve hematological remission before alloSCT. A significant concern with CXCR4-directed ERT is the potential destruction of the stem cell niche, caused by the so-called “cross-fire effect”, which could impair engraftment. Preclinical studies using a humanized murine leukemia model have demonstrated that CXCR4-directed ERT can effectively eliminate leukemic cells. Interestingly, the number of mesenchymal stromal cells (MSCs), the primary component of the stem cell niche, remained

unchanged, although their proliferative capacity was impaired. Nonetheless, their overall functionality after ERT was not significantly affected, as post-alloSCT engraftment occurred within the expected timeframe [27]. This may be partly explained by the low CXCR4 expression in MSCs and their resistance to radiation-induced cell death [37].

Retrospective analyses incorporating CXCR4-directed ERT in chemotherapy-based conditioning before auto- or alloSCT in hematologic malignancies such as T- and B-cell lymphomas, multiple myeloma and leukemia have confirmed adequate engraftment [33, 38-40]. Our data reveal that combining CXCR4-directed ERT with TBI-based conditioning is also viable, with engraftment occurring within the expected range for all but one patient, who died prior to engraftment. Importantly, this retrospective analysis demonstrates for the first

time in a patient setting that the mesenchymal compartment of the BM-niche retains its ability to support transplanted stem cells post-CXCR4-directed ERT.

Eradicating leukemic blasts is paramount in treating r/r AML patients to enable stem cell transplant engraftment and provide time for the immunogenic graft versus leukemia effect to develop. In this analysis, all but one patient achieved at least MLFS.

Other studies have demonstrated that lymphoid neoplasms exhibit higher CXCR4 expression compared to myeloid neoplasms such as AML [41]. Integrating CXCR4-directed ERT into auto- or alloSCT strategies for heavily pretreated lymphoproliferative disorders, including lymphomas and multiple myeloma, has achieved remissions in all evaluated patients [33, 38-40]. In myeloid neoplasms like AML, significant variability exists in the CXCR4 expression as determined by PET imaging, thus limiting the suitability of this approach to a specific subgroup [25]. Notably, the two patients in this study who achieved long-term survival exhibited the highest CXCR4 expression within their AML manifestations. Consequently, *in vivo* CXCR4 expression, as assessed by PET imaging, may serve as a predictive marker for the therapeutic efficacy of CXCR4-directed ERT. However larger cohorts are needed to further support this observation.

In the context of AML comparable disease control and long-term survival rates have been reported in studies using chemotherapy-based conditioning regimens, such as clofarabine [42]. Historical data show that patients with r/r AML have dismal prognosis with median OS of less than 12 months and for patients relapsing after alloSCT outcome is even worse [43, 44]. However, most patients with AML who fail to respond to several lines of therapy either die before or are considered non-eligible for alloSCT, in consequence data on multi-refractory AML patients are scarce.

The observed toxicity profile highlights the aggressive nature of conditioning regimens for r/r AML, which, while potentially curative, pose considerable morbidity and mortality risks. Complications such as uncontrolled infection and organ damage rapidly accumulate in patients with uncontrolled disease who undergo multiple lines of treatment, often leading to a rapid decline in the patient's condition resulting in the inability to receive further intensive treatments. Given this context, some of the observed toxicities may be attributed to this difficult-to-treat cohort, in which all patients were heavily pre-treated and all but one patient had uncontrolled disease at the time of conditioning.

Thus, the promising response rates observed in this cohort raise the question, whether applying this treatment approach in earlier lines, might yield better outcomes and lower toxicity rates when patients have undergone fewer previous lines of treatment. Consequently, a CXCR4-directed ERT approach in earlier therapy lines may not only enhance objective outcomes but also improve the quality of life for high-risk AML patients, as it counteracts the decline associated with multiple treatment lines and their long-term toxicities [45]. However, long-term experience with other ERT approaches, such as PSMA- and SSTR-directed ERT, suggests that these therapies rarely can lead to hematological toxicities, including myelodysplastic syndromes, or organ dysfunctions such as renal insufficiency [46, 47]. Although no such side effects have been observed in the studied cohort, the small number of patients and brief follow-up period preclude any definitive conclusions regarding long-term toxicity and efficacy following CXCR4-directed ERT.

Discussing other ERT strategies requires consideration of the pharmacokinetics and dynamics of the radiopharmaceuticals involved. ¹⁸⁸Rhenium-labeled anti-CD66 and ¹³³Iodine-labeled anti-CD45 have been successfully used in alloSCT conditioning for myeloid malignancies [48-50]. The SIERRA phase III trial demonstrated superior response rates and survival with ¹³³I-anti-CD45 compared to standard of care [51]. A significant challenge is the timing between ERT and stem cell transplantation to ensure radiation decay and prevent harm to healthy stem cells. Conversely, shorter-lived ERT agents might be excreted faster and thus reduce toxicity but could compromise efficacy against deeply situated or protected leukemic cells. The use of Pentixather labelled with ⁹⁰Y with its shorter half-life compared to ¹⁷⁷Lu has been feasible and has significantly reduced the interval between ERT and SCT [33]. However, ⁹⁰Y has higher beta minus energies and a longer range that lead to increased irradiation of surrounding tissue. For example, ¹⁷⁷Lu presents with a short beta minus maximum range of 2 mm, while ⁹⁰Y presents with an 11 mm maximum range. Therefore, higher kidney damage could occur and has been observed in the treatment of neuroendocrine tumors with ⁹⁰Y-DOTATATE compared to ¹⁷⁷Lu-DOTATATE [52]. The pharmacokinetics and potential benefits of other ERT strategies must therefore be carefully evaluated for the clinical setting of conditioning regimens to prevent additional toxicities.

Conclusion

Data from this retrospective analysis indicate that CXCR4-directed ERT in sequential combination

with TBI and chemotherapy offers an individually tailored treatment approach for patients with multi-refractory AML. No acute high-grade toxicities clearly attributed to ERT in addition to the conventional conditioning could be observed and stem cell engraftment was not impaired. Clearance of AML blasts was reached in the majority of patients (6 out of 7 patients). Future studies should refine CXCR4-directed ERT by dose optimization, patient selection and incorporation in earlier therapy lines for high-risk AML patients.

Supplementary Material

Supplementary methods, figure and tables.
<https://www.thno.org/v15p0019s1.pdf>

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Institutional review board statement

The study was conducted in accordance with the Declaration of Helsinki. It was conducted according to the principles of the Declaration of Helsinki. It was approved by the Medical Ethics Committee, vote 423/17 S, of the Klinikum rechts der Isar of the Technical University of Munich.

Informed consent statement

Informed consent was obtained from all subjects involved in the study.

Author contributions

Conceptualization K.B., M.E., M.V. and P.H.; data curation, visualization, writing original draft preparation, K.B., T.L., M.R., M.E. and P.H.; methodology K.B., T.L., J.C.P., A.A., J.B.L., C.D.A., M.R., M.E. and P.H.; formal analysis, writing-review and editing, K.B., T.L., M.H., K.K., K.N., K.S.G., W.W., J.B.L., C.D.A., F.B., M.R., M.E., M.V. and P.H. All authors have read and agreed to the published version of the manuscript.

Competing Interests

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References

- Stone RM, Mandrekar SJ, Sanford BL, Laumann K, Geyer S, Bloomfield CD, et al. Midostaurin plus Chemotherapy for Acute Myeloid Leukemia with a FLT3 Mutation. *N Engl J Med*. 2017; 377: 454-64.
- DiNardo CD, Jonas BA, Pullarkat V, Thirman MJ, Garcia JS, Wei AH, et al. Azacitidine and Venetoclax in Previously Untreated Acute Myeloid Leukemia. *N Engl J Med*. 2020; 383: 617-29.
- Montesinos P, Recher C, Vives S, Zarzycka E, Wang J, Bertani G, et al. Ivosidenib and Azacitidine in IDH1-Mutated Acute Myeloid Leukemia. *N Engl J Med*. 2022; 386: 1519-31.
- Issa GC, Aldoss I, DiPersio J, Cuglievan B, Stone R, Arellano M, et al. The menin inhibitor revumenib in KMT2A-rearranged or NPM1-mutant leukaemia. *Nature*. 2023; 615: 920-4.
- Döhner H, Wei AH, Appelbaum FR, Craddock C, DiNardo CD, Dombret H, et al. Diagnosis and management of AML in adults: 2022 recommendations from an international expert panel on behalf of the ELN. *Blood*. 2022; 140: 1345-77.
- Duval M, Klein JP, He W, Cahn JY, Cairo M, Camitta BM, et al. Hematopoietic stem-cell transplantation for acute leukemia in relapse or primary induction failure. *J Clin Oncol*. 2010; 28: 3730-8.
- Yerushalmi Y, Shem-Tov N, Danylesko I, Canaan J, Avigdor A, Yerushalmi R, et al. Second hematopoietic stem cell transplantation as salvage therapy for relapsed acute myeloid leukemia/myelodysplastic syndromes after a first transplantation. *Haematologica*. 2023; 108: 1782-92.
- Yanada M, Konuma T, Yamasaki S, Kondo T, Fukuda T, Shingai N, et al. Relapse of acute myeloid leukemia after allogeneic hematopoietic cell transplantation: clinical features and outcomes. *Bone Marrow Transplant*. 2021; 56: 1126-33.
- Abou Dalle I, Labopin M, Kröger N, Schroeder T, Finke J, Stelljes M, et al. Impact of disease burden on clinical outcomes of AML patients receiving allogeneic hematopoietic cell transplantation: a study from the Acute Leukemia Working Party of the European Society for Blood and Marrow Transplantation. *Bone Marrow Transplant*. 2023; 58: 784-90.
- Jethava YS, Sica S, Savani B, Socola F, Jagasia M, Mohty M, et al. Conditioning regimens for allogeneic hematopoietic stem cell transplants in acute myeloid leukemia. *Bone Marrow Transplant*. 2017; 52: 1504-11.
- Karpova D, Bonig H. Concise Review: CXCR4/CXCL12 Signaling in Immature Hematopoiesis—Lessons From Pharmacological and Genetic Models. *Stem Cells*. 2015; 33: 2391-9.
- Zlotnik A, Yoshie O. Chemokines: a new classification system and their role in immunity. *Immunity*. 2000; 12: 121-7.
- Zou YR, Kottmann AH, Kuroda M, Taniuchi I, Littman DR. Function of the chemokine receptor CXCR4 in haematopoiesis and in cerebellar development. *Nature*. 1998; 393: 595-9.
- Nagasawa T, Hirota S, Tachibana K, Takakura N, Nishikawa S, Kitamura Y, et al. Defects of B-cell lymphopoiesis and bone-marrow myelopoiesis in mice lacking the CXC chemokine PBSF/SDF-1. *Nature*. 1996; 382: 635-8.
- Sugiyama T, Kohara H, Noda M, Nagasawa T. Maintenance of the hematopoietic stem cell pool by CXCL12-CXCR4 chemokine signaling in bone marrow stromal cell niches. *Immunity*. 2006; 25: 977-88.
- Orimo A, Gupta PB, Sgroi DC, Arenzana-Seisdedos F, Delaunay T, Naeem R, et al. Stromal fibroblasts present in invasive human breast carcinomas promote tumor growth and angiogenesis through elevated SDF-1/CXCL12 secretion. *Cell*. 2005; 121: 335-48.
- Burger JA, Kipps TJ. CXCR4: a key receptor in the crosstalk between tumor cells and their microenvironment. *Blood*. 2006; 107: 1761-7.
- Spoo AC, Lübbert M, Wierda WG, Burger JA. CXCR4 is a prognostic marker in acute myelogenous leukemia. *Blood*. 2007; 109: 786-91.
- Domanska UM, Kruizinga RC, Nagengast WB, Timmer-Bosscha H, Huls G, de Vries EG, et al. A review on CXCR4/CXCL12 axis in oncology: no place to hide. *Eur J Cancer*. 2013; 49: 219-30.
- Guo F, Wang Y, Liu J, Mok SC, Xue F, Zhang W. CXCL12/CXCR4: a symbiotic bridge linking cancer cells and their stromal neighbors in oncogenic communication networks. *Oncogene*. 2016; 35: 816-26.
- Scala S. Molecular Pathways: Targeting the CXCR4-CXCL12 Axis—Untapped Potential in the Tumor Microenvironment. *Clin Cancer Res*. 2015; 21: 4278-85.
- Meads MB, Gatenby RA, Dalton WS. Environment-mediated drug resistance: a major contributor to minimal residual disease. *Nat Rev Cancer*. 2009; 9: 665-74.

23. Demmer O, Gourni E, Schumacher U, Kessler H, Wester HJ. PET imaging of CXCR4 receptors in cancer by a new optimized ligand. *ChemMedChem*. 2011; 6: 1789-91.
24. Schottelius M, Osl T, Poschenrieder A, Hoffmann F, Beykan S, Hänscheid H, et al. [(177)Lu]pentixather: Comprehensive Preclinical Characterization of a First CXCR4-directed Endoradiotherapeutic Agent. *Theranostics*. 2017; 7: 2350-62.
25. Herhaus P, Habringer S, Philipp-Abbrederis K, Vag T, Gerngross C, Schottelius M, et al. Targeted positron emission tomography imaging of CXCR4 expression in patients with acute myeloid leukemia. *Haematologica*. 2016; 101: 932-40.
26. Cho BS, Zeng Z, Mu H, Wang Z, Konoplev S, McQueen T, et al. Antileukemia activity of the novel peptidic CXCR4 antagonist LY2510924 as monotherapy and in combination with chemotherapy. *Blood*. 2015; 126: 222-32.
27. Habringer S, Lapa C, Herhaus P, Schottelius M, Istvanffy R, Steiger K, et al. Dual Targeting of Acute Leukemia and Supporting Niche by CXCR4-Directed Theranostics. *Theranostics*. 2018; 8: 369-83.
28. Peci F, Dekker L, Pagliaro A, van Boxtel R, Nierkens S, Belderbos M. The cellular composition and function of the bone marrow niche after allogeneic hematopoietic cell transplantation. *Bone Marrow Transplant*. 2022; 57: 1357-64.
29. Shen S, Meredith RF, Duan J, Macey DJ, Khazaeli MB, Robert F, et al. Improved prediction of myelotoxicity using a patient-specific imaging dose estimate for non-marrow-targeting (90)Y-antibody therapy. *J Nucl Med*. 2002; 43: 1245-53.
30. Hindorf C, Glatting G, Chiesa C, Lindén O, Flux G. EANM Dosimetry Committee guidelines for bone marrow and whole-body dosimetry. *Eur J Nucl Med Mol Imaging*. 2010; 37: 1238-50.
31. Andersson M, Johansson L, Eckerman K, Mattsson S. IDAC-Dose 2.1, an internal dosimetry program for diagnostic nuclear medicine based on the ICRP adult reference voxel phantoms. *EJNMMI Res*. 2017; 7: 88.
32. Bodei L, Mueller-Brand J, Baum RP, Pavel ME, Hörsch D, O'Dorisio MS, et al. The joint IAEA, EANM, and SNMMI practical guidance on peptide receptor radionuclide therapy (PRRT) in neuroendocrine tumours. *Eur J Nucl Med Mol Imaging*. 2013; 40: 800-16.
33. Maurer S, Herhaus P, Lippenmeyer R, Hänscheid H, Kircher M, Schirbel A, et al. Side Effects of CXC-Chemokine Receptor 4-Directed Endoradiotherapy with Pentixather Before Hematopoietic Stem Cell Transplantation. *J Nucl Med*. 2019; 60: 1399-405.
34. Gyurkocza B, Sandmaier BM. Conditioning regimens for hematopoietic cell transplantation: one size does not fit all. *Blood*. 2014; 124: 344-53.
35. Salhotra A, Stein AS. Role of Radiation Based Conditioning Regimens in Patients With High-Risk AML Undergoing Allogeneic Transplantation in Remission or Active Disease and Mechanisms of Post-Transplant Relapse. *Front Oncol*. 2022; 12: 802648.
36. Hänscheid H, Schirbel A, Hartrampf P, Kraus S, Werner RA, Einsele H, et al. Biokinetics and Dosimetry of (177)Lu-Pentixather. *J Nucl Med*. 2022; 63: 754-60.
37. Sugrue T, Lowndes NF, Ceredig R. Mesenchymal stromal cells: radio-resistant members of the bone marrow. *Immunol Cell Biol*. 2013; 91: 5-11.
38. Buck AK, Grigoleit GU, Kraus S, Schirbel A, Heinsch M, Dreher N, et al. C-X-C Motif Chemokine Receptor 4-Targeted Radioligand Therapy in Patients with Advanced T-Cell Lymphoma. *J Nucl Med*. 2023; 64: 34-9.
39. Lapa C, Hänscheid H, Kircher M, Schirbel A, Wunderlich G, Werner RA, et al. Feasibility of CXCR4-Directed Radioligand Therapy in Advanced Diffuse Large B-Cell Lymphoma. *J Nucl Med*. 2019; 60: 60-4.
40. Lapa C, Herrmann K, Schirbel A, Hänscheid H, Luckerath K, Schottelius M, et al. CXCR4-directed endoradiotherapy induces high response rates in extramedullary relapsed Multiple Myeloma. *Theranostics*. 2017; 7: 1589-97.
41. Buck AK, Serfling SE, Lindner T, Hänscheid H, Schirbel A, Hahner S, et al. CXCR4-targeted theranostics in oncology. *Eur J Nucl Med Mol Imaging*. 2022; 49: 4133-44.
42. Le Bourgeois A, Labopin M, Marçais A, de Latour RP, Blaise D, Chantepie S, et al. Sequential allogeneic hematopoietic stem cell transplantation for active refractory/relapsed myeloid malignancies: results of a reduced-intensity conditioning preceded by clofarabine and cytosine arabinoside, a retrospective study on behalf of the SFGM-TC. *Ann Hematol*. 2020; 99: 1855-62.
43. Schmäler AK, Ngoya M, Galimard JE, Bazarbachi A, Finke J, Kröger N, et al. Continuously improving outcome over time after second allogeneic stem cell transplantation in relapsed acute myeloid leukemia: an EBMT registry analysis of 1540 patients. *Blood Cancer J*. 2024; 14: 76.
44. Yanada M, Yamasaki S, Kondo T, Kawata T, Harada K, Uchida N, et al. Allogeneic hematopoietic cell transplantation for patients with acute myeloid leukemia not in remission. *Leukemia*. 2024; 38: 513-20.
45. Korol EE, Wang S, Johnston K, Ravandi-Kashani F, Levis M, van Nooten F. Health-Related Quality of Life of Patients with Acute Myeloid Leukemia: A Systematic Literature Review. *Oncol Ther*. 2017; 5: 1-16.
46. Steinhilber L, Lunger L, Cala L, Pfob CH, Lapa C, Hartrampf PE, et al. Long-Term Nephrotoxicity of (177)Lu-PSMA Radioligand Therapy. *J Nucl Med*. 2024; 65: 79-84.
47. Kennedy KR, Turner JH, MacDonald WBG, Claringbold PG, Boardman G, Ransom DT. Long-term survival and toxicity in patients with neuroendocrine tumors treated with (177) Lu-octreotate peptide radionuclide therapy. *Cancer*. 2022; 128: 2182-92.
48. Bunjes D, Buchmann I, Duncker C, Seitz U, Kotzerke J, Wiesneth M, et al. Rhenium 188-labeled anti-CD66 (a, b, c, e) monoclonal antibody to intensify the conditioning regimen prior to stem cell transplantation for patients with high-risk acute myeloid leukemia or myelodysplastic syndrome: results of a phase I-II study. *Blood*. 2001; 98: 565-72.
49. Pagel JM, Appelbaum FR, Eary JF, Rajendran J, Fisher DR, Gooley T, et al. 131I-anti-CD45 antibody plus busulfan and cyclophosphamide before allogeneic hematopoietic cell transplantation for treatment of acute myeloid leukemia in first remission. *Blood*. 2006; 107: 2184-91.
50. Koenecke C, Hofmann M, Bolte O, Gielow P, Dammann E, Stadler M, et al. Radioimmunotherapy with [188Re]-labelled anti-CD66 antibody in the conditioning for allogeneic stem cell transplantation for high-risk acute myeloid leukemia. *Int J Hematol*. 2008; 87: 414-21.
51. Gyurkocza B, Seropian S, Choe H, Litzow M, Abboud C, Koshy N, et al. S248: SIERRA TRIAL RESULTS WITH A TARGETED RADIOTHERAPY, IOMAB-B, A MYELOABLATIVE CONDITIONING WITH REDUCED INTENSITY TOLERABILITY YIELDS HIGH CR, LONG TERM SURVIVAL IN HSCT INELIGIBLE ACTIVE R/R AML. *HemaSphere*. 2023; 7: e2941723.
52. Bodei L, Kidd M, Paganelli G, Grana CM, Drozdov I, Cremonesi M, et al. Long-term tolerability of PRRT in 807 patients with neuroendocrine tumours: the value and limitations of clinical factors. *Eur J Nucl Med Mol Imaging*. 2015; 42: 5-19.