

A Bifunctional Fusion Membrane- Based Biocompatible Nanovaccine to  
Potentiate Cancer Immunotherapy

*Wei Fu<sup>a,f,1</sup>, Xing Cai<sup>b,1</sup>, Jinru Yang<sup>b,1</sup>, Lian Yang<sup>c,d,\*</sup>, Yaoyu Pan<sup>e,\*</sup>, Zhan Tuo<sup>f,\*</sup>*

<sup>a</sup>Chongqing Key Laboratory of Molecular Oncology and Epigenetics, The First Affiliated Hospital of Chongqing Medical University, Chongqing 400016, China.

<sup>b</sup>Department of Radiation and Medical Oncology, Zhongnan Hospital of Wuhan University, Wuhan 430071, P.R. China.

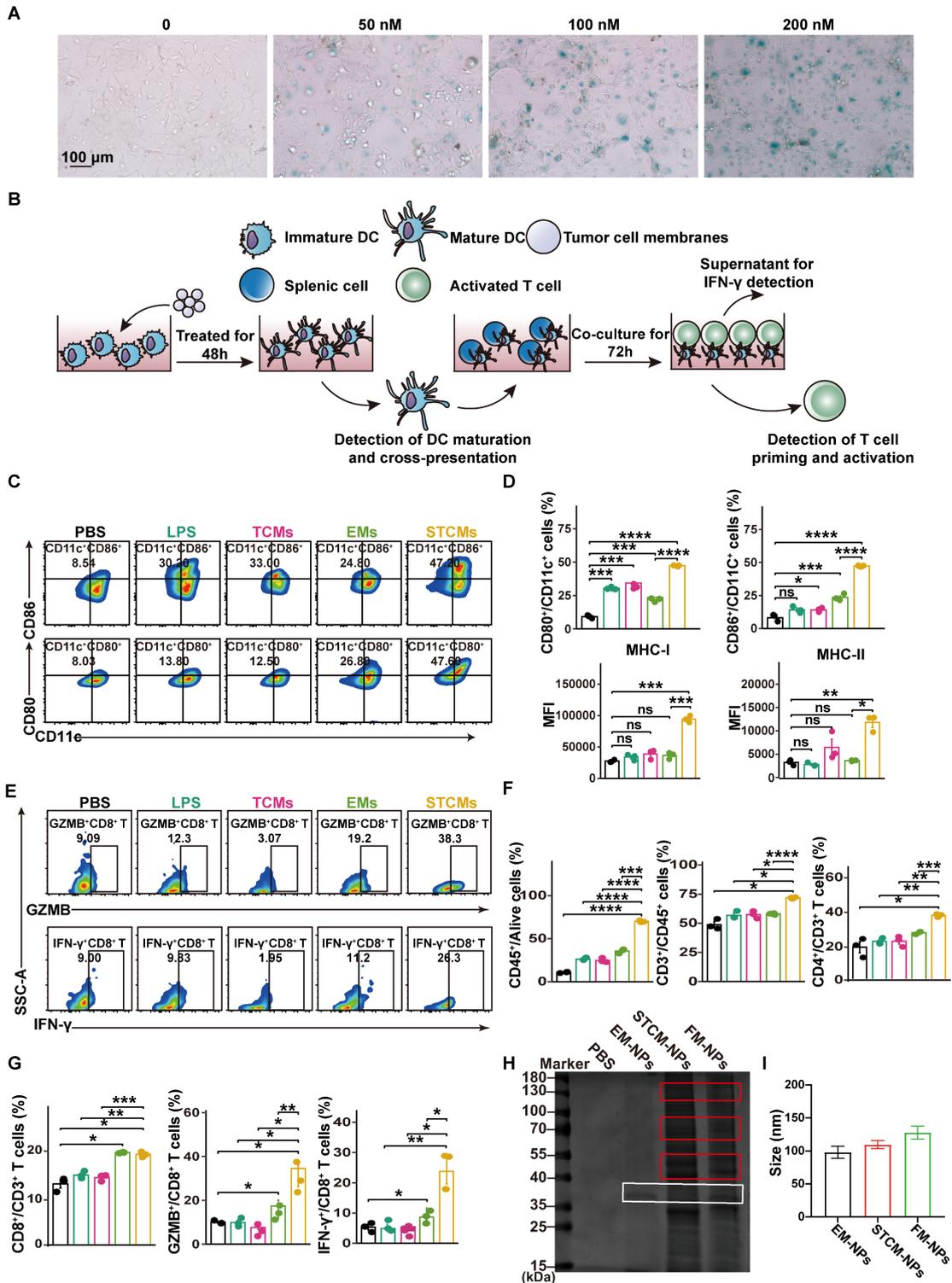
<sup>c</sup>Department of Radiology, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, 430022, China.

<sup>d</sup>Hubei Key Laboratory of Molecular Imaging, Wuhan, 430022, China.

<sup>e</sup>Department of Polymer, School of Material Science and Engineering, Hubei University, Wuhan, Hubei 430062, China.

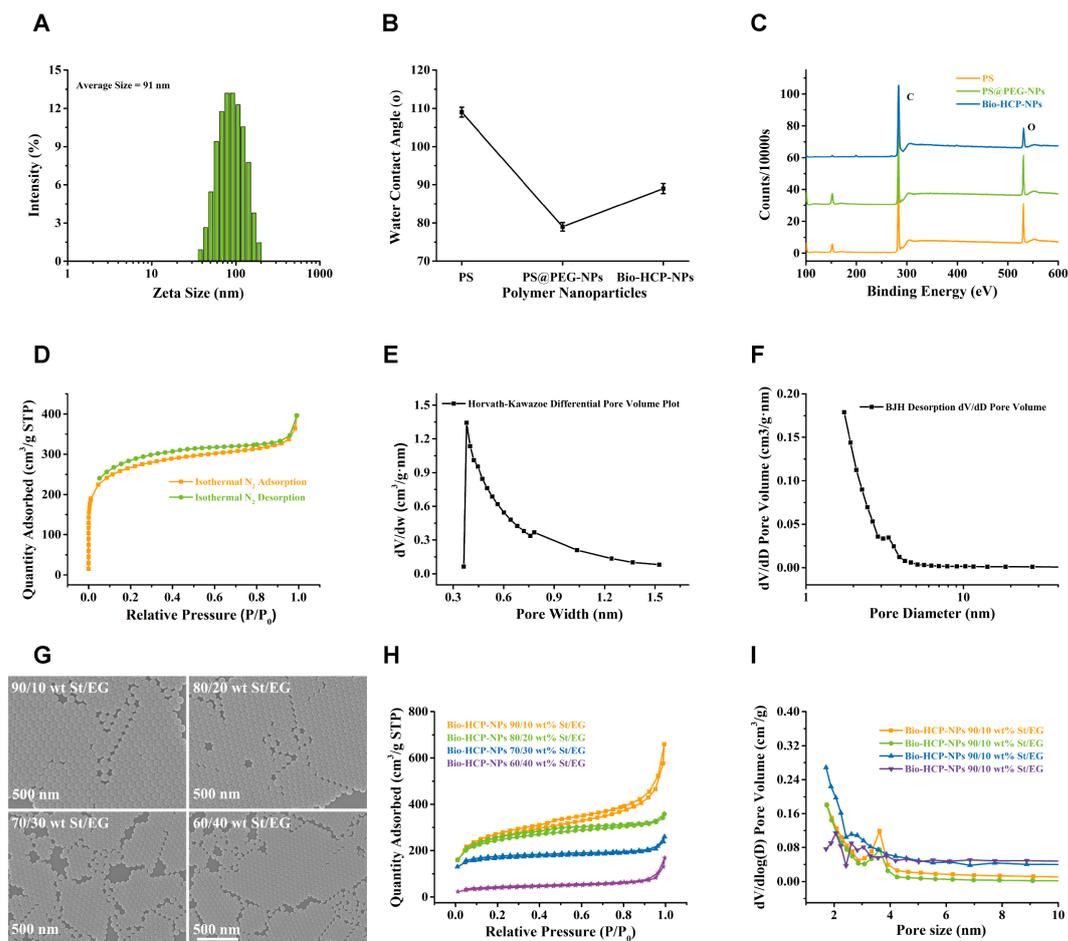
<sup>f</sup>The Affiliated Cancer Hospital of Zhengzhou University & Henan Cancer Hospital, Zhengzhou 450008, China.

\*E-mail address: tztuozhan@163.com (Z. Tuo).



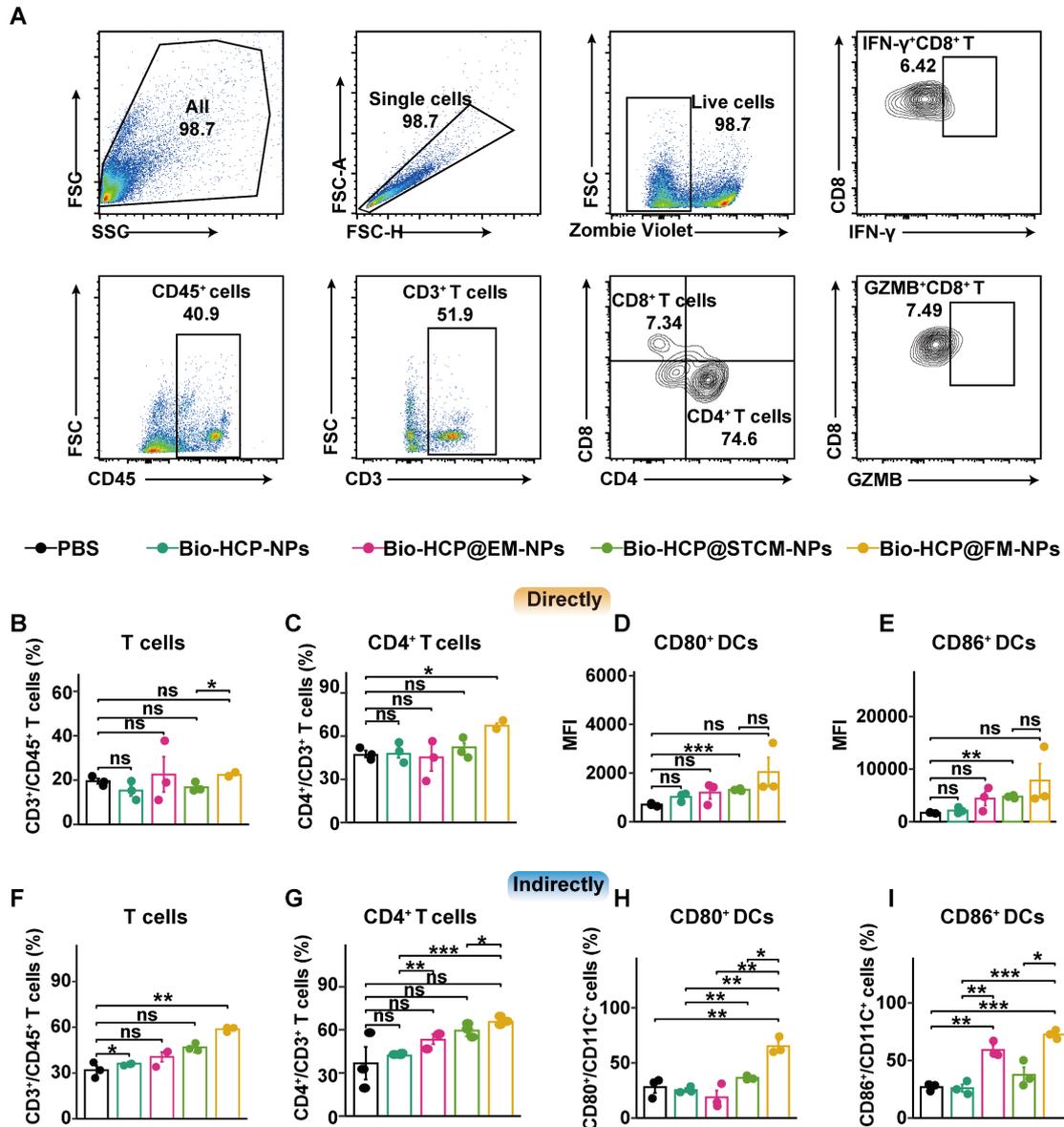
**Supplementary Figure 1. STCM-NPs promote antigen presenting by DCs and activation of T cells, related to Figure 1.** (A) SA $\beta$ G staining of B16-F10 tumor cells following senescence induction via treatment with different concentrations of cisplatin for 6 days. Scale bar, 100  $\mu$ m. (B) Sketch depicting the co-culture assays. (C)

Representative flow cytometry images of mDCs (CD11c<sup>+</sup> CD80<sup>+</sup> and CD11c<sup>+</sup> CD86<sup>+</sup>) co-incubated with different groups (PBS, LPS, TCMs, EMs, and STCMs) for 48 h *in vitro* (n = 3). (D) The proportion of mDCs and the levels of MHC-I and MHC-II on mDCs after being co-cultured with various formulations for 48 h *in vitro* (n = 3). (E) Representative flow cytometry images of IFN- $\gamma$ <sup>+</sup> CD8<sup>+</sup> T cells (lower) and GZMB<sup>+</sup> CD8<sup>+</sup> T cells (upper) after being co-cultured with BMDCs pre-treated with various formulations for 72 h *in vitro*. (F) Percentages of CD45<sup>+</sup> T cells, CD3<sup>+</sup> T cells and CD4<sup>+</sup> T cells (n = 3) after being co-cultured with BMDCs pre-treated various formulations for 72 h *in vitro*. (G) Percentages of CD8<sup>+</sup> T cells, GZMB<sup>+</sup> T cells and IFN- $\gamma$ <sup>+</sup> CD8<sup>+</sup> T cells (gated from CD3<sup>+</sup> CD8<sup>+</sup> cells; n = 3). (H) The SDS-PAGE image of STCM-NPs (red rectangle), and EM-NPs (black rectangle). The amounts of protein in gels were 10 ug. (I) Diameter (n = 3) of EM-NPs, STCM-NPs and FM-NPs. Data are presented as mean  $\pm$  SD. One-way ANOVA with subsequent multiple comparison tests were conducted, where ns indicates no significance, \* $p$  < 0.05, \*\* $p$  < 0.01, \*\*\* $p$  < 0.001 and \*\*\*\* $p$  < 0.0001.



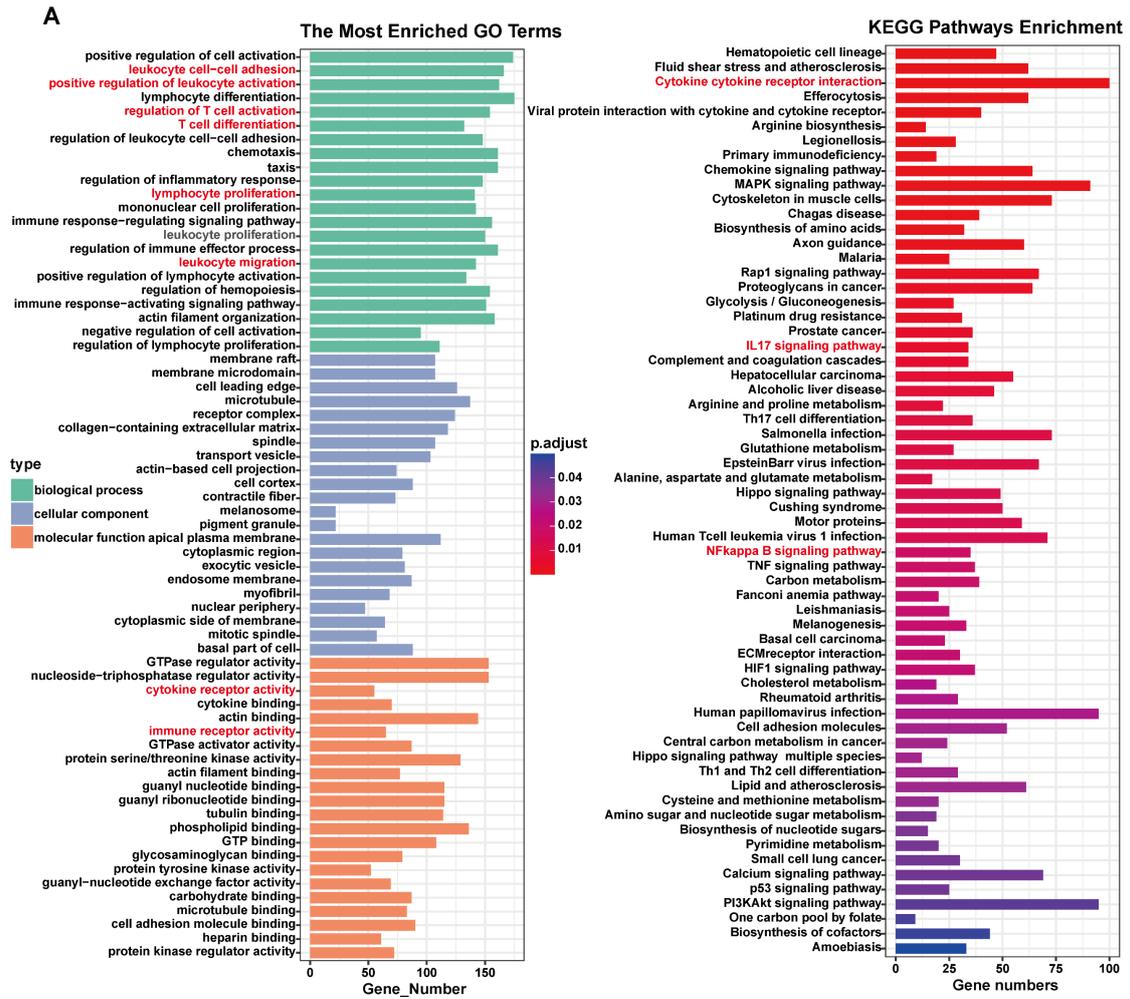
**Supplementary Figure 2. Additional analysis and characterization of Bio-HCP-**

**NPs.** (A) Zeta size curves of Bio-HCP-NPs. (B) WCA analysis of PS, PS@PEG-NPs and Bio-HCP-NPs (C) XPS surface elements analysis of PS, PS@PEG-NPs and Bio-HCP-NPs. (D) Isothermal N<sub>2</sub> absorption and desorption curves of Bio-HCP-NPs in total-pore BET testing. (E) Horvath-Kawazoe differential pore volume plot of Bio-HCP-NPs in total-pore BET testing. (F) BJH pore size curves of Bio-HCP-NPs in total-pore BET testing. (G) FESEM images of series Bio-HCP-NPs formed with different proportion of monomers. (H) Additional morphological and structural characterization of Bio-HCP-NPs in 90/10 wt% St/EG ratio. (I) Pore sizes distribution curves of series Bio-HCP-NPs formed with different proportion of monomers.



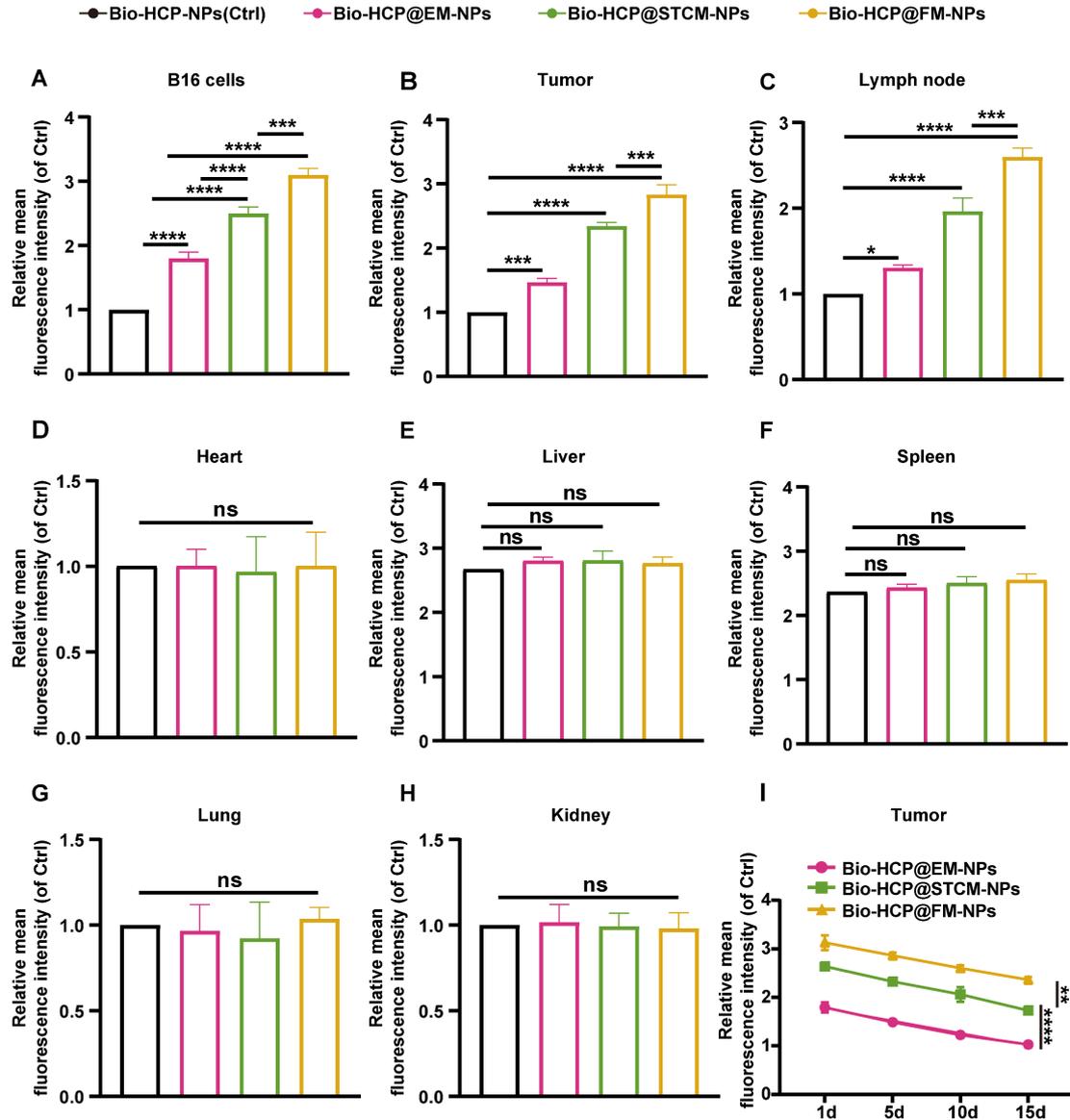
**Supplementary Figure 3. Bio-HCP@FM-NPs activate DCs, related to Figure 3. (A)**

Flow cytometry gating strategy for measuring the percentages of T cells, IFN- $\gamma$ <sup>+</sup> T cells and GZMB<sup>+</sup> T cells (n = 3). (B-C) Percentages of T cells, CD4<sup>+</sup> T cells in direct co-culture assay (n = 3). (D-E) Expression of CD86 and CD80 on DCs in direct co-culture assay (n = 3). (F-I) Percentages of T cells, CD4<sup>+</sup> T cells, CD80<sup>+</sup> DCs and CD86<sup>+</sup> DCs in indirect co-culture assay (n = 3).



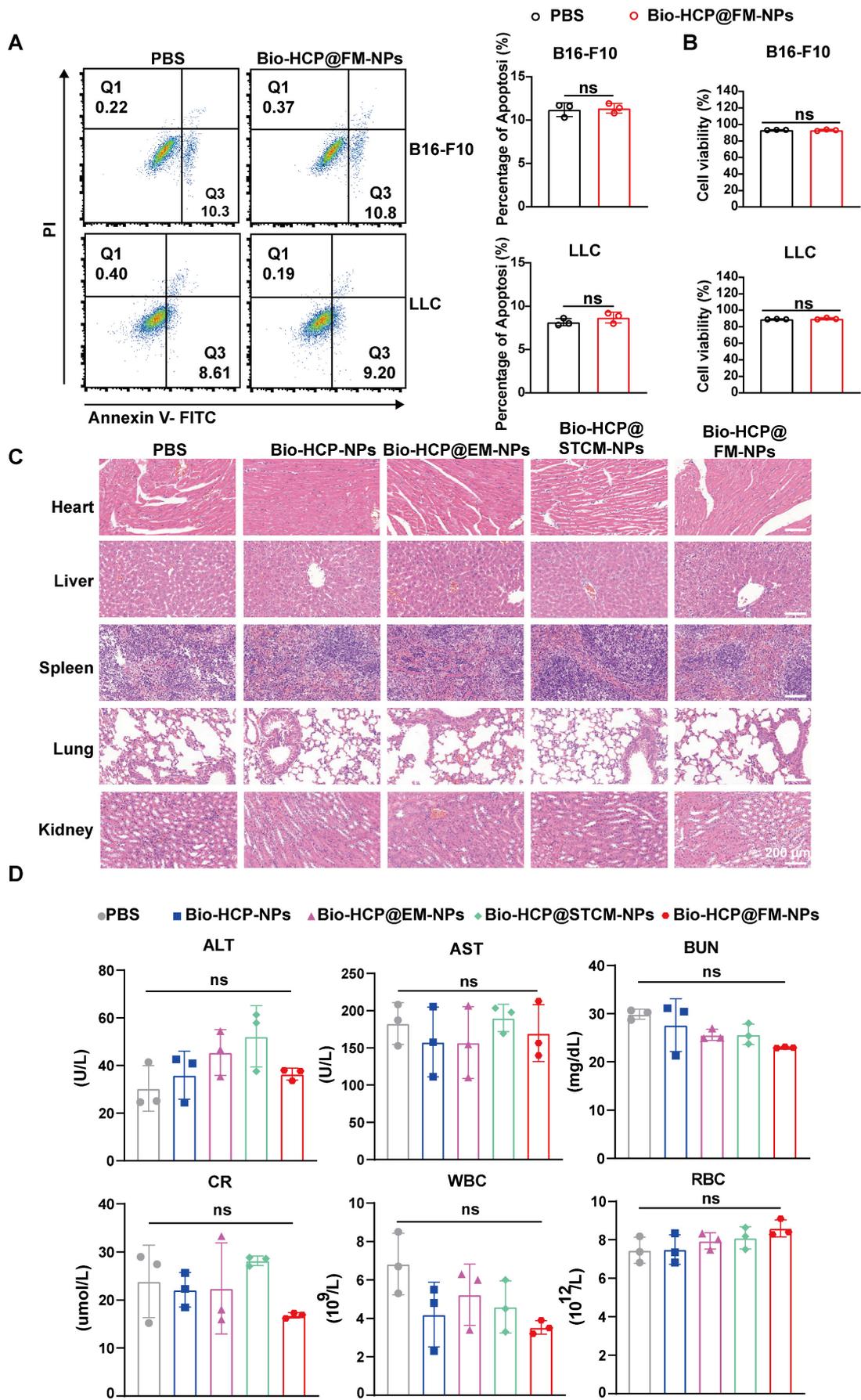
**Supplementary Figure 4. GO and KEGG analysis of BMDCs, related to Figure 3.**

(A) GO and KEGG analysis of BMDCs.



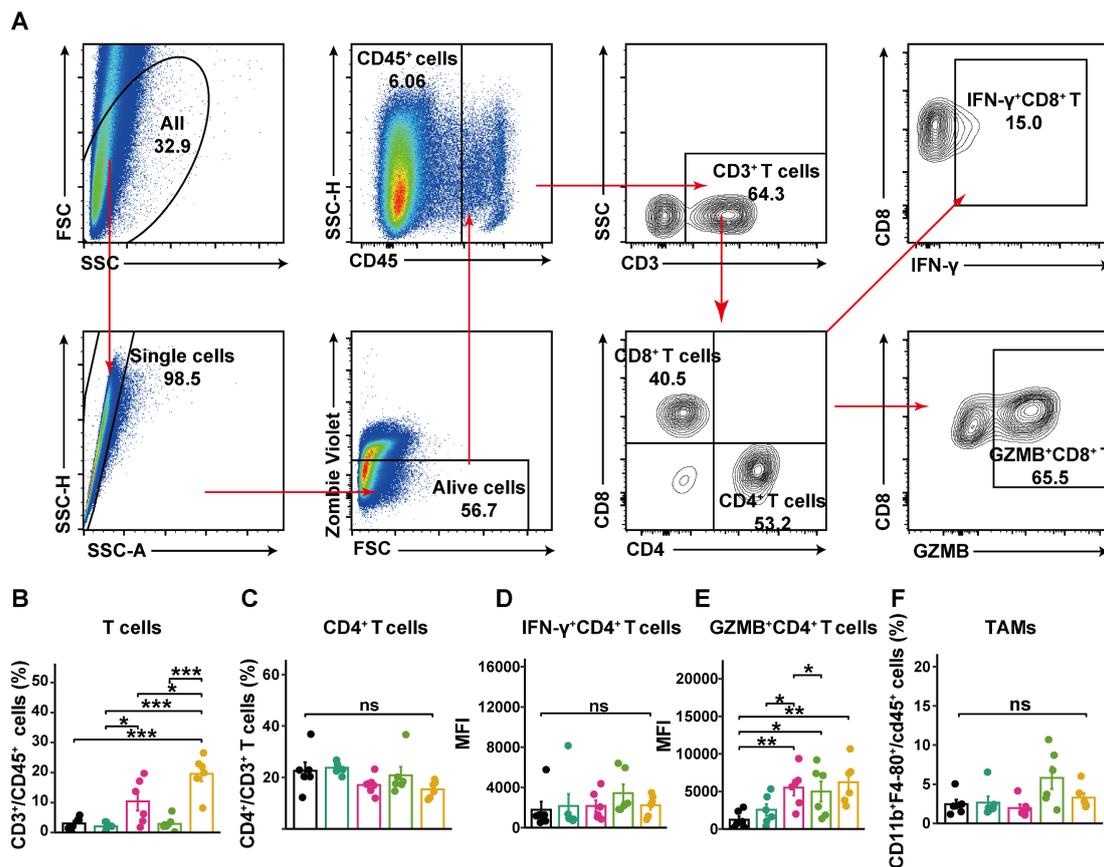
**Supplementary Figure 5. Assessment of biodistribution and targeting capability of Bio-HCP@FM-NPs vaccination *in vitro* and *in vivo*.** (A) Quantitative analysis of fluorescence signals from FITC-labeled nanoparticles in DCs at 24 h post-injection. (B-H) The quantitative analysis of nanoparticle accumulation in major organs, tumor and lymph nodes from Bio-HCP@EM-NPs, Bio-HCP@STCM-NPs and Bio-HCP@FM-NPs injected B16-F10 cells orthotopic tumor-bearing mice *in vivo*. (I) Retention ability of Bio-HCP@EM-NPs, Bio-HCP@STCM-NPs and Bio-HCP@FM-NPs in tumor tissues of B16-F10 cells orthotopic tumor-bearing murine model quantified by flow

cytometry at different time points. Data are presented as mean  $\pm$  SD, with  $n = 3$  per group. Statistical significance is indicated as follows: n.s. (no significance),  $*p < 0.05$ ,  $**p < 0.01$ ,  $***p < 0.001$ ,  $****p < 0.0001$ .



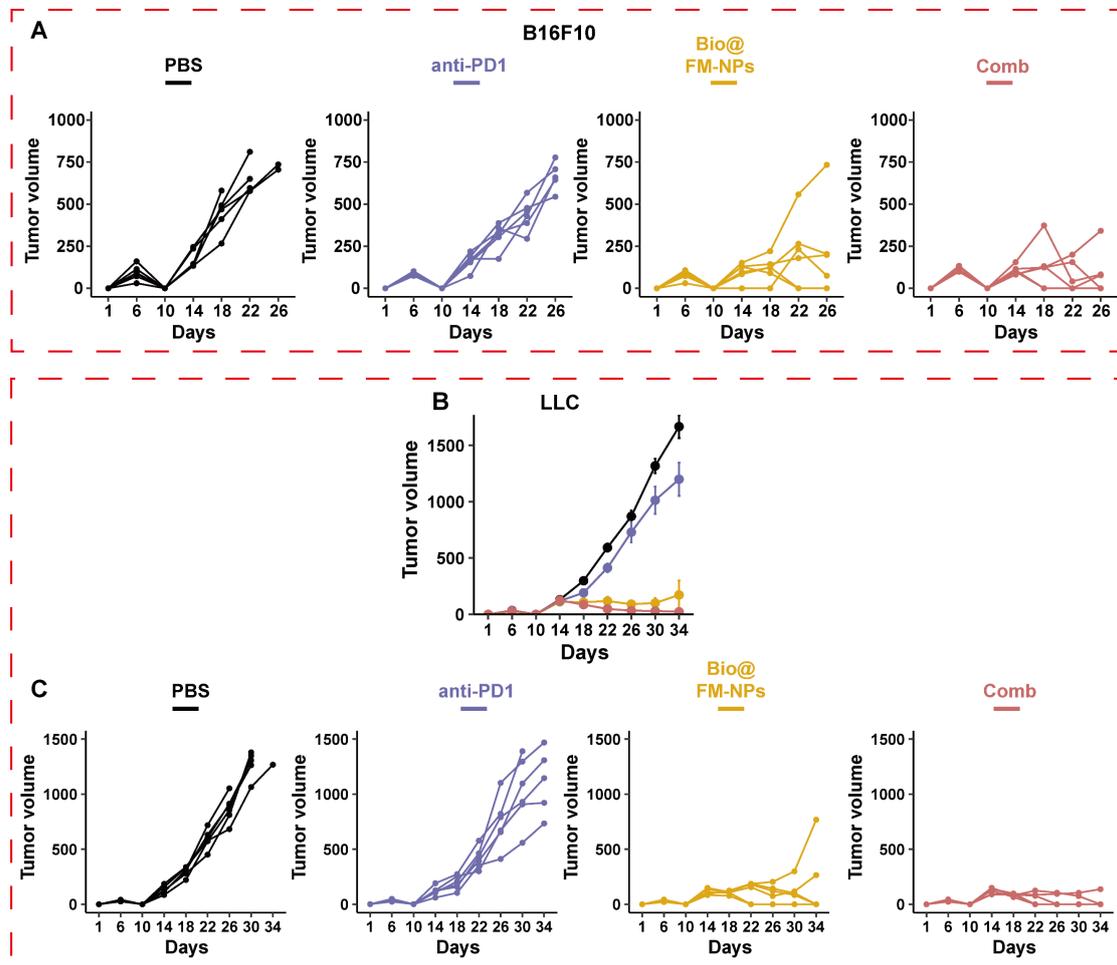
Supplementary Figure 6. Bio-HCP@FM-NPs show good biosafety. (A-B) The

cytotoxicity of Bio-HCP@FM-NPs against B16-F10 and LLC cells after co-incubation with nanovaccines. (C) H&E staining of heart, liver, spleen, lung, and kidney in B16-F10 mouse melanoma tumor model on day 7 after treatment. The scale bar is 200  $\mu\text{m}$ . (D) Hemanalysis was performed on blood drawn from mice on day 7 after treatment. Aspartate transaminase (AST), white blood cell (WBC), alanine transaminase (ALT), red blood cell (RBC), blood urea nitrogen (BUN), and creatinine (CR) are presented as the means  $\pm$  SEM. All data are mean  $\pm$  SD;  $n = 3$ . \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ , and ns, not significant.



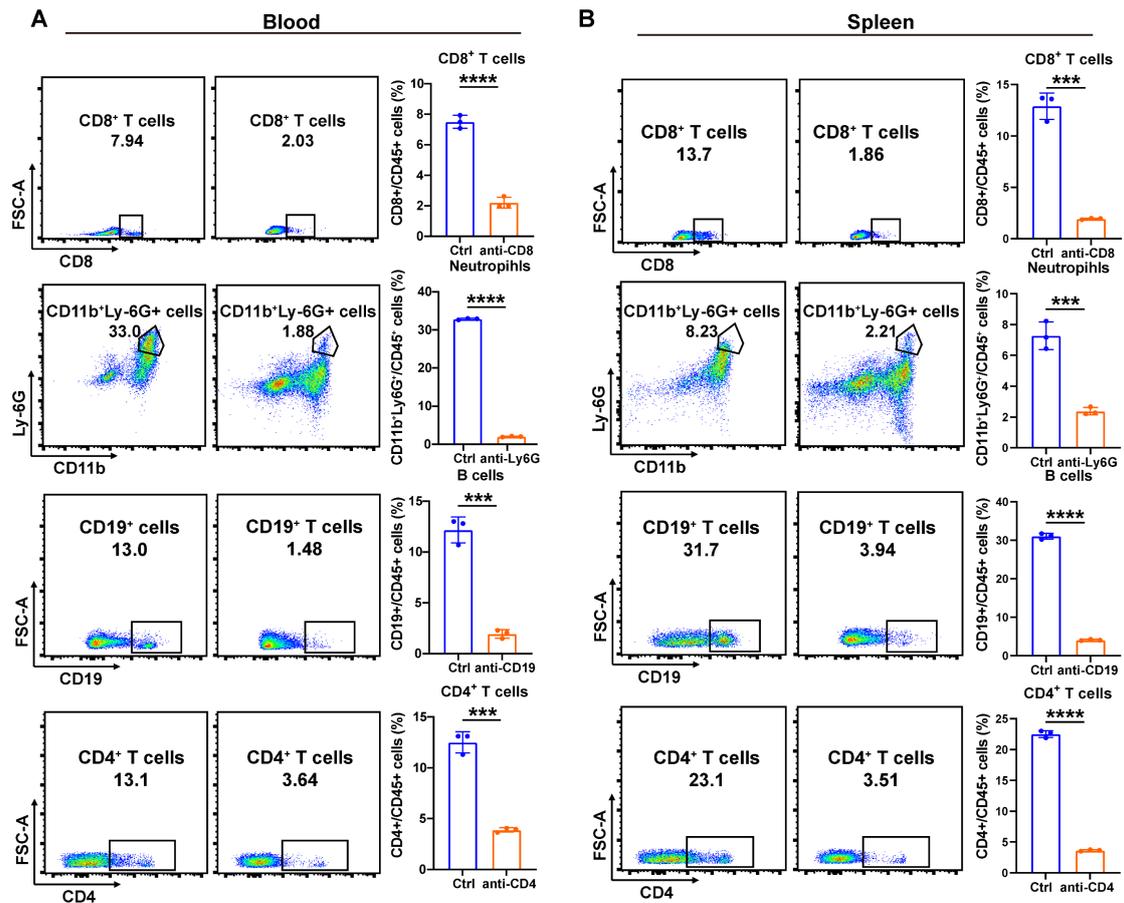
**Supplementary Figure 7. Bio-HCP@FM-NPs increase tumor immune response in the TME, related to Figure 5. (A)** Flow cytometry gating strategy for measuring the percentages of T cells, IFN- $\gamma^+$  and GZMB $^+$  T cells in tumor tissue. (B-C) Percentages

of T cells and CD4<sup>+</sup> T cells in tumor tissue. (D-E) Expression of IFN- $\gamma$  and GZMB on CD4<sup>+</sup> T cells in tumor tissue (n = 3). (F) Percentages of TAMs in tumor tissue.

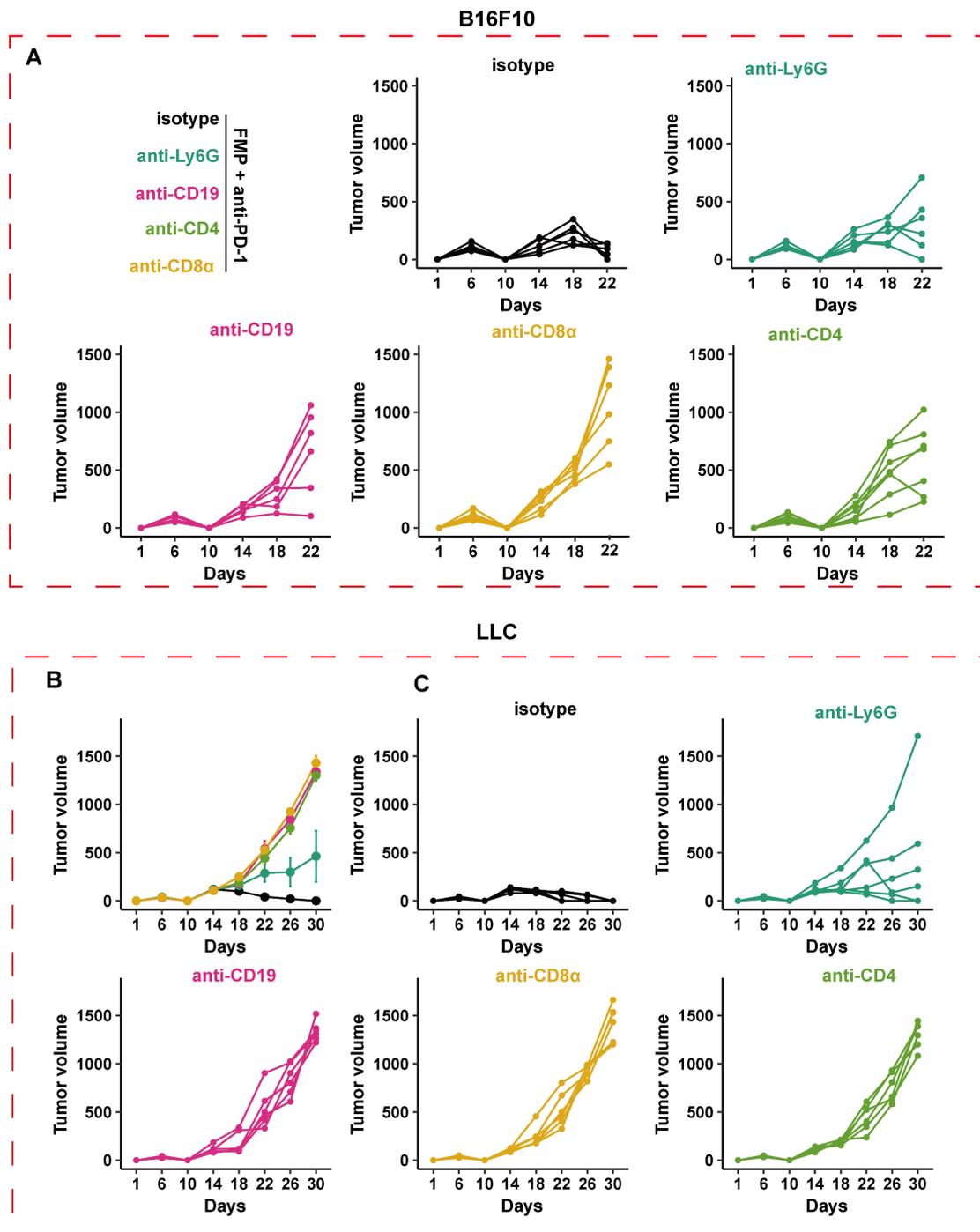


**Supplementary Figure 8. Combinatorial efficacy of personalized Bio-HCP@FM-NPs vaccination and anti-PD-1 against postsurgical tumor recurrence, related to Figure 7.**

(A) B16F10 tumor-growth curves for each mouse in different groups (n = 6). (B) Average tumor-growth curves of *C57BL/6J* mice bearing LLC tumor with different treatments as indicated (n = 6). (C) LLC tumor-growth curves for each mouse in different groups (n = 6).



**Supplementary Figure 9. Validation of the immune cell depletion efficiency, related to Figure 7. (A-B) Flow cytometry analysis of changes in immune cells from peripheral blood and spleen of mice subjected to different immune cell depletion treatments as indicated (n = 6-8 per group).**



**Supplementary Figure 10. Both innate and adaptive immunity are required for tumor recurrence after personalized Bio-HCP@FM-NPs vaccination, related to Figure 7. (A) B16F10 tumor-growth curves for each mouse in different groups (n = 6). (B) Average tumor-growth curves in LLC-bearing mice pre-treated with different monoclonal antibodies, before the combination treatment of Bio-HCP-NPs vaccination**

and anti-PD-1 (n = 6). (C) LLC tumor-growth curves for each mouse in different groups (n = 6).

**Table S1.** Elemental analysis of XPS in series polymer nanoparticles.

Sample	C (Atom %)	O (Atom %)
PS	82.16	17.84
PS@PEG-NPs	79.02	20.98
Bio-HCP-NPs	91.58	8.42

**Table S2.** Specific surface area (SBET), average pore size (DBJH) and pore volume ((Vp)) of series Bio-HCP-NPs before and after loaded drug.

Sample	$S_{BET}$ (m <sup>2</sup> /g)	$D_{BJH}$ (nm)	$V_P$ (cm <sup>3</sup> /g)
Bio-HCP-NPs	8.08	24.45	0.06
PS@PEG-NPs	907.05	6.65	1.02
Bio-HCP-NPs@GM-CSF	349.33	7.50	0.57

**Table S3.** Specific surface area (SBET), average pore size (DBJH) and pore volume ((Vp)) of series Bio-HCP-NPs formed from different proportions of monomers.

Sample	$S_{BET}$ (m <sup>2</sup> /g)	$D_{BJH}$ (nm)	$V_P$ (cm <sup>3</sup> /g)
90/10 St/EG	907.05	6.65	1.02
80/20 St/EG	873.29	3.73	0.55
70/30 St/EG	624.26	6.31	0.40

<b>60/40 St/EG</b>	406.74	11.96	0.78
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