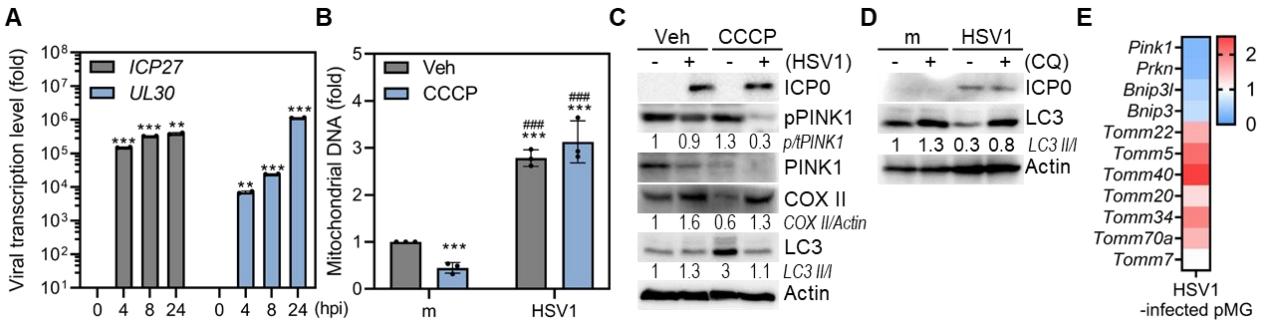




28 **Figure S1.**

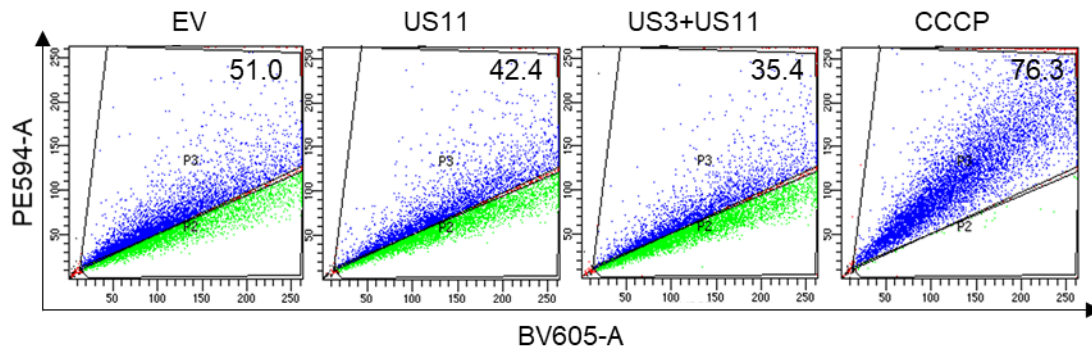


29

30 **Fig. S1. Suppression of mitophagy by HSV1 in murine microglia**

31 **A** Murine microglial cell line (BV2) were infected with HSV1 (MOI 1). At indicated time points,  
 32 RNA was isolated from cells, and *HSV1 ICP27* and *UL30* expression levels were analyzed by  
 33 qRT-PCR. **B, C** BV2 were infected with HSV1 at MOI 1 for 24 h, followed by vehicle control  
 34 (Veh) or 20  $\mu$ M CCCP treatment for 2 h. Mitochondrial DNA copies were quantified by PCR (B)  
 35 and an immunoblot was performed to determine protein expression of HSV1 ICP0, phospho-  
 36 PINK1 (p-PINK1), PINK1, Mitochondria-encoded cytochrome C oxidase II (COX II), LC3, and  
 37 Actin (C). Band quantification relative to Actin is shown below the blot. **D** BV2 were infected  
 38 with HSV1 (MOI 1) for 24 h, followed by 10  $\mu$ M chloroquine (CQ) treatment for 8 h.  
 39 Immunoblot analysis was performed to examine the levels of HSV1 ICP0, LC3, and Actin. **E** A  
 40 heatmap was generated using gene expression profiles from bulk RNA-sequencing of HSV1-  
 41 infected pMG compared with mock-infected pMG (MOI 10). Data are presented as means  $\pm$  SD  
 42 (n = 3 biological replicates; *p* values were calculated by Student's *t* test). \*\* *p* < 0.01; \*\*\**p* <  
 43 0.001, compared with mock (m)-infected Veh-treated group. ####*p* < 0.001, compared with mock-  
 44 infected CCCP-treated group.

45 **Figure S2.**



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47

48 **Fig. S2. Suppression of microglial mitophagy by HSV1-encoded US11**

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50 HMC3-mt-Keima were transfected with either empty vector (EV), US3, US11, or US3+US11-

51 encoding plasmids and mitophagy activity were assessed by flow cytometry. Cells treated with

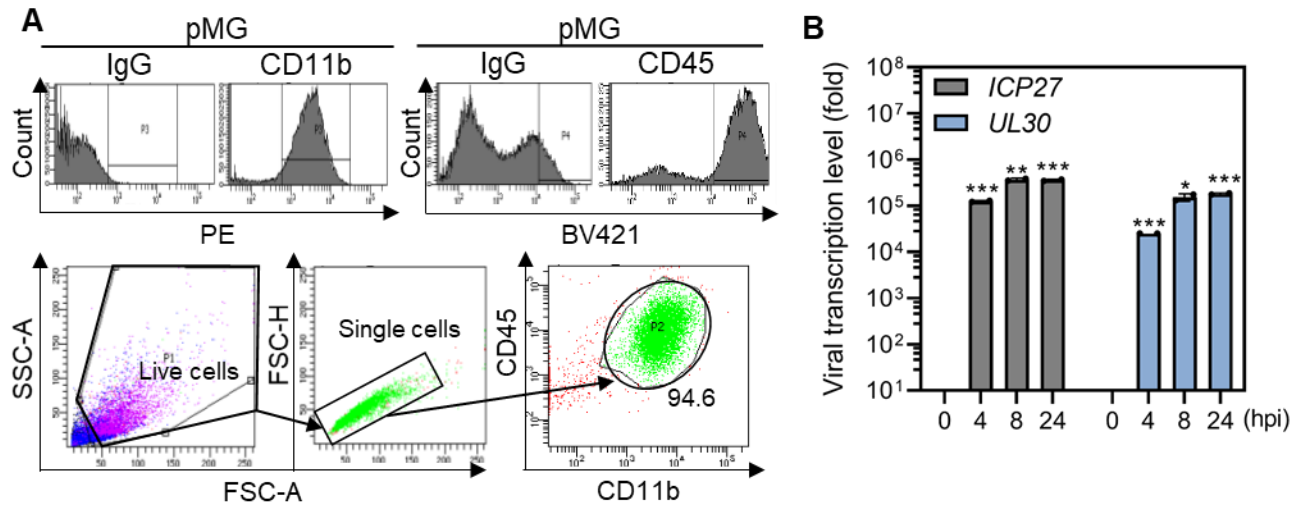
52 20  $\mu$ M CCCP for 2 h were used as positive control for mitophagy induction, as detected by mt-

53 Keima.

54

55 **Figure S3.**

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58 **Fig. S3. Characterization of isolated primary murine microglia (pMG)**

59 A Mixed glial cells (MxG) were isolated from mouse pups and grown for 14 days. After 14 days,  
60 primary murine microglia were isolated from MxG and stained with anti-CD11b and anti-CD45  
61 antibodies for flow cytometry. B pMG were infected with HSV1 at MOI 10. At indicated time  
62 points, RNA was isolated from cells, and qRT-PCR was performed to examine the level of *HSV1*  
63 *ICP27* and *UL30*. Data are presented as means  $\pm$  SD ( $n = 3$  biological replicates;  $p$  values were  
64 calculated by Student's  $t$  test). \*\*  $p < 0.01$ ; \*\*\*  $p < 0.001$ , versus mock ( $m$ )-infected group.

65

66

67



70 **Fig. S4. Alterations in HSV-1-associated host antiviral signaling pathway in microglia**

71 **A, B** KEGG pathway map show upregulated (red) and downregulated (blue) genes in HSV1-  
72 infected pMG (MOI 10) compared with mock-infected pMG. Herpes simplex virus 1 infection  
73 (KEGG #; mmu05168) (A) and Cytokine-Cytokine receptor interaction (KEGG #; 04060) (B)  
74 were generated.

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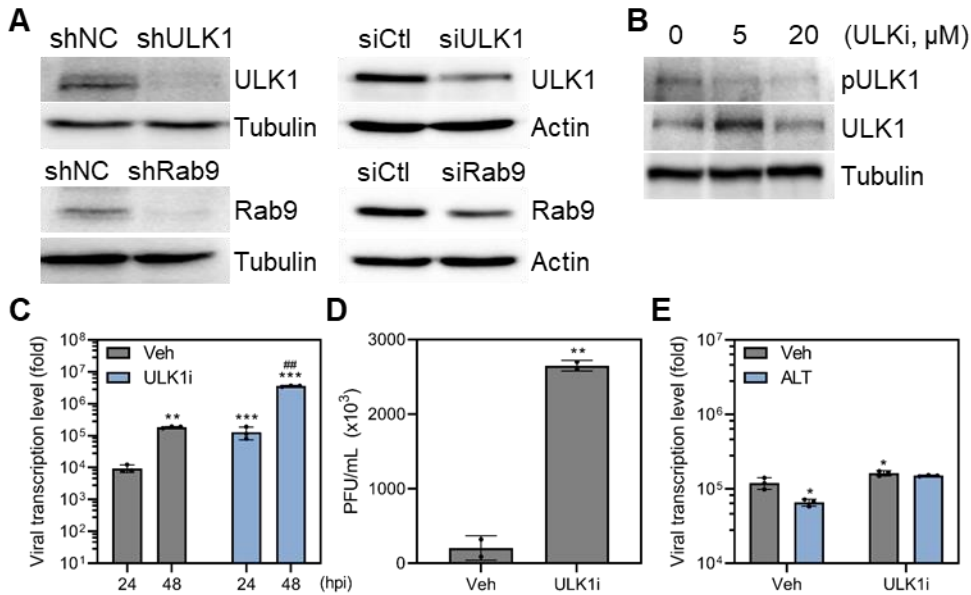
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87 **Figure S5.**



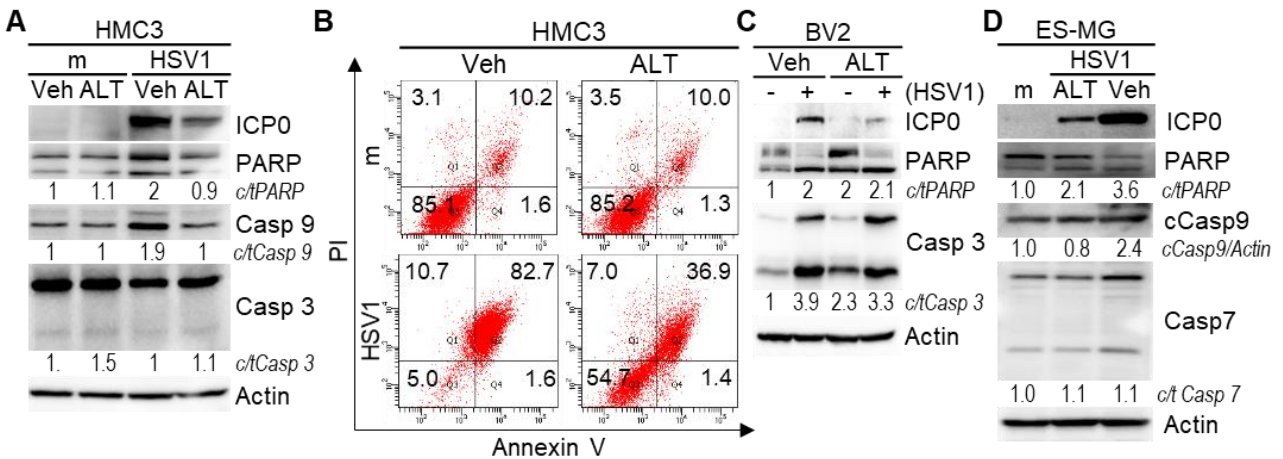
88

89 **Fig. S5. ULK1 modulates HSV1 replication in microglia**

90 **A** HMC3 were transfected with scrambled negative control (shNC), ULK1-specific shRNA  
 91 (shULK1), and Rab9-specific shRNA (shRab9) or control siRNA (siCtl), ULK1-specific siRNA  
 92 (siULK1), and Rab9-specific siRNA (siRab9). The knockdown efficiency of shRNA or siRNA  
 93 was validated by immunoblot analysis showing protein levels of ULK1, Rab9, Actin, and  
 94 Tubulin. **B** HMC3 were treated with 20 μM ULK1 inhibitor (SBI-0206965, ULK1i) for 48 h and  
 95 a representative immunoblot analysis of phospho-ULK1 (p-ULK1), ULK1, and Tubulin is  
 96 shown. **C, D** HSV1 was inoculated in HMC3 (MOI 1) and cells were treated with 20 μM ULK1i.  
 97 *HSV1 ICP27* mRNA levels were analyzed by qRT-PCR (**C**) and a plaque assay were performed  
 98 to determine HSV1 titer (**D**). \* $p < 0.05$ ; \*\* $p < 0.01$ ; \*\*\* $p < 0.001$ , versus vehicle (*Veh*)-treated  
 99 group at 24hpi. ### $p < 0.01$ , versus *Veh*-treated at 48hpi. **E** HSV1 was inoculated in HMC3 at  
 100 MOI 1 and cells were treated with 30 μM ALT001 (ALT) and 20 μM ULK1i. At 48 hpi, RNA  
 101 was isolated from cells, and *HSV1 ICP27* mRNA levels were analyzed by qRT-PCR. \* $p < 0.05$ ,  
 102 versus *Veh*-treated group. Data are presented as means ± SD (n = 3 biological replicates;  $p$  values  
 103 were calculated by Student's  $t$  test).

104 **Figure S6.**

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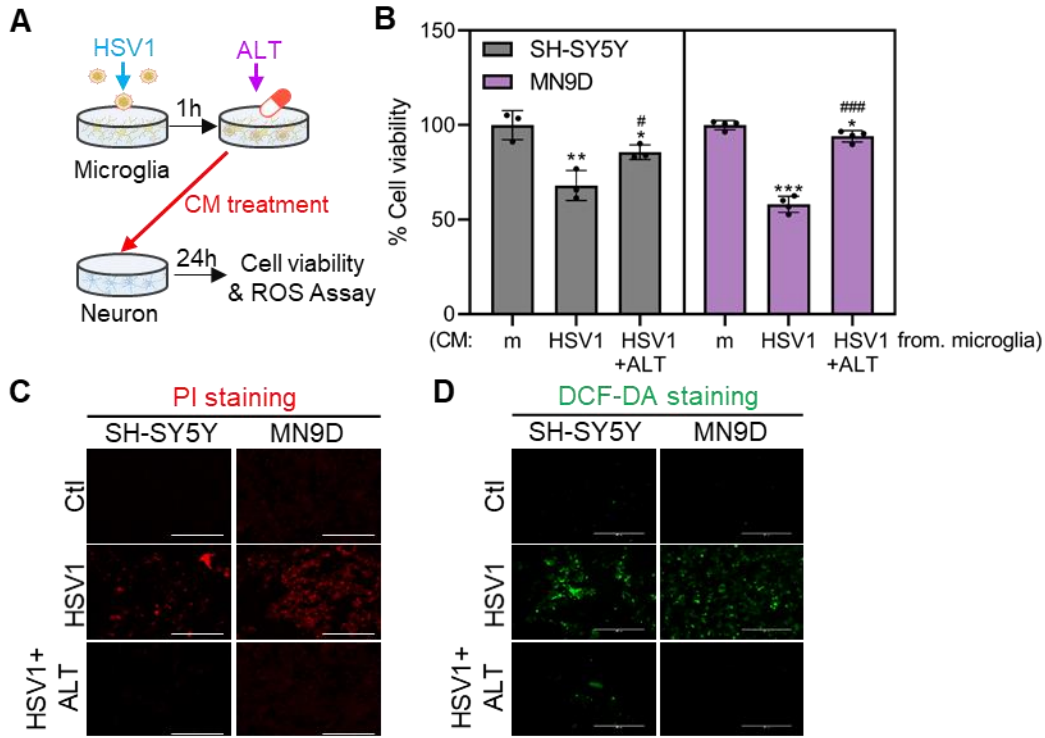
107 **Fig. S6. ALT001 suppresses HSV1-induced apoptosis in microglia**

108 **A, B** HSV1 were inoculated in HMC3 (MOI 1) and cells were treated with vehicle (Veh) or 30  
109  $\mu$ M ALT001 (ALT). At 48 hpi, HSV1 ICP0, cleaved (c) PARP/total (t) PARP, cleaved caspase  
110 9/total caspase 9 and cleaved caspase 3/total caspase 3 expression is shown by immunoblot  
111 analysis (A). Actin was used as a protein loading control. Numbers below the blot represent  
112 quantified band intensity by densitometric analysis. Cells were stained with Annexin V-FITC/PI  
113 and apoptosis was analyzed by flow cytometry (B). **C, D** BV2 (C) or ES-MG (D) cells were  
114 infected with HSV1 (MOI 1) in the presence or absence of 30  $\mu$ M ALT. HSV1 ICP0, cleaved  
115 PARP/total PARP, and cleaved caspase 3/total caspase 3 is shown by immunoblot analysis.  
116 Images are representatives of three independent experiments. Band quantification relative to  
117 Actin is shown below the blot.

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119 **Figure S7.**

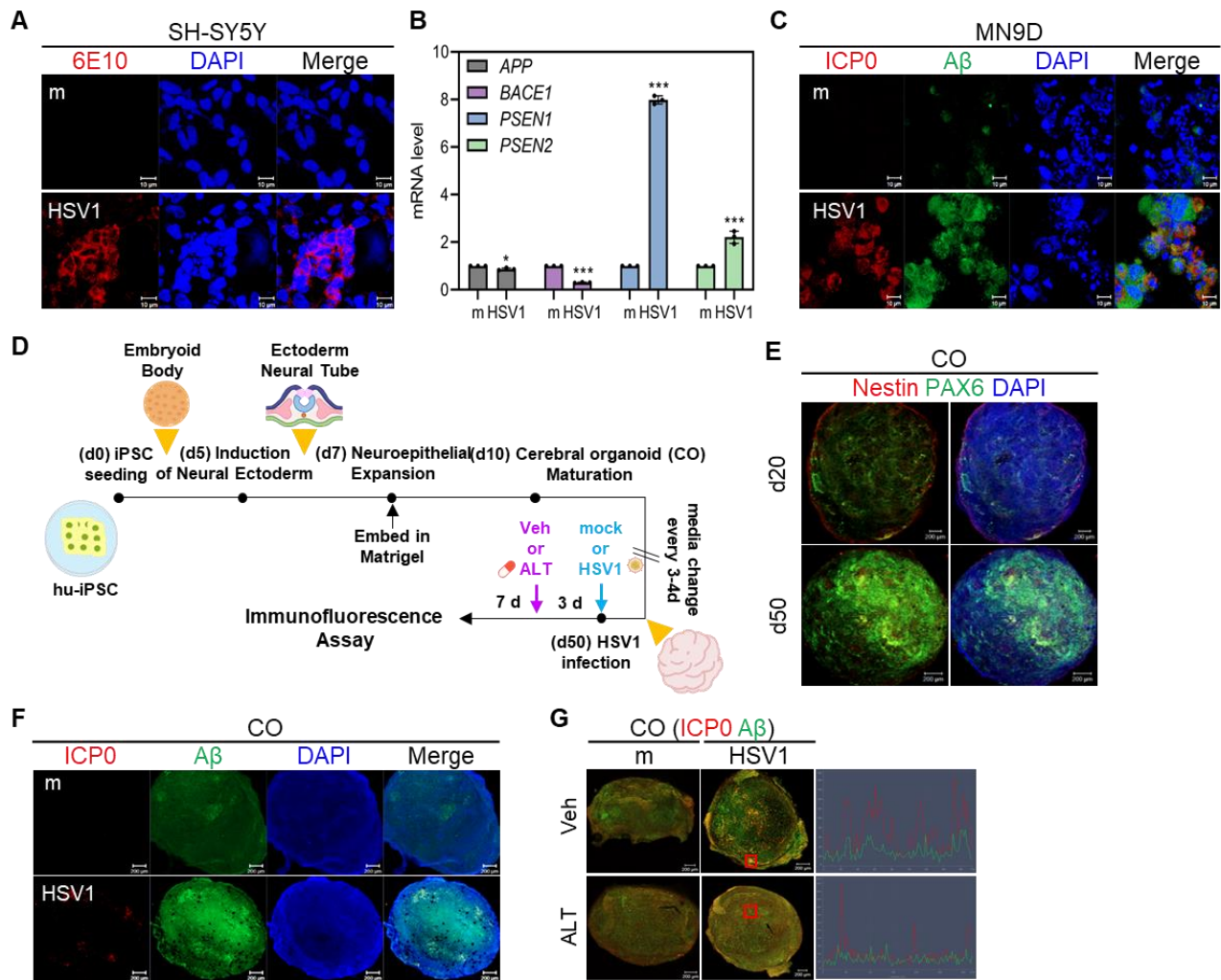


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121 **Fig. S7. ALT001 suppresses HSV1-mediated cell death in neuronal cells**

122 **A** A schematic representation of conditioned media (CM) treatment from microglia to neuronal  
 123 cells is shown. **B** Human and mouse neuronal cells (SH-SY5Y and MN9D) were treated with  
 124 CM collected from mock (m)-, HSV1-, or HSV1-infected with ALT001 (HSV1+ALT) microglia.  
 125 After 24 h, CCK8 assay was performed to measure cell viability. Statistical analysis: one-way  
 126 ANOVA with Dunnett's post-hoc correction. \* $p < 0.05$ ; \*\* $p < 0.01$ ; \*\*\* $p < 0.001$ , versus CM  
 127 from m-infected group. # $p < 0.05$ , ### $p < 0.001$ , versus CM from Veh-treated HSV1-infected  
 128 group. **C, D** CM-treated neuronal cells were stained with propidium iodide (PI) (red) or 2',7'-  
 129 Dichlorofluorescein Diacetate (DCF-DA) (green). Stained cells were analyzed by EVOS FL  
 130 Auto 2 Imaging system. Scale bar = 200  $\mu$ m.

131 **Figure S8.**



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133 **Fig. S8. HSV1 triggers Aβ plaque formation in neuron and cerebral organoids (CO)**

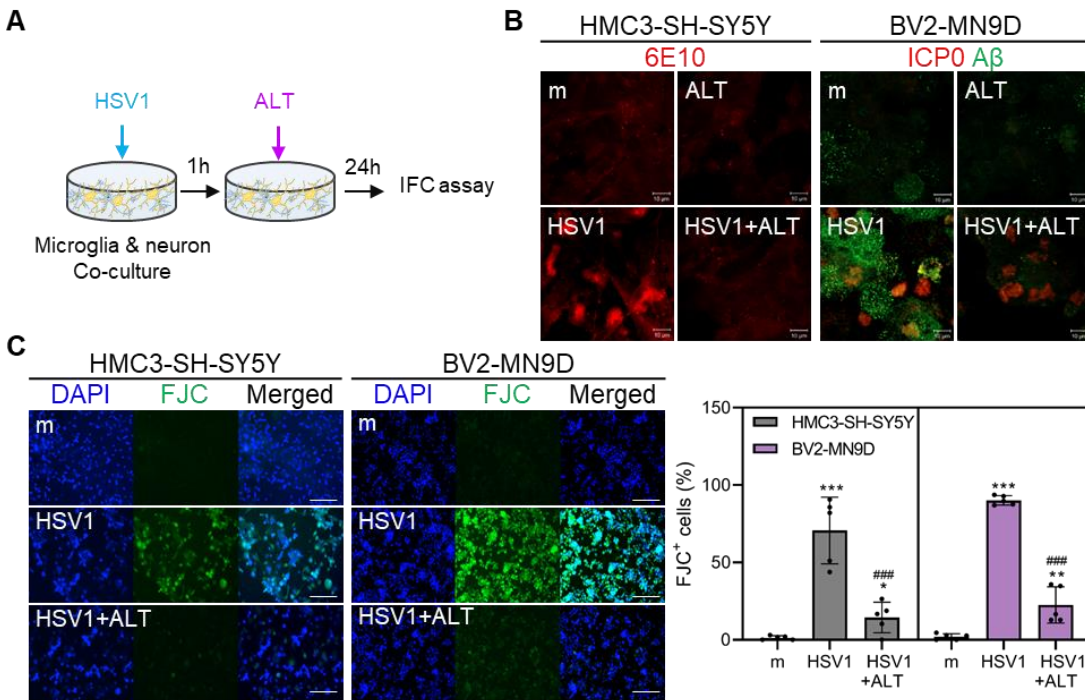
134 **A, B** Human neuroblastoma cells (SH-SY5Y) were infected with HSV1 (MOI 1) for 48 h and  
 135 stained against 6E10 (red) antibodies for confocal microscopy analysis (A) and mRNA levels of  
 136 *amyloid beta precursor protein (APP)*, *beta-secretase 1 (BACE1)*, *presenilin 1 (PSEN1)*, and  
 137 *presenilin 2 (PSEN2)* was measured by qRT-PCR (means ± SD; n = 3) (B). Statistical analysis;  
 138 Student's t test. \* $p < 0.05$ ; \*\*\* $p < 0.001$ , versus mock (m)-infected group. Scale bar=10 μm. **C**

139 Mouse dopaminergic neuronal cells (MN9D) were infected with HSV1 (MOI 1) for 48 h and  
140 were stained with HSV1 ICP0 (red) and A $\beta$  (green) for confocal microscopy analysis. Scale bar  
141 =10  $\mu$ m. **D** Schematic representation of HSV1 infection in cerebral organoid (CO) model is  
142 shown. **E** On day 20 or 50 after CO generation, CO were stained with neuronal marker, Nestin  
143 (red) and PAX6 (green). Scale bar=200  $\mu$ m. **F** On day 50 of CO generation, CO were infected  
144 with HSV1 ( $10^5$  PFU) for 3 days followed by immunofluorescence staining against HSV1 ICP0  
145 (red) and A $\beta$  (green) for confocal microscopy analysis. Scale bar =200 $\mu$ m. **G** CO were infected  
146 with HSV1 ( $10^5$  PFU) for 3 days and incubated with vehicle (Veh) or 30  $\mu$ M ALT001 (ALT) for  
147 7 days. CO were stained with HSV1 ICP0 (red) or A $\beta$  (green) for confocal microscopy analysis.  
148 The graph on the right represents the quantification of fluorescence intensity in the red box area  
149 of images. Scale bar =200  $\mu$ m.

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151

152 **Figure S9.**



153

154 **Fig. S9. ALT001 reduces degeneration of neurons in microglia-neuron co-culture model**

155 A A schematic representation of microglia & neuron co-culture system is shown. HSV1 was  
 156 inoculated in human microglia-neuron (HMC3-SH-SY5Y) or murine microglia-neuron (BV2-  
 157 MN9D) co-culture model and these cells were treated with 30  $\mu$ M ALT001 (ALT). **B** At 24 hpi,  
 158 cells were stained with 6E10 (red), A $\beta$  (green) or HSV1 ICP0 (red) for confocal microscopy  
 159 analysis. Scale bar =10  $\mu$ m. **C** cells were stained with FJC (green). % FJC-positive cells were  
 160 calculated using at least 5 different images. Scale bar=200  $\mu$ m. Data are expressed as the means  
 161  $\pm$  SD of at least three independent experiments. Statistical analysis: one-way ANOVA with  
 162 Dunnett's post-hoc correction. \* $p < 0.05$ ; \*\* $p < 0.01$ ; \*\*\* $p < 0.001$ , versus vehicle (Veh)-treated  
 163 mock (m)-infected group. # $p < 0.05$ ; ### $p < 0.001$ , versus Veh-treated HSV1-infected group.

164

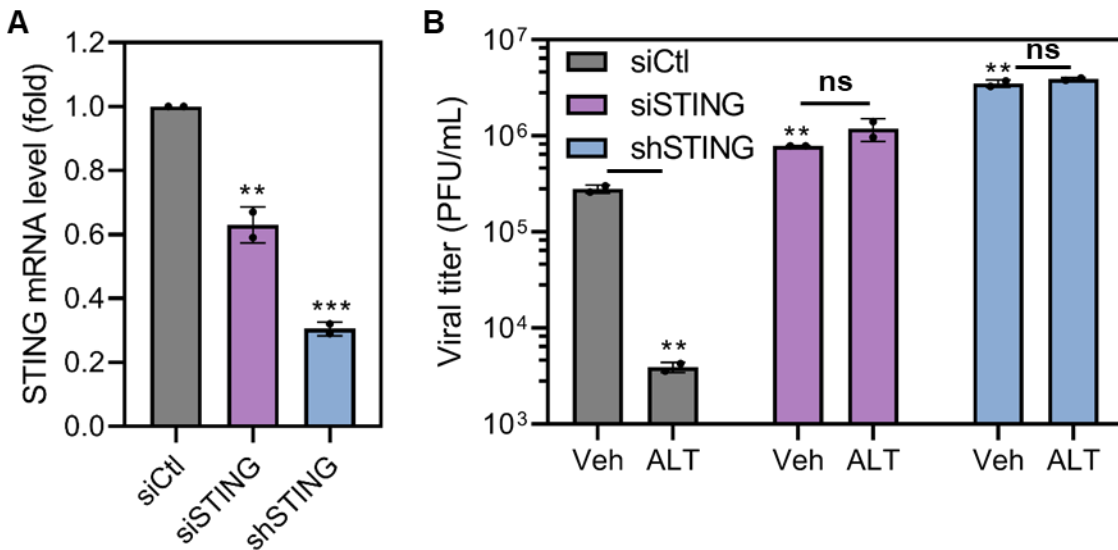
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168 **Figure S10.**

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170

171 **Fig. S10. ALT001-mediated mitophagy exerts antiviral response against HSV1 via STING**

172 A STING mRNA expression level was measured in control siRNA-transfected HMC3 (siCtl),

173 STING-specific siRNA-transfected HMC3 (siSTING), and shSTING-stably expressing HMC3

174 (shSTING) by qRT-PCR. B STING knockdown cells by siRNA or shRNA introduction were

175 infected with HSV1 at MOI 1 in the presence or absence of vehicle (Veh) or 30  $\mu$ M ALT001

176 (ALT). Supernatant was collected at 48 hpi and a plaque assay was performed in Vero cells using

177 the supernatant. Statistical analysis: one-way ANOVA with Dunnett's post-hoc correction.

178 \*\* $p < 0.01$ ; \*\*\* $p < 0.001$ , versus siCtl-transfected Veh group. ns=non-significant.

179