1 Table S1. Sequences of knocked-down genes.

Name	Sequence (5'-3')
shTMBIM1#1	CCGTTTCCCATGGAACATCAT
shTMBIM1#2	GCTGTCTACTACGTGTCCTAT
siYBX1#1	GGAGGCAGCAAAUGUUACA
siYBX1#2	UGUAACAUUUGCUGCCUCC
siYBX1#3	CCACGCAAUUACCAGCAAA

3 Table S2. Coding Sequences for Overexpression Plasmids (TMBIM1 and CCL2)

Name	Sequence (5'-3')
	gaattcATGTCCAACCCCAGCGCCCCACCACCATATGAAGACCGCAAC
	CCCCTGTACCCAGGCCCTCCGCCCCTGGGGGGCTATGGGCAGCCAT
	CTGTCCTGCCAGGAGGGTATCCTGCCTACCCTGGCTACCCGCAGCC
	TGGCTACGGTCACCCTGCTGGCTACCCACAGCCCATGCCCCCACC
	CACCCGATGCCCATGAACTACGGCCCAGGCCATGGCTATGATGGG
	GAGGAGAGAGCAGTGAGTGATAGCTTCGGGGCCTGGAGAGTGGGAT
	GACCGGAAAGTGCGACACACTTTTATCCGAAAGGTTTACTCCATCA
	TCTCCGTGCAGCTGCTCATCACTGTGGCCATCATTGCTATCTTCACC
	TTTGTGGAACCTGTCAGCGCCTTTGTGAGGAGAAATGTGGCTGTCT
	ACTACGTGTCCTATGCTGTCTTCGTTGTCACCTACCTGATCCTTGCC
	TGCTGCCAGGGACCCAGACGCCGTTTCCCATGGAACATCATTCTGC
I MBINI I - F lag	TGACCCTTTTTACTTTTGCCATGGGCTTCATGACGGGCACCATTTCC
	AGTATGTACCAAACCAAAGCCGTCATCATTGCAATGATCATCACTG
	CGGTGGTATCCATTTCAGTCACCATCTTCTGCTTTCAGACCAAGGT
	GGACTTCACCTCGTGCACAGGCCTCTTCTGTGTCCTGGGAATTGTG
	CTCCTGGTGACTGGGATTGTCACTAGCATTGTGCTCTACTTCCAAT
	ACGTTTACTGGCTCCACATGCTCTATGCTGCTCTGGGGGGCCATTTGT
	TTCACCCTGTTCCTGGCTTACGACACAGCTGGTCCTGGGGAACC
	GGAAGCACCATCAGCCCCGAGGACTACATCACTGGCGCCCTGC
	AGATTTACACAGACATCATCTACATCTTCACCTTTGTGCTGCAGCT
	GATGGGGGGATCGCAATGATTACAAGGATGACGACGATAAGTAAgga
	tcc
CCL2	gaattcATGAAAGTCTCTGCCGCCCTTCTGTGCCTGCTGCTCATAGCAG
	CCACCTTCATTCCCCAAGGGCTCGCTCAGCCAGATGCAATCAAT
	CCAGTCACCTGCTGTTATAACTTCACCAATAGGAAGATCTCAGTGCA
	GAGGCTCGCGAGCTATAGAAGAATCACCAGCAGCAAGTGTCCCAA
	AGAAGCTGTGATCTTCAAGACCATTGTGGCCAAGGAGATCTGTGCT
	GACCCCAAGCAGAAGTGGGTTCAGGATTCCATGGACCACCTGGAC
	AAGCAAACCCAAACTCCGAAGACTTGAggatcc

5 Table S3. Primers used

Name	Sequence (5'- 3')		
β-ACTIN-F	CATGTACGTTGCTATCCAGGC		
β-ACTIN-R	CTCCTTAATGTCACGCACGAT		
TMBIM1-F	GAGAGAGCGGTGAGTGATAGC		
TMBIM1-R	ACCTTTCGGATAAAAGTGTGTCG		
CD274-F	GCTGCACTAATTGTCTATTGGGA		
CD274-R	AATTCGCTTGTAGTCGGCACC		
CCL2-F	CAGCCAGATGCAATCAATGCC		
CCL2-R	TGGAATCCTGAACCCACTTCT		
PD-L1-Chip-M1F1	CCAACTTCGGGAACTTTGGG		
PD-L1-Chip-M1R1	GCATGGAGTTCTCTTTGGCC		
PD-L1-Chip-M1F2	CACCCAAACTTACAGTCACCA		
PD-L1-Chip-M1R2	GCTGACACTGCCTTGATTTG		
PD-L1-Chip-M1F3	CTTACAGTCACCAAAATTGCTCT		
PD-L1-Chip-M1R3	TGGGAAATTATTGAGGCTGACAC		
PD-L1-Chip-M2F1	TTTGGAAGCAAATGTCATAACCA		
PD-L1-Chip-M2R1	CCTGTGTGCTCCCTTTTCTT		
PD-L1-Chip-M2F2	TGGAAGCAAATGTCATAACCAAT		
PD-L1-Chip-M2R2	CCTCAAGTGATCCGCCAAAG		
PD-L1-Chip-M2F3	GTGTCAGCCTCAATAATTTCCCA		
PD-L1-Chip-M2R3	TGGACTCTTCGTTGTTTGCC		
PD-L1-Chip-M1F1	AGGAAAGGCAAACAACGAAGA		
PD-L1-Chip-M1R1	GGTGCTCTCTTTTCTCGAACTC		
PD-L1-Chip-M1F2	GGCAAACAACGAAGAGTCCA		
PD-L1-Chip-M1R2	TATCCCGCGCTGAACTTCTA		
PD-L1-Chip-M1F3	AAGAAAAGGGAGCACACAGG		
PD-L1-Chip-M1R3	AATGGGCCCAAGATGACAGA		
CCL2-Chip-M1F1	ACACTGGTTGGGGAGAAAAG		
CCL2-Chip-M1R1	AGTGGCCATTTCAGACTCTG		
CCL2-Chip-M1F2	TGGTTGGGGGAGAAAAGGAGT		
CCL2-Chip-M1R2	GAGAATCCCAGAGCAGAGACT		
CCL2-Chip-M1F3	GGAGTAACTAGTGAGATTCAGGC		
CCL2-Chip-M1R3	GAGAATCCCAGAGCAGAGACT		
CCL2-Chip-M1F4	GCAGGGCTCGAGTTGATTTG		
CCL2-Chip-M1R4	TCTCTTCTTCCTGGGACTAGAC		
CCL2-Chip-M2F1	AGTTTCCTCGCTTCCTTCCT		
CCL2-Chip-M2R1	TCCATTCACTGCTGAGACCA		
CCL2-Chip-M2F2	CTGCAGTTTTCGCTTCAGAGA		
CCL2-Chip-M2R2	AGTAGGAAAGGGAAGCAGGG		
CCL2-Chip-M2F3	TGTGGTCAGTCTGGGCTTAA		
CCL2-Chip-M2R3	TCCATGAGTGATAAGTGGGCT		
CCL2-Chip-M2F4	TGTGGTCAGTCTGGGCTTAA		
CCL2-Chip-M2R4	CAAGCAGGAGGAGGAGGATCTT		
CCL2-Chip-M3F1	TAATGCATTGTCAGGGAGCC		

CCL2-Chip-M3R1	TGCAGAAAAGGAAGGAAGCG
CCL2-Chip-M3F2	TAATGCATTGTCAGGGAGCC
CCL2-Chip-M3R2	TTTCTCTGAAGCGAAAACTGC
CCL2-Chip-M3F3	CCTTGGAATGTGGCCTGAAG
CCL2-Chip-M3R3	TTAAGCCCAGACTGACCACA
musACTB-F	GGCTGTATTCCCCTCCATCG
musACTB-R	CCAGTTGGTAACAATGCCATGT
musINOS-F	GTTCTCAGCCCAACAATACAAGA
musINOS-R	GTGGACGGGTCGATGTCAC
musARG1-F	CTCCAAGCCAAAGTCCTTAGAG
musARG1-R	AGGAGCTGTCATTAGGGACATC
musCCL2-F	TTAAAAACCTGGATCGGAACCAA
musCCL2-R	GCATTAGCTTCAGATTTACGGGT
musTMBIM1-F	AATCTTCACCTTTGTGGAACCAG
musTMBIM1-R	CAGGCAAGGGTCAGGTAGG
musCD274-F	GCTCCAAAGGACTTGTACGTG
musCD274-R	TGATCTGAAGGGCAGCATTTC

7 Table S4. Antibodies used.

Name	Source	Identifier
TMBIM1	Abcam	ab121358
CCL2	Abclonal	A23288
CCL2	Proteintech	66272-1-Ig
PD-L1	Abcam	ab205921
β-ΑCΤΙΝ	Proteintech	HRP-66009
a-TUBULIN	Proteintech	11224-1-AP
Flag	Sigma	F1802
P-YBX1	CST	2900
YBX1	Santacruz	sc-101198
Lamin b1	Proteintech	12987-1-AP
CD33	Abcam	ab270942
CD11B	Proteintech	66519-1-Ig
Ki67	Abcam	ab15580
anti-mouse PD-1	BioXCell	BE0146
IgG2a	BioXCell	BE0146
InVivoMAb anti-mouse/human/rat CCL2 (MCP-1)	BioXCell	BE0185
APC-Cy7 Rat Anti-Mouse CD45(30-F11)	BD Pharmingen	557659
Alexa Fluor 700 Hamster anti-Mouse CD3e(500A2)	BD Pharmingen	557984
FITC Rat Anti-Mouse CD8a (53-6.7)	BD Pharmingen	553030
PE anti-human/mouse Granzyme B Recombinant	Biolegend	396406
Purified Rat Anti-Mouse CD16/CD32 (Mouse BD Fc Block) (2.4G2)	BD Pharmingen	553141
PerCP-Cy5.5 Rat Anti-CD11b(M1/70)	BD Pharmingen	550993
PE-Cy7 Rat Anti-Mouse CD4(RM4-5)	BD Pharmingen	552775
PE CD33 WM53	BD Pharmingen	555450
Ms Ly-6G Ly-6C BV650 RB6-8C5 50ug	BD Pharmingen	740454

9 Table S5. ELISA kits used.

Name	Source	Identifier
CCL2 ELISA human	JINGMEI BIOTECHNOLOGY	JM-1401H1
CCL2 ELISA mouse	JINGMEI BIOTECHNOLOGY	JM-03003M2
PD-L1 ELISA human	JINGMEI BIOTECHNOLOGY	JM-5676H1
PD-L1 ELISA mouse	JINGMEI BIOTECHNOLOGY	JM-11722

- 11 Table S7. Molecular docking results.

TMBIM1(red)	YBX1(green)
ASN-3	THR-7
GLN-44	ARG-282
GLY-46	ARG-282
ALA-51	ARG-279
GLN-55	ARG-247
PRO-56	ARG-247
PRO-54	ARG-253
PRO-62	TYR-241
MET-65	TYR-238







17 TMBIM1 expression levels and prognostic significance across various cancer types.

18 (A) TMBIM1 expression across different cancer types based on TCGA dataset. The box plots represent the

19 transcriptional expression of TMBIM1 (TPM) in tumor (T) and adjacent normal (N) tissues. Tumor types

with significant differences in TMBIM1 expression are highlighted in green (decreased in tumor) and red 20 (increased in tumor) compared to normal tissues (P < 0.05). (B) TMBIM1 expression across pancreatic 21 cancer and other selected cancer cell lines from the HPA database. Pancreatic cancer cells exhibit some of 22 the highest TMBIM1 expression levels compared to other cancer types, as indicated by TPM values. (C) 23 Forest plots showing the prognostic value of TMBIM1 expression in various cancers in TCGA database for 24 Disease-Free Interval (DFI), Disease-Specific Survival (DSS), Progression-Free Interval (PFI), and Overall 25 Survival (OS). Hazard ratios (HR) with 95% confidence intervals (CI) were calculated for each cancer type. 26 Higher TMBIM1 expression is significantly associated with worse prognosis in multiple cancers, including 27 pancreatic cancer, as indicated by the red boxes. Statistical significance is marked by P-values < 0.05. 28 29

30 Figure S2



32 TMBIM1 expression and diagnostic significance in pancreatic cancer.

(A) ROC curve analysis of TMBIM1 expression for distinguishing pancreatic cancer from normal tissues using TCGA and GTEx data (AUC = 0.900, 95% CI: 0.863–0.933). (B) Boxplot showing TMBIM1 expression across different tumor stages in TCGA-PAAD, indicating a positive correlation (Spearman: rho = 0.213, p = 0.00448). (C) Boxplot showing TMBIM1 expression across different tumor grades in TCGA-PAAD (Spearman: rho = 0.199, p = 0.00805).

38



41 Quantification of mRNA and protein expression of TMBIM1 in pancreatic cancer cell lines and stably
 42 transfected cell lines.

(A-B) Relative TMBIM1 mRNA (A) and protein (B) expression levels in human pancreatic cell lines 43 44 (HPDE, Capan-1, CFPAC-1, MiaPaCa-2, SW1990, AsPC-1, and PANC-1) compared with HPDE cells as a control. tatistical significance was determined using Student's t test. (F). (C-D) Validation of TMBIM1 45 knockdown efficiency in Capan-1 shRNA constructs (shTMBIM1#1 and shTMBIM1#2) at the mRNA levels 46 (C) and protein (D). (E-F) Overexpression of TMBIM1 in CFPAC-1 cells transfected with a Flag-TMBIM1 47 48 vector, confirmed at the mRNA levels (E) and protein (F). Statistical significance was determined using Student's t test (C, E). The data are presented as the mean \pm standard deviation (SD). *P < 0.05; **P < 0.01; 49 ***P < 0.001; ns, not significant. 50

51

52 Figure S4



54 Functional assays assessing the impact of TMBIM1 in pancreatic cancer cell lines.

(A-C) CCK8 assays measuring cell viability of Capan-1, PANC-1, and CFPAC1 cells following treatment 55 with TMBIM1 knockdown or overexpression were performed over 4 days. (D) Colony formation assays in 56 Capan-1 and PANC-1 cells, comparing shNC and shTMBIM1. (E) Colony formation assays in CFPAC1 57 cells, comparing vector control and TMBIM1-OE. (F-G) Edu incorporation assays evaluating DNA 58 synthesis and cell proliferation in Capan-1, PANC-1, and CFPAC1 cells, scale bar, 100 µm. (H) Transwell 59 migration assays measuring the migratory ability of Capan-1, PANC-1 following treatment with shTMBIM1, 60 scale bar, 100 µm. All assays were performed in triplicates, with statistical significance determined by 61 Student's t-test and indicated for each comparison to the vector control group. The data are presented as the 62 means \pm SDs.(n.s., not significant, *p < 0.05, **p < 0.01, ***p < 0.001). 63

64 Figure S5.





66 Knockdown of TMBIM1 inhibits pancreatic tumor growth in vivo.

67 (A) Tumor volume in mice injected with PANC-1 cells transfected with either shNC (control) or shTMBIM1 68 was measured over 35 days. (B) Representative images of excised tumors from the shNC and shTMBIM1 69 groups.(C) Tumor weight comparison between control (vector) and shTMBIM1 groups. (D) Representative 70 Ki67 IHC staining of tumor sections, showing reduced proliferation in shTMBIM1 tumors compared to 71 controls, scale bar, 100 μ m. Quantification of Ki67 IHC scores is shown on the right. Data are presented as 72 mean \pm SD. *p<0.05, ***p<0.001.

74 Figure S6



В

Α

Multi-Dimensional analysis of cell types, gene expression, and TMBIM1-driven immune microenvironment in pancreatic cancer

Principal component analysis plot displaying the overall transcriptomic differences between cell types in the 78 PDAC tumor microenvironment, based on CRA001160 and GSE212966. (B) t-SNE plot of PDAC tissue 79 displaying clustering of distinct cell types, including T cells, MDSCs, macrophages, and malignant cells, on 80 the basis of their transcriptomic profiles. (C) Dot plot showing cell type-specific gene expression across 81 various cell populations, including immune, stromal, and malignant cell types, in PDAC, highlighting the 82 distribution of key markers. (D) A Venn diagram illustrating the overlap of genes identified from bulk 83 RNA-seq, single-cell RNA-seq (scRNA-seq), and a curated chemokine list (from TISDB). A total of 40 84 genes were shared between RNA-seq and scRNA-seq, while only one gene, CCL2, was found to be 85 common across all three datasets, highlighting its potential significance in the tumor microenvironment of 86 pancreatic cancer. (E) Dot plot showing cell type-specific gene expression across various cell populations, 87 including mast, PMN-MDSCs, M-MDSCs, B, Macrophages, Monocytes, Treg, and CD8⁺ T cells in all 88 CD45⁺ cells. (F) tSNE analysis illustrating immune cell distribution in TMBIM1-high and TMBIM1-low 89 groups in pancreatic cancer (CD8⁺ T cells (red), M-MDSCs (orange), and PMN-MDSCs (blue) in the 90 TMBIM1-high group (left panel) and TMBIM1-low group (right panel)). 91

93 Figure S7





Ε

200

0

Vector

shTMBIM1

CCL2-OE CCL2-OE+shTMBIM1

Capan-1

95 Functional analysis of CCL2 overexpression (CCL2-OE) and shTMBIM1 in pancreatic cancer cells.

(A) Western blot analysis of CCL2 and GAPDH expression in Capan-1 and PANC-1 cells treated with 96 vector control (SHT), TMBIM1 knockdown (BLAM1), CCL2 overexpression (CCL2-OE), and combined 97 TMBIM1 knockdown and CCL2 overexpression (BLAM1 + CCL2-OE). (B) EDU incorporation assays to 98 assess cell proliferation in Capan-1 and PANC-1 cells across the four treatment groups, scale bar, 100 µm. 99 (C) CCK8 assays evaluating cell viability in the four treatment groups for both Capan-1 and PANC-1 cells. 100 (D) Colony formation assays assessing proliferative capacity of Capan-1 and PANC-1 cells in the four 101 treatment groups. (E) Transwell migration assays measuring the migratory ability of Capan-1 and PANC-1 102 cells in the four treatment groups, scale bar, 100 µm. All assays were performed in triplicates, with statistical 103 significance determined by Student's t-test and indicated for each comparison to the vector control group. 104 The data are presented as the means \pm SDs.(n.s., not significant, *p < 0.05, **p < 0.01, ***p < 0.001). 105

107

108 CD8⁺ T cell migration in co-culture with tumor cells: Impact of TMBIM1 knockdown and rCCL2
 109 supplementation.

(A-B) Migration of CD8⁺ T cells was assessed in Capan-1 tumor cell groups (A) and PANC1 tumor cell group (B) under three conditions: control (NC), shTMBIM1, and shTMBIM1 + recombinant CCL2 (rCCL2). Migration was assessed using a transwell assay. Data are presented as mean±SD. No significant differences in T-cell migration were observed among the three groups, despite triplicate experiments. Statistical analysis was performed using a Student's t-test. (n.s., not significant, *p < 0.05, **p < 0.01, ***p < 0.001).

117 Structural interaction between YBX1 and TMBIM1.

121 Schematic representation of the YBX1 binding motif in the CCL2 and PD-L1 promoter regions.

122 Wild-type (WT) and mutant (MUT) sequences for the CCL2 (-1871 to -1862) and PD-L1 (-1286-1277)

- 123 binding sites are shown.

Western blot analysis of TMBIM1 expression in Pan02 cells. Left panel: Tmbim1-OE compared to vector
control. Right panel: TMBIM1 knockdown using two independent shRNAs (shTMBIM1#1 and
shTMBIM1#2) compared to the negative control (shNC). β-Actin serves as the loading control for all
experiments.

140 Figure S12. Flow cytometry gating strategy for immune cell analysis.

- 141 The gating strategy of multiple immune cells (CD8⁺ T cells, CD4⁺ T cells, MDSCs, Macrophages). The
- 142 frequency of tumor-infiltrating immune cells in tumor bearing mice were used by flow cytometry.

144

145 Immune cell profiling in Vector control and TMBIM1 overexpression groups.

This figure shows immune cell markers in two groups: Vector and TMBIM1 overexpression. The upper three panels represent immune cells with significant differences between the two groups: CD11b⁺GR1⁺ cells, CD8⁺ T cells, CD8⁺/GZMB⁺ T cells; The lower three panels show immune cells with no significant differences between the two groups: CD4⁺ T cells, CD4⁺GZMB⁺ T cells, CD11b⁺F4/80⁺ macrophages, with no statistical difference observed. Statistical analysis for all comparisons was performed using a Student's t-test.

154 Comparison of ips CTLA-4(-) + PD-1(-) scores in low and high TMBIM1 expression groups based on

155 TCIA databases.

153

156 This figure shows the comparison of low TMBIM1 and high TMBIM1 expression groups with respect to

157 IPS CTLA-4 (-) and PD-1 (-) status, using data from the TCIA database. Significant differences were

158 observed between the 2 groups. Statistical analysis was performed using a Student's t-test.

(A) Immunohistochemical staining of TMBIM1 in pancreatic cancer tissues from the FUSCC cohort showing various levels of expression: negative, weakly positive, moderately positive, and strongly positive. (B) Scatter plot of TMBIM1 expression levels in the TCGA-PAAD cohort, stratified by survival time, demonstrating a correlation between higher TMBIM1 expression and shorter survival. (C) Kaplan-Meier survival curve showing OS probability in the TCGA cohort, with higher TMBIM1 expression associated with significantly poorer OS (Log-rank P = 0.0052). (D) Kaplan-Meier survival curve showing PFS

probability in the TCGA cohort, indicating worse PFS in patients with higher TMBIM1 expression 168 (Log-rank P = 0.0011). (E) Kaplan-Meier survival curve for DSS in the TCGA cohort, revealing that higher 169 TMBIM1 expression correlates with significantly poorer DSS (Log-rank P = 0.0041). (F) ROC curves for 1-, 170 3-, and 5-year OS prediction based on TMBIM1 expression in the TCGA cohort, with AUC values of 0.598, 171 0.685, and 0.725, respectively. (G) Kaplan-Meier survival curve of OS probability in the GSE79668 dataset, 172 demonstrating that higher TMBIM1 expression is significantly associated with reduced OS (Log-rank P < 173 0.001). (H) Kaplan-Meier survival curve of OS probability in the CPTAC dataset, showing significantly 174 shorter OS in patients with higher TMBIM1 expression (Log-rank P = 0.0196). 175