Supplemental Materials

A specific and adaptable approach to track CD206⁺ macrophages by molecular MRI and fluorescence imaging

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Figure S1. High resolution mass spectrometry (HR-MS) of MR1-cy5.



Figure S2. High resolution mass spectrometry (HR-MS) of MR2-cy5.



Figure S3. High resolution mass spectrometry (HR-MS) of Mann2-DTPA-Gd.



Figure S4: Relaxivity (r_1) of Mann2-DTPA-Gd.



Figure S5. Validation of the differentiation of CD206⁺ and CD206⁻ macrophages using IFNgamma and IL-4, respectively, from RAW 264.7 cells by flow cytometry (n = 3, unpaired t-test, **, p = 0.0038).

CD206+ macrophages



Fig. S6. Gating strategy for the differentiation RAW cells to CD206⁺ and CD206⁻ macrophages.



Figure S7. Mouse model of wound healing on days 1, 4, and 7.



Figure S8. H&E staining of wound tissues on days 1, 4, and 7. The arrows indicate the infiltrated immune cells around the wound areas. Images: 10x, scale is 100 μm.



Figure S9: Contrast-to-noise ratio (CNR) of wildtype mouse at 45 min post-injection on day 7 was about 2- to 3-fold higher compared to that on day 4 after wound injury.



Figure S10. Gating strategy for flow cytometric analysis in the wound healing model.



Figure S11. Van Steensel's cross-correlation function (CCF) between CD206 and MR2-cy5 in stroke.



Figure S12. High resolution mass spectrometry (HR-MS) of MannGdFish.



Figure S13: Relaxivity (r₁) of MannGdFish.



Figure S14: Blood half-life of MannGdFish detected by ICP-MS using a two-way exponential model was 0.3 min for the fast phase and 6.1 min for the slow phase.