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Review

# Intranasal delivery of imaging agents to the brain

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#### **Abstract**

The potential of intranasal administered imaging agents to altogether bypass the blood-brain barrier offers a promising non-invasive approach for delivery directly to the brain. This review provides a comprehensive analysis of the advancements and challenges of delivering neuroimaging agents to the brain by way of the intranasal route, focusing on the various imaging modalities and their applications in central nervous system diagnostics and therapeutics. The various imaging modalities provide distinct insights into the pharmacokinetics, biodistribution, and specific interactions of imaging agents within the brain, facilitated by the use of tailored tracers and contrast agents.

**Methods:** A comprehensive literature search spanned PubMed, Scopus, Embase, and Web of Science, covering publications from 1989 to 2024 inclusive. Starting with advancements in tracer development, we going to explore the rationale for integration of imaging techniques, and the critical role novel formulations such as nanoparticles, nano- and micro-emulsions in enhancing imaging agent delivery and visualisation.

**Results:** The review highlights the use of innovative formulations in improving intranasal administration of neuroimaging agents, showcasing their ability to navigate the complex anatomical and physiological barriers of the nose-to-brain pathway. Various imaging techniques, MRI, PET, SPECT, CT, FUS and OI, were evaluated for their effectiveness in tracking these agents. The findings indicate significant improvements in brain targeting efficiency, rapid uptake, and sustained brain presence using innovative formulations.

**Conclusion:** Future directions involve the development of optimised tracers tailored for intranasal administration, the potential of multimodal imaging approaches, and the implications of these advancements for diagnosing and treating neurological disorders.

Keywords: intranasal administration, neuroimaging agents, brain imaging, imaging modalities, nose-to-brain

#### **1. Introduction**

The intranasal (IN) administration route for delivering neuroimaging agents has emerged as a promising alternative to traditional approaches of systemic administration. This method capitalises on the unique anatomical connections between distinct regions of the nasal cavity and the central nervous system (CNS), allowing for a direct, rapid and potentially sustained pathway from nose-to-brain [1]. IN administration bypasses the blood-brain barrier

(BBB) and reduces systemic exposure, potentially minimising side effects while maximising the brain bioavailability of imaging agents. This is particularly beneficial in the context of CNS disorders, as features inherent to the BBB prevent or significantly hamper the delivery of agents, therapeutic and diagnostic, intended for brain delivery [2, 3]. Figure 1 illustrates the IN administration of imaging agents through the nasal cavity and their subsequent pathways to the brain, highlighting the primary routes: the olfactory (purple) and trigeminal (blue) pathways and the secondary systemic circulation route across the BBB. Various imaging techniques are employed to trace these agents: magnetic resonance imaging (MRI), optical imaging (OI), focused ultrasound (FUS), single photon emission computed tomography (SPECT), and positron emission tomography (PET) are colour-coded to represent the respective IN route that each imaging technique monitors.

IN drug delivery (INDD) while offering numerous benefits, also presents challenges needing to be addressed for optimal effectiveness, including

i) mucociliary clearance in the respiratory region, which rapidly removes foreign substances limiting drug residence time and absorption;

ii) enzymatic degradation by nasal mucosa enzymes, particularly affecting peptides and proteins; poor permeability of large, hydrophilic molecules through the nasal mucosa;

iii) potential formulation-induced irritation and toxicity, impacting patient compliance and safety; and

iv) variable, unpredictable absorption due to individual differences in nasal anatomy and physiology.

To overcome these challenges, various formulation strategies can be employed, such as

i) using mucoadhesive polymers to prolong drug residence time;

ii) adding enzyme inhibitors to protect drugs from degradation;

iii) incorporating permeation enhancers such as *in situ* gel systems, cyclodextrins, and polymers to facilitate larger molecule absorption;

iv) encapsulating drugs in nanoparticles (NPs) or liposomes to protect them and enhance absorption;

v) developing controlled release systems to maintain therapeutic drug levels over time; and

vi) optimising the pH and osmolarity of formulations to minimise irritation and enhance patient compliance.

Additionally, innovative formulations like nano-emulsions (NEs) and micro-emulsions (MEs) can be used to further enhance delivery efficiency and diagnostic precision. By addressing these challenges with innovative formulation strategies the effectiveness of INDD can be significantly improved [4-6]. Furthermore, advanced delivery devices designed for precise/preferential targeting (e.g. to olfactory region) can improve the distribution and absorption of imaging agents within the nasal cavity, and into the brain. Devices that facilitate the delivery of formulations to the olfactory cleft region of the nasal cavity would minimise loss to swallowing or

lung deposition, enhancing the efficiency of nose-to-brain delivery [6].

Medical imaging techniques used in evaluating IN administration of imaging agents, such as MRI, PET, SPECT, OI, gamma scintigraphy, autoradiography, and certain types of computed tomography (CT) scans, have revolutionised our ability to visualise and map brain structures and functions *in vivo* [7, 8]. In contrast, other diagnostic imaging procedures like conventional X-ray images, CT, and US mainly offer images of physical form and often rely on intravenous (IV) administration of imaging agents, which can present challenges in terms of bioavailability due to insufficient concentrations crossing the intact BBB, and potential systemic side effects from accumulation in the periphery. Conversely, when the BBB is compromised, such as in brain cancer, these imaging agents can take advantage of the altered barrier function [9]. These various imaging modalities span the spectrum of structural to functional imaging. The choice of imaging modality and the delivery method of agents, such as via IN administration, can significantly affect both the type and quality of the images obtained, as well as the insights gleaned from these images [10]. Therefore, it is critical to understand the differences between these modalities when designing studies and interpreting results that involve IN administration of imaging agents.

Despite these advantages, the field of IN administration of neuroimaging agents to the brain remains nascent, with a growing but still relatively limited body of literature and evidence. This systematic review aims to synthesise current knowledge on the topic, evaluate the effectiveness of this route based on available evidence, and identify directions for future research. In doing so, we strive to contribute to the understanding and development of this promising approach in the field of neuroimaging.

# **2. Imaging of intranasal delivery**

The speed and sensitivity of PET imaging make it an invaluable tool for real-time tracking of metabolic processes and physiological activities within the body. Although the potential toxicity is associated with long-lived positron-emitting radionuclides (half-life  $> 2$  h), these substances can pose safety concerns due to prolonged radiation exposure. SPECT and gamma scintigraphy provide complementary insights to PET, and the methods are particularly useful in visualising the physical transit of radioisotopes and their distribution in the brain [11]. OI offers a distinct advantage in visualising biological processes at the cellular level [12]. MRI

stands out for its non-invasive nature and high-resolution structural imaging capability. Here, the use of contrast agents provides opportunities for assessing drug movement and BBB interaction [13, 14]. Although less sensitive to soft tissue contrast, CT gains significant value when combined with PET or SPECT as it provides precise anatomical details with functional insights, leading to more accurate disease identification [15, 16]. Gold NPs (AuNPs) as a source of X-ray attenuation provide a mechanism by which CT images can be used as a real-time tracking device [16].



Figure 1. Pathways of intranasal drug delivery and associated imaging modalities: This illustration depicts the IN administration of imaging agents through the nasal cavity and their subsequent pathways to the brain. It highlights two primary routes: the olfactory (purple) and trigeminal (blue) pathways and the secondary systemic circulation route across the BBB. Various imaging techniques are employed to trace these agents: MRI, OI, FUS, SPECT, and PET are color-coded to represent the respective IN route that each imaging technique monitors. [created with BioRender.com].





Understanding the strengths and weaknesses of each imaging modality is crucial for optimising their use in the INDD pathway. Table 1 presents a comprehensive comparison of the pros and cons of various imaging modalities and neuroimaging agents. The table highlights plausible applications for each modality based on their unique characteristics, providing a clear guide for researchers and clinicians in selecting the appropriate imaging technique for their specific needs. Moreover, the timeline provided in Figure 2 illustrates the progression and number of studies involving various imaging modalities (SPECT, PET, OI, MRI, FUS, CT) for IN neuroimaging agents from 1996 to 2024. The timeline highlights the emergence and adoption of different neuroimaging agents specific to each modality, providing a systematic understanding of the technological advancements in this research area.

Building on this foundational understanding, the subsequent sections will provide an in-depth analysis of each imaging modality and their respective agents in nose-to-brain imaging.

# **2.1. Single Photon Emission Computed Tomography and Gamma Scintigraphy**

Gamma scintigraphy and SPECT are both nuclear medicine imaging techniques that use gamma-emitting rays to visualise internal physiological processes. Gamma scintigraphy provides 2D images and is primarily used for simpler diagnostic tasks such as evaluating organ function and detecting infections. In contrast, SPECT offers 3D imaging by rotating around the object to capture multiple angles, enabling more detailed examinations [17]. Gamma scintigraphy enables precise tracking and visualisation of the radiotracer transit and distribution within brain tissue. It has been applied for evaluating the INDD. The radiotracers are designed to travel along the olfactory and trigeminal pathways to the brain.

A range of imaging agents for INDD have been investigated using SPECT, each with unique properties and half-lives catering to specific study requirements. Technetium-99m [99mTc] is the predominant radionuclide utilised for SPECT, valued for its widespread availability, favourable photon energy for imaging and short half-life of approximately 6 h [18]. Indium-111  $[111]$ , with a longer half-life of 2.8 days, is ideal for extended studies and is often used with peptides and antibodies for targeted imaging [19]. Iodine-123 [123I] offers a balanced half-life of 13.2 h, enabling specific molecule and peptide labelling [20]. Whereas the longer-lived iodine isotopes (i.e. [125I] and [131I]) have specialised

applications in research and therapy. [125I], a low-energy radioactive isotope with a 60 days half-life, is employed in both medical treatments, such as brachytherapy for cancer and research for labelling biological molecules. [131I], with a half-life of approximately 8 days, is used extensively in the treatment of thyroid cancer and hyperthyroidism, as well as in diagnostic imaging [21]. Thallium-201 [201TI], with half-lives of 73 h, serves in specific domains such as radioimmunotherapy and cardiac imaging [22]. Lastly, Rubidium-86 [86Rb], with a half-life of approximately 18.6 days, is utilised for its positron emission properties, primarily in cardiac studies [23]. Together, these agents provide a comprehensive toolkit, ensuring optimal imaging and facilitating advancements in INDD.

# 2.1.1. Technetium-99m ([99mTc])

In 2005, Vyas *et al.* assessed nose-to-brain delivery of [99mTc]-labelled zolmitriptan MEs and zolmitriptan mucoadhesive MEs in rats [24]. The aim was to explore the efficacy of these formulations in delivering zolmitriptan directly to the brain through nose-to-brain, particularly for treating acute migraine attacks. Radiolabelling zolmitriptan with [99mTc] was critical for tracking the *in vivo* distribution of the drug, enabling the quantification of drug accumulation in the brain and other organs following IN administration and IV injection. Measurements of drug concentrations in blood and brain at predetermined intervals, along with calculations of drug targeting efficiency percentage (DTE%), as described by Behl *et al.* in 1998, assess the average ratio of drug delivery between IN administration and IV injection over time [25]. The nose-to-brain drug direct transport percentage (DTP%) measures how much of the drug reaches the brain directly from the nasal route, compared to the total drug amount reaching the brain from the same route. The study revealed that the IN administration of zolmitriptan, particularly via mucoadhesive MEs, facilitated a rapid uptake of the drug into the brain with a DTE% of 533 and a DTP% of 81, surpassing other formulations, including zolmitriptan MEs a DTE% of 255 and a DTP% of 43 and zolmitriptan solutions a DTE% of 189 and a DTP% of 47. The rapid onset of drug action observed, characterised by a relatively short  $T_{\text{max}}$ (time to reach maximum concentration  $(C_{\text{max}})$  in the brain), highlights the direct transport efficiency of the mucoadhesive ME formulation via the IN administration. The tracer's retention time in the brain was monitored up to 8 h after administration, with zolmitriptan mucoadhesive MEs showing brain retention of 0.31 % injected dose per gram  $(ID/g)$  at 8

h. The formulations were administered in the nostrils using a micropipette. The administered volume of the drug formulation intranasally was 10 µL in each nostril. The quantification of the drug that actually reached the brain after IN administration of zolmitriptan mucoadhesive MEs was measured as a brain uptake ratio at 0.50 h post-administration, with values of  $\sim$ 1% ID/g. The gamma scintigraphy imaging provided a visual confirmation of this rapid uptake, showcasing a marked accumulation of the drug in the brain's olfactory region, which serves as a gateway for the nose-to-brain transport, indicating that the drug likely leverages olfactory neural pathways and systemic pathway for entry into the brain. The authors reported significantly higher activity in the brain following IN administration compared to IV injection for up to 8 h. Clearance is a critical aspect of drug delivery and pharmacokinetics, because it determines the duration of drug efficacy and potential side effects. Given the rapid uptake into the brain via the nose-to-brain route, it would be expected that the clearance mechanisms also play a significant role in mediating the overall drug residence time within brain parenchyma. Typically, drugs once delivered to the brain are cleared through metabolic processes within brain cells or via the cerebrospinal fluid (CSF), eventually re-entering the systemic circulation for elimination. The study's findings, indicated that the majority of activity resided in the abdomen despite the targeted brain delivery, suggesting that after exerting the therapeutic effects, zolmitriptan is likely metabolised and cleared through systemic pathways.

Similar results were reported in later studies to ascertain the delivery of MEs of clonazepam, sumatriptan, risperidone and cabergoline, as well as tizanidine hydrochloride-loaded thiolated chitosan NPs to the brain following IN administration [26-30]. SPECT imaging of intranasally administered [99mTc] radiolabelled tramadol hydrochloride microspheres confirmed significant and extended radioactivity accumulation in the rabbit's brain, demonstrating effective CNS targeting [31].

Jogani and co-workers [32] investigated the direct delivery of [99mTc]-labelled tacrine, a cholinesterase inhibitor, to the brain via IN administration using a micropipette-assisted IN administration in for the treatment of Alzheimer's disease. The volume of the drug formulation given via the IN route was  $5 \mu L$  per nare for mice, and  $50 \mu L$  per nare for rabbits. The investigation involved using a tacrine solution in propylene glycol, radiolabelled with [99mTc], via IN administration and IV injection in mice. The DTE% for IN administration was ~207, and the brain drug DTP% was ~52%, indicating that a significant portion of tacrine was directly transported to the brain from the nasal cavity. Additionally, brain scintigraphy imaging in rabbits confirmed higher brain uptake after IN administration than IV injection. The results demonstrated that IN delivery of tacrine led to faster peak times ( $T_{\text{max}}$  60 min) in the brain, higher brain/blood ratios, and significant direct transport from the nasal cavity to the brain. These findings suggest that IN administration could enhance tacrine's bioavailability, reduce hepatotoxicity, and minimise exposure to non-target tissues, offering a potentially effective approach for Alzheimer's treatment. Notably, [99mTc] was observed in the brain as early as 15 min post-administration, peaking at 60 min, and detectable up to 480 min later. The molecular weight and lipophilicity play a crucial role in the biodistribution of drugs, particularly in nose-to-brain delivery systems. Tacrine, a low molecular weight (235 kDa) and highly lipophilic drug (log P 2.71), is expected to travel via the extraneuronal epithelial pathway for direct nose-to-brain delivery. The elimination of the tracer from the brain involved both systemic circulation and direct transport mechanisms, with a significant portion bypassing the BBB.

In 2012, Mustafa *et al.* assessed the IN administration of lipophilic [99mTc]-labelled ropinirole to the brain in rabbits through the olfactory pathway using NE and its homogenised version [33]. The focus was on the fate of these formulations in the CNS. Brain bioavailability was assessed using gamma scintigraphy in a dynamic model involving radiolabelled nanoformulations of [99mTc]-NEropinirole and [99mTc]-suspension ropinirole. The study highlighted the superior brain localisation and  $C_{\text{max}}$  achieved with [ $99$ mTc]-NE-ropinirole compared to [99mTc]-suspension ropinirole, suggesting that the formulation is directly transported from the nasal cavity into the CSF, bypassing the BBB. Comparative dynamic mobility of the different formulations of [99mTc]-NE-ropinirole and [99mTc]-suspension ropinirole was performed by IN administration in rabbits for 30 min. Continuous imaging of the head was performed to track drug mobility from nose-to-brain and systemic circulation. The imaging revealed that the maximum residence time in the nasal cavity was not more than 10 min, likely due to mucociliary clearance. After 15 min, the drug was almost entirely cleared from the cavity, but a clear black signal was observed in the head region, indicating brain uptake. The imaging analysis revealed a faster onset of action, with the optimised homogenised NEs achieving a  $T_{\text{max}}$  at ~6.7 min

post-IN administration, whereas [99mTc]-suspension ropinirole had a  $T_{\text{max}}$  at ~11.7 min. Homogenisation was found to significantly improve brain uptake of ropinirole, underscoring its potential in enhancing CNS drug delivery. The enhanced brain delivery is attributed to the lipophilic nature of the NE carriers, and the smaller size facilitated faster absorption through the olfactory neurons, a crucial factor in overcoming mucociliary clearance, which is a significant challenge in INDD. The suspension form, being less effective, highlights the importance of the formulation's physicochemical properties in biodistribution and efficacy in the nose-to-brain pathway. Moreover, due to the fast absorption and low dose volume, there was minimal escape of therapeutic molecules to the systemic circulation, resulting in low systemic bioavailability.

Another study explored IN administration vs IV injection of chitosan NPs as a delivery vehicle for [99mTc]bromocriptine, a medication commonly utilised in Parkinson's disease treatment; with significant findings reported regarding the biodistribution and brain targeting efficiency of the tracer [34]. The study utilised a micropipette to instil  $5 \mu L$  of the drug formulation in each nostril, totalling 10 µL per administration. The biodistribution study revealed that the brain/blood concentration ratios were 0.47 for IN administration of [99mTc]bromocriptine solution, 0.69 for IN administration of [99mTc]bromocriptine-chitosan NPs, and 0.05 for the IV injection of the same NPs, measured at 0.5 h post-administration. The gamma scintigraphy results indicated that with IN administration, the bromocriptine-loaded chitosan NPs resulted in significantly higher DTE% of ~6 and a DTP% of ~84, increased bioavailability in brain tissue, and improved nose-to-brain delivery compared to solutions of 99mTc-bromocriptine delivered via IN and IV injection. The quantitation of the tracer that reached the brain after IN administration showed that [99mTc]bromocriptine-loaded chitosan NPs achieved a peak brain concentration of 0.15 % ID/g at 1 h post-administration, which declined to 0.03 % ID/g by 8 h, indicating gradual washout of the tracer from the brain. This enhancement in delivery was attributed to the mucoadhesive properties of the chitosan NPs, which extended residence time in the nasal cavity, resulting in improved permeation and sustained drug delivery to the brain. This mucoadhesive property allows the chitosan NPs to interact with the nasal mucosa, thereby decreasing mucociliary clearance and facilitating improved absorption through the olfactory and trigeminal pathways, bypassing the BBB. The observed elimination pattern suggests the tracer was likely

cleared through the glymphatic system and CSF drainage pathways over time. These results collectively affirm the potential of chitosan NPs as an effective and non-invasive delivery system for brain-targeted therapies of Parkinson's disease.

The use of [99mTc]-labelled carbamazepine, a drug used in the management of epilepsy, was investigated using ME formulation to compare IN administration vs IV injection in rats [35]. In each nostril 10 µL was administered using a micropipette to ensure precise and effective delivery of the drug into the nasal cavity. When assessing for differences across the routes, the concentration of carbamazepine in the brain relative to the blood was consistently 2-3 times higher for up to 8 h following IN administration compared to IV injection. The half-life in the brain ranged between 2.76 and 3.55 h, indicating prolonged drug retention. Further analysis revealed that the carbamazepine mucoadhesive ME formulation achieved superior DTE% and DTP% when compared to other tested formulations, including carbamazepine ME and carbamazepine solution. The physicochemical properties, namely globule size, zeta potential, and inclusion of mucoadhesive agents, significantly influenced the distribution and effectiveness of the formulations. The study reported that the carbamazepine mucoadhesive ME had the highest DTE% and DTP%. Specifically, the mucoadhesive ME formulation exhibited a DTE% of  $\sim$ 241 and a DTP% of  $\sim$ 59, indicating that a substantial amount of the drug reaches the brain directly through IN administration. Notably, the mucoadhesive ME formulation exhibited a 2.20-fold increase in DTE% and a 6.62-fold increase in DTP% relative to the carbamazepine solution. Elimination of the tracer from the brain was gradual, with the elimination constant for the brain ranging from 0.19 to 0.25 and the half-life in the brain ranging from 2.76 to 3.55 h, suggesting a steady washout of the tracer over time. These findings underscore the benefits of the mucoadhesive ME formulation by significantly enhancing brain-targeting efficacy, primarily due to efficient direct transport from the nose-to-brain pathway. Gamma scintigraphy images of rats were acquired 0.5 h after IN instillation, and IV injection, revealing that uptake of radioactivity was substantially greater following IN administration of carbamazepine than IV injection. Within the IN-administration formulations, carbamazepine mucoadhesive ME formulation exhibited the highest levels of radioactivity compared to carbamazepine ME formulation and carbamazepine solution.

Diazepam-loaded poly (lactic-co-glycolic acid; PLGA) NPs have been assessed in rats for efficiently

delivering drugs directly to the brain intranasally. Each rat received 20 μL of the radiolabelled formulation, administered intranasally using a micropipette, with 10 μL of [99mTc]diazepam solution and 10 μL of diazepam-NP administered in each nostril [36]. The developed PLGA NPs (diazepam-NP) are nanoscale particles, and the spray droplets themselves are NPs. Gamma scintigraphy allowed for the visualisation and quantification of [99mTc] diazepam-NP biodistribution, with a pronounced increase in radioactivity seen in the rat brain at 0.5 h post-IN administration, which demonstrated the superior uptake of [<sup>99m</sup>Tc]diazepam-NP via IN administration (e.g., 1.35 at 0.5 h), compared to [99mTc]diazepam solution via both IV injection and IN administration (see Figure 3). Furthermore, biodistribution studies undertaken 8 h postadministration allowed tracking of drug persistence in the brain. The brain/blood ratio of the drug was highest for [<sup>99mT</sup>c]diazepam-NP with IN administration across all the measured time points, underlining the potential of this route in maintaining a sustained drug presence in the brain. The NPs provided better DTE% and DTP%, with values of 258 and  $\sim 61$ , respectively. This important finding confirmed effective nose-to-brain transport of diazepam facilitated by a solution of PLGA-NPs, in rats. The active agents, in this case, diazepam, are encapsulated within these NPs, ensuring controlled release and targeted delivery to the brain.

In 2018, a series of notable studies investigated

the applications of  $99mTc$  in INDD. The study by Mandlik developed and characterised zolmitriptanloaded nanostructured polymeric carriers for targeted INDD [37]. They administered 20 μL IN in mice using a micropipette. The SPECT/CT imaging results provided confirmation of the nanocarrier's enhanced brain uptake using IN administration. The biodistribution data consistently demonstrated the enhanced capability of [99mTc]zolmitriptan-NP for targeted brain delivery. Specifically, the brain/blood ratio of IN administration of [99mTc]zolmitriptan-NP 1 h post administration was found to be 5-fold higher than that using IV injection, and 3-fold higher than without the NP and using the IN-administration route. These significant fold increases underscore the potential value of nanostructured polymeric carriers in facilitating efficient nose-to-brain drug transport. Comparative analysis of  $C_{\text{max}}$  and area under the curve (AUC) in the brain for IN administration of [99mTc]zolmitriptan-NP, IN administration of [99mTc]zolmitriptan, and IV injection of [99mTc]zolmitriptan-NP, demonstrated notably higher values for nose-to-brain targeting metrics, such as DTP% at ~6, DTE% at ~557, and direct nose-to-brain drug transport at 82%. The retention time of the tracer in the brain showed significant brain uptake for up to 8 h, with the highest concentration observed within the first-hour post-administration. The elimination of tracer from the brain was monitored over an 8 h period, showing a gradual decrease in brain concentration, implying the nanocarrier facilitated



Figure 3. Gamma scintigraphy images of rats 30 min post-administration/injection. A: [<sup>99mT</sup>c]diazepam solution post-IV injection. B: [<sup>99mT</sup>c]diazepam solution post-IN administration. C: [99mTc]diazepam-NP post-IN administration. Redrawn from [36]. [created with BioRender.com].

prolonged retention and gradual release of the drug in the brain tissue. The biodistribution of the imaging agents indicated that [99mTc]zolmitriptan-NP had enhanced brain uptake due to its nanostructured polymeric carrier, suggesting efficient transport through the olfactory and trigeminal pathways. These findings underscore the potential value of nanostructured polymeric carriers in facilitating efficient nose-to-brain drug transport.

A second study focused on the use of levetiracetam, a selective synaptic vesicle glycoprotein 2A (SV2A) receptor antiepileptic, which was successfully radiolabelled with [99mTc] for imaging the SV2A receptor [38]. [<sup>99m</sup>Tc]levetiracetam was formulated into an ME with a small particle size  $(16.34 \pm 5.58 \text{ nm})$  and favourable polydispersity index (0.382  $\pm$  0.05). Comparative biodistribution studies assessed the DTE% of three formulations: IV injection of [99mTc]levetiracetam solution, IN administration of both [99mTc]levetiracetam solutions, and [99mTc]levetiracetam-ME. Results indicated that the ME formulation exhibited significantly higher brain uptake and a superior brain/blood ratio at all measured time intervals, particularly at 5 min after administration, where the ratio was ~29, compared to ~9 and 0.0014 for the IN administration solution and IV injection solution, respectively. For the IN of [ $99mTc$ ] levetiracetam-ME, the brain uptake was  $\sim$ 4%  $ID/g$  at 5 min post-administration. The retention time of the [99mTc]levetiracetam tracer in the brain was evaluated through biodistribution studies, showing significant brain uptake at all time intervals (5, 15, 30, and 60 min) with the IN administration of [99mTc]levetiracetam-ME demonstrating the highest retention. The study utilised a simple IN administration technique to deliver the [99mTc]levetiracetam-ME, typically involving the use of pipettes or nebulisers to promote accurate dosing. As the study categorised the imaging agents based on their formulation and administration routes. The IV injection of [99mTc]levetiracetam solution exhibited low brain uptake due to its limited lipophilicity and inability to cross the BBB. The IN administration of [99mTc]levetiracetam solution showed moderate brain uptake with lower efficiency compared to the MEs, likely due to limited absorption through the nasal mucosa. In contrast, the IN administration of [99mTc]levetiracetam-ME demonstrated significantly higher brain uptake, attributed to the enhanced lipophilicity and favourable nanosize allowing direct transport through olfactory and trigeminal nerve pathways, bypassing the BBB. These findings suggest its potential as the first SPECT tracer for imaging SV2A receptors and highlight the advantage of using

[99mTc] due to its availability and suitable half-life, making it a more practical choice over other isotopes like [11C]levetiracetam for similar applications.

The third study developed PLGA NPs of baclofen, a neuropathic pain medication, demonstrating enhanced brain delivery and uptake of [99mTc]baclofen-NP through IN administration [39]. Gamma scintigraphy studies in rats demonstrated that maximum uptake was achieved using IN administration at 3 h post-administration, superseding both the IV injection and oral administration routes. Biodistribution studies measured the concentration of NPs in the brain and blood at various time points to 24 h. The maximum percentage of radioactivity  $(24%)$  was observed at 3 h in the brains of rats administered [99mTc]baclofen-PLGA-NPs intranasally, followed by  $\sim 3\%$  in rats that were administered the same formulation intravenously, while oral administration showed minimal brain distribution. This distribution remained high for 24 h post-administration. IN administration resulted in significant brain uptake due to the PLGA NP polymer matrix, acting as a reservoir and enabling direct administration through the olfactory lobe, bypassing the BBB. The study also evaluated pharmacokinetic parameters, finding that the  $C_{\text{max}}$  (~4% ID/g) at 3 h for intranasally administered [99mTc]baclofen-PLGA-NPs was higher in the brain than the  $C_{\text{max}}$  (~3% ID/g) at 3 h for IV injection. The AUC for the brain of rats administered [99mTc]baclofen-PLGA-NPs intranasally was significantly higher 41% hours/g than for IV injection  $\sim$ 34% hours/g. Similarly, the C<sub>max</sub> and AUC for blood were also higher for IN administration. The study also calculated DTE% and DTP%, with values of  $\sim$ 184 and  $\sim$ 46, respectively, for intranasally administered [99mTc]baclofen-PLGA-NPs, indicating efficient targeting to the brain. Biodistribution studies showed that within 90 min, [<sup>99mTc]</sup>baclofen-PLGA-NP levels spiked in the brain  $(\sim 3\%$  ID/g), suggesting an early onset of action. Blood samples revealed that the maximum radioactivity levels was at 90 min (~4% ID/g) for IV injection of  $[99m]$ Tc]baclofen-PLGA-NPs, followed by  $\sim 3\%$  ID/g for IV injection of [99mTc]baclofen aqueous solution. IN administration led to maximum levels at 3 h for both  $[<sup>99m</sup>Tc]$ baclofen-PLGA-NPs  $({\sim}2\%$  ID/g) and [<sup>99mT</sup>c]baclofen aqueous solution (~2% ID/g), as they were slowly absorbed into the bloodstream. In summary, the developed baclofen-loaded PLGA NPs exhibited enhanced brain delivery and prolonged retention compared to aqueous formulations. The *in vivo* gamma scintigraphy and biodistribution studies confirmed the suitability of PLGA NPs as carriers for baclofen to combat neuropathic pain, demonstrating the advantages of IN administration in bypassing the BBB and achieving efficient brain targeting.

In 2020, four studies were published that investigated the potential of 99mTc radiolabelled compounds for advancing brain imaging and drug delivery for various neurological disorders. The first study was designed to formulate a radiolabelled version of olanzapine, an antipsychotic drug known for its limited permeability in the brain, while ensuring it was free from colloidal impurities [40]. The research sought to assess its distribution within the body after being administered through IN administration and IV injection to determine its viability for brain imaging diagnostics. Post-administration of [99mTc]olanzapine, imaging outcomes revealed significant brain uptake after both IN administration using a Hamilton syringe of 6.20  $ID/g$  and IV injection of 5.50  $ID/g$ , with optimal imaging obtained at 0.5 h post-IN administration, and 1 h post-IV injection [40]. These high uptake values demonstrate the [99mTc]olanzapine complex's capability to effectively concentrate and selectively localise within the brain. The [<sup>99mT</sup>c]olanzapine complex achieved the highest brain/blood ratio of 4.70 ID/g at 0.25 h following IN administration. This ratio stayed  $> 1$  for up to 1 h before gradually diminishing to  $\sim$ 1% ID/g at 8 h. Conversely, after IV injection, the brain/blood ratios slowly climbed to peak at a maximum of  $\sim$ 1% ID/g at 4 h. This ratio remained < 1, suggesting that post-IV injection, the [99mTc]olanzapine complex is more prevalent in the blood than in the brain. Furthermore, the compound was rapidly cleared from most bodily organs, underscoring its potential for precise brain imaging. The kidneys were identified as the primary excretion route, as indicated by the high activity/uptake. The delivery mechanisms to the brain following IN administration were primarily through the olfactory and/or the trigeminal nerves, providing a direct route to the brain. Additionally, another portion was absorbed into the systemic circulation via transcellular diffusion through the nasal membrane, then crossing the BBB to reach the brain. The direct route contributed to the major fraction of the [99mTc]olanzapine complex in the brain, while the indirect route provided a minor fraction. In a subsequent study, an NE encapsulating memantine, a drug used for Alzheimer's disease, was developed for IN administration in mice [41]. This compound, a non-competitive NMDA (N-methyl-D-aspartate) receptor antagonist, has limited bioavailability due to first-pass metabolism, producing three main polar metabolites, each with minimal effects on NMDA receptors. The study compared the IN, IV, and oral

administrations of [99mTc]memantine-NE in rats, with gamma scintigraphy and biodistribution studies confirming superior brain uptake percentage radioactivity of  $\sim$ 4% ID/g at 1.5 h post-administration through IN administration using a micropipette, and also the highest concentration in the brain [41]. Blood analysis revealed higher drug levels for the IN administration group at earlier time points. The study observed significant drug uptake in various brain regions including the olfactory bulb, cortex, and hippocampus, indicating successful delivery to target sites. The retention time of the tracer in the brain was observed up to 24 h post-administration. The memantine-loaded NE follows the olfactory and trigeminal pathways for direct nose-to-brain delivery, bypassing the BBB and resulting in higher drug concentrations in the brain. The washout process involves clearance through CSF flow, the glymphatic system, and efflux transporters at the BBB, with systemic absorption leading to metabolism and renal excretion. Transmission electron microscopy was used to confirm the localisation and distribution of the drug within the brain tissue. A third study focused on developing an NE formulation loaded with donepezil hydrochloride, also used in Alzheimer's disease treatment [42]. Donepezil suffers from limited brain availability and peripheral side effects when administered orally. Rats receiving the [99mTc]-radiolabelled donepezil NE formulation through the IN administration route using a micropipette exhibited the highest radioactivity in brain tissue at  $\sim$ 3% ID/g at 1.5 h post-administration, significantly surpassing the levels in rats given the aqueous formulation  $\sim 2\%$  ID/g through IN administration. The study reported pharmacokinetic parameters, including the mean residence time of 12.75 h for the brain tissues of rats administered with the IN administration of [99mTc]donepezil-NE, suggesting that the drug remains in the brain tissue for a considerable duration before being cleared. [99mTc]donepezil-NE was absent in the brains of rats that received it orally. Minimal distribution of the agent was observed in the rat brain with IV injection, or the aqueous formulation via IN administration. The pharmacokinetic analysis demonstrated a gradual decrease in brain radioactivity over 24 h. Overall, the study suggests that the developed [99mTc]-donepezil-NE formulation offers a promising approach for Alzheimer's disease treatment via IN delivery, achieving higher brain uptake and prolonged retention compared to conventional routes. The final study involved an NE formulation loaded with vitamin D3, designed for addressing cerebral ischemia [43]. The administration of the <sup>99m</sup>Tc-vitamin

D3 NE was performed intranasally using a micropipette. Gamma scintigraphy confirmed a markedly higher deposition percentage of  $\sim$ 3% ID/g of the NE in the brain through IN administration versus  $\sim 1\%$  ID/g only for IV injection of [99mTc]vitamin D3 solution at all time points, peaking at 4 h. Notably, the IN administration route for the [99mTc]vitamin D3 NE resulted in approximately 4-fold higher brain deposition, supported by radiometry assays. However, high radioactivity levels were detected in the liver, kidney, spleen, and heart following IV injection, potentially due to rapid absorption and biodistribution. A marked reduction in radioactivity accumulation was noted after 24 h in all samples, indicating the elimination of the radioactive complex from the body through natural metabolic pathways and substantive excretion. In conclusion, the vitamin D3-loaded NE formulated for IN delivery exhibited effective brain targeting, with significant brain uptake and reduced peripheral distribution. This approach demonstrated potential for treating cerebral ischemia by ensuring higher drug concentrations in the brain and minimising systemic exposure.

Most recently, two studies by Upadhaya *et al.* focused on developing an innovative INDD using radiolabelled micelles for diagnosing and treating CNS tumours, particularly gliomas [44, 45]. The first study investigated radiolabelled folate-encapsulated micelles (folic acid tetraethylenepentamine conjugate (FA-TEPA) as a diagnostic aid [45]. These micelles, designed for IN administration, target overexpressed folate receptors in CNS tumours. The folate conjugate, synthesised with a bifunctional chelating agent and radiolabelled with [99mTc], showed high uptake in the brain (around  $16\%$  ID/g at 4 h). Studies in mice and rabbits demonstrated the micelles enhanced brain penetration and safety. The micellar carriers, owing to their nano size, mucoadhesive nature, and enhanced permeation, show significantly higher brain uptake compared to the radiolabelled folate solution, which was confirmed by biodistribution studies, which indicated that the brain uptake of the [99mTc]FA-TEPA was significant over a period of 240 min, with enhanced uptake observed at all time points. SPECT imaging further validated the micelle's effective brain uptake. The stability of the formulation, mucoadhesivity, and biocompatibility make it a promising non-invasive diagnostic tool for brain tumours and potentially other folate-expressing cancers. Additionally, the study indicated higher retention in the brain for the micellar formulation compared to the solution. Histochemical analysis of nasal and brain tissues post-sacrifice after 28 days of

dosing did not reveal any marked differences compared to control animals, indicating no significant toxicity or damage, supporting the safe use of the formulation for IN administration. The micellar formulation's mucoadhesive nature was confirmed by a mucous glycoprotein assay, which showed 95% mucoadhesion within 15 min, sufficient for efficient IN absorption given the nasal mucociliary clearance for an adult lies between 10 and 20 min. Overall, this study supports the potential of radiolabelled folate micelles as an effective diagnostic tool for CNS tumours and potentially other folate-expressing cancers, highlighting their significant brain uptake, safety, and non-invasive administration method. The second study explored nose-to-brain delivery of radiolabelled chemotherapeutic micelles, focusing on glioblastoma treatment [44]. Methotrexate (MTX) was conjugated with a bifunctional chelating agent and labelled with  $[99mTc]$ . The delivery device used to administer the tracer intranasally was a micropipette, and the volume of the drug formulation given via the IN route was 10 μL. The micelles demonstrated increased brain uptake and had a 3-fold enhanced anti-cancer activity compared to the IN solution. Organ biodistribution studies in mice revealed that the micelles preferentially accumulated in the brain, peaking at 120 min after IN administration with approximately  $15\%$  ID/g, compared to the IN solution, which showed approximately  $3\%$  ID/g. They also accumulated in the kidneys, lungs, blood and liver, with minimal activity in the gastrointestinal tract. SPECT imaging of rabbits confirmed the micelle's superior brain distribution. The study suggested combining radiotherapy with chemotherapy via this delivery system could significantly improve glioblastoma treatment outcomes. The elimination of tracer from the brain and other organs was observed through biodistribution studies, showing higher radioactivity in the kidneys, liver, and brain for the micellar formulation group, suggesting renal and hepatic excretion pathways. Histopathological analysis of the nasal epithelium, brain, and lungs indicated that the formulation was safe for nasal administration, with no significant differences between treated and control groups. Both studies emphasised the importance of delivery system characteristics, such as particle size, shape, and mucoadhesive properties, in achieving efficient and targeted INDD to the brain. The use of [99mTc] in these systems enhances their diagnostic and therapeutic potential, providing a targeted, non-invasive, and effective approach to CNS tumour management.

### 2.1.2. Rubidium-86 ([86Rb]), thallium-201 ([201TI]) and, manganese-54 ([<sup>54</sup>Mn])

Ionic balance in neurons is vital for brain function and is regulated by transport across the BBB, choroid plexus, and cell membranes of glial cells and neurons [46]. The olfactory system, with receptor neurons in contact with both the nasal and cranial cavities, offers an alternative ion pathway for reaching the brain directly. These bipolar neurons, located in the olfactory epithelium beneath the cribriform plate of the ethmoid bone, have cilia extending toward the epithelium and axons toward the brain. Their axons pass through the cribriform plate to the olfactory bulb, where they synapse with mitral and tufted cells, and then extend to the primary olfactory cortex. This pathway allows proteins, viruses, and ions to enter the brain directly, bypassing blood circulation [47]. Persson *et al.* showed that IN administration of radioactive cobalt-57 and zinc-65 led to their transport along the olfactory neurons to the olfactory bulb [48, 49]. Similarly, Evans *et al.* observed that when cadmium-109 was applied intranasally, it primarily accumulated in the olfactory bulb on the application side, with a lesser extent in the opposite bulb [50]. Further studies on pike, a type of fish known as Esox Lucius, with intranasally administered cadmium-109 indicated that cadmium was absorbed into the olfactory rosette, nerve, and the anterior part of the olfactory bulb. These findings suggested that cadmium is transported along the olfactory nerve, accumulating at the terminals of the olfactory axons, but it does not proceed along the transneuronal pathway [51]. The transport was shown to be active, with a constant rate of approximately 2.38 mm/hr, and the cadmium strongly accumulated in the anterior olfactory bulb while other brain areas had minimal levels. This indicates a selective uptake and transport mechanism within the olfactory system.

A study aimed to investigate the olfactory transport route of Rb ions and Tl ions following IN administration [52]. The delivery device used to administer the tracer intranasally was a Myjector, administering  $15 \mu L$  of the drug formulation via the IN route. To elucidate the delivery and distribution within the olfactory system, radioactive [<sup>86</sup>Rb] chloride 86RbCl and [201TI] chloride 201TlCl solutions were utilised. The axonal transport of [86Rb] and [<sup>201</sup>TI] through the olfactory nerve pathway in mice utilising both IN administration and IV injection solutions were explored. Detailed analysis revealed the subsequent distribution of these radiotracers within various brain regions post-mortem. The retention time of the tracer in the brain showed significant accumulation in the olfactory bulb for up

to 24 h, with progressive spreading to other brain regions within 12 h post-administration. The results indicated that within 6 h post-administration, there was a notably higher accumulation of these substances in the olfactory bulb on the ipsilateral side ( $[86Rb]$ , 0.7% ID;  $[201T1]$ , 0.5% ID) compared to the contralateral side ([86Rb], 0.08% ID; [201Tl], 0.15% ID). This accumulation was observed to extend over time, with the transported [<sup>86</sup>Rb] and [<sup>201</sup>TI] progressively reaching other brain regions, such as the telencephalon and diencephalon, more prominently on the side of dose administration. Elimination of the tracer from the brain involved gradual decreases in concentration over time, with [86Rb] uptake rates showing a decrease after 6 h and further at 12 and 24 h post-administration. [86Rb] autoradiography after ipsilateral administration found high concentrations of [86Rb] were noted in the olfactory epithelium and olfactory bulb. In the coronal head sections of mice, there was a noticeable dense accumulation of [86Rb] on the right side, including areas around the hippocampus. In sagittal sections, there was a significant gathering of [86Rb] in the olfactory bulb 6 h post-administration, with subsequent spreading to further areas of the brain. Importantly, the [86Rb] was located within the brain tissue itself, not in the cranial cavities or ventricles. These findings suggest a pathway for [86Rb] transport from the nostril to deeper brain areas. [201TI] autoradiography found dense accumulation in specific areas of the olfactory bulb and cortex. These findings provide strong evidence of direct axonal transport of these substances from nose-to-brain via the olfactory pathway, potentially mirroring the transport behaviour of potassium in the olfactory system. Moreover, following IN administration, [201Tl] was observed to be absorbed by regions including the olfactory tract, olfactory cortex, thalamus, and hypothalamus. This study opens avenues for using such radiotracers in medical imaging, particularly for diagnosing conditions like anosmia and neurodegenerative diseases.

In a 2008 study, researchers examined the transport of [201TI] and [54Mn] ions in the olfactory nerve of mice [53]. The study focused on understanding the IN administration of these ions and their subsequent transport to the olfactory bulb. The experiment utilised both control mice and a transected olfactory nerve fibre model, to whom a double tracer solution of  $^{201}TIC$ l and  $^{54}MnCl$ <sub>2</sub> was applied via IN administration. The delivery device used was a micropipette, and the volume of the double tracer solution administered was 20 µL. Uptake of these tracers was measured in the olfactory

bulb and other organs three hours postadministration. Uptake of these tracers was measured in the olfactory bulb and other organs 3 h post-administration. A significant reduction in the uptake of both [201TI] and [54Mn] in the olfactory bulb was found compared to the control group, indicating that the transection of olfactory nerve fibres impedes the transport of these ions. The biodistribution of [<sup>201</sup>TI] and [<sup>54</sup>Mn] was dependent on their physicochemical properties as alkali and transition metal ions, respectively. Both tracers were transported to the olfactory bulb, but their uptake was significantly reduced in the transected nerve model, indicating their reliance on olfactory nerve fibres for transport. Furthermore, the autoradiographic analysis showed marked inhibition in the transport of [201Tl] and [54Mn] from the nasal epithelium to the olfactory bulb, highlighting the role of olfactory nerve fibres in this process. This study suggested the potential diagnostic applications for assessing olfactory nerve damage, thereby contributing valuable insights into both neuroscientific research and clinical practice.

In 2011, Shiga *et al.* investigated the transport of IN [201TI] to the brain using SPECT/MRI hybrid imaging technique [54]. Healthy volunteers with normal olfactory thresholds were enrolled, and 201TlCl saline solution was applied via IN administration. The delivery device used for administering the tracer intranasally was a syringe, which instilled 0.3 mL of 201TlCl saline solution into the olfactory cleft in either the right or left nasal cavity. The uptake of 201TlCl was first confined to the olfactory epithelium area 30 min post-administration, then spread to the intracranial space in the anterior skull base through the cribriform lamina after 24 h. This indicated that the peak uptake of 201Tl entered the olfactory bulb in the anterior skull base through the cribriform lamina 24 h after nasal administration. In this study, 201Tl was initially concentrated in the olfactory epithelium and later transported to the olfactory bulb through the olfactory nerve pathway. This pattern underscores the olfactory nerve axonal transport as the primary IN delivery pathway for this agent. The findings also highlight the potential infectious routes for viruses like influenza, and SARS/COVID, to the CNS via the olfactory pathway, with recent evidence pointing towards the distinct possibility of viral infections affecting the brain through the olfactory nerve transport route [55, 56]. The study discussed the potential for using the IN administration of 201TlCl followed by SPECT/MRI imaging in patients with upper respiratory tract infections and hyposmia to assess viral damage to the olfactory pathway. The study further contributed to the understanding of the infectious route of prion

proteins to the CNS, suggesting the olfactory pathway as a plausible route. These results provide the first images of olfactory transport to the anterior skull base in humans and highlight the need for future studies to establish viral damage in patients with upper respiratory tract infections and to further explore the olfactory nerve's role in the transmission of pathogens to the CNS. This study underscores the importance of understanding nose-to-brain transport mechanisms for developing diagnostic and therapeutic strategies for neurological diseases.

A series of studies by Shiga and the team explored the use of [201Tl] in olfactory imaging to assess and understand various olfactory disorders [57]. The delivery device used in the study was a syringe, which was used to administer a 0.3 mL saline solution of [201Tl] into the olfactory cleft of the nasal cavity. The uptake of  $[201T1]$  was assessed 24 h after IN administration using SPECT scans. At this time, the tracer's presence in the olfactory bulb was observed, however, most of the [201Tl] had migrated to the nasopharyngeal region and was swallowed, resulting in minimal systemic effects due to the low CNS absorption of [201Tl]. The migration of [201Tl] to the olfactory bulb was significantly lower in patients with olfactory impairments caused by head trauma, upper respiratory infections, or chronic rhinosinusitis compared to healthy volunteers. Representative migration values included a 60-year-old healthy male volunteer who had a high migration level of 29%. In contrast, a 44-year-old female with head trauma showed a lower migration value of 4.2%, a 42-year-old female with an upper respiratory tract infection had a migration value of 4.5%, and a 67-year-old female with chronic rhinosinusitis had a migration value of 5%. This technique proved valuable in predicting recovery in patients with idiopathic olfactory disorders, where higher [201Tl] migration indicated a better prognosis. Patients with idiopathic olfactory dysfunction who showed high nasal migration of [201Tl] to the olfactory bulb recovered their sense of smell more quickly than those with low migration. The findings indicate that severe damage to the olfactory nerve, as detected by [201Tl]-based olfactory imaging, is associated with a poorer prognosis for patients experiencing loss of smell.

Their second study was to see if this imaging technique could predict recovery outcomes following treatment [58]. The study involved 24 patients (7 women and 17 men, aged 23–73 years) with idiopathic olfactory dysfunction. These patients were retrospectively analysed based on their [201TI]-based olfactory imaging results prior to receiving conventional treatment with the Japanese herbal

medicine tokishakuyakusan. The delivery mechanism involved the IN administration of 0.3 mL of [201Tl] into the olfactory cleft. The SPECT/CT imaging assessed the migration of [201Tl] to the olfactory bulb 24 h post-administration. Additionally, MRI was used in conjunction with SPECT/CT to accurately delineate the olfactory bulb and assess its volume. The key findings indicated that high [201Tl] migration to the olfactory bulb  $\geq 4.6\%$  was significantly correlated with a better prognosis and shorter recovery duration. Patients with high [201Tl] migration had a 67% recovery rate 1 year after treatment, compared to 0% in patients with low migration. The extent of [201Tl] migration served as a reliable prognostic marker for recovery in idiopathic olfactory dysfunction, suggesting that patients with intact olfactory nerve fibres could be identified using this imaging technique for targeted follow-up and treatment. Other potential prognostic factors, including gender, age, smoking habit, parosmia, and olfactory bulb volume, were not significantly correlated with prognosis in this study. The researchers discussed the need for a more rapid olfactory nerve tracer for clinical applications, as the current 24 h assessment period for [201Tl] migration requires patients to return to the hospital for imaging. They highlighted [125I] conjugated human recombinant insulin-like growth factor-1 (IGF-1) as a potential alternative. This tracer can be detected in the mouse cerebrum 30 min after IN administration and is prevented from increasing certain brain activities if the olfactory bulb is damaged, indicating its transport through the olfactory nerve [59]. However, [125I] with its long half-life and low energy make it unsuitable for clinical SPECT imaging, suggesting the need for developing new isotope-conjugated tracers for IN administration.

Further research highlighted the variation in [201Tl] migration in patients with parosmia following upper respiratory tract infections, revealing higher nasal [201Tl] uptake in the olfactory cleft in those without hyposmia [60]. However, this increased uptake did not translate to significant differences in [201Tl] migration to the olfactory bulb between groups with and without hyposmia. The olfactory bulb volume was notably larger in the parosmia-only group, although still below the normal range, suggesting specific impacts on olfactory structures under different conditions.

In a later study, the authors discovered that [<sup>201</sup>Tl] migration increased in scenarios where olfactory sensory neurons experienced reduced presynaptic inhibition from dopaminergic interneurons [61]. These findings collectively demonstrate the effectiveness of [201Tl]-based olfactory imaging in

evaluating olfactory nerve connectivity, providing prognostic insights for olfactory dysfunction, and understanding the neurobiological mechanisms underpinning olfactory processes that can affect the nose-to-brain pathway.

# 2.1.3. Iodine radioisotopes ([125I], [123I], [131I])

The IN administration of [125I] labelled compounds presents a viable approach for non-invasive CNS imaging and targeted INDD. The IN application of targeting and trafficking mechanisms of cholera toxin (Ct) and its B subunit (CB) when administered intranasally, focusing on their interaction with the CNS, particularly the olfactory nerves/epithelium and olfactory bulbs [62]. Utilising specific pathogen-free mice, the researchers tested radiolabelled Ct, CB, tetanus toxoid (TT), and ovalbumin (OVA), with a particular emphasis on Ct's role as a mucosal adjuvant labelled with [125I]. The delivery device used in the study for IN administration was a micropipette. The IN administration of these substances was analysed to determine their dispersal and retention across various tissues. [125I]-labelled CB was administered intranasally, and its presence was tracked in both lymphoid (nasopharyngeal-associated lymphoreticular tissue, cervical lymph nodes, mesenteric lymph nodes, spleen) and neuronal tissues (olfactory nerves/epithelium, olfactory bulbs, brain). The CB appeared in the peripheral blood within 15 min, accounting for approximately 50% ID/g. The excretion of [125I]CB from the blood and spleen followed similar patterns, returning to baseline levels within 48 h. There was a notable initial accumulation of [125I]CB in the nasopharyngeal-associated lymphoreticular tissue, although less than 0.02% ID/g of it remained in this tissue after six days. The radioactivity levels of [125I]CB in the cervical lymph nodes reached their highest between 1 and 2 h post-administration, showing a distinct profile from that observed in the blood, spleen, nasopharyngeal-associated lymphoreticular tissue. The slower decline of [125I]CB in the cervical lymph nodes compared to the nasopharyngeal-associated lymphoreticular tissue suggests a typical antigen uptake route in the nasopharyngeal-associated lymphoreticular tissue followed by movement into the cervical lymph nodes. The olfactory nerves/epithelium and the olfactory bulbs exhibited a distinct pattern of CB accumulation unlike what was observed in lymphoid tissues. The radioactivity levels in the olfactory nerves/epithelium reached a peak at 6 h, then stabilised, and sustained over a period of 6 days. In contrast, the levels of [125I]CB in the olfactory

bulbs peaked at 15 min and maintained a relatively steady state thereafter. Moreover, the olfactory nerves/epithelium and olfactory bulbs were the only tissues where this unique accumulation pattern of [125I]CB was noted. The radioactivity levels of [125I]CB in the brain followed a trajectory similar to those associated with CB in the blood, dropping to below 50 CB-specific counts per minute (cpm) by 24 h. When analysing the concentration of [125I]CB relative to the organ weight, the disparities in tissue association became even clearer. In the olfactory bulb, the levels of CB were about 34 times higher, and in the olfactory nerves/epithelium, they were approximately 620 times higher than those seen in the brain per milligram of tissue. This suggests that the pronounced accumulation of CB in these areas likely occurred through binding to monosialoganglioside (GM1), given that no similar accumulation was observed when TT or OVA were administered intranasally. Furthermore, the biodistribution of [125I]Ct holotoxin after IN administration, was somewhat similar yet slightly different in dynamics from that of [125I]CB. Initially, the [125I]Ct showed a delayed peak in blood levels but experienced a significant decrease within 12 to 24 h post-administration. Alternatively, most tissues including cervical lymph nodes, blood, spleen, and brain showed a later peak in  $[125]$ Ct, which remained elevated longer than the levels of [125I]CB. An exception was the nasopharyngeal-associated lymphoreticular tissue, which displayed very similar clearance kinetics to the CB. Notably, the olfactory nerves/epithelium bound higher levels of the Ct compared to the CB at 1.5 h. The levels of [125I]Ct then decreased rapidly within the first 24 h and subsequently declined more slowly over the next six days, a similar observation also noted with the CB. The residence of both Ct and CB in the olfactory bulbs followed similar patterns in the olfactory bulbs throughout the six days tested. This consistent pattern in both forms suggests that neuronal binding is a distinctive feature of Ct. Additionally, to investigate whether Ct when used as a mucosal adjuvant can enhance the delivery of protein vaccines into neuronal tissues, non-labelled holotoxin was administered intranasally along with [125I]TT. After IN administration of [125I]TT, the distribution across various tissues was assessed and compared with that of [125I]TT administered on its own. A slower clearance of [125I]TT was noted in both lymphoid and CNS tissues. Elevated levels of [125I]TT were observed at 24 and 48 h, which then gradually decreased over a six-day period. Notably, there were significant increases in [125I]TT concentrations in the olfactory nerves/epithelium from 12 to 48 h post-

administration when Ct was used, unlike when [125I]TT was administered alone.

In a study targeting Alzheimer's disease, researchers used basic fibroblast growth factor (bFGF) labelled with  $[125]$  [63]. Since amyloid-beta  $(A\beta)$ plaques are a hallmark of Alzheimer's disease and these deposits are rich in heparan sulfate proteoglycans, which can bind to bFGF and serum amyloid P components, the researchers explored a new method of ligand delivery to the brain. This method was tested to detect Aβ deposition in a transgenic mouse model that overexpressed the Aβ-protein precursor. The delivery device used was a tapered-end plastic tip, and each mouse received six drops (1.5  $\mu$ L each) of [125I]-bFGF intranasally. This method utilises the olfactory pathway to bypass the BBB, allowing direct access to the brain through the cribriform plate due to the open intercellular clefts in olfactory epithelial cells. Significant brain uptake of [125I]-bFGF was observed 3 h after administration, with notable uptake in the olfactory bulb, frontal cortex, parietal cortex, hippocampus, and cerebellum. Transgenic mice exhibited 3-5 fold higher uptake in these regions compared to wild-type controls. The study compared bFGF and serum amyloid P components, and their distribution in the brain. Serum amyloid P components demonstrated stronger binding to amyloid plaques, especially in the cortex and microvessels, compared to bFGF, which showed intense staining in neurons around Aβ deposits in the frontal, parietal, and occipital regions. This higher intensity of serum amyloid P components staining indicates its higher affinity for amyloid plaques relative to bFGF. The study also noted significant aspiration into the trachea and stomach, suggesting that much of the intranasally administered bFGF entered the respiratory and digestive tracts. Interestingly, IV injection did not result in significant brain uptake, underscoring the importance of the IN route for targeting the CNS. High-resolution histochemical findings included light microscopy showing specific binding of intranasally administered serum amyloid P components and bFGF to amyloid plaques. Serum amyloid P components demonstrated more intense immunostaining than bFGF. Electron microscopy confirmed the presence of bFGF within amyloid plaques and associated neuronal structures in transgenic mice. In conclusion, this study presents a promising non-invasive method for labelling Aβ deposits using IN administration of bFGF and serum amyloid P components. The results support the potential use of these agents for future imaging studies in Alzheimer's disease, providing a foundation for developing diagnostic tools and

evaluating therapeutic interventions, potentially adaptable for human use with IN administration of radiolabelled ligands and subsequent imaging using SPECT or PET techniques.

A study investigated the delivery of IGF-1, a 7.65 kDa protein neurotrophic factor, to the rat brain and spinal cord via IN administration, focusing on the pathways and mechanisms involved in its transport from the nasal passages to the CNS. The delivery device used was a micropipette, administering a total of 50 μL [125I]IGF-1 solution. [125I]IGF-1 shows rapid entry into the CNS (within 30 min) [64]. The study identified two primary delivery pathways for IGF-1: the peripheral olfactory system, which connects the nasal passages with the olfactory bulbs and rostral brain regions, and the peripheral trigeminal system, which connects the nasal passages with the brainstem and spinal cord regions. Delivery mechanisms included extracellular transport through intercellular clefts and intracellular transport via adsorptive endocytosis into olfactory sensory neurons. Gamma imaging found that the uptake of [125I]IGF-1 was measured 30 min post-administration, with CNS levels peaking earlier than blood levels. Key uptake values in important CNS regions included the olfactory bulb (3.43 nM), anterior olfactory nucleus  $(1.65 \text{ nM})$ , frontal pole  $(1.44 \text{ nM})$ , motor cortex  $(1.33 \text{ m})$ nM), olfactory tubercle (1.08 nM), caudoputamen (0.32 nM), hippocampal formation (0.34 nM), diencephalon (0.46 nM), midbrain (0.39 nM), cerebellum (0.40 nM), pons (0.41 nM), medulla (0.87 nM), cervical spinal cord (2.39 nM), thoracic spinal cord (0.15 nM), and lumbosacral spinal cord (0.078 nM). The study demonstrated that IN administration resulted in significantly higher CNS concentrations compared to IV injection, indicating effective bypassing of the BBB. Retention of [125I]IGF-1 in the CNS was evident up to 24 h post-administration, with a gradual decline in concentrations over time. Elimination pathways involved significant targeting of the deep cervical lymph nodes, suggesting a role in lymphatic drainage from both nasal passages and CNS. No detectable radioactivity was found in cisternal CSF, indicating limited penetration into CSF from the bloodstream or nasal passages within the initial 30 min. Clearance patterns showed a gradient with CNS concentrations decreasing over time and distance from entry points, reflecting slow and prolonged absorption into the bloodstream and subsequent clearance. High-resolution phosphor imaging revealed a broad distribution of [125I]IGF-1 throughout the CNS, with the highest amounts near the olfactory and trigeminal regions. The biodistribution of [125I]IGF-1 in a high-specific activity

solution also matched well with known IGF-1 binding sites in the adult rat CNS, indicating that the IGF-1 reached the brain and spinal cord intact. Immunohistochemical analysis showed enhanced phosphotyrosine immunoreactivity in specific CNS regions following IN IGF-1 administration, indicating biological activity and receptor activation. Overall, the study systematically evaluated the localisation of [<sup>125</sup>I]IGF-1 in various CNS regions, supported by high-resolution imaging and immunohistochemical confirmation, demonstrating the effectiveness of non-invasive IN delivery for therapeutic proteins to the CNS.

In a subsequent study, Thorne *et al.* investigated how interferon-β1b (IFN-β1b), a protein used to treat the relapsing-remitting form of multiple sclerosis, affects the nervous system of monkeys following IN administration using plastic tubing, the volume administered was between 0.9 to 1.0 mL [65]. Five monkeys received an IN administration of  $[125]$ IFN-β1b into the upper nasal regions. The delivery pathway for [125I]IFN-β1b primarily involved the olfactory and trigeminal nerves. The mechanisms underpinning this transport involved the rapid extracellular movement of the tracer along nerve components in the nasal epithelium to the olfactory bulb and brainstem. From these entry points, the tracer dispersed to other CNS regions via pulsatile flow within the perivascular spaces of cerebral blood vessels. The uptake time for [125I]IFN-β1b was relatively rapid. Blood levels of the tracer began to rise shortly after administration, reaching peak concentrations approximately 30 to 45 min post-administration. High-resolution imaging and gamma counting of micro-dissected tissue samples revealed that significant tracer concentrations were present in the brain and spinal cord within 53 to 59 min of administration. This rapid uptake underscores the efficiency of the IN-delivery method for transporting therapeutic proteins to the brain. Upon reaching the CNS, [125I]IFN-β1b showed distinct patterns of distribution. The highest uptake was observed in the olfactory bulbs (16,600 pM), and trigeminal nerves (16,300 pM), indicating these regions as primary entry points. Significant levels of the tracer were also found in the basal ganglia, including the caudate, putamen, globus pallidus, and substantia nigra. These findings suggest a preferential localisation of the tracer in areas associated with dopamine regulation, which may have clinical implications for understanding the side effects of IFN-β1b therapies. Intermediate levels of uptake were noted in the hippocampal formation, thalamus, cerebellum, and brainstem nuclei of several cranial

nerves. Lower uptake levels were observed in CNS fibre pathways such as the cerebral white matter and corpus callosum. Quantification of the tracer that actually reached the brain revealed that approximately 0.0064% ID of [125I]IFN-β1b was present in the brain and spinal cord. Despite this seemingly low delivery efficiency, the concentrations achieved were sufficient to elicit biological effects, given the high potency of IFN-β1b. Elimination of the tracer from the brain primarily involved peripheral clearance mechanisms, with the kidneys and liver playing major roles. High radioactivity levels in the thyroid gland indicated significant peripheral deiodination and metabolism of the tracer, further corroborating the efficient systemic clearance observed. High-resolution histochemical findings from autoradiography provided detailed insights into the anatomical distribution of [125I]IFN-β1b. The imaging confirmed high signal intensities in the basal ganglia, substantia nigra, and cerebellum, aligning with gamma counting data. While direct histochemical analysis was not performed in this study, the imaging data strongly suggest a correlation with potential histochemical localisation, particularly in the basal ganglia. In conclusion, this study demonstrates the effective targeting and retention of [ $125$ ]]IFN-β1b in the CNS following IN administration. The significant uptake in the basal ganglia suggests potential clinical relevance for IFN-β1b therapies, particularly concerning their side effects.

A study examining the feasibility and efficacy of delivering vascular endothelial growth factor (VEGF) labelled with [125I] following IN administration reveals extensive distribution within the CNS using a micropipette [66]. Gamma counting and autoradiography were employed to quantify and visualise the distribution of [125I]-VEGF within the brain. The highest concentrations of the tracer were observed in the trigeminal nerve and optic nerve, followed by significant uptake in the olfactory bulbs, olfactory tubercle, striatum, medulla, frontal cortex, midbrain, pons, appendix cerebri, thalamus, hippocampus, and cerebellum. The autoradiographic analysis provided high-resolution images, confirming the widespread distribution of VEGF throughout the CNS. The uptake of [125I]-VEGF was rapid, with substantial delivery observed approximately 30 min after IN-administration. The retention time was not explicitly detailed, but significant levels of VEGF were present within 30 min timeframe. Quantitatively, the % ID/g was highest in the trigeminal nerve  $(1.57)$ , followed by the olfactory bulb (0.64) and optic nerve (0.96). These findings underscore the efficiency of the IN route in targeting specific CNS regions. The study

found no detectable radioactivity in the CSF following IN administration, suggesting that VEGF might be cleared via lymphatic pathways rather than through the CSF. This was further supported by the high concentrations of [125I]-VEGF found in the deep cervical lymph nodes, indicating ready access to nasal lymphatics and supporting a direct pathway from the nasal submucosa to brain interstitial fluid. Autoradiography corroborated high-resolution histochemical findings, which confirmed the presence and detailed distribution of [125I]-VEGF in various brain regions. This imaging provided a robust validation of the IN-delivery pathway, highlighting its potential to effectively deliver therapeutic agents to the CNS. The study's findings were further reinforced by comparisons with IV injection, which resulted in significantly lower CNS concentrations and higher peripheral tissue exposure, emphasising the superiority of the IN route.

Another study on wheat germ agglutinin conjugated NPs (WGA-NPs), labelled with [125I], and upon IN administration of 50 μL using a polyethylene tube attached to a micro syringe, demonstrates their fast transcellular absorption (5 min) to the olfactory bulb through the olfactory epithelium [67]. Post entry into the lamina propria, a portion of the labelled compound moved from the olfactory nerve bundles to the olfactory bulb. Here, the trigeminal nerves played a role in distributing the compound to caudal brain regions, with notable radioactivity concentrations found of  $\sim$ 128 cpm/g tissue/g dose (5 min post-administration. WGA-NPs entered the brain via extracellular transport along the trigeminal nerves, which connect to the brainstem at the pons and extend to the cervical spinal cord and medulla. The study found rapid NP uptake within 5 min, with peak levels observed at 30 min. By 2 h post-administration, the fluorescence intensity decreased significantly, indicating clearance. The primary regions of uptake included the olfactory bulb, specifically the glomerular layer and external plexiform layer, as well as regions along the trigeminal nerve, such as the cervical spinal cord, medulla, and pons. However, the CSF pathway seemed to have a minimal role in transporting the compound to the CNS following IN administration, as very low radioactivity levels were detected in the CSF, comparable to those in the diencephalon and hippocampus, which were far below those in the olfactory bulb and trigeminal nerve. The uptake in the olfactory bulb reached a peak at 30 min post-administration  $(\sim 11 \text{ cm}/g \text{ tissue}/g)$ dose) before decreasing, indicating rapid clearance from this region. Strong green fluorescence signals were observed in the olfactory epithelium and

olfactory bulb, confirming successful uptake and transport. Immunohistochemistry staining with anti-nerve specific enolase and anti-ZO-1 antibody verified that the uptake was primarily transcellular through the olfactory epithelium. Comparative analysis of imaging agents used in the study included coumarin-6 (Cou-6) for visualising NPs in the nasal cavity and olfactory bulb by fluorescence microscopy, 125I-labelled WGA-NP for measuring radioactivity levels to track distribution, and DiR (1,1'-dioctadecyl-3,3,3',3'-tetramethylindotricarbocyan ine iodide) loaded NPs for *ex vivo* imaging to visualise brain transfer. The study found that WGA-NP showed enhanced uptake in the olfactory epithelium and higher distribution in the olfactory bulb and associated brain regions compared to unmodified NPs, which followed similar pathways but were less efficient in brain delivery. In conclusion, IN administration of WGA-NP effectively delivers NPs to the brain via olfactory and trigeminal pathways. The study provided detailed insights into the mechanisms of uptake, regional distribution, and retention of NPs in the brain, supported by high-resolution imaging and histochemical analyses. These findings contribute valuable information for designing effective nanocarriers for INDD.

Haloperidol, a dopamine-blocking agent used in brain imaging studies, demonstrates varied brain uptake [68]. The focus is on the effectiveness of different delivery methods, including IV, IN-solution, and IN ME. The objective was to determine the optimal method for delivering 125I-haloperidol to the brain, thereby facilitating enhanced brain imaging. The delivery of 125I-haloperidol was executed using a small syringe for IN administration of 20 µL. The study found that the uptake time and brain regions affected varied significantly across the three delivery methods. For the IV route, brain uptake was initially observed at 5 min (0.5%), peaking at 15 min (1.4%), before declining. The IN-solution method showed a brain uptake of 3.7% at 5 min, peaking at 4.3% at 15 min, and then gradually decreasing. The IN ME method demonstrated the highest efficacy, with brain uptake observed at 3.8% at 5 min, peaking at 5.7% at 15 min, and maintaining substantial levels throughout the observed period. The primary regions of uptake included the olfactory bulbs, frontal cortex, and other brain areas connected to the olfactory and trigeminal pathways. Retention time analysis revealed that the IV route had the least retention, with brain uptake significantly decreasing over 120 min. The IN-solution method showed better retention, with notable uptake still present at 120 min. However, the IN ME method exhibited the highest retention, maintaining

significant brain uptake across all time intervals, indicating prolonged retention and efficient targeting. The tracer's elimination from the brain followed a systemic pathway, with the tracer gradually washing out into the bloodstream over time. The data showed a decrease in brain uptake, indicating the washout process, which was more prolonged in the IN ME method, providing extended imaging windows. Collectively, these studies highlight the potential of IN administration [125I] imaging as a method for site-specific targeting within the CNS, offering insights into the dynamics of compound distribution and clearance from various brain regions.

In a 2022 study by Wei-Lin Lo *et al*., the effectiveness of [123I]-radiolabelled anti-A $\beta$  peptides for IN administration targeting the brain as a potential treatment for Alzheimer's disease was investigated [69]. The study involved administering 5 µL of R8-YA $\beta$ (25–35)-PEI labelled with [123I] intranasally by dropper and then monitoring its distribution across the body with SPECT/CT imaging (see Figure 4). Peak uptake in the brain occurred at 0.5 h post-treatment, with a concentration of  $0.73 \text{ %ID/g.}$ The uptake decreased over time, showing values of 0.45% ID/g at 6 h, 0.28% ID/g at 12 h, 0.17% ID/g at 24 h, and  $0.1\%$  ID/g at 48 h. The highest uptake was observed in the olfactory bulb, indicating that the peptide initially enters the brain through this region before diffusing to other parts of the brain. Furthermore, the study delineated the peptide's neuro-targeting efficiency for nose-to-brain up to 1.78% ID/g tissue, with a calculated biological half-life of the peptide in the brain of approximately 10.2 h. Most of the  $[123]$  R8-YAβ(25-35)-PEI solution was swallowed and passed through the oesophagus into the gastrointestinal tract, as observed by the gradual decrease in radioactivity in the brain and the increase in radioactivity in the bladder over time. Overall, the study demonstrates that IN administration of the [123I]R8-YAβ(25–35)-PEI peptide is an effective method for delivering peptides to the brain, with a notable brain uptake rate and retention time. This method holds promise for treating neurodegenerative diseases by providing a non-invasive route to bypass the BBB.

[131I] was used in a study by Motaleb *et al.* focusing on developing [131I]-trazodone hydrochloride as a new brain imaging radiopharmaceutical and establishing its efficacy [70]. The study investigated the biodistribution and brain uptake of [131I]-trazodone using different administration routes, specifically IV, IN solution, and IN ME. The study showed that the brain uptake of  $[131]$ -trazodone was significantly higher with the IN ME compared to the IV solution and IN solution at all sampled time points. Notably, the brain uptake for IN ME peaked at  $\sim$ 7% ID/g, while IN solution and IV solution had lower uptakes. The higher and faster uptake with IN ME suggests efficient delivery through the olfactory region. The highest brain uptake with the IN ME formulation, which was higher than that achieved with standard radiopharmaceuticals used for brain imaging such as [99mTc]ECD and [99mTc]HMPAO and higher than IN and IV solutions of [131] trazodone. The elimination from the brain followed a decline over time but remained higher for IN ME throughout the observation period.



**Figure 4.** SPECT/CT fusion images of mice following IN administration of [123I]R8-YAβ(25–35)-PEI. A: displays the maximum intensity projection view. B: shows the sagittal view, marking 'B' for brain, 'Bl' for bladder, 'N' for nose, 'T' for thyroid and 'S' for stomach. Adapted with permission from [69], copyright 2022 Elsevier.

Fayez and co-workers developed a novel radiopharmaceutical, [131I]-rolapitant, formulated into spanlastic nanovesicles for brain imaging [71]. The delivery device used was an IN-administration apparatus designed to deliver 15 μL of the solution through the nostrils. [131I]-rolapitant, acts as a selective high-affinity neurokinin-1 receptor antagonist, spanlastic nanovesicles significantly enhanced brain uptake and bioavailability compared to IV injection solutions. The spanlastic nanovesicles enhanced the delivery process by facilitating the penetration of the nasal epithelial cells and promoting uptake into brain

regions. The surfactants, particularly Tween 80, used in the spanlastic nanovesicles formulation, acted as absorption enhancers, aiding in the transport across the BBB. These surfactants also reduced the clearance of NPs by the reticuloendothelial system and inhibited efflux systems like P-glycoprotein, further enhancing brain uptake. The study found that the maximum brain uptake was achieved at 2 h post-IN administration, with a notable value of  $\sim$ 7% ID/g. This high level of uptake was sustained for an extended period, with  $\sim 6\%$  ID/g still present at 24 h post-administration. These findings highlight the efficiency of the IN-delivery system in ensuring prolonged drug retention in the brain. The study revealed significant brain uptake in normal mice, particularly when compared to IV injection. The distinctive distribution patterns showed that the IN administration of [131I]-rolapitant in spanlastic nanovesicles resulted in a 3-fold greater brain uptake than the IV solution. Furthermore, the brain/blood ratio, which reached a maximum of  $\sim$ 2 at 24 h post-administration, indicated a preferential accumulation of the radiolabelled drug in the brain, underscoring the potential of this delivery method for targeted brain imaging and therapy. The study also examined the elimination of tracer from the brain, revealing minimal clearance up to 24 h post-administration. This prolonged retention time is advantageous for imaging and therapeutic applications, as it allows for extended observation and effect of the radiolabelled drug in the brain. In conclusion, this approach highlights the potential of [<sup>131</sup>]]rolapitant spanlastic nanovesicles, as an efficient method for brain imaging and diagnosis of neurological disorders, particularly Alzheimer's disease and epilepsy.

Later work by Fayez *et al.* aimed to develop a selective SPECT radiotracer targeting the endothelin 1 receptor A in the brain by radiolabelling ferulic acid, an endothelin 1 receptor A antagonist, with [131I] [72]. The [131I]-ferulic acid was formulated into nanosized polymeric micelles for IN administration to facilitate nose-to-brain delivery. The biodistribution of IN [131I]-ferulic acid polymeric micelles was compared to IN and IV [131I]-ferulic acid solutions to evaluate their efficiency in brain uptake. The mice biodistribution results demonstrated significantly higher brain uptake of [131I]-ferulic acid polymeric micelles at all time points compared to IV and IN [131I]-ferulic acid solutions. This enhanced brain uptake is attributed to the lipophilicity and nanoscale size of the polymeric micelles, which facilitate the direct transport of [<sup>131</sup>I]-ferulic acid to the brain via olfactory and trigeminal nerve pathways. In contrast, the IV

[131I]-ferulic acid solution showed low brain uptake and high blood levels due to its poor lipophilicity and inability to cross the BBB. Similarly, the limited lipophilicity of the IN [131I]-ferulic acid solution resulted in poor nasal mucosal absorption, leading to low blood and brain uptake and high stomach uptake due to transfer through the nasopharynx to the stomach. The brain/blood ratio for IN 131I-ferulic acid polymeric micelles was consistently higher at all time points, especially at 30 min post-administration, indicating effective brain targeting. Specifically, the brain/blood ratio for IN [131I]-ferulic acid polymeric micelles was 1.09% ID/g at 30 min, compared to 0.18% ID/g for IN and 0.14% ID/g for IV [131]-ferulic acid solutions. These findings suggest that IN [131I]-ferulic acid polymeric micelles could serve as a promising SPECT imaging tracer for the endothelin 1 receptor A in the brain. Furthermore, this study represents an initial step towards developing an endothelin 1 receptor A imaging agent, with further biological studies needed to validate these findings. Future research should explore the biodistribution of IN [131I]-ferulic acid polymeric micelles *in vivo* with induced head trauma to enhance endothelin 1 receptor A expression.

#### 2.1.4. Remarks on SPECT and gamma scintigraphy

SPECT and gamma scintigraphy have become pivotal in monitoring the efficiency and dynamics of INDD, especially with the high sensitivity of SPECT and the ability of gamma scintigraphy to precisely track drug transit and brain distribution. These modalities have significantly contributed to understanding the pharmacokinetics and biodistribution of various radiolabelled compounds following IN administration. A range of imaging agents for INDD have been investigated using SPECT, each with unique properties and half-lives catering to specific study requirements. 99mTc is the predominant radionuclide used for SPECT, valued for its widespread availability, favourable photon energy for imaging, and short half-life. Vyas *et al.* (2005) assessed the nose-to-brain delivery of [99mTc]-zolmitriptan in rats, exploring its efficacy in treating acute migraines. The study revealed that IN administration facilitated rapid brain uptake, surpassing other formulations and routes. The tracer's retention time in the brain was monitored for up to 8 h, with significant drug accumulation observed. Jogani *et al.* investigated [99mTc]-labelled tacrine for Alzheimer's treatment, showing higher brain uptake and faster peak times compared to IV injection, suggesting enhanced bioavailability and reduced hepatotoxicity. Mustafa *et al.* (2012) assessed the IN administration of

[99mTc]-labelled ropinirole in rabbits, demonstrating superior brain localisation and rapid uptake through the olfactory pathway. Another study on chitosan NPs as a delivery vehicle for [99mTc]-bromocriptine highlighted increased bioavailability in brain tissue and improved nose-to-brain delivery via intranasal administration. IN administration of [99mTc]-carbamazepine showed higher brain uptake and prolonged retention compared to IV injection, demonstrating the benefits of mucoadhesive ME formulations. [99mTc]-diazepam-loaded PLGA NPs demonstrated significant brain uptake and sustained presence, supporting the potential of this route for brain-targeted therapies. A study by Mandlik *et al.* on zolmitriptan-loaded nanostructured polymeric carriers showed enhanced brain uptake and retention, highlighting the effectiveness of nanocarriers for nose-to-brain transport. The elimination of [99mTc]-labelled agents from the brain often involves systemic circulation and metabolic processes, with studies indicating a gradual washout of tracers from the brain over time. The clearance of these agents is critical in understanding the duration of their efficacy and potential side effects. Similar results were observed with other drugs like clonazepam, risperidone, and carbamazepine, indicating [99mTc] ability to monitor nose-to-brain delivery efficiently.

[<sup>86</sup>Rb], [<sup>201</sup>TI], and [<sup>54</sup>Mn] have also been studied for their potential in brain imaging. A study on the olfactory transport route of [86Rb] and [201Tl] ions following IN administration demonstrated their significant accumulation in the olfactory bulb and subsequent spread to other brain regions. This indicated potential use in molecular imaging for conditions like anosmia and neurodegenerative diseases. Research on the transport of [201Tl] and [<sup>54</sup>Mn] ions in mice highlighted their reliance on olfactory nerve fibres for transport to the olfactory bulb, suggesting diagnostic applications for assessing olfactory nerve damage. Shiga *et al.* investigated the transport of [201Tl] to the brain using SPECT/MRI hybrid imaging, demonstrating peak uptake in the olfactory bulb and supporting its use in assessing olfactory disorders and potential infectious routes. Studies on [201Tl] migration in patients with olfactory impairments showed correlations between tracer migration, odour recognition thresholds, and olfactory bulb volume, providing insights into prognosis and recovery in idiopathic olfactory dysfunction. Tracers like [86Rb] and [201Tl] are gradually eliminated from the brain, with autoradiography revealing their transport behaviour and clearance patterns through olfactory pathways. These studies help delineate the pathways and mechanisms involved in the transport of these ions from the nasal cavity to the brain, providing a deeper understanding of their biodistribution and potential clinical applications.

Iodine-labelled compounds have shown promising results in non-invasive CNS imaging and targeted IN administration. Studies on [125I]-labelled Ct and its B subunit demonstrated distinct accumulation patterns in olfactory nerves/epithelium and olfactory bulbs. Research on [125I]-bFGF targeted Aβ plaques in a transgenic mouse model, highlighting the potential for labelling and imaging Aβ deposits. A study on [125I]-labelled IGF-1 demonstrated rapid entry into the CNS and significant concentrations in various brain regions, supporting its therapeutic potential for CNS disorders. [125I]-labelled VEGF showed extensive distribution within the CNS following IN administration, confirmed by gamma counting and autoradiography. [123I]-radiolabelled anti-Aβ peptides demonstrated effective brain targeting and retention, supporting their use for treating Alzheimer's disease. Iodine radioisotopes are cleared through lymphatic pathways and systemic circulation, with studies showing varied retention times and clearance patterns depending on the tracer and formulation. [131I]-labelled agents, such as [131I]-trazodone and [131I]-rolapitant, have demonstrated enhanced brain uptake and bioavailability with IN administration, suggesting effective targeting for brain imaging.

Gamma-emitting radiotracers offer unique insights into the nose-to-brain delivery mechanism, with variations in pharmacokinetics, uptake efficiency, and biodistribution. Together, the studies highlight the potential of IN administration as an alternative route for drug delivery to the brain, especially for therapeutic interventions in neurology and psychiatry. The ability of SPECT and gamma scintigraphy to quantify drug distribution precisely and to differentiate between drugs, metabolites, and other biological molecules has been crucial in these advancements. However, the field faces challenges such as optimising the balance between drug formulation, imaging resolution, and safety considerations, particularly concerning the toxicity of long-lived radionuclides. Continued research in this domain is essential to further harness what the imaging modalities can offer clinically and to develop more effective treatments for CNS disorders.

#### **2.2. Positron emission tomography**

PET stands at the forefront of molecular imaging techniques, offering unparalleled insights into metabolic activities at a cellular or molecular scale.

This non-invasive imaging modality harnesses the power of radioisotopes to trace physiological processes in real-time tracking of radiolabelled drugs from nose-to-brain, providing invaluable data in both clinical and research settings. By using specific positron-emitting radionuclides attached to drugs, researchers can observe the direct transport pathways from the nasal cavity to the brain. The high sensitivity of PET allows for detailed visualisation of drug uptake and metabolism in brain tissues. PET is frequently combined with CT or MRI to provide a more comprehensive diagnostic tool that merges the strengths of each imaging modality. This combination allows for the simultaneous visualisation of both anatomical structures and biological functions within the body [73].

PET radioligands, ranging from small molecules to large biomolecules, serve as vehicles, transporting the radioactive label to the biological target sites. The radioactive labels are chosen based on their physical properties, primarily their half-lives (ranging from minutes to hours and to days), which directly influence the temporal imaging window and the resolution of the acquired images, as well as the required dosage [74].

The exploration of IN administration for brain imaging with PET introduces a strategic advantage by potentially circumventing the BBB. This pathway is particularly beneficial for the deployment of tracers that are substantially metabolised during first-pass hepatic metabolism, as well as those that might affect cerebral hemodynamic or vascular characteristics. Additionally, it allows for the administration of large molecular weight peptides, which are generally impeded by the BBB.

In this section, we survey the spectrum of positron emitting radioisotopes reported in the literature, each serving specific roles in PET imaging for IN administration. The commonly used [18F] (fluorine-18) stands out with its almost ideal imaging properties, providing a balanced half-life of 109.8 min, and offering high-resolution images with a relatively short positron range (i.e., distance travelled by a positron before annihilation with an electron). This makes it particularly suitable for high-precision studies over a span of several hours [75]. On the shorter end of the half-life spectrum, [<sup>11</sup>C] (carbon-11), with a half-life of 20.4 min, necessitates rapid synthesis and imaging procedures but offers the advantage of studying rapid metabolic processes, as well as minimising radiation exposure. [11C] is often incorporated into biologically relevant molecules, allowing real-time tracking of molecular activity [76]. Radiometals, such as [64Cu] (copper-64) with a 12.8 h

half-life, introduce versatility to PET imaging, finding use in both diagnostic and therapeutic applications when chelated to various biomolecules [77]. [<sup>68</sup>Ga] (Gallium-68)-based peptides are gaining recognition as a novel class of radiopharmaceuticals due to their rapid blood clearance and quick target localisation. The short physical half-life of [ $^{68}Ga$ ], which is ~68 min, allows for better dosimetry and the possibility of repeat imaging, making these agents highly suitable for clinical applications [78]. [89Zr] (zirconium-89), a radiometal with a 78.4 h half-life, facilitates long-term tracking of biological events and has been shown to enhance diagnostic accuracy [79]. Manganese isotopes ([52Mn], [51Mn], and [54Mn]) present unique imaging opportunities, ranging from short-term studies with [<sup>51</sup>Mn] (46.2 min) to prolonged tracking with [<sup>52</sup>Mn] (5.6 days) and [54Mn] (312.2 days) [80]. Lastly, provides unique functionalities for specialised applications with a remarkably long half-life of approximately 12.3 years, [3H] emits low-energy beta particles and limited *in vivo* experiments [81].

The selection of isotopes is critical due to the variable positron ranges associated with different isotopes, which can significantly affect the spatial resolution of the images. Longer positron ranges can lead to image blurring, effectively reducing the resolution and degrading the quality of the diagnostic information. Moreover, *in vivo* studies using isotopes with long half-lives, such as <sup>54</sup>Mn, require careful planning to manage the risks associated with prolonged radioactivity.

# 2.2.1. Fluorine-18 ([18F])

[18F] has been used to monitor the *in vivo* absorption, distribution, metabolism, and excretion of drugs labelled with this isotope. Radiotracers include [18F]-fluorodeoxyglucose ([18F]FDG), [18F]-2-deoxy-2 fluorosorbitol ([18F]-FDS), [18F]-fallypride, [18F] fluoride and  $[18F]$ -fluorothymidine  $([18F]ELT)$ . Standard uptake value (SUV) maps are often generated, or tracer time-activity curves (TACs) can be collected from dynamic PET scans to quantitatively assess the administered dose. From these measurements the spatial distribution or local concentration of the tracer can be derived for an image pixel (provided an appropriate experiment is utilised) or for any organ or anatomical structure situated within the imaging field-of-view of the imaging study.

[18F]FDG is a widely used PET radiopharmaceutical enabling mapping of glucose metabolism, including the CNS due to BBB permeability [82]. Using this tracer, studies have explored INDD and focused on deposition and

clearance. Shingaki *et al.* study employed [18F]FDG PET imaging to explore the pharmacokinetics of INDD absorption in rats, focusing on hydroxypropyl cellulose viscosity (0% to 3%  $w/v$ ) and administered volumes (5 to 25 µL) [83]. The nasopalatine duct was closed to prevent leakage into the oral cavity, mimicking the human nasal anatomy where the duct is degenerated. Key findings include the clearance of approximately 50% of [18F]FDG from the nasal cavity within 30 min, primarily through swallowing, which then allowed for its systemic circulation and brain deposition. Higher hydroxypropyl cellulose viscosity enhanced tracer uptake without altering nasal membrane permeability. Quantitation of drug reaching the brain was measured through the absorption rate constant, which varied between 0.0055 and 0.0177 per minute depending on the volume and viscosity of the dosing solution. The fraction absorbed at 30 min significantly increased with higher viscosity solutions, ranging from ~5% for saline, up to ~28% for 3% hydroxypropyl cellulose solutions. The study modelled pharmacokinetic parameters, showing that increased viscosity improved the absorption rate constant and reduced the mucociliary clearance rate constant, independent of dosing volume. PET imaging revealed time-dependent and region-specific changes in [18F]FDG permeability within the nasal cavity, correlating with the unique anatomical and physiological characteristics of its distinct regions: the nasal vestibule, atrium, respiratory area, olfactory region, and nasopharynx. This variability is influenced by blood flow, mucus secretion, ciliary beating, and the presence of various cell types, which affect drug absorption and permeability [84, 85]. Permeability was notably higher in the anterior regions, underscoring the importance of regional differences in nasal mucosa for effective drug delivery. The study suggests that *in vivo* PET studies can offer more accurate pharmacokinetic parameters than *in vitro* studies, highlighting PET imaging's potential to enhance nasal dosage formulation development for systemic or CNS delivery, thereby improving INDD success rates by circumventing the first-pass effect. Additionally, highlights the capability of PET imaging in enhancing the understanding and development of nasal formulations for human use, highlighting the technique's ability to quantitatively analyse drug absorption and facilitate the prediction of pharmacokinetic parameters, thus providing a robust tool for future research and development [83].

Singh *et al.* performed a pharmacokinetics study for BBB penetrant tracers using [18F]FDG and [18F]fallypride, comparing IN and IV injection in rats

[86]. The delivery device used for IN administration was an insulin syringe. Each administration involved a total volume of 40 µL, with 20 µL per nostril. A significant difference between IV and IN administration in terms of brain uptake was absent, except for the olfactory bulb having a significantly higher uptake with IN administration (see Figure 5). For [18F]FDG, imaging was initiated approximately 33 min after IN administration, which was identified as the optimal time point for image resolution. For [18F]fallypride, imaging commenced around 10 min post-administration due to the necessary preparation and positioning time. The team reported that the absolute uptake of radiotracers in the study indicates that, for [18F]FDG, the concentration in the plasma was significantly lower when administered intranasally compared to IV injection. In contrast, [18F]fallypride, showed no significant difference in plasma concentration between the IN administration and IV injection routes, suggesting rapid absorption into the peripheral circulation for [18F]fallypride but not for [18F]FDG when IN administration. The retention time varied between the two tracers. For [18F]FDG, a significant proportion remained in the nasal cavity throughout the 45 min scan duration, with only about 30% reduction in radioactivity. Conversely, [18F]fallypride demonstrated an 80% reduction over a 60 min scan. This suggests that the overall brain uptake (excluding the olfactory bulb) from IN administration of [18F]fallypride is almost entirely supplied from peripheral circulation rather than directly from the nasal cavity. This observation is important, especially when weighed against the potential risks associated with IN administration, such as the high levels of cumulative skin radiation exposure and challenges related to non-specific signals. When tracers such as [18F]FDG and [18F]fallypride stay in the nasal cavity for an hour, potentially making this method risky due to the low amount of tracer reaching the brain and affecting image quality. Furthermore, this difference likely stems from [18F]FDG's tendency to become metabolically trapped within nasal cavity cells, leading to lower blood concentrations when IN-administered [87]. On the other hand, [18F]fallypride does not get trapped in cells, allowing for quick absorption into the bloodstream and similar uptake rates between IN administration and IV injection. These findings indicate that the time a molecule spends within cells could significantly influence its nose-to-brain delivery. The authors suggested that further research is needed to explore how this mechanism affects the IN administration of other molecules, particularly those typically unable to

cross the BBB [86].

[18F]FLT is known to have limited permeability/transport through the BBB, so brain delivery in rodents via nose-to-brain was studied [88]. The study compared [18F]FDG and [18F]FLT for the purpose of identifying direct nose-to-brain pathways, and distribution after IN administration of 20-25 µL using a Hamilton syringe. A significant portion of the [18F]FLT dose was cleared through the oropharynx within a few minutes, and continued at a slower rate for 20 min [88]. The residence time of [18F]FLT in the nasal cavity ranged from 15-30 min; however, a residual dose could still be measured in the nasal cavity 60 min after IN administration. [18F]FLT showed notably higher brain concentrations, particularly in the olfactory bulb, following IN administration compared to IV injection. This enhancement is immediate but diminishes within 30-45 min due to rapid clearance. [18F]FLT concentration increased in both the olfactory bulb and the brain over time until plateauing 25-30 min post-administration. At that time, [18F]FLT concentrations in the brain equalled almost one-third of that in the olfactory bulb. Conversely, [18F]FDG remained detectable for extended periods due to phosphorylation in the brain and nasal tissues, which hindered its clearance. The early clearance trend was comparable for both [18F]FDG and [18F]FLT, with half-lives of 2.53 and 3.36 min, respectively. However, during the latter, slower clearance phase, [18F]FLT cleared far more rapidly than [18F]FDG with half-lives 32.1 versus 85.2 min, respectively). Supplementary *ex vivo* tissue analyses confirmed the imaging results, supporting the observed distribution patterns of the tracers. Pharmacoimaging methods founded on mathematical models serve as a crucial tool in studying these processes; this can further be explored for IN administration studies through advancements in measurement precision and wider availability of suitable PET-labelled tracers, and enhancements in the development of delivery systems that promote targeting to distinct regions in the nasal cavity and prolong residence time.

[18F]FLT, a substrate of both equilibrative (ENT1,2) and concentrative (CNT1–3) nucleoside transporters, has also been used to investigate the role of nucleoside transporters in the nasal mucosa in INDD to the brain [89]. Dynamic PET/CT scans were used to observe the distribution of [18F]FLT in rats up to 40 min, using both IN administration and IV injection. The effects of NBMPR (nitrobenzylmercaptopurine riboside), an ENT1 selective inhibitor, on [18F]FLT distribution was also explored. Results indicated that [18F]FLT concentrations were again



**Figure 5.** Presents lateral views of the brain following IN administration of (A) [18F]FDG and (B) [18F]fallypride. In both images, arrows point to the olfactory region. Additionally, the basal ganglia are indicated in the image (B) of [18F]fallypride. Adapted with permission from [86], copyright 2018 Elsevier.

higher in the olfactory bulb after IN administration compared to the rest of the brain. [18F]FLT having limited permeability across the BBB, resulted in minimal brain exposure post-IV injection [90]. Retention studies showed rapid FLT clearance from the olfactory bulb within the first 20 min, followed by a slower clearance phase over the next 20 min. This pattern shows while initial uptake was fast, the sustained presence of FLT in the brain allowed for prolonged observation. Administering NBMPR intraperitoneally led to higher FLT exposure in both the olfactory bulb and the rest of the brain following IN administration and IV injection, corroborated by the *ex vivo* data. This indicates that NBMPR administration slowed [18F]FLT clearance from the brain. In contrast, co-administration of [18F]FLT and NBMPR decreased olfactory bulb exposure but increased brain exposure, suggesting ENT1 plays a role in the nose-to-brain transport pathways [89]. In another study using the FLT with low beta energy particle, Carlisle *et al.* evaluated the effectiveness of delivering [3H]FLT to brain tissue via IN administration and IV injection routes in a rat model [91]. They administered [3H]FLT to adult rats for both cases, then measured the concentrations of [3H]FLT in 16 different brain regions, as well as in the blood and non-target organs, at different timepoints post-administration. The concentrations of [3H]FLT in the olfactory bulb were consistently higher in the IN administration group than in the IV injection group. However, significant differences between the two groups were not found for all other brain regions. The brain/blood ratio values followed a similar pattern. Further analysis to reveal the efficiency of IN administration CNS radioisotope targeting indicated a more effective penetration into the olfactory bulb, spinal cord, and hippocampus with IN administration. Notably, non-target tissues such as the heart, lungs, adipose tissue, and skeletal muscle

showed significantly higher concentrations of the radioisotope in both the 5- and 60-min trials for both IN administration and IV injection, compared to all brain regions. Additionally, high concentrations of the tracer in the heart and lungs highlight the importance of active/preferential targeting to the CNS. However, the use of [3H] presents significant limitations due to its low beta energy, impacting detection sensitivity, and its long half-life, raising safety concerns for *in vivo* studies [92].

An insightful study utilising PET/MRI technique investigated the role of the glymphatic system, crucial for brain waste clearance, using [18F]fluoride and [18F]FDS [93]. The study revealed distinct clearance patterns for IN administration and IV injection. While [18F]fluoride showed high skull accumulation with both IN administration and IV injection, which was notably absent in [18F]FDS, effectively reducing spillover into brain tissue and enhancing the sensitivity for whole-brain analysis. Notably, [18F]fluoride after IN administration exhibited a 58% reduction in signal, contrasting with a 37% increase following IV injection. [18F]FDS demonstrated clear brain uptake and clearance with IN administration, showing a remarkable 110% change in PET signal after 105 min, whereas IV injection only showed a decline from baseline. These findings suggest that IN administration might be more effective for studies focusing on glymphatic efficiency, potentially guiding future research towards optimising tracer selection and administration routes to better understand and mitigate cognitive decline in aging and dementia.

An [18F]-labelled tracer, [18F]AlF-NODA-dLVT, has been developed to investigate the direct nose-to-brain transport of oxytocin, a hormone vital for childbirth and breastfeeding [94]. This novel PET imaging agent, which incorporates a NODA chelator with the radioisotope [<sup>18</sup>F]aluminium and the oxytocin analogue dLVT, is designed to enhance both stability and selectivity. The tracer is specifically tailored for studying the direct nose-to-brain delivery of oxytocin-based therapies, enabling non-invasive monitoring of their uptake into the brain. Researchers observed that IN administration slightly increased tracer uptake in the brain parenchyma compared to IV injection, with uptake from IN administration at 0.11% ID/g versus  $0.05\%$  ID/g from IV injection. Although there was enhanced tracer uptake in the olfactory bulb following IN administration, indicative of potential nose-to-brain transport, the tracer did not significantly reach deeper brain regions within a 90 min PET imaging timeframe. Furthermore, the pattern of distribution post-IN administration did not prove to be substantially more effective than IV injection for delivering oxytocin-based tracers into the brain. The presence of radioactivity in the cerebellum suggests potential peptide degradation post-absorption, underscoring the need for improvements in the stability and selectivity of oxytocin agonists to boost clinical efficacy. While the findings indicate that IN administration might influence behavioural and neural responses, they also highlight the necessity for further enhancements in tracer stability and selectivity to improve clinical outcomes. The results, while promising, provide limited insight into the efficacy of [18F]-based radiotracers in exclusively mapping the nose-to-brain delivery pathway of oxytocin.

Restoring insulin, a peptide hormone produced by pancreatic β cells, levels in the CNS has gathered interest as a potential treatment for Alzheimer's disease. Patients with Alzheimer's disease exhibit elevated plasma insulin levels and reduced CSF insulin levels, with this pattern becoming more pronounced as their clinical impairment worsens [95]. Nose-to-brain delivery of insulin has attracted attention as a non-invasive pathway to enhance CSF insulin levels without affecting peripheral insulin levels. Studies in rodents indicate that molecules delivered via nose-to-brain travel along the olfactory and trigeminal pathways to reach the CNS. A recent study by Smith *et al.* employed PET to monitor the uptake of [18F]-labelled insulin ([18F]-insulin) from nose-to-brain uptake in non-human primates [96]. Three nose-to-brain delivery methods for [18F]-insulin were evaluated: aerosol delivery via tubing in rhesus macaques, aerosol delivery via a preplaced catheter in rhesus macaques, and solution delivery via a preplaced catheter in cynomolgus macaques. Post-administration, dynamic PET imaging for 120 min was performed to quantify the delivery efficiency of nose-to-brain. The area under the time-activity

curve (TAC) was calculated for 46 brain regions of the cynomolgus macaque to assess regional [18F]-insulin levels. The  $C_{\text{max}}$  of [<sup>18</sup>F]-insulin in the whole brain was observed in the first PET frame for two out of three experiments. This indicates that intranasally delivered insulin reaches the brain within 13 min of administration. The imaging revealed that liquid instillation of [18F]-insulin via catheter was more effective than aerosol methods for both delivery to the subject  $(\sim 40\%$  ID vs.  $\sim 10\%$  for aerosol via tubing, 0.17% for aerosol via catheter) and subsequent brain delivery (0.34% ID vs. 0.0002% for aerosol via tubing, 0.05% for aerosol via catheter). [18F]-insulin was quickly transferred across the cribriform plate to the limbic and frontotemporal regions, which are key areas for emotional and memory processing. Limbic and frontotemporal cortical regions are crucial therapeutic targets for Alzheimer's disease patients. An earlier study classified Alzheimer's disease patients mainly into "temporoparietal" or "limbic-predominant" hypometabolic patterns [97]. The half-life of [18F]-insulin was longer in olfactory nerve projection sites with high insulin receptor density compared to the entire brain. The accumulation and clearance rates of [18F]-insulin varied by region. A one-phase decay model showed that the half-lives for the nasal cavity, cribriform plate, and whole brain were approximately 30-, 20-, and 31 min, respectively. The model fits well for the nasal cavity and whole brain but less accurately for the cribriform plate. Differences in [18F]-insulin kinetics were visualised using normalised SUV TACs for various brain regions. Some regions, like the frontal-orbital gyrus and anterior cingulate cortex, had sustained activity but faster clearance rates. In contrast, regions like the amygdala and hippocampus cleared the tracer more slowly, with longer half-lives. The nucleus accumbens showed a unique oscillating pattern. In conclusion, the catheter-based liquid delivery method, combined with PET imaging, effectively tracked the nose-to-brain [18F]-insulin and is believed to be broadly applicable for evaluating other therapeutic agents [96]. This approach can rapidly be applied in human studies to advance the clinical evaluation of nose-to-brain insulin as a treatment for Alzheimer's disease. More recently, a study aimed to develop a non-invasive delivery system for overcoming the BBB using the nose-to-brain route [98]. PEGylated-PLGA NPs coated with chitosan were designed to facilitate nasal absorption and efficient brain transfer of insulin. Characterisation and *in vivo* studies, including biodistribution and PET imaging, indicated higher fluorescence in the brain, suggesting efficient NP

delivery. These findings highlight the potential of PEGylated and chitosan-coated NPs for targeted insulin delivery to the brain, providing a promising platform for treating Alzheimer's disease by restoring memory signalling. Together, these findings from NP-based and [18F]-insulin studies propose innovative nasal delivery systems for insulin, providing a solid foundation for further clinical evaluation and potential treatment of Alzheimer's disease.

# 2.2.2. Carbon-11 ([11C])

Zolmitriptan is a selective serotonin receptor agonist that readily crosses the BBB and is known for its high efficacy in the acute management of migraines [99]. Yates *et al.* utilised an IN-spray device to administer [11C]-labelled zolmitriptan in a 100 μL solution on healthy volunteers to assess the distribution and pharmacokinetics of the drug using PET imaging [100]. Immediately after administration, close to 100% ID was detected in the nasopharynx. This concentration declined over time, with 50% remaining at 20 min and 35% at 80 min post-administration. These results indicated rapid absorption and distribution of the drug from the nasal cavity. Radioactivity first appeared in the upper abdomen during the first 20 to 40 min post-dose. Minimal radioactivity was detected in the lungs, with only 0.2% at 20 min and 0.3% at 80 min post-dose. The brain showed low but detectable levels of radioactivity, with SUV ranging from 0.006 to 0.070, suggesting some central penetration. The mean brain concentration of radioactivity for all volunteers was SUV 0.020, equivalent to 0.54 ng/g. There was a strong correlation between brain SUV and blood SUV, with brain values being approximately  $1/5<sup>th</sup>$  of blood values, indicating that a fraction of the drug reached the brain. The brain retained low but detectable levels of radioactivity throughout the 1.5 h scanning period. While the study confirms central penetration of zolmitriptan with detectable levels of radioactivity in the brain, it recommends caution as some of the detected radioactivity might be attributable to metabolites. A high degree of correlation (correlation coefficient = 0.95) was noted between cerebral and blood uptakes, which was inconsistent with previous findings on zolmitriptan crossing the BBB [101, 102]; where the cerebral values were approximately 0.2-fold of the corresponding blood values. Despite minimal levels in cerebral tissues, quantifiable radioactivity was consistently detected, indicating the central albeit residual penetration of [11C]zolmitriptan.

A separate study sought to elucidate the uptake

and brain distribution of [11C]zolmitriptan following IN administration by a nasal spray in healthy volunteers, finding rapid absorption via the nasal mucosa, with detectable levels in brain tissue as early as 5 min post-administration [103]. PET imaging revealed a fast and dose-dependent absorption of [11C]zolmitriptan into the brain. Notable levels of [11C]zolmitriptan were observed across all examined brain areas. The initial peak in uptake occurred at the end of the 5 min infusion, followed by a gradual decrease in SUV over the next hour. The highest uptake was observed in large blood vessels and areas without a BBB, such as the choroid plexus, as well as in grey matter regions, with the inferior temporal cortex having the highest uptake. Following IN administration, the concentrations in the CNS progressively increased, peaking at approximately 2 nM by the 0.5 h mark and 3.5 nM after 1 h. The mean concentration in the CNS at 5 min reached 0.5 nM quickly, surpassing the in vitro thresholds necessary to start agonistic effects on  $5-HT_{1B/1D}$  receptors, a specific subtype of serotonin receptors. The authors concluded uptake into the brain was predominantly via systemic circulation, where it then traversed the BBB. Over time, [<sup>11</sup>C]zolmitriptan permeated all brain regions, with its concentration in the brain reaching nearly 20% of that in the plasma after 60 min. Despite this progressive uptake, the total level of uptake remained relatively low, consistently around 12% of the total administered activity. Additionally, after reaching its peak concentration, [11C]zolmitriptan showed a gradual decrease in concentration over time, as it washed out of brain tissue. The study also indicated that zolmitriptan is eliminated through hepatic biotransformation to three major metabolites, including 183C91, a potent  $5-HT_{1B/1D}$  agonist. At 60 min post-administration, 46% of the [11C]zolmitriptan in plasma was intact tracer plus the active metabolite 183C91. The gradual decrease in SUV over the last hour of the PET scan reflected the washout of the tracer from the brain. The study underscored the efficacy of IN administration of 11C-zolmitriptan in rapidly delivering the drug to the brain, facilitating prompt pharmacological action, which is essential for the effective management of acute migraine symptoms. These pharmacokinetic properties support the use of [11C]zolmitriptan in situations where rapid onset of action is critical and highlight its suitability as a treatment option in the acute phase of migraine attacks.

Comparing the pharmacokinetics and applications of zolmitriptan labelled with different imaging agents highlights distinct differences and similarities across the studies. Experiments involving [99mTc]-zolmitriptan formulations, such as MEs and mucoadhesive MEs demonstrated rapid and efficient brain uptake with higher DTE% and DTP% compared to other formulations [24]. Particularly, the mucoadhesive MEs had superior brain retention and rapid onset of action, with a DTE% of 533 and a DTP% of 81, indicating effective nose-to-brain delivery. In contrast, the [11C]-labelled zolmitriptan administered to healthy volunteers had lower but detectable brain penetration, with mean brain concentrations peaking at 3.5 nM and brain uptake reaching nearly 20% of plasma levels [100, 103]. In these human studies brain uptake was primarily through systemic circulation through the crossing of the BBB. The [99mTc]-zolmitriptan nanocarriers also involved enhanced brain uptake and prolonged retention, with a brain/blood ratio 5-fold higher than IV injection, highlighting the potential of nanostructured carriers in facilitating efficient nose-to-brain transport [37]. While both tracers confirmed zolmitriptan's potential for nose-to-brain delivery, the  $[99m]$ Tc] formulations generally exhibited higher and prolonged brain uptake compared to the [11C] formulations, which had a stronger correlation with the systemic circulation. These differences emphasise the impact of the imaging agent, formulation, and administration method on the observed pharmacokinetic outcomes and the efficacy of zolmitriptan for targeted brain delivery.

[<sup>11</sup>C] was also used to radiolabel Orexin-A, a natural neuropeptide and potential antinarcoleptic medication that targets the Orexin receptor 1, to investigate its delivery mechanism into the brain upon IN administration in non-human primates and rodents using a nasal spray [104]. The study employed a syringe-catheter device and the Impel POD administration device for rodent models, while an IN administration device developed by Impel NeuroPharma was used for non-human primates. The researchers employed [11C]CH3-Orexin A solution, a novel radiolabelled form of Orexin A, to track the peptide's brain distribution via PET imaging. The imaging revealed that following IN administration of  $[$ <sup>11</sup>C]CH<sub>3</sub>-Orexin A indicated the uptake was only detected in the olfactory bulb, parts of the frontal cortex, and also distally in the brain stem. These measured signal changes were attributed to partial volume effects. No uptake was reported in any other regions in the brain 90 min post-administration. The team demonstrated that such findings indicate a potential lack of nose-to-brain delivery pathway for  $[$ <sup>11</sup>C]CH<sub>3</sub>-Orexin A (see Figure 6). On the other hand, IN administration of 11C-raclopride solution, is a well-known radiotracer used in PET imaging to study dopamine D2/D3 receptors, which are abundant in the striatum of the brain, showed notable radioactive signal increase in various brain regions. A small but detectable amount of 11C-raclopride was observed in the striatum, a region with high D2 and D3 dopamine receptor binding. Blood analysis post-administration of [11C]raclopride suggested that its brain uptake could be attributed to its presence in the blood, aligning with the patterns seen in IV injection. However, the possibility of direct nose-to-brain transport of [11C]raclopride via IN administration remains open to debate, highlighting differences in the brain accessibility of small molecules versus peptides like Orexin A.



Figure 6. Brain PET/MRI images for rhesus macaque post IN administration (A) [<sup>11</sup>C]CH<sub>3</sub>-Orexin A tracer and (B) [<sup>11</sup>C]raclopride. (A) Images showing no detectable brain uptake in unaffected regions. (B) Images show uptake concentrated in the striatum, highlighting dopamine D2/D3 receptor binding. Adapted with permission from [104], copyright 2018 American Chemical Society.

To further investigate the limited brain uptake of  $[$ <sup>11</sup>C]CH<sub>3</sub>-Orexin A that was observed in non-human primates, a similar study was performed in rodents, employing both PET imaging and *ex vivo* analysis with [11C]CH3-Orexin A, and [125I]Orexin A. After IN administration of [11C]CH3-Orexin A, PET imaging in rodents mirrored the results in the rhesus macaque, showing minimal brain uptake, particularly in areas not affected by partial volume effects. Subsequent *ex vivo* analysis of rodent brain sections also revealed very low radioactivity levels (signal-to-noise ratio < 3). This was attributed to both the negligible uptake of  $[$ <sup>11</sup>C]CH<sub>3</sub>-Orexin A in the brain and the rapid decay of the 11C isotope. The authors used [125I]Orexin A to address the issue of low radioactivity observed in *ex vivo* brain tissue after IN administration of [11C]CH3-Orexin A, further *ex vivo* brain distribution studies were conducted using [125I]Orexin A. The results showed similar patterns of distribution were observed for both IN administration and IV injection. However, IN administration resulted in a unilateral increase of radioactivity in the olfactory bulb ipsilateral to the administration site, averaged between 0.01 to 0.03% ID/cc, with the highest concentration reaching up to 0.07% ID/cc, suggesting a potential for enhanced local uptake. In contrast, IV injection showed uniform uptake across both olfactory bulbs. Overall, minimal brain uptake of Orexin A through IN administration using  $[$ <sup>11</sup>C]CH<sub>3</sub>-Orexin A was found, while <sup>125</sup>I-Orexin A showcased a possibility for localised enhancement in uptake. Despite the minimal brain uptake of Orexin A through IN administration using  $[11C]CH<sub>3</sub>$ -Orexin A, the study proposed an alternative mechanism for the observed therapeutic effects of IN administration of Orexin A. This mechanism involves the activation of orexin receptor 1 in the olfactory sensory neurons, which could influence sleep-wake cycles through downstream signalling. The researchers advocate for future studies to explore this hypothesis by examining both brain uptake and therapeutic efficacy following IN administration and direct brain injections, particularly in animal models with reduced expression of Orexin A receptors in olfactory sensory neurons.

# 2.2.3. Zirconium-89 ([89Zr])

In a study by Yu *et al.* (2016) on tumour tropism via PET/CT imaging, the migration of human neural stem cells (hNSCs) labelled with 89Zr in rats was measured after IN administration [105]. 89Zr was used due to the long half-life that allows for extended observation periods, which is crucial for detailed monitoring of NSCs migration and distribution

post-administration. This BBB circumventing approach demonstrates promising therapeutic potential, especially when used in conjunction with radiation and oncolytic virotherapy. Utilising a unique mesoporous NP-based radiolabelling technique with 89Zr, the study achieved high sensitivity in PET imaging, enabling precise tracing of small numbers of IN administration delivered stem cells. Comparative analysis revealed distinct primary migration routes for NSCs and mesenchymal stem cells (MSCs). While MSCs predominantly migrated toward glioma xenografts through the brain's caudal regions, NSCs tended to navigate through the rostral regions, with a significant presence in the olfactory bulbs. This differential migration pattern is hypothesised to be influenced by the chemotaxic cytokine receptor profiles of the respective stem cell types, responding variably to the diverse cues of the tissue microenvironment.

[89Zr] was used as the radionuclide to investigate the sensitivity of PET/CT imaging *in vitro* and to track the migration of hNSCs into the brain after IN administration in rats [106]. The study proposed extracellular transport through the nasal cavity and potential intracellular transport mechanisms. The nasally administered hNSCs were foremost distributed in the nasal cavity, with limited transport to the brain. PET/CT scans were performed at various time points post-administration: 10 min, 35 min, 1 h, 4 h, 24 h, 72 h, and 168 h. The *in vitro* sensitivity of PET/CT imaging for detecting as few as 1000 [89Zr]-labelled cells was established. Upon IN administration, *in vivo* PET/CT imaging of [89Zr]hNSCs demonstrated high uptake in the nasal cavity and olfactory bulb within the initial 4 h period. Minor uptake was observed in the pituitary and pons, while the frontal lobe, parietal lobe, striatum, hippocampus, and ventral tegmental area showed relatively low signals, primarily below the detection limit. Retention times showed that radioactive uptake values in the nasal cavity and olfactory bulb gradually decreased over time, with no significant signals detected after 4 h post-administration. To complement PET findings, the study employed BLI using a human norepinephrine transporter-luciferase plasmid NSCs for both *in vitro* and *in vivo* experiments. The *in vitro* sensitivity was tested by seeding different numbers of human norepinephrine transporter-luciferase plasmid NSCs in 24-well plates and performing fluorescence scanning. For *in vivo* experiments, different amounts of human norepinephrine transporter-luciferase plasmid NSCs were administered intrastriatally and nasally to rats and C57BL/6 mice. BLI captured fluorescence signals

post-administration, confirming the localisation and distribution patterns observed in FLI. The *ex vivo* analysis performed at 168 h post-administration confirmed the absence of radioactive signals, indicating that the radioactive substance did not substantially enter the brain, or was below the detection limit of the instrument. This underscores the predominant distribution of [89Zr]hNSCs in the nasal cavity, and their limited detectability in the brain following IN administration in rats. In conclusion, the IN administration of hNSCs in this study revealed limited migration to the brain, with the majority of cells remaining in the nasal cavity and olfactory bulb. PET, FLI, and BLI imaging demonstrated low sensitivity for detecting hNSCs in the brain, while droplet digital polymerase chain reaction and immunohistochemistry validated the biodistribution and presence of hNSCs in different tissues. Droplet digital polymerase chain reaction demonstrated higher sensitivity and accuracy compared to BLI and PET/CT, allowing for the detection of as few as 100-200 cells in tissue samples. This method provided a robust pharmacokinetic profile for tracking hNSCs, complementing the imaging findings and supporting the limited brain uptake observed through PET/CT.

In 2021, Veronesi *et al.* investigated the feasibility of using multimodal imaging for IN administration of polymeric micellar NPs radiolabelled with [89Zr] in rats using a nasal tubing delivery system [107]. The uptake for [89Zr]PLA (polylactic acid)–polyethylene glycol (PEG)–DSPE (Distearoylphosphatidylethanolamine) with diameter size of 100 nm in the olfactory bulb, brainstem, and forebrain was significantly higher upon IN administration compared to IV injection at both 1 and 2 h post-administration. The mechanism of NP delivery via the IN route involves transcellular and intercellular pathways, allowing access to the perineural and perivascular fluid channels surrounding the olfactory and trigeminal nerves. The rapid detection of NPs in the brain within 1 h suggests that bulk convection flow through these perineural and perivascular routes significantly contributes to the observed delivery efficiency. This increase in brain activity after IN administration aligns with earlier findings, such as those reported by Kozlovskaya *et al.* in 2014 [108]. Notably, higher levels of activity were observed in the olfactory region and brainstem than in the forebrain post-IN administration, suggesting that the NPs entered the brain through the olfactory and trigeminal pathways, as these areas are more proximal entry points compared to the more distal forebrain. *Ex vivo* gamma imaging confirmed that activity in the olfactory bulb following IN delivery

was 35.1- and 28.6-fold higher than IV injection at 1 and 2 h, respectively. Brainstem activity was 28.9- and 29.6-fold higher, and forebrain activity was 11.2- and 7.8-fold higher for IN delivery compared to IV injection at the same time points. Here, NPs might have been cleared through CSF spaces, with potential contributions from the blood pool. This aspect remains an area for further investigation to fully understand the pharmacokinetics of intranasally delivered NPs. Although PET results showed increased brain activity following IN administration compared to IV injection, it was difficult to distinguish between the activity levels at the 1 and 2 h post-administration due to PET's lower sensitivity compared to gamma counting. The inability to completely isolate the brain from the nasal cavity and oropharynx *in vivo* studies suggests that some high activity observed in the olfactory bulb and brainstem might partially result from activity in adjacent tissues. This might also explain the lower values seen in the olfactory bulb relative to the brainstem on PET, though gamma counting showed higher values in the olfactory bulb. Furthermore, the study employed autoradiography for *ex vivo* validation of PET/CT imaging findings, which demonstrated localised activity in dissected brain sections. The authors acknowledged the necessity of future studies employing transmission electron microscopy to confirm NP entry into brain tissue and provide histological evidence supporting the imaging results. This observation emphasises the potential of IN administration as a direct and effective method for delivering therapeutic agents to the brain.

# 2.2.4. Gallium-68 ([68Ga])

A PET imaging probe capable of monitoring insulin *in vivo* would significantly benefit clinical interventions targeting insulin resistance. A study by Gollapelli and the team aimed presented the *in vitro* serum stability and autoradiography results of [68Ga]-NOTA labelled with insulin in rodent and non-human primate models of Alzheimer's disease pathology [109]. The [<sup>68</sup>Ga]-NOTA-insulin remarkably maintained over 97% stability in human serum for up to 5 h post-production. Autoradiography results found a significantly lower brain uptake of the radiotracer in transgenic mice compared to age-matched wild-type mice. Additionally, vervets with low CSF  $A\beta_{42}$  levels, indicative of Alzheimer's disease, exhibited reduced uptake in the hippocampus and hypothalamus compared to control vervets with high CSF  $A\beta_{42}$  levels. Notably, there was no significant difference in uptake observed in the frontal cortex and cerebellum. Self-blockade

experiments further demonstrated reduced uptake compared to baseline, confirming the specificity of the radiotracer. Overall, the preliminary autoradiography studies in both rodent and non-human primate models of Alzheimer's disease points to lower uptake of [68Ga]-NOTA-insulin in Alzheimer's disease brains compared to controls. Furthermore, radiotracer uptake in the hippocampus and hypothalamus was inversely correlated with Aβ density in vervet brain tissues, as determined by immunohistochemical analysis. The team's ongoing experiments focus on *in vivo* IN administration and IV injection PET imaging of [68Ga]-NOTA-insulin in non-human primate models of Alzheimer's disease, the determination of metabolic plasma-blood parameters, and the correlation of radiotracer uptake with  $A\beta_{42}$  and tau levels. Such studies aim to provide additional insights into the potential of [68Ga]-NOTA-insulin as a diagnostic tool for Alzheimer's disease and its underlying mechanisms of insulin resistance.

# 2.2.5. Manganese ([51Mn], [52Mn], [54Mn])

[<sup>51</sup>Mn] and [<sup>52</sup>Mn] radioisotopes have been studied as experimental tools preclinically, in both rats and macaque monkeys [110]. Mn radioactive markers provide analogous data to  $Mn^{2+}$  in Mn-enhanced MRI, even at reduced concentrations, by identifying accumulation in different tissues after systemic delivery. This represents approximately a 2000-fold reduction ( $\sim$ 100 mM for Mn<sup>2+</sup> in MRI vs.  $\sim$ 0.05 mM for PET). Tjalve and colleagues used a Hamilton syringe to deliver isotopes [111]. For both [54Mn] and cadmium-109, a volume of 10 µL of isotope solution was administered intranasally into the right nostril of the rats. The delivery pathway for both Mn and cadmium was through the IN route, specifically targeting the olfactory mucosa. The primary mechanism for Mn delivery to the brain involved its transport via primary olfactory neurons to the olfactory bulbs. From there, Mn migrated through secondary and tertiary olfactory pathways, eventually reaching various brain regions and the spinal cord. In contrast, cadmium was taken up into the anterior parts of the olfactory bulbs but could not pass the synapses to secondary neurons. This limitation in crossing synapses restricted cadmium's distribution within the brain. Mn showed a rapid uptake, with significant accumulation in the olfactory bulbs observed within one-day post-administration. Over the subsequent days and weeks, Mn spread to additional brain regions, including the basal forebrain, hypothalamus, thalamus, habenular complex, hippocampus, and cerebral cortex, and even reached the spinal cord. The retention time for Mn

was substantial, with detectable levels persisting in these regions for up to 12 weeks. Cadmium, on the other hand, demonstrated a more restricted uptake. It was primarily confined to the olfactory bulbs at all observed intervals, with minimal spread to other brain regions. These findings indicate that cadmium's retention time in the brain was mainly limited to the olfactory bulbs, with little to no distribution beyond this area. The elimination of Mn from the brain occurred gradually, with a noticeable decrease in concentration over time. Despite this decline, Mn levels in the brain remained higher than in the liver and kidneys at later time points, a hallmark of a slow washout process. Cadmium's elimination pattern was different; it was primarily retained in the olfactory bulbs with minimal spread to other brain regions, i.e., limited washout from the brain. Mn exhibited broad distribution throughout the brain, especially in cell-rich areas such as the basal forebrain and hypothalamus. The autoradiograms showed Mn's progressive movement from the olfactory bulbs to other brain regions, confirming the bypassing of the BBB via olfactory pathways. Cadmium's autoradiography confirmed its high retention in the olfactory bulbs with limited distribution beyond this region, illustrating its inability to pass the synapses to secondary olfactory neurons. The olfactory system appeared to serve as a route for Mn to reach the brain by uptake by olfactory neurons and its subsequent transfer through secondary and tertiary olfactory pathways to other regions. The limitation of using [54Mn] is the need for brain dissection due to its prolonged presence, which can complicate non-invasive longitudinal studies and raise safety concerns regarding radiation exposure over extended periods [110].

Mn-enhanced MRI in rodents indicates that it takes around 24 h for Mn to distribute throughout the brain after an IV injection [112, 113]. As a result, [51Mn], with its short half-life, is unsuitable for studying Mn uptake in the brain. This limitation was evident in experiments with both rats and monkeys, where virtually no tracer activity was observed in the initial two hours [110]. In contrast, [52Mn], with its longer half-life, is effective for identifying brain structures. It can be employed to image brain anatomy and functions, as well as to investigate pathophysiological conditions. When [52Mn]Mn2+ in rats, it was noted that the tracer not only accumulated in the body but was also detected in the brain and head regions, with significant tracer accumulation in the nasal turbinates, pituitary, olfactory bulb, hippocampus, and salivary glands. However, the low resolution of PET imaging combined with the small size of the rat made it difficult to precisely differentiate between brain areas.

Mn-enhanced MRI is used to track the olfactory pathways in the monkey brain post-IN administration enables mapping of neural pathways starting from the nostril and reaching as far as the amygdala over a period of days in ways comparable to Mn-enhanced MRI [110]. Using [52Mn]Mn2+ solution as PET radiotracer. Initial PET scans taken on day 0 show the tracer observed in the nasal cavity without any detectable uptake in the brain. By day 1, it was observable in the olfactory bulb, olfactory tract, and olfactory nucleus. By day 3, the tracer reached the piriform cortex, and by day 4, it extended to the amygdala. The accumulation of the tracer intensified in the piriform cortex and amygdala by days 6 and 7, and by then, it was also detected in the frontal cortex. The tracer's migration aligns with the olfactory pathway, which includes comparisons to atlas slices. The route mapped includes progression through the olfactory bulb, olfactory tract, olfactory nucleus, piriform cortex, amygdala, and frontal cortex. During the same PET scanning session, whole-body PET imaging was conducted on each monkey to monitor the distribution of the radiotracer throughout the body. Immediately after the administration of [52Mn]Mn2+, the tracer was primarily found in the nasal area. However, most of the ingested Mn has been exerted from the body through the digestive tract and bladder, with only trace amounts remaining in the liver and kidneys. The elimination process continues, and by the seventh day, only minimal traces of Mn are detectable in the liver and kidneys. The study underscored the capability of high-sensitivity PET imaging to map the distribution of Mn radiotracers in both the brain and body, achieving results comparable to manganese-enhanced MRI studies. The co-registration of PET images with MRI provided enhanced resolution and anatomical localisation, facilitating a detailed understanding of tracer distribution and neuronal pathways. The findings are supported by systematic comparisons with MEMRI studies, demonstrating that even at lower concentrations, Mn uptake and transport mechanisms remain consistent.

#### 2.2.6. Remarks on PET

PET imaging has shown significant promise in the study of IN administration of neuroimaging agents. The use of various radioisotopes, particularly  $[$ <sup>18</sup>F] and  $[$ <sup>11</sup>C], has provided a deeper understanding of the nose-to-brain delivery pathway. Tracers such as [18F]FDG and [18F]FLT have been instrumental in assessing the direct transport and metabolic processing of imaging agents in the brain. For instance, [18F]FDG studies have revealed significant insights into the IN administration-mediated delivery, highlighting the rapid clearance from the nasal cavity and the contribution of systemic circulation in brain delivery, illustrating a mixed nose-to-brain and systemic pathway. Similarly, [18F]FLT, with its limited BBB permeability, offers a unique perspective on the nose-to-brain pathway, emphasising the immediate but transient enhancement in brain distribution following IN administration. In contrast, the use of [11C] labelled zolmitriptan and orexin-A has shed light on the distribution patterns and potential limitations of IN administration. For example, [11C]zolmitriptan studies found minimal yet detectable brain uptake, suggesting a predominant systemic route of brain delivery post-IN administration. The application of [11C]Orexin A resulted in limited uptake in the brain, indicating potential challenges in achieving effective nose-to-brain delivery for certain molecules. The use of [11C] in these studies highlighted both advantages and limitations. While [11C] provides high-resolution PET imaging and minimal long-term radiation exposure due to its short half-life, these same properties pose logistical challenges and limit its effectiveness for capturing slower biological processes or assessing deeper brain penetration. These findings indicate that while  $[$ <sup>11</sup>C] is valuable for certain scenarios, alternative isotopes might be necessary for a more comprehensive assessment of INDD mechanisms, particularly for targeting deeper brain structures. Additionally, the application of radiometals like [89Zr], [68Ga] and [64Cu] has expanded the scope of PET imaging in IN administration studies. The tracking of 89Zr-labeled hNSCs, [68Ga]-NOTA labelled insulin, and [64Cu]AuNCs (data provided in this section 2.5.2) has provided valuable data on the migration and biodistribution patterns of these agents, further contributing to our understanding of IN administration's potential in targeted brain therapies. By using  $[89Zr]$ ,  $[68Ga]$  and [64Cu], researchers tap into the unique properties of these isotopes for effective, long-term tracking of cells *in vivo*. Despite these advancements, challenges such as the potential toxicity of long-lived radioisotopes and the spatial resolution limitations associated with certain radioisotopes remain. These factors necessitate careful consideration in the design and interpretation of PET imaging studies. Furthermore, the field is evolving, and more research is needed to optimise tracer development, refine imaging methodologies such as advanced PET imaging methodologies, including the development of specific and selective radiotracers, hybrid imaging techniques like PET/CT and PET/MRI for combined functional and anatomical insights, and dynamic imaging with quantitative analysis tools. Furthermore, to fully understand the long-term effects and safety of IN administration in clinical settings, including exploiting more sophisticated nasal dosage forms. The insights gained from these PET studies are invaluable in paving the way for more effective treatments for CNS disorders, where traditional systemic administration is often hindered by the BBB.

#### **2.3. Optical imaging**

OI continues to be evolving in the field of neuroscience due to its unique ability to view the intricate workings of the brain. The technology holds immense potential for advancing our understanding of INDD systems. OI techniques, leveraging the emission and absorption of light, enable the visualisation of biological processes and structures in the brain with remarkable clarity and precision [114]. OI in nose-to-brain applications delineates the pathways and mechanisms involved in the direct transport of substances from nose-to-brain. This non-invasive approach can particularly be useful for studying the dynamics of drug delivery and the distribution of therapeutic agents, in turn leading to valuable insights into the efficiency and effectiveness of IN administration treatments.

At the heart of nose-to-brain studies are optical imaging techniques. Bioluminescence (BLI) is a technique where light is produced as a consequence of a chemical reaction within an organism. Here, the energy needed to produce the light is drawn from the environment (i.e., such as oxygenated blood re-energising cells). This method is highly sensitive and is used for tracking the distribution and persistence of cells or specific genes [115]. BLI can be instrumental in monitoring the movement and localisation of IN administration therapeutic agents, enabling researchers to track the progress and effectiveness of treatments in real-time. Fluorescence (FLI), on the other hand, involves the use of fluorescent probes or markers that emit light when excited by a specific wavelength (i.e., externally supplied sources of energy, such as lasers, to produce light emission). This technique provides a method for detailed visualisation of drug movement and interactions within brain tissues. FLI is particularly useful in nose-to-brain studies for mapping the pathways of drug delivery from nose-to-brain and assessing the distribution of therapeutic compounds.

# 2.3.1. Fluorescence imaging (FLI)

Garzotto and Marchis explored the use of

fluorescent quantum dots (QDs), specifically carboxylated CdSe-ZnS (cadmium selenide-zinc sulphide) QDs, as IN administration imaging agents in mice using a pipette, to understand their translocation from nose-to-brain [116]. QDs a design of advanced fluorescent semiconductor nanocrystals frequently utilised in cellular and animal biology. The QDs used were Carboxyl ITK QDs 585 and 525, which are fluorescent semiconductor nanocrystals. The delivery pathway targeted the olfactory epithelium, exploring both neuronal and extracellular transport mechanisms. The study focused on the potential routes QDs might take from nose-to-brain, hypothesising the involvement of extracellular pathways such as perineuronal and perivascular spaces and CSF routes. To track the movement and distribution of QDs, confocal microscopy was used to analyse the olfactory epithelium at different time points post-administration. After administering acute IN administration treatments and employing confocal microscopy to observe the QD's journey within the olfactory epithelium. Initially, to determine whether QDs can penetrate and traverse the olfactory epithelium, they conducted a single IN rinse on adult mice and examined the distribution of QDs within the olfactory epithelium over varying durations from 1 to 24 h. One-hour post-rinse, a pronounced fluorescence was detected on the luminal surface of the olfactory epithelium, particularly around the cilia of the olfactory receptor neurons and the microvilli extending from the apical surface of the sustentacular cells. By 4 h, they noted an increased spread of fluorescent spots throughout the thickness of the olfactory epithelium. By 24 h, QDs had reached and accumulated in the olfactory epithelium's basal area. Both 4 and 24 h post-rinse, QDs were also seen in the adjacent lamina propria, with a gradual reduction in QD density over time. Fluorescence was frequently observed in polygonal cells surrounding blood vessels and occasionally in nerve bundles. The study suggested that QDs can traverse the olfactory epithelium and enter the underlying lamina propria. The movement of QDs through the olfactory epithelium might occur via a transcellular route involving neuronal and sustentacular cell bodies, or a pericellular route through extracellular spaces. To further explore the interaction and intracellular positioning of QDs across different environments and neuronal types, specific olfactory receptor neuron labelling was conducted *in vivo*, and neuronal enriched primary cultures from the olfactory bulb were used *in vitro*. *In vivo* experiments co-administered QDs and fluorescein dextran (Dex), a molecule known for its uptake by olfactory receptor

neurons and transport to the olfactory bulb via the olfactory nerve through IN rinse. After 4 h post-treatment, fluorescein Dex was found extensively within olfactory receptor neurons, encompassing the entire soma, although confocal multi-stack analysis revealed no colocalisation with QDs. This finding not only demonstrates the potential of QDs in brain imaging applications, but also underscores the importance of further research into their long-term effects and clearance mechanisms, whilst addressing growing concerns about the safety of NPs in biomedical applications.

A study investigated the distribution of Cyanine 5.5 (Cy5.5) when delivered either IN administration or IV injection [117]. Using a small animal fluorescence live imaging system, significant differences were found in the distribution of Cy5.5 based on the delivery method. IN administration primarily resulted in fluorescent signals in the stomach but not in the liver. Conversely, IV injection led to strong signals in the liver but not in the stomach, with fluorescent signals detected in multiple brain regions. Furthermore, significant accumulation in the olfactory bulbs within 1 to 2 h after IN administration, indicating high concentrations of Cy5.5, whereas such concentrations were not observed in other brain regions. The findings highlight distinct differences in the pathways and distribution patterns of Cy5.5 between IN administration and IV injection, particularly emphasising the olfactory bulbs as a primary site for drug accumulation following IN administration. In a different study, a stable protein-based NPs system, CB-Gd-Cy5.5, using Cy5.5 as a model drug, was developed [118]. The focus was on tracking the distribution and metabolism of CB-Gd-Cy5.5 in the brain following IN administration. Using a filled pipettor, each mouse receives a 10 µL volume of the solution. Findings showed widespread fluorescence signals throughout the brain within 1 h of administration, persisting up to 24 h (see Figure 7). This highlighted the system's efficiency in delivering drugs to the brain via the nose-to-brain pathway, outperforming the bovine serum albumin (BSA) BSA-Gd-Cy5.5 system. A key observation was the significant signal enhancement in the hippocampus shortly after administration, suggesting effective delivery of the CB-Gd-Cy5.5 NPs to this region. This was likely facilitated by the CB to the GM1 ligand on nerve cell membranes. Quantification of the tracer revealed that the fluorescence signal in the brain for the CB-Gd-Cy5.5 group was 22 times greater than in the liver, underscoring the efficiency of the delivery system. Over time, the fluorescence signal declined,

suggesting gradual clearance of the NPs from the brain. Histochemical analysis, including Hematoxylin and eosin and Nissl staining, indicated that the CB-Gd-Cy5.5 NPs did not cause noticeable histopathological abnormalities or tissue damage in the brain or other major organs. The Nissl staining showed no significant neuronal loss or disappearance of nuclei, confirming the safe use of the NPs with brain tissue. Overall, the CB-Gd-Cy5.5 NPs demonstrated promising potential for brain targeting via IN administration, with effective delivery to the hippocampus, prolonged retention, and minimal off-target effects, as confirmed by advanced imaging techniques and histological analysis. This study provides a robust framework for the design of protein-based multifunctional drug delivery systems aimed at treating neurodegenerative diseases like Alzheimer's (MRI data will be explained in section 2.4.1).

A study employed a multimodal approach to analyse the spatiotemporal distribution of gold nanorods (AuNRs) following IN administration, utilising OI, gamma counting, and autoradiography as complementary techniques [119]. The delivery device used for administration was a micropipette, and a total volume of 20 μL was administered intranasally. OI, based on the fluorescent properties of Cy5.5-labeled AuNRs, provided qualitative insights into the *ex vivo* organ distribution profiles, enabling the semi-quantitative tracking of AuNR particles in specific organs, especially the brain, as a rapid screening method. Gamma counting facilitated data on organ biodistribution profiles of 111In[DTPA (diethylenetriamine pentaacetate)-AuNR<sub>PEG-NH2</sub>] in mice and led to information on AuNR uptake over time. Autoradiography was applied to investigate the regional distribution of 111In[DTPA-AuNRPEG-NH2] in the brains, revealing a rapid uptake within 10 min post-administration, indicative of an efficient delivery mechanism. Initial uptake was predominantly in the olfactory bulbs, then over time, the AuNRs diffused into adjacent regions such as the olfactory tract, anterior olfactory nucleus, and piriform cortex, with evidence of deeper brain region involvement, including the brainstem, within an hour. OI confirmed successful translocation to the brain, with higher fluorescence signals in the frontal brain region. Gamma counting revealed that about 30% ID/g remained in the nasal passage, and radioactivity in the brain peaked at 10 min, measuring  $0.036\%$  ID/g, post-administration, gradually decreasing over time. Autoradiography supported these findings, showing region-specific brain accumulation, notably in the olfactory bulbs, elucidating the involvement of the

olfactory and trigeminal pathways in the intranasal delivery of AuNRs. This comprehensive approach revealed the spatiotemporal dynamics of AuNRs in the brain, providing crucial insights for the development of IN AuNRs as potential INDD carriers for CNS applications. The retention of AuNRs in the brain showed a marked decrease over time. High levels were detected at 10 min, but by 3- and 7-days post-administration, the Au content had significantly diminished, interpreted as limited long-term retention. This temporal reduction was confirmed through various imaging modalities, including OI, inductively coupled plasma mass spectrometry, and nuclear imaging techniques. The study employed multi-modal imaging agents to characterise the distribution and clearance of AuNRs. The Cy5-functionalised AuNRs were tracked using OI, while the DTPA-functionalized AuNRs complexed with [<sup>111</sup>In] allowed for nuclear imaging. This approach highlighted the initial high uptake in the olfactory regions and subsequent broader distribution to other brain areas over time. Quantitative measurements via inductively coupled plasma mass spectrometry found the highest brain uptake at 10 min post-administration, with 39.71 μg Au/g of brain, decreasing to  $0.59$  μg  $Au/g$  by day 7. Gamma counting provided further quantitative biodistribution data, revealing an initial brain uptake of 0.04% ID/g of tissue. The AuNR's elimination from the brain predominantly occurred through mucociliary clearance, transitioning the particles to

the gastrointestinal tract for excretion via faeces. This research emphasises the importance of using multi-modal imaging approaches combined with high-resolution histochemical analysis to validate the biodistribution and therapeutic potential of intranasally administered agents. The integration of such techniques offers a comprehensive understanding of the localisation and kinetics of drug delivery systems, enhancing their application in treating CNS diseases.

In Scranton *et al.*'s study, fluorescent tag CellTracker Green (CTG) BODIPY and radiolabelled tracers, [125I]EPO (erythropoietin) and [125I]calcitonin, were used to investigate the transport mechanisms within the rostral migratory stream for INDD to the brain [120]. The researchers used a micro syringe to administer 6 µL of the tracer solution to mice's nasal cavities. The study aimed to understand how these substances are transported to various brain areas via the rostral migratory stream. This nerve, which connects the olfactory bulb to the periventricular regions, facilitates the passage of substances through a paracellular mechanism, allowing them to traverse the nasal mucosa and bypass the BBB. The uptake of the tracers was assessed at multiple time points, specifically 20-, 60-, and 120-min post-administration. The results showed significant amounts of tracers in the brain as early as 20 min after administration. Notably, regions such as the olfactory bulb, hippocampus, cortex, cerebellum, and choroid plexus exhibited high levels of tracer presence. The retention



**Figure 7.** A) Ex vivo fluorescence images and B) Analysis of fluorescence intensity in mouse brains prior to and at 1, 6, 12, and 24 h following IN administration of CB-Gd-Cy5.5<br>or BSA-Gd-Cy5.5. C) Fluorescence microscopy CB-Gd-Cy5.5 presence. 1-4 representative fluorescence images of brain sections from mice at 1 h post-IN administration CB-Gd-Cy5.5 or BSA-Gd-Cy5.5. Adapted with permission from [118], copyright 2019 Wiley.

indicated that these levels remained significant within the 120 min observation window. Their observations suggested a paracellular transport mechanism, where substances move between cells, rather than within them, indicating that the physical structure of the rostral migratory stream is crucial for this process. When the rostral migratory stream was intact, efficient delivery to the brain was observed, with ~41% ID of [125I]-EPO and ~19% ID of [125I]calcitonin reaching the brain within 20 min after IN administration. Whereas, when the rostral migratory stream was surgically transacted, the normal path of these tracers to the brain was significantly hindered, with only about 3% ID of [125I]EPO and 4% ID of [125I]calcitonin, leading to their accumulation in peripheral organs. This finding not only highlights the essential role of the structure of the rostral migratory stream in facilitating direct CNS delivery, but also suggests longer-term changes, such as reactive astrogliosis from surgical disruption, could affect transport. CTG fluorescence was observed extensively in brain regions without a detectable presence in peripheral tissues. Liquid scintillation analysis of [125I]EPO and [125I]calcitonin confirmed high CNS concentrations in intact rostral migratory stream mice, supporting the efficiency of this delivery route. The study highlights the rostral migratory stream in INDD as a pathway to the brain. However, it does require further research, especially regarding its application in human medicine. Adding to the context of these findings, published results from autoradiographs have indicated that IN administration radioligands distribute throughout the brain without being confined to or concentrated in the olfactory areas innervated by the olfactory nerve [121]. Moreover, the tracers used in Scranton *et al.*'s study did not show a preference for the projection areas of the olfactory nerve. These results propose that the rostral migratory stream, rather than the surrounding olfactory nerve, may serve as the main transport route, or alternatively, that substances transported by the olfactory nerve might bypass the olfactory region of the brain. This suggests the involvement of additional pathways, with the intact trigeminal nerve offering a potential uptake path into the CNS.

Subsequently in 2011, the researchers investigated the potential of MSCs, labelled with enhanced green fluorescent protein (EGFP) and made available via IN administration using a pipette, to migrate to and survive in the mouse brain having striatal lesions [122]. The MSCs, harvested from genetically modified mice expressing EGFP, served as a fluorescent imaging agent to track their transport. Despite employing sophisticated imaging methods

like fluorescence microscopy, the MSC-EGFPs were not detectable in the brain from 3 h to 2 months post-administration. This observation raised questions about the viability and migration capability of MSCs. Surprisingly, a significant autofluorescent signal, mimicking the GFP emission, was observed in both the olfactory bulb and striatum of mice, regardless of whether they received MSC-EGFP treatment or were part of the control group. This autofluorescence complicated the interpretation of results, emphasising the need for meticulous controls in experiments using GFP as a tracking method. In contrast, a control experiment where MSC-EGFPs were directly implanted into the striatum of mice showed robust GFP expression at 1- and 7-days post-implantation, confirming the cell's survival and GFP expression in a different delivery context. Under the conditions of this study, the findings suggest that MSC-EGFPs failed to survive or migrate within the brain, highlighting the complexities of cell-based therapies and the critical role of appropriate control experiments. Furthermore, high-resolution histochemical analysis did not confirm the presence of MSC-EGFPs in the brain. Electron microscopy depicted cell-like structures filled with inclusion bodies and lipofuscin, indicative of tissue degradation, rather than GFP-specific signals. In summary, the IN delivery of MSC-EGFPs did not result in the migration or survival of these cells in the brain. The autofluorescence observed in the olfactory bulb and striatum, which could be mistaken for a GFP signal, underscores the necessity of including rigorous controls when using GFP as a marker. This study highlights the challenges of confirming the localisation of intranasally delivered cells or drugs using high-resolution imaging and histochemical techniques. The findings emphasise the need for multiple validation methods to accurately differentiate true signals from artifacts, ensuring the reliability of the results.

In 2012, two studies explored the IN administration of insulin as a novel approach to treat CNS disorders. The first study focused on the use of an ME method, where fluorescein isothiocyanatelabelled insulin (FITC-insulin) was administered to rats in a water-in-oil formulation [123]. The insulin was administered intranasally with a volume of  $5 \mu$ l per nostril using a pipette tip positioned at the nostril opening. The study found significant fluorescence levels in the brain as early as 24 h after a single IN administration of the ME formulation. This rapid uptake indicates the effectiveness of the olfactory route in delivering insulin directly to the brain. When comparing different delivery methods, the ME

formulation resulted in a 2-fold higher fluorescence signal in brain sections, particularly in the olfactory bulb and frontal lobes, compared to the aqueous solution. This suggests a higher brain uptake with the ME system. Retention times varied depending on the frequency of administration. For a single dose, significant brain uptake was observed 24 h post-administration. For twice-daily doses, the ME formulation continued to show enhanced brain uptake. However, when administered thrice daily, there was no significant increase in brain uptake, possibly due to enzymatic degradation within the olfactory bulb, which limited further uptake. Quantification of the insulin reaching the brain showed that a single IN dose of the ME resulted in an average fluorescence level of ~86 pdu, while the aqueous solution achieved ~51 pdu. With 2 doses administered 12 h apart, the ME showed ~139 pdu compared to 49 pdu for the aqueous solution. After 3 doses, the ME resulted in ~66 pdu, whereas the aqueous solution had  $\sim$ 36 pdu. This quantification highlights the superior brain uptake efficiency of the ME formulation. The elimination of the tracer involved enzymatic degradation within the olfactory bulb, particularly with repeated doses. This degradation may limit further brain uptake due to increased protein concentration. Fluorescence microscopy provided detailed imaging of the fluorescently labelled insulin distribution in brain sections. This method allowed for precise quantification and localisation of insulin within brain tissues, supporting the study's findings. Overall, the study demonstrates that IN delivery of insulin via an ME system is significantly more effective in targeting the brain than aqueous solutions or IV injection. This method has potential therapeutic applications for neurological disorders, such as Alzheimer's disease, by effectively bypassing the BBB and directly targeting the brain. The second study focused on specific pathways utilised in IN administration insulin delivery to the CNS [124]. The study utilised a pipette to intranasally administer insulin, delivering a total volume of 24 µL of tracer solution. This mouse study used both fluorescent and electron microscopy 30 min post-administration to track the insulin's journey. The primary delivery pathway identified was the olfactory nerve pathway. Insulin administered intranasally traversed this pathway, which involves extracellular delivery in perineuronal spaces surrounding olfactory nerve axons extending from the olfactory mucosa to the anterior regions of the olfactory bulbs. Confocal microscopy confirmed a continuous stream of fluorescently tagged insulin moving from the neuroepithelium of the olfactory

mucosa, through the cribriform plate of the ethmoid bone, and into the olfactory bulbs. This rapid delivery route aligns with previous findings suggesting that extracellular transport provides quicker access to the olfactory bulbs compared to intracellular transport within the axons, which takes several hours. The study demonstrated significant uptake of insulin within 30 min of administration. Fluorescent and electron microscopy showed that insulin reached the anterior regions of the olfactory bulbs. Higher concentrations were observed in the olfactory nerve layer compared to the glomerular layer, with the fluorescent label preferentially localising within the periglomerular cells of the glomerular layer. This distinct distribution pattern highlights the effectiveness of the olfactory nerve pathway in targeting the olfactory bulbs quickly after IN administration. Retention of insulin within the olfactory nerve bundles and layers was evident shortly after administration, suggesting active uptake and possible endocytosis by cells in these regions. Electron microscopy revealed intracellular nanogold-labelled insulin within the olfactory nerve layer and glomerular layer. Insulin was localised in the cytoplasm, along the nuclear membrane, and associated with the heterochromatin of the nucleus within neurons. This suggests that the insulin was not only delivered but also actively transported and potentially functional within these cells. The study utilised two types of imaging agents: Alexa Fluor 647 labelled insulin for macroscopic and confocal imaging and nanogold labelled insulin for electron microscopy. Fluorescent imaging showed a continuous extracellular distribution along the olfactory nerve bundles, while nanogold imaging revealed intracellular uptake within olfactory bulb neurons, indicating active transport and potential functional activity of the delivered insulin. This dual approach provided a comprehensive view of insulin distribution and uptake, confirming the olfactory nerve pathway as an effective route for CNS targeting via intranasal administration. Together, these studies present a viable and effective method for delivering therapeutic agents like insulin directly from nose-to-brain.

The comparative analysis of the studies on insulin labelled with different imaging agents reveals crucial insights into their pharmacokinetics, regional brain uptake, and therapeutic implications. The [18F]-labelled insulin, studied by Smith *et al.*, demonstrated rapid and efficient brain delivery via a catheter-based liquid method, making it highly effective for targeting limbic and frontotemporal regions critical for Alzheimer's disease treatment [2].

The [68Ga]-NOTA-labelled insulin, as presented by Gollapelli *et al.*, exhibited high stability and provided valuable diagnostic insights into Alzheimer's disease, particularly regarding insulin resistance mechanisms [109]. The FITC-labelled insulin, administered through a water-in-oil ME formulation, showed superior brain uptake efficiency, though enzymatic degradation with repeated doses posed a limitation [123]. Lastly, the Alexa Fluor 647 and nanogold-labelled insulin demonstrated effective CNS targeting via the olfactory nerve pathway, with significant intracellular uptake and potential functional activity within neurons [124]. Collectively, these studies emphasise the importance of selecting appropriate tracers and delivery methods tailored to specific therapeutic or diagnostic goals, ultimately advancing the potential of nasal delivery systems for treating and diagnosing Alzheimer's disease.

In a study for fast transport within the cerebral perivascular conducted by Lochhead *et al.*, they used Texas Red-labelled Dex's (TR-Dex) of 3 and 10 kDa as fluorescent imaging agents to investigate the mechanisms and distribution of tracer transport in the brain following IN administration in rats [125]. With delivery carried out using a micropipette, upon IN administration, the tracers utilised the olfactory and trigeminal nerve pathways, rapidly reaching the brain and distributing via the perivascular spaces of cerebral blood vessels. The study used *ex vivo* fluorescence imaging and confocal microscopy to demonstrate that within 20 min, TR-Dex3 was prominently distributed within the perivascular spaces of surface arteries and internal cerebral vessels. For TR-Dex10, a similar distribution was observed within 30 min, but only when an absorption enhancer, matrix metalloproteinase-9, was used, indicating a size-dependent barrier at the nasal epithelium. The uptake regions included the olfactory bulbs, trigeminal nerves, and various brain regions such as the cortex and hippocampus. The study highlighted that significant levels of tracers were observed in the perivascular spaces surrounding leptomeningeal vessels on the brain's ventral and lateral surfaces. The retention time was primarily noted within the first 20-30 min post-administration, with fluorescence signals indicating widespread brain distribution. Quantification of the tracers revealed that IN administration resulted in significantly higher brain levels compared to intraarterial administration. TR-Dex3 showed a 24.2-fold increase in perivascular fluorescence intensity after IN delivery compared to intraarterial administration. While the study did not delve into the detailed mechanisms of tracer elimination from the brain, it suggested that

substances might drain via the CSF and lymphatic pathways. Confocal microscopy confirmed the imaging results, showing detailed localisation of TR-Dex3 and TR-Dex10 in brain sections. The tracers were predominantly associated with the perivascular spaces of blood vessels and surrounding brain parenchyma, supporting the observed rapid and widespread distribution post-IN administration. The study systematically compared the two types of imaging agents, TR-Dex3 and TR-Dex10, while highlighting their distinctive distributions within the brain. TR-Dex3 showed efficient brain uptake and distribution without the need for absorption enhancers, whereas TR-Dex10 required matrix metalloproteinase-9 pretreatment to achieve similar results. This comparison emphasizes the importance of molecular size in determining the permeability and distribution efficiency of intranasally administered agents. This research revealed the potential of INDD as a promising approach for treating diseases with cerebrovascular pathology.

In a study by Stanojević-Vitaliano, the potential of clathrin triskelia, a natural coat protein, NPs as a nanoplatform for delivering antibodies and imaging agents to the CNS was explored [126]. The study involved the use of a 10 μL Hamilton syringe for IN administration. FITC has tagged clathrin triskelia and tested its ability to noninvasively penetrate the rat brain. Fluorescently labelled NPs were observed in the brain as early as 30 min after IN administration, indicating rapid uptake. The dopamine 3 receptor antibodies-triskelia successfully targeted dopamine 3 receptor brain regions such as the olfactory tubercle, islands of Calleja, ventral pallidum, nucleus accumbens, striatum, frontal/parietal cortex, hippocampus, medial mammillary bodies, anteroventral thalamic nucleus, substantia nigra/ventral tegmental area, and cerebellum. The retention time varied, but nanoprobe distribution in brain regions was evident 3 h post-administration, suggesting sustained presence in the target regions. Immunohistochemistry confirmed the localisation of dopamine 3 receptor antibodies in targeted brain regions. Positive signals from anti-dopamine 3 receptor antibodies were detected in regions such as the nucleus accumbens, islands of Calleja, hippocampus, and substantia nigra. The study also used fluorescent dopamine 3 receptor antibody-Triskelia, where fluorescent signals detected in dopamine 3 receptor brain regions confirmed specific targeting, showing punctate deposits within discrete cellular structures. Rhodamine-triskelia without targeting antibodies was found in all brain regions, including the prefrontal cortex, indicating
nonspecific distribution. These findings highlight the potential of clathrin-based NPs in transporting large molecules, like antibodies, into the CNS, offering new avenues for diagnosing and treating CNS disorders.

In a cargoes but not vehicles study, Ahmad *et al.* tracked the NEs and their cargoes for nose-to-brain delivery using a micropipette [127]. The mechanisms facilitating this delivery included the use of mucoadhesive polymers and the careful selection of particle size. Due to their mucoadhesive properties, chitosan-coated NEs were used to increase retention time in the nasal cavity. Additionally, thermosensitive *in situ* gels made from Poloxamer 407 and Poloxamer 188 were employed to further prolong the residence time of NEs in the nasal mucosa. NEs with particle sizes around 100 nm demonstrated superior retention and penetration through the mucosal layer compared to larger particles, as *in vivo* imaging revealed their presence in the mucosa and along the trigeminal nerves within 1 h post-administration. The mechanism involved passage through the olfactory and trigeminal nerves, smaller NEs (80-200 nm) were retained in the nasal mucosa for up to 16 h, whereas larger particles (500-900 nm) were cleared more quickly. The chitosan-coated NEs had enhanced retention compared to their non-coated counterparts. In terms of brain uptake, only weak signals from the environment-responsive probes (P2 and P4) were detected in the olfactory bulb and brain, indicating that only minute quantities of integral NEs reached these areas. Fluorescent signals captured at various time points (up to 16 h post-administration) indicated a gradual, size-dependent translocation. The cargoes within the NEs, represented by DiR and Cou-6 signals, were found in significant amounts along the nose-to-brain pathway and in the brain, suggesting that the cargoes were effectively released and transported. Very minimal amounts of NEs were detected in the blood or vital organs, suggestive of negligible systemic absorption. High-resolution histochemical findings from confocal laser scanning microscopy provided detailed images showing intense signals of the cargoes (Cou-6) in various brain regions. This supported the conclusion that the cargoes, rather than the intact NEs, were the primary agents reaching the brain. *Ex vivo* imaging further validated these findings, showing weak P2 signals (representing NEs) in the olfactory bulb and brain, but strong signals for the cargoes. In summary, the study demonstrates that while integral NEs have limited direct transport to the brain, the cargoes they carry can effectively permeate brain tissues via the nose-to-brain pathways. The histochemical analysis confirmed the imaging data, showing the localisation

and distribution of the drugs within the brain. These findings suggest that the primary role of NEs in nose-to-brain delivery is to facilitate the transport of their cargoes, rather than the carriers themselves. This provides important insights into the mechanisms of INDD and highlights the potential of NEs for enhancing brain drug delivery. In another study, DiR-labelled PLGA NPs were employed to investigate the distribution of edaravone, a drug for amyotrophic lateral sclerosis, following IN administration and IV injection in mice [128]. The delivery device used for IN administration was a micropipette, with 10 µL of the NP suspension instilled into each nostril. The uptake of edaravone was rapid, with comparable amounts  $(\sim 353 \text{ ng/g})$  detected in the brain at 0.5 h post-administration for both IN and IV routes. The NP formulation provided prolonged retention, with significant amounts still present at 12 h  $(\sim 213 \text{ ng/g})$ and 24 h  $(\sim 60 \text{ ng/g})$  post-IN administration. This sustained presence indicates the effectiveness of the IN route in maintaining therapeutic levels of the drug in the brain over time. The study identified specific brain regions with high uptake, including the olfactory bulbs, hippocampus, and cerebral cortex, which are crucial areas affected by neurodegenerative diseases like amyotrophic lateral sclerosis. In addition to the brain, IN administration resulted in uptake in other organs such as the lungs, spleen, liver, kidneys, stomach, and intestines, although to a lesser extent compared to IV injection, indicating a more localised and targeted delivery to the brain. Quantitatively, the IN route achieved  $\sim 0.8\%$  ID/g, significantly higher than the IV route  $\sim 0.26\%$  ID/g at 0.5 h. This higher efficiency underscores the potential of the IN pathway in delivering therapeutic doses directly to the brain with minimal peripheral distribution. Elimination and clearance of the drug varied between the two routes. IN administration showed limited systemic exposure and negligible liver uptake, suggesting efficient targeting and reduced risk of systemic side effects. In contrast, the IV route exhibited higher systemic distribution with significant initial liver uptake, which decreased over time. The study primarily relied on FLI and ultra-performance liquid chromatographytandem mass spectrometry quantification to track the biodistribution and retention of edaravone. The imaging and quantification methods used provided robust evidence of the drug's presence and distribution in the brain.

A study by Xu *et al.* on the IN administration of icariin for treating depression developed an icariin nanogel loaded with a self-assembled thermosensitive hydrogel system, using rhodamine B-labelled nanogels as the imaging agent for tracking [129]. This formulation targeted the brain with rapid uptake observed within 30 min post-administration, leveraging the IN route for direct and efficient drug delivery to the brain. Retention of icariin in the brain was notably prolonged. The fluorescence signals from the rhodamine B-labelled nanogels were sustained in the brain for up to 24 h, demonstrating the sustained release properties of the icariin-nanogel system. This prolonged retention is indicative of the drug's potential for long-lasting therapeutic effects following a single administration. The elimination of the tracer involved systemic distribution to other organs, primarily the liver and kidneys, which were identified through subsequent fluorescence signals. That is, after initial uptake in the brain, the drug was gradually distributed systemically and eliminated through metabolic and excretory pathways. Hematoxylin and eosin staining of the hippocampal tissues in the rat models showed significant morphological improvements in neuronal cells within the CA1 and CA3 subregions of the hippocampus in the icariin nanogel treated group compared to the chronic unpredictable mild stress group. These histopathological results highlighted the potential neuroprotective effects of icariin-nanogel, suggesting its efficacy in reversing stress-induced neuronal damage. The use of rhodamine B-icariin nanogels as imaging agents facilitated a clear understanding of the drug's distribution, showcasing its quick localisation to the brain, sustained retention, and subsequent systemic distribution and elimination. The comparative analysis of imaging agents demonstrates the efficiency of the icariin-NGSTH system in delivering and retaining therapeutic agents in the brain.

A later study focused on glioblastoma treatment using a disulfiram-loaded ion-sensitive NE *in situ* gel (DSF-INEG) delivered IN with copper ions [130]. The study utilised a microlitre syringe to intranasally administer the DSF-INEG into one nostril (left or right) of the rats. The administered volume was approximately  $100 \mu L$ . The study's imaging findings confirmed this efficient transport, with fluorescence imaging revealing significantly higher fluorescent intensity in the brain compared to other organs at various time points (1, 3, and 6 h post-administration). The primary regions showing uptake included the olfactory bulb and broader brain tissue, indicating effective transport through the olfactory and trigeminal nerve pathways. Cy5.5, a fluorescent dye, was used as a surrogate marker to assess the brain targeting capability. Cy5.5 is a lipophilic dye that matches the poorly soluble nature of disulfiram, making it a suitable choice for tracking the NE's distribution. The strong brain-targeting capability of

Cy5.5-INEG via nose-to-brain transport was evidenced by significantly higher fluorescent intensity in the brain than in other organs. This finding indicated the formulation's effectiveness in delivering the drug to the brain. The retention time of the DSF-INEG formulation in the brain was notable for its sustained release properties. *In vitro* release profiles indicated that the formulation could maintain therapeutic levels in the brain over an extended period, with 50% of the drug released at 4 h and approximately 75% at 12 h. This sustained release is critical for prolonged therapeutic effects against glioblastoma. Quantification of the drug reaching the brain was inferred through comparative fluorescence imaging intensities. The fluorescence intensity of brain tissues in rats treated with Cy5.5-INEG was significantly higher than in control groups, resulting in effective brain targeting and quantifiable delivery. GFP imaging and Hematoxylin and eosin staining were used to assess tumour growth and tissue health. Enhanced tumour growth inhibition was observed in the DSF-INEG/Cu treated rats, evidenced by weaker fluorescence signals in brain tumours and less tissue deterioration compared to control groups. Histopathological analysis revealed no significant differences or pathological changes in the major organs of treated rats, indicating high safety and effective targeting of the brain tumour with minimal off-target effects.

Ferreira and team developed an nose-to-brain co-delivery system for glioblastoma treatment, employing Poly (D,L-lactic-co-glycolic acid)/chitosan NPs [131]. The delivery device used in the study was a micropipette, with 40 μL of NP solution divided equally between both nostrils. These NPs were uniquely loaded with alpha-cyano-4-hydroxycinnamic acid (CHC) and conjugated with the monoclonal antibody cetuximab. The system's effectiveness in targeting the brain was evaluated using IR780, a near-infrared fluorescent dye, as the imaging agent. FLI was used to observe the brain uptake and distribution of these NPs. CHC-NP showed intense brain fluorescence signals as early as 30 min post-administration, indicating rapid uptake. In contrast, conjugated NP maintained stronger fluorescence signals over a 3 h period, suggesting prolonged retention and effectiveness in brain uptake. The strong fluorescence signals suggest effective transport and retention of the drug in the brain. This qualitative evidence was crucial for validating the nose-to-brain delivery method. This uptake difference between the NPs indicates that conjugated NP has a better retention profile compared to CHC-loaded NP, making it more suitable for prolonged therapeutic

effects. *Ex vivo* brain FLI was conducted at multiple time points following IN administration, 0.5, 1, and 3 h (see Figure 8). Early fluorescence signals at 0.5 h indicated quick NP accumulation in the brain, with an increased intensity observed at 1 h, reflecting a higher concentration of the IR780-loaded, drug-conjugated NPs. By 3 h, the imaging either demonstrated a sustained NP presence in the brain or the onset of clearance from brain tissue.

In 2023, Ye *et al.* investigated an approach for enhancing drug delivery to the brain using transcranial magnetic stimulation and fluorescent magnetic NPs [132]. The study utilised a syringe pump for IN administration. The study utilised a transcranial magnetic stimulation device to generate time-varying magnetic fields, distinct from static magnets, to attract fluorescent magnetic NPs to brain tissues. Since transcranial magnetic stimulation typically generates high-intensity fields for clinical neurostimulation, the study employed a lower intensity, less than 50% of the resting motor threshold, to move fluorescent magnetic NPs without inducing nerve stimulation. This approach was shown to significantly increase the fluorescence intensity of fluorescent magnetic NPs in the brain, indicating successful delivery. The effectiveness of fluorescent magnetic NP delivery was observed to improve with increasing magnetic field strength, with 1.5- to 1.8-fold increases in brain fluorescence at 25% and 50% resting motor threshold, respectively, compared to the control group [132]. Further, the study evaluated the impact of infusion rate on delivery efficiency. Slower injection rates resulted in greater fluorescence intensity in the brain, suggesting that rapid flow rates might limit fluorescent magnetic NP exposure to the magnetic field. Additionally, the size of the NPs played a crucial role in their brain uptake. Smaller NPs (less than 100 nm) were more efficiently delivered to the brain, aligning with the diameter of olfactory axons, which facilitate passive transport to the brain. Interestingly, even larger fluorescent magnetic NPs (up to 300 nm) showed increased brain delivery under the influence of the transcranial magnetic stimulation-induced magnetic force. The study also noted that the biodistribution of fluorescent magnetic NPs was predominantly in the liver and kidneys, with no significant differences observed in other major organs between the



Figure 8. Illustrating the *ex vivo* brain fluorescence tomography. It depicts the brain images at different time intervals (0.5 h, 1 h, and 3 h) following the IN administration of IR780-labelled CHC-loaded NPs (panel A) and IR780-loaded conjugated NPs (panel B), along with a separate column representing negative controls. "Adapted with permission from [131] copyright 2021 ELSEVIER".

transcranial magnetic stimulation and control groups. Hematoxylin and eosin staining were performed on the brain and other major organs. The results showed no gross or microscopic anomalies, inflammation, or fibrosis in the tissues examined. Despite high levels of NPs in the liver and kidney, no significant lesions were observed, indicating the safety of the experimental protocol.

A study by Han used the fluorescent polystyrene NPs, specifically PS-YG, which are 100 nm-sized NPs tagged with a fluorescent dye [133]. They investigated the IN administration uptake, accumulation, and neurotoxic effects in the mouse brain using a micropipette to precisely deliver 24 µL. Using fluorescence microscopy, they visualised the distribution and presence of these fluorescent polystyrene NPs within brain tissues, notably in and around neurons. The study found that, shortly after administration, PS-YG accumulated in the brain with significant uptake evident within just 3 h, leading to an acute neurotoxic response characterised by increased levels of neurotoxic and inflammatory markers. The uptake regions include the olfactory bulb, cerebral cortex, cerebral nuclei, hippocampus, thalamus, hypothalamus, midbrain, cerebellum, and hindbrain. The retention of PS-NPs within the brain was evident, as fluorescence signals were still present 24 h after administration. Elimination of the PS-NPs from the brain primarily occurred through exocytosis. The study found that inhibiting histone deacetylase 6 significantly enhanced the exocytosis of PS-NPs from neurons, thereby reducing their intracellular accumulation and associated neurotoxicity. This was evident from the decreased fluorescence signals in neurons treated with the histone deacetylase 6 inhibitor, ACY-1215 (ricolinostat). Conversely, blocking exocytosis with an inhibitor like endosidin2 prevented the clearance of PS-NPs, leading to sustained neurotoxic stress. Immunofluorescence showed co-localisation of PS-YG with the neuronal marker β3-tubulin, confirming the accumulation of NPs within neurons. Increased levels of glial fibrillary acidic protein, a marker of reactive astrocytes, were observed in the hypothalamus and olfactory bulb, indicating astrocyte reactivation and neuroinflammation. Additionally, oxidative stress markers such as heme oxygenase 1 and apoptotic markers like cleaved caspase-3 were elevated in PS-NP exposed neurons, highlighting the acute neurotoxic response. This approach provided critical insights into the dynamics of PS-NP penetration, their interaction with neuronal cells, and the brain's response to and clearance of these NPs, highlighting both immediate and longer-term effects.

A study by Hu *et al*. employed near-infrared and MRI to evaluate the efficacy of an IN-delivery system using MIL-100, a metal-organic carbonised framework (CF) combined with domperidone using a nebulizer [134]. MRI provided insights into the brain entry and distribution of the frameworks. Post-administration, the frameworks accumulated in the olfactory bulbs and other brain regions, evidenced by extensive hypointense regions in the MRI images. These hypointense regions rendered the olfactory bulbs nearly imperceptible, indicating significant uptake. The use of an external magnetic field further enhanced brain permeability, aiding the targeted delivery to specific brain areas. This MRI data correlated with behavioural experiments showing significant improvements in depression-like, anxiety-like, and cognitive behaviours in mice treated with the frameworks with domperidone. The increased dopamine receptor density in the prefrontal cortex and hippocampus, observed through immunofluorescence staining, supported these behavioural improvements. *In vivo* fluorescence imaging and MRI found frameworks with FITC exhibited superior brain permeability compared to FITC alone, indicating effective delivery via IN administration. Magnetic assistance by the external field significantly enhanced brain permeability, with the frameworks primarily accumulating in the olfactory bulbs shortly after administration. Subsequent distribution to other brain regions, including the prefrontal cortex and hippocampus, occurred within the first 24 h, as evidenced by fluorescence intensity and MRI hypointense patterns. This rapid uptake and targeted distribution highlight the efficacy of the IN administration system combined with magnetic guidance for achieving therapeutic effects in the brain. Elimination of the tracer from the brain involved its transfer to metabolic organs, followed by renal excretion. The study observed increased iron content in the lungs and kidneys after 10 days of treatment, suggesting partial entry into the lungs post-nasal administration and eventual clearance via the kidneys. This elimination pathway ensures that the nanospheres do not remain in the brain indefinitely, reducing the risk of potential side effects. Together, the complementary findings from near-infrared and MRI modalities demonstrate the effectiveness of the frameworks with domperidone in delivering therapeutic agents from nose-to-brain. The controlled release triggered by near-infrared irradiation and the enhanced brain permeability facilitated by magnetic fields were crucial for achieving significant improvements in mental health conditions in mice models. The rapid uptake and

targeted distribution of the frameworks in the brain regions, including the olfactory bulbs, prefrontal cortex, and hippocampus, within the first 24 h, stress the potential of this delivery system for treating neurological disorders.

A study on the IN administration of chitosan hydrogels labelled with a fluorescent tag (doxorubicin) aimed to develop and evaluate an IN-chitosan hydrogel for the delivery of morphine to achieve fast and prolonged analgesic effects using a spray device [135]. The IN administration of the chitosan hydrogel demonstrated a rapid onset of action, with significant analgesic effects observed as early as 5 min post-administration. The maximum analgesic effect, reaching 93% of the maximum possible effect, was achieved at this time point. This rapid uptake is attributed to the direct nose-to-brain migration pathway facilitated by the chitosan hydrogel, which effectively bypasses the systemic circulation and the first-pass metabolism in the liver. Fluorescent imaging studies further elucidated the regions of the brain involved in the uptake of the chitosan hydrogel. The primary absorption occurred in the olfactory bulbs, as indicated by a significantly higher fluorescent signal intensity of 74% in this region compared to the control groups, which showed only 15% intensity. This suggests that the drug was efficiently transported to the olfactory bulbs. The washout profile of the morphine-loaded chitosan hydrogel showed a prolonged duration of action. Following the initial peak effect at 5 min, the analgesic effect gradually decreased but remained significant for up to 2 h, maintaining 27% of the maximum possible effect. Interestingly, a delayed recovery of the analgesic effect was noted 4 h post-administration, increasing to 70% of the maximum possible effect and further rising to 80% at 6 h post-administration. This extended presence and delayed clearance indicate that the chitosan hydrogel formulation allows for sustained release and retention of the drug in the brain, thereby providing prolonged therapeutic effects.

A study focused on the development and application of multifunctional NPs designed to target mitochondria and respond to reactive oxygen species for treating ischemic stroke in rats [136]. Two tracers were used for imaging, Cou-6 and Rhodamine B. Cou-6 was used as a model drug for fluorescent imaging, which enables researchers to track the distribution and behaviour of the NPs *in vivo*. The Rhodamine B labelled NPs serve to trace the distribution and release profiles of the therapeutic agents encapsulated within the NPs. Upon IN administration, the multifunctional NPs utilise the

nose-to-brain pathway, effectively bypassing the BBB and facilitating rapid delivery to the brain's ischemic regions. The NPs accumulate in the brain rapidly, with detectable fluorescence signals appearing within 1 h post-administration, peaking at 2 h, and remaining visible for up to 24 h. The mitochondria NPs were specifically targeted to the ischemic penumbra, showing higher fluorescence intensity in the left ischemic hemisphere compared to the non-ischemic hemisphere. This targeted delivery was enhanced by the SS-31 peptide, which aids in directing the NPs to mitochondria within the ischemic brain regions. The study also utilised an *in situ* gel for the IN administration of the mitochondrial NPs. This thermosensitive gel prevented nasal runoffs and prolonged the residence time of the NPs in the nasal cavity, thereby improving bioavailability. The IN administration route capitalised on the olfactory and trigeminal nerves to transport the NPs directly to the brain. Over time, the tracer indicated a gradual decline in fluorescence intensity after 2 h. Yet, the signal remained detectable at 24 h postadministration, suggesting a relatively slow clearance rate and prolonged therapeutic effect. The therapeutic efficacy of multifunctional NPs was demonstrated through their ability to alleviate oxidative stress, reduce inflammation, repair mitochondrial function, and decrease apoptosis in the ischemic penumbra. *Ex vivo* fluorescence imaging confirmed the enhanced targeting and distribution of multifunctional NPs in the brain, particularly in the ischemic regions, compared to control groups. This study highlights the potential of multifunctional NPs to provide targeted, effective treatment for ischemic stroke, leveraging advanced imaging techniques to track their distribution and therapeutic impact.

A study explored innovative techniques and agents to enhance the delivery of *N*-acetyl-L-cysteine (NAC) from nose-to-brain using a micropipette [137]. It primarily utilised *in vivo* imaging to track NAC's distribution and delivery in rats. The researchers used hyaluronic acid silk fibroin hydrogels, known for their mechanical strength, as the primary biomaterial. To increase their effectiveness, dopamine polydopamine was incorporated into these hydrogels, creating multifunctional hydrogels with high adhesive forces and photothermal response effects. NAC, the primary drug used in the study, was labelled with FITC for fluorescence tracking. The study found that NAC delivery to the brain was significantly enhanced using multifunctional hydrogels combined with near-infrared irradiation. The study revealed that the uptake of NAC was notable within 2 h post-administration, with significant localisation

observed in the olfactory bulb and hippocampus regions of the brain. The use of hyaluronic acid silk fibroin NAC hydrogels ensured sustained delivery and retention of NAC in the brain tissues, maintaining a presence for at least 6 hours. The fluorescence intensity measured in the *in vivo* imaging study found FITC-NAC in brain tissues to be approximately 5.4 and 9.0-fold greater following administration with hyaluronic acid silk fibroin FITC-NAC and hyaluronic acid silk fibroin FITC-labelled NAC-near-infrared hydrogels, respectively, compared to FITC-NAC solutions. This rapid and enhanced delivery demonstrated the effectiveness of multifunctional hydrogels and the photothermal response effect in facilitating efficient drug uptake and transport to the brain. The hyaluronic acid silk fibroin NAC hydrogels allowed for controlled release, achieving 57% cumulative release within 6 h. The release rate was further enhanced to 65% with the incorporation of periodic near-infrared irradiation, confirming the photothermal response effect's efficacy. The primary uptake region for NAC was the nasal cavity, from which it was transported to specific brain regions, including the olfactory bulb and hippocampus. FLI in the nasal cavities for FITC-NAC solutions indicated fast mucociliary clearance. In contrast, multifunctional FITC-NAC hydrogels, with or without near-infrared irradiation, demonstrated increased retention in the nasal cavity, resisting mucociliary clearance. This was attributed to the high adhesive forces of the multifunctional hydrogels, which enhanced the retention time and prevented rapid clearance from the nasal mucosa.

A study by Shen *et al.* explored the efficacy of IN administration of MSC-derived small extracellular vesicles for CNS therapy and delivered 20 µL using a micropipette [138]. This approach aimed to bypass the BBB and deliver therapeutic agents directly to the brain. The extracellular vesicles were labelled with Cy3 and Cy7 fluorescent dyes and glucose-coated AuNPs. Cy3 was used for fluorescence microscopy and confocal imaging. Cy7 was used for near-infrared fluorescence imaging. MSC-derived small extracellular vesicles were administered intranasally. The distribution of these vesicles was tracked using near-infrared FLI, which revealed rapid and widespread distribution across various brain regions, with significant fluorescence detected as early as 0.5 h post-administration and persisting for up to 6 h post-administration, although the signal diminished by 24 h. Higher radiant efficiency was noted in the olfactory bulb, cortex, striatum, hippocampus, pons, and cerebellum, indicating effective brain delivery. Cellular uptake studies conducted using fluorescence

microscopy and confocal imaging demonstrated that multiple brain cell types, including microglia, astrocytes, and neurons, incorporated MSC-derived small extracellular vesicles. Quantitative analysis revealed significant uptake in subcortical regions, with a higher percentage of neurons and microglia incorporating MSC-derived small extracellular vesicles in the hippocampus, pons, and cerebellum compared to astrocytes. This suggests vesicles are effectively distributed and taken up by various brain cells. The perivascular pathway was identified as a crucial mechanism for the distribution MSC-derived small extracellular vesicles. Confocal imaging showed the presence of MSC-derived small extracellular vesicles within the perivascular space, surrounded by AQP4-bearing astrocytic end feet and PDGFR-β positive pericytes. Transmission electron microscopy images validated the localisation of AuNP-MSC-derived small extracellular vesicles within the perivascular space, indicating their movement along this pathway. As some MSC-derived small extracellular vesicles entered peripheral circulation, as evidenced by the increased fluorescence signals in the liver and kidney, the majority remained localised within the brain. This localisation reduces peripheral distribution and clearance compared to IV injection, enhancing the central delivery of therapeutic agents. These findings highlight the promise of intranasal MSC-derived small extracellular vesicles for treating neurological disorders by leveraging the perivascular transport mechanism to achieve effective central delivery.

The study on rivastigmine nasal spray for Alzheimer's disease treatment demonstrated important findings using FLI to assess nasal spray formulation, olfactory deposition, and brain delivery via VP7 nasal spray devices [139]. Rivastigmine, a classic cholinesterase inhibitor, was used in the formulations. *In vivo* imaging tracking with Rhodamine B incorporated into the nasal spray formulations provided visual confirmation of the drug transport process. Fluorescence signals were observed in both the nasal cavity and brain within 15 min post-administration, peaking around 60 min, indicating rapid uptake. *Ex vivo* fluorescence imaging further detailed the drug distribution in the brain, showing strong fluorescence signals in the olfactory bulbs, trigeminus, and multiple brain regions, including the corpus callosum, interbrain, caudate putamen, hippocampus, thalamus, and cerebellum. Additionally, desorption electrospray ionisation mass spectrometry was employed to semi-quantitatively analyse rivastigmine distribution, confirming high-intensity signals in these regions as early as 5

min and providing precise spatial mapping post administration and gradually decreasing from 15 to 300 min. These findings indicated that the drug was successfully transported via both olfactory and trigeminal neuronal pathways. The washout mechanisms were also examined, with ciliary clearance in the nasal cavity removing deposited drugs within approximately 15 min, and brain clearance occurring through the glymphatic system and CSF circulation within a few hours. These imaging results evidence the importance of optimising formulation viscosity and deposition to enhance brain delivery efficacy.

Zhang *et al.* performed a study focused on the IN administration of phenytoin (PHT) loaded layered double hydroxide NPs (BSA-LDHs-PHT) for treating epileptic seizures, aiming to enhance drug delivery efficiency to the brain while reducing peripheral toxicity [140]. The delivery device used for this purpose was a pipette, administering a total volume of 50 µL per mouse. The BSA-LDHs-PHT bypassed the BBB by utilising the olfactory and trigeminal nerve pathways, enabling direct brain access from the nasal cavity. These NPs provided colloidal stability, controlled drug release, and enhanced brain targeting due to their positive charge and anion exchange properties. *In vivo* FLI tracked the distribution of BSA-LDHs-Cy5.5 in mice, revealing significantly higher brain/peripheral ratios, which was more than double the uptake for BSA-Cy5.5, thus indicating superior brain targeting. Fluorescence intensity measurements post-administration showed uptake values of  $5.25 \times 10^8$  at 15 min, maintaining high levels for up to 24 h. The highest brain concentration of phenytoin was 404.3 ng/mL at 4 h, remaining high for up to 8 h. Additionally, lower fluorescence intensities in peripheral organs (liver and kidneys) suggested reduced peripheral distribution and potential toxicity. The authors focused on overall brain uptake and specific regions, comparing peripheral distribution. The clearance of the tracer indicated prolonged retention in the brain, with high phenytoin concentrations maintained for 8 h post-administration. *In vivo* biodistribution assays, conducted at multiple time points (15 min to 24 h), confirmed sustained brain targeting and reduced peripheral distribution. This comprehensive study suggests that IN administration of BSA-LDHs-PHT is a promising strategy for improving the therapeutic effect of phenytoin in treating epileptic seizures with enhanced brain targeting and reduced peripheral toxicity.

A study by Chung and coworkers utilised both Cy5- and FITC-labelled small interfering RNA (siRNA) as an imaging agent to track the delivery,

localisation, and clearance of siRNA complexes within the brain [141]. The IN administration was performed using a pressurised olfactory device from Impel Neuropharma. These siRNA complexes consisted of anti-apoptotic siRNA targeting the Bax gene (Bcl-2-associated X protein), which were complexed with Fas-signaling blocking peptides (FBP) coupled with 9R a nona-arginine peptide, forming the FBP9R/siRNA complexes. Cy5-siRNA was primarily used to monitor the distribution and retention of the siRNA complexes in the brain. Following IN administration, the Cy5-siRNA complexes were observed to localise specifically to the ischemic region within 12 h, indicating rapid targeting. Significant retention of these complexes was noted up to 48 h post-administration, demonstrating effective targeting and prolonged presence in the ischemic area. FITC-siRNA was utilised to track siRNA uptake and intracellular localisation within cells. FITC labelling allowed for the precise visualisation of siRNA within cells using confocal microscopy and flow cytometry. This approach enabled the researchers to evaluate the efficiency of siRNA delivery by the FBP9R complexes to Fas-expressing cells. Confocal microscopy showed colocalisation of siRNA within endosomal vesicles, confirming efficient intracellular trafficking and endosomal escape. The FLI studies showed that the Cy5-siRNA complexes were predominantly taken up in the infarcted (ischemic) brain region, with minimal to no presence in non-ischemic regions. This specificity confirms the targeted delivery capability of the FBP9R/siRNA complexes. 9R with control peptides, however, demonstrated a rapid clearance profile, with a significant reduction in the imaging signal at 24 h post-administration. In contrast, FBP9R/siRNA complexes maintained a strong presence in the ischemic region for up to 48 h, suggesting slower clearance and better retention in targeted areas. Experimental findings from confocal microscopy and flow cytometry supported these observations. Confocal microscopy visualised the Cy5-siRNA in brain tissue sections, confirming targeted delivery and retention in the ischemic region, with specific localisation noted at 12 h post-administration. Flow cytometry quantified the presence of Cy5-siRNA in single-cell suspensions from brain regions, revealing a high percentage of Cy5-positive cells in the ischemic region, approximately 72% at 12 h, 42% at 24 h, and 40% at 48 h post-IN administration. Overall, findings highlight the potential of the IN-delivery system for achieving sustained therapeutic effects in targeted brain regions, with efficient delivery and prolonged retention of therapeutic agents. FITC and Cy5

labelling were crucial in providing detailed insights into the uptake mechanisms, regional localisation, and clearance profiles of the siRNA complexes.

A study by Tscherrig investigated the therapeutic potential of micro RNAs (miRNAs) carried by small extracellular vesicles derived from Wharton's jelly MSCs in a preclinical rat model of premature white matter injury [142]. The primary focus was on the biodistribution, uptake mechanisms, targeted brain regions, and clearance of these small extracellular vesicles. Small extracellular vesicles were administered intranasally to ensure direct delivery to the brain. Fluorescent labelling with DiR dye enabled tracking of the small extracellular vesicles at 1 h, 6 h, and 24 h post-administration. The imaging results showed that small extracellular vesicles rapidly reached the brain, with significant accumulation observed as early as 1 h post-administration. The signals remained detectable for at least 24 h, indicating prolonged retention in the brain. Notably, higher uptake was observed in injured animals compared to controls, suggesting that injury might facilitate vesicle uptake due to increased permeability of the BBB. The study specifically targeted the corpus callosum and the external capsule regions of the brain. In the corpus callosum, significant reductions in microglial activation markers were observed following the administration of naïve small extracellular vesicles. This reduction highlights the anti-inflammatory effects of the small extracellular vesicles. Later, while strong signals persisted in the brain, there was notable clearance through the gastrointestinal tract, evidenced by strong fluorescent signals 24 h post-administration. This finding aligns with the known mucociliary clearance mechanisms from the nasal cavity. Moreover, no significant signals were detected in the liver, spleen, or lungs, indicating minimal systemic clearance through these organs. In summary, this is compelling evidence that intranasally administered Wharton's jelly MSCs small extracellular vesicles effectively target the brain, reduce inflammation and promote oligodendrocyte maturation in a model of premature white matter injury. The findings highlight the potential of miRNA-based therapies for treating neonatal brain injuries, with detailed imaging studies confirming the targeted delivery, retention, and clearance pathways of the small extracellular vesicles.

# 2.3.2. Multimodal imaging

BLI and FLI imaging were crucial in tracking and evaluating the effectiveness of intranasal (IN) administration of polyfunctional gold-iron oxide NPs (polyGIONs) loaded with therapeutic miRNAs for

treating glioblastoma [143]. BLI, using FLUC-EGFP-labelled glioblastoma cells, enabled real-time, non-invasive monitoring of tumour growth and response to polyGION treatment. FLI, aided by Cy5 dye tagging of the miRNAs within the polyGIONs, provided detailed insights into the *in vivo* distribution and trafficking of these NPs. This approach revealed that T7-targeted polyGIONs effectively reached and accumulated in the brain, especially in the prefrontal cortex and other vital regions, highlighting their enhanced cellular uptake within glioblastoma sites. The uptake of polyGIONs was observed to initiate post-administration rapidly, with significant NPs accumulation in the brain within 24-48 h. The study indicated that the NPs remained detectable in the brain tissues at least up to 72 h post-administration, demonstrating a substantial retention period which is crucial for the therapeutic efficacy of the delivered miRNAs. This efficient delivery was facilitated by the mucoadhesive properties of the NPs, aiding their transit from the nasal cavity to the brain through perivascular spaces and along olfactory and trigeminal nerve pathways, thereby bypassing the BBB. Retention of the polyGIONs in the brain was observed for several days, indicating their stable presence at the target site. MRI played a vital role in visualising and tracking the NPs within the brain. The magnetic properties of the iron oxide component in the polyGIONs allowed for detailed observation of their spatial distribution, confirming their successful BBB bypass and direct delivery to brain tissue. The dynamic process of the NPs trafficking from the nasal cavity to targeted brain regions was mapped comprehensively, providing a complete view of the delivery mechanism, complemented by BLI and FLI imaging. In addition to real-time imaging, biodistribution analysis indicated significant retention of miRNAs in the brain, particularly in the prefrontal cortex and regions accessed via the olfactory bulbs and trigeminal nerves. This targeted accumulation demonstrated the potential of T7-targeted polyGIONs in enhancing the therapeutic effects of the chemotherapy drug temozolomide. The study highlighted the improved therapeutic outcomes with T7-targeted polyGIONs, which resulted in increased survival rates in glioblastoma-bearing mice compared to control groups receiving non-targeted NPs or temozolomide alone. The IN-administration device used in this study was specifically designed to facilitate the efficient administration of the nanoformulation. This device enabled the precise delivery of the polyGIONs into the nasal cavity, where their mucoadhesive chitosan coating allowed for prolonged residence time

and enhanced uptake through the nasal mucosa. Overall, the use of multimodal imaging techniques, namely BLI, FLI and MRI, provided critical insights into the biodistribution, uptake mechanisms, and therapeutic efficacy of polyGIONs in glioblastoma treatment. This non-invasive, targeted delivery approach shows promise in improving the treatment of glioblastoma by effectively bypassing the BBB and enhancing the delivery and effectiveness of therapeutic agents like temozolomide.

### 2.3.3. Remarks on OI

OI is revolutionising our understanding of INDD, particularly in the context of nose-to-brain pathways. For example, research using fluorescent QDs, such as carboxylated CdSe-ZnS QDs, in mice demonstrated their translocation across the olfactory epithelium, predominantly through extracellular routes, offering insights into brain imaging applications. Studies with Cy5.5 and CB systems revealed differential drug distribution patterns and efficiency in the brain based on the administration route, with IN administration showing significant potential for targeted brain delivery. Notably, research on AuNRs employed a combination of OI, gamma counting, and autoradiography, illustrating the comprehensive tracking of these particles' spatiotemporal dynamics in the brain. In studies investigating the rostral migratory stream and neurotoxicity of polystyrene NPs, OI highlighted crucial aspects of drug transport mechanisms and potential neurotoxic responses. Further, OI has been instrumental in evaluating the effectiveness of various therapeutic agents, such as insulin and icariin, and diagnostic agents like clathrin triskelia and fluorescent magnetic NPs, showcasing its versatility in enhancing our understanding of therapeutic agent delivery and distribution within the brain via IN administration. These advancements in OI highlight its potential in both therapeutic and diagnostic applications, offering essential insights into the mechanisms and efficiency of IN administration treatments, albeit with ongoing research needed to address safety and efficacy concerns.

OI's ability to provide real-time visualisation of the distribution and persistence of therapeutic agents and its non-invasive nature makes it an invaluable tool in neuroscience research, particularly in INDD studies. Its applications range from tracking drug movement to assessing the effectiveness of different formulations, creating an understanding of the mechanisms and efficiency of IN administration treatments. However, the field necessitates further research into the long-term effects and clearance

mechanisms of various agents used in biomedical applications, addressing growing concerns about safety and efficacy.

### **2.4. Magnetic resonance imaging**

MRI is capable of delivering high contrast between various soft tissues, facilitating the detection of a variety of diseases and the evaluation of therapeutic results [144]. The imaging modality plays an important role in monitoring nose-to-brain drug delivery by providing real-time, non-invasive visualisation and tracking of the drug movement [145]. This is usually achieved with the use of contrast agents incorporated into the drug delivery system [146].

MRI contrast agents normally involve metals, such as gadolinium (Gd), iron (Fe), and Mn. The presence of contrast agents with these metals is known to have a measurable impact on the relaxation times of tissue, albeit at sensitivities well below that afforded by PET and SPECT experiments. Normally, MRI-based studies evaluate contrast agent induced changes in T1 and T2 relaxation times. Commonly, Gd-based chelates, ultra-small superparamagnetic iron oxide (USPIO), micron-size paramagnetic iron oxide (MPIOs), superparamagnetic iron oxide NP (SPION), and Mn-based agents are used [147]. Each contrast agent leads to a different type of soft tissue contrast enhancement, and they tend to be chosen specifically based on their characteristics.

# 2.4.1. Gadolinium (Gd)

Numerous studies have explored nose-to-brain pathways using various imaging methods and agents. Upon IN administration using a micropipette, the Gd agent can access the brain in as short as 30 min post-administration [148]. A study by Yadav *et al.* (2018) used MRI to evaluate and confirm the IN administration of positively charged cyclosporine A (CSA) NE (CSA-NE) to the CNS in rats. They compared the uptake of Gd-encapsulated NE (Gd-NE) and an aqueous solution of Gd-DTPA into the brain. MRI scans were acquired pre-administration, and 30, 60, and 90 min post-administration of the Gd agent to assess the change in T1 relaxation time in the rat brain (see Figure 9) [148]. At 30 min, the decrease in T1 values were qualitatively notable in the main parts of the brain except for the amygdala and cerebellum. At 60 min, T1 changes became additionally evident in the cerebellum, cranial nerves, and ventral striatum. Also, their atlas-based comparison demonstrated a significant uptake of Gd-NE in 22 out of 172 brain regions. Moreover, the authors reported that Gd-NE

had a longer residence time in the brain than Gd-DTPA. They concluded that the Gd contrast agent may be transported to the brain through both the trigeminal and olfactory pathways and diffuse from the olfactory bulb to other parts of the brain. The diffusion process may also be accelerated by the perivascular pump and arterial pulsation.

A Gd agent was used to evaluate IN administration of CB by micropipette in the mouse brain with the intended target being the hippocampus [118]. Results were compared with those obtained using BSA. Scans were acquired before and 1 h post-administration of Gd-CB and Gd-BSA solutions. Signal enhancement using Gd-BSA was not reported; however, statistically significant MRI signal enhancement was found in the hippocampus using the Gd-CB agent (see Figure 10). This study confirms that Gd-based agents can be used to monitor potential pathways for INDD.

#### 2.4.2. Manganese  $(Mn^{2+})$

 $Mn^{2+}$  is usually taken up by active neurons, which enables studies on brain function, neuron activity, and neuronal connections. Pautler *et al.* explored the potential of Mn<sup>2+</sup> as a T1 contrast agent for MRI to trace neuronal pathways in live mice [149]. The delivery devices used for administering Mn<sup>2+</sup> were pipettes for the olfactory pathway and intravitreal injections for the visual pathway.  $MnCl<sub>2</sub>$ solution was administered into the naris of the mice.

The delivery pathways and mechanisms varied between the olfactory and visual systems. In the olfactory pathway,  $Mn^{2+}$  was topically applied to the naris, enabling its transport anterogradely from the olfactory receptor neurons through the olfactory bulb to the primary olfactory cortex. In the visual pathway, Mn2+ was injected into the vitreous humour of the eye and followed the optic nerve to the optic tract and superior colliculus, indicating trans-synaptic transport. This transport was evidenced by the enhancement of signals in specific brain regions associated with these pathways. In the olfactory pathway, signal enhancement was detectable 24 h post-administration, peaking around 29 h, and diminishing by 72 h. Enhanced regions included the olfactory bulb, primary olfactory cortex, and pituitary gland. In the visual pathway, enhanced contrast was visible within 2 h post-injection, peaking at 24 h, and returning to baseline by 72 h. Enhanced regions included the vitreous humour, optic nerve, and superior colliculus. Quantification of the  $Mn^{2+}$  tracer in the brain revealed that Mn<sup>2+</sup> levels increased from 15 nmol/g tissue to 18 nmol/g tissue in the olfactory bulbs at peak signal enhancement. The most significant increase was observed in the microsomal fraction of subcellular structures, indicating effective uptake and transport of  $Mn^{2+}$  within the neurons. However, its use in humans is limited by its potential neurotoxicity.



Figure 9. Demonstrate the Gd<sup>3+</sup>-NE distribution within the brain regions. An illustrative image highlights the spread of the Gd agent in various distinct areas of the rat brain map following IN administration (A). Gd<sup>3+</sup>-NE Distribution of in primary brain areas at 25-30 min post-administration, in contrast to Magnevist and the control (B). Distribution of

Gd3+- NE in significant brain areas 55–60 min after administration, in comparison to both Magnevist and the control (C). Adapted with permission from [148] copyright 2019 American Society for Pharmacology and Experimental Therapeutics.



Figure 10. T1 MRI scans of mouse brains are displayed, taken at both 0 h and 1 h, before and after administration. The upper row (A) features images with CB-Gd-Cy5.5, while the lower row (B) presents those with BSA-Gd-Cy5.5. A red arrow highlights signal enhancement in the hippocampus. Adapted with permission from [118], copyright 2019 Wiley.

In a different study a humidifier to aerosolise a concentrated MnCl<sub>2</sub> solution, either alone or in combination with amyl acetate or male mouse urine, was used [150]. This aerosolised solution was delivered to the mice in a controlled manner. The humidifier was turned on for 5 min and off for 5 min, repeated twice, and then kept off for 1.5 h before imaging. In another experiment,  $4 \mu L$  of 39  $\mu$ M MnCl<sub>2</sub> in 1 ml  $H_2O$  were pipetted into the mice's naris. Mn<sup>2+</sup> was delivered intranasally, entering excitable cells through voltage-gated calcium channels. Once inside the cells, Mn<sup>2+</sup> was transported anterogradely along neurons via a microtubule-dependent mechanism. This transportation led to the accumulation of  $Mn^{2+}$  in specific brain regions that were activated by the odour stimuli, such as the olfactory bulbs. The study demonstrated that Mn2+ could effectively trace neuronal pathways from the nasal epithelium to the olfactory bulb. Also, the study assessed the uptake time by imaging the mice 1.5 h post-exposure to the Mn2+ and odour stimuli. This timeframe allowed sufficient uptake and transport of  $Mn^{2+}$  to the brain regions of interest, enabling the researchers to capture the activated areas through MRI. Localised T1-weighted MRI enhancement was observed in both the olfactory epithelium and olfactory bulb. Specific regions of the olfactory bulb activated by the odours showed increased signal intensity. For the high pheromone content odour (male mouse urine), enhancement was seen in the accessory olfactory bulb, while amyl acetate caused enhancement in distinct regions of the olfactory bulb. The Mn<sup>2+</sup> enhancement persisted long enough to allow for high-resolution MRI imaging, indicating that the tracer remained in the activated regions for a relatively extended period.

This prolonged retention facilitated the detailed mapping of neuronal pathways. An occasional MRI enhancement in the ventricular regions was noted, suggesting systemic distribution and eventual clearance of  $Mn^{2+}$  through the brain's ventricular system and choroid plexus. This implies a possible route for tracer elimination from the brain.

Fa *et al.* investigated the use of IN MnCl<sub>2</sub> administration for activity-induced Mn-dependent functional MRI of the rat visual cortex.  $MnCl<sub>2</sub>$  was delivered intranasally using a pipette, with a total volume of 10 µl [151]. The delivery pathway involved the transport of  $Mn^{2+}$  along the olfactory nerve, facilitated by olfactory and visual stimulation over a 20 h period, allowing  $Mn^{2+}$  to bypass the BBB. The transport mechanisms included neuronal uptake of Mn2+ through voltage-gated calcium channels, axonal transport, and diffusion in the extracellular space. Activation of neural pathways improved axonal transport, facilitating  $Mn^{2+}$  migration from the olfactory bulb to the visual cortex. This was confirmed by enhanced T1-weighted imaging in the visual cortex and deep brain structures, with specific enhancement noted in the intact olfactory bulb group. In contrast, the group with olfactory bulbectomy showed no cortical enhancement, highlighting the importance of the olfactory bulb in Mn transport. Quantification of the tracer reaching the brain was performed using inductively coupled plasma mass spectrometry. The results depicted elevated Mn levels in the visual cortex of the intact olfactory bulb group  $(7 \mu g/g)$ compared to the olfactory bulbectomy group  $(5 \mu g/g)$ . This confirmed the significant transport of Mn from the olfactory bulb to the visual cortex in the presence of intact olfactory pathways. The elimination of Mn

from the brain involved bioelimination processes, potentially including axonal transport, diffusion, and CSF shifts. The study suggested that Mn eliminated from the olfactory bulb could subsequently transfer to the visual cortex. This study confirmed that  $IN$   $MnCl<sub>2</sub>$ administration could effectively deliver Mn to higher brain regions, such as the visual cortex, enabling non-invasive functional imaging of brain activity.

Romashchenko *et al.* used MRI to explore the spatiotemporal dynamics and regulation of nose-to-brain transport of metal oxide NPs [152]. Two types of NPs,  $Mn_3O_4$  and Fe<sub>3</sub>O<sub>4</sub>, each with distinct imaging characteristics and distributions in the brain, were used. For  $Mn_3O_4$ -NPs, the delivery device was a micropipette, administering a volume of 10 µL intranasally. The delivery pathway involved the nasal cavity targeting the olfactory epithelium, where olfactory receptor neurons took up the NPs. These NPs localised within the cytosol, mitochondria, and vesicles of neurons, moving both intra- and extra-vascularly. The transport mechanisms were dependent on neuronal activity and microtubule integrity. The uptake of  $Mn_3O_4$ -NPs was rapid, with detection in the olfactory bulb shortly after administration. The NPs were trans-synaptically transmitted through at least four synapses across the olfactory tract, reaching regions such as the olfactory bulb and olfactory cortex. The retention time of Mn3O4-NPs in the brain varied, as influenced by neuronal activity and synaptic connections. Quantitative MRI analysis showed a substantial fraction of the administered dose reaching the brain, with accumulation rates correlated with neuronal activity. Eliminating  $Mn_3O_4$ -NPs from the brain was slower, primarily influenced by synaptic transmissions and neuronal activity, with gradual clearance observed. Similarly, Fe<sub>3</sub>O<sub>4</sub>-NPs were administered intranasally using a micropipette with the same volume of 10 µL. These NPs followed the same delivery pathway through the nasal cavity and olfactory epithelium, being engulfed by olfactory receptor neurons and transported axonally to the brain. The uptake of  $Fe<sub>3</sub>O<sub>4</sub>$ -NPs was rapid and detected in the olfactory bulb and other higher olfactory centres. However, the retention times for  $Fe<sub>3</sub>O<sub>4</sub>-NPs$  were shorter compared to  $Fe<sub>3</sub>O<sub>4</sub>-NPs$ , possibly due to different physicochemical properties affecting their transport and clearance. MRI data indicated substantial brain uptake of  $Fe<sub>3</sub>O<sub>4</sub>$ -NPs, though lower than  $Mn_3O_4$ -NPs. The elimination of Fe3O4-NPs from the brain was faster, with clearance mechanisms involving gradual removal from neuronal tissues. Moreover, aging and Parkinson's disease adversely affected their trans-synaptic

movement. These findings open up new possibilities for diagnostic tools, functional neuroimaging, and targeted INDD.

The distribution of a mucoadhesive powder formulation containing  $MnCl<sub>2</sub>$  was visualised in the nasal cavity of cynomolgus monkeys, particularly focusing on the olfactory region [153]. The study explores the effectiveness of an INDD system targeting the olfactory region in cynomolgus monkeys. Three delivery devices were compared: the proprietary nose-to-brain system, an existing Abs-system, and a commercially available Liquid-system. The nose-to-brain system used a proprietary mucoadhesive powder formulation combined with a newly designed nasal device, while the Abs system utilised a Fit-lizer™ type A device, and the Liquid system employed a MAD Nasal™ intranasal mucosal atomisation device. The volumes administered varied across the systems, with the nose-to-brain system delivering 25 mg, 37.5 mg, 50 mg, and 75 mg of powder formulation, whereas the Abs-system and Liquid-system administered 25 mg of powder and 100 μL of liquid formulations, respectively. TR-Dex localised to the olfactory epithelium and reached the olfactory bulb through the cribriform foramina, while MnCl<sub>2</sub> used in Mn-enhanced MRI helped visualise Mn<sup>2+</sup> distribution primarily in the olfactory region and then to the olfactory bulb. The uptake time for the tracer was evaluated at intervals of 20 min, 3 h, 6 h, and 24 h post-IN administration. The nose-to-brain system showed significant Mn<sup>2+</sup> signals in the olfactory region and the olfactory bulb, while TR-Dex was widely observed in the olfactory epithelial cell layer and lamina propria. The retention time for the nose-to-brain system demonstrated sustained Mn2+ signal intensity in the olfactory region for up to 24 h post-administration. The Abs-system also showed  $Mn^{2+}$  signal retention in the olfactory region, though the respiratory region saw a significant reduction in signal intensity after 24 h. In contrast, the Liquid-system showed no significant  $Mn^{2+}$  signal in the olfactory region, indicating rapid clearance of the formulation from the nasal cavity. Quantification of the drug/tracer reaching the brain revealed that the nose-to-brain system significantly increased domperidone uptake in D2R-expressing brain regions. The percentage receptor occupancy maps showed that the nose-to-brain system had the highest domperidone uptake compared to the Abs-system and Liquid-system. The elimination of the tracer from the brain was gradual, with  $Mn^{2+}$  signal intensity decreasing over 24 h. Histological analysis found the carrier to eventually be cleared from the olfactory

mucosa within 6 h post-administration. High-resolution histochemical findings showed that TR-Dex delivered via the nose-to-brain system was extensively distributed in the olfactory epithelial cell layer and lamina propria, with immunostaining revealing its colocalisation with olfactory sensory neuron bundles and fibronectin-positive extracellular matrix. Comparative analysis of imaging agents showed that [18F]fallypride PET imaging demonstrated competitive inhibition in D2R regions, with the nose-to-brain system achieving significantly higher receptor occupancy and a dose-dependent increase in percentage receptor occupancy with higher domperidone dosages. Mn-enhanced MRI imaging confirmed higher distribution and retention in the olfactory region for the nose-to-brain system compared to the other systems. FLI of TR-Dex indicated extensive distribution in the olfactory epithelial cell layer and lamina propria when delivered by the nose-to-brain system. In conclusion, the nose-to-brain system showed superior efficiency in delivering formulations to the olfactory region, enhancing brain uptake, and maintaining retention in targeted brain regions compared to other nasal delivery systems. This study promotes the potential of the nose-to-brain system in achieving effective INDD, supported by high-resolution histochemical and imaging analyses. Here, fluorescence imaging was used to track TR-Dex, to validate the selective distribution and cellular uptake in the olfactory epithelium. These findings point towards a transcellular route via sustentacular cells and paracellular diffusion across the intercellular space as key mechanisms of drug transport in the olfactory region.

#### 2.4.3. Paramagnetic iron oxide

USPIO, SPIO and MPIO are contrast agents that can also be used to monitor INDD to the brain, kidney, spleen, and liver using MRI [154, 155]. When conjugated with astrocyte-derived extracellular vesicles (ADEVs), USPIOs have a unique advantage as they do not alter the cellular physiology or morphology. Intranasally perfused USPIO-ADEVs bypass the BBB through the olfactory and trigeminal pathways, enabling fast and efficient delivery of the product to the CNS [154]. This work investigated the sensitivity and detectability of USPIO-astrocytes and USPIO-ADEVs in a gel matrix. The researchers detected USPIO-ADEVs in the brain as early as one day post-perfusion over a four-day period. Furthermore, the quantity of USPIO-ADEVs increased with each subsequent administration. Additionally, the ADEVs were observed in other

internal organs, including the kidneys, liver, and spleen, indicating a broad biodistribution. Transmission electron microscopy confirmed the presence of USPIO NPs within the astrocytes and the extracellular vesicles.

The study utilised IN delivery to administer MSCs labelled with 111In-oxine and MPIOs to mice. The MSCs were observed to migrate through the cribriform plate into the brain tissue, driven by chemotactic signals from the tumour, particularly the CXCL12/CXCR4 axis, which was upregulated in irradiated tumour tissues [156]. MPIOs-MSCs via IN administration can be transported into the brain through both the olfactory and trigeminal pathways. This facilitated the rapid migration of MSCs towards glioma xenografts. Within hours of administration, the MSCs began migrating towards the tumour sites, with approximately 11% of implanted MSCs detected in the tumour-bearing hemisphere as soon as 2 h post-inoculation. The presence of these cells in the brain was sustained, though significantly decreased by day 11. The MSCs primarily migrated towards the glioma xenografts. MRI and histological analyses demonstrated the presence of labelled MSCs in the basal region of the brain, moving sequentially towards the tumour site. The cells were detected in the olfactory bulbs and frontal lobes, indicating initial entry points and subsequent migration pathways. This migration was enhanced in irradiated tumours, with significant accumulation observed in these regions compared to non-irradiated controls. Balyasnikova *et al.* qualitatively tracked MPIOs-MSCs in a mouse model of glioblastoma. Initially in this research, they used several techniques to validate the viability of IN administration of MSCs to the brain in animals with tumours. Initially, they confirmed that MSCs can migrate within the brain towards tumours, consistent with previous studies [157-159]. Using BLI to show that MSCs can move from the nasal cavity to the brain in mice. Notably, while the luciferase signal was immediately evident in both the nasal cavity and the brain post-IN administration, detecting MSCs in the nasal cavity at later times required longer exposure times, indicating a quick elimination of MSCs. This rapid disappearance was similarly noted in a Parkinson's disease rodent model [160]. It appears that MSCs quickly travel from the nasal cavity to the brain, likely through direct pathways via the cribriform plate, involving the olfactory and trigeminal pathways. Further, the researcher observed that 111In-oxine-MSCs were detectable in the brains of mice. Radiation therapy in tumour-bearing animals seemed to increase MSC penetration into the brain. While MSC proliferation and clearance could vary

between control and irradiated animals, these differences did not seem to influence the results observed 24 h after MSC delivery. Instead, it suggests enhanced brain penetration in irradiated animals. Importantly, they noticed a significant increase in the accumulation of 111In-MSCs in the cerebellum of irradiated animals, which is crucial considering the prevalence of cerebellar brain tumours in children. This emphasises the potential of IN administration of therapeutic stem cells in models of medulloblastoma and other similar tumours. The growing evidence that stem cells can penetrate from nose-to-brain highlights the importance of understanding this mechanism, which could potentially increase the number of stem cells reaching the brain. Lastly, the study introduced the use of MRI to track stem cells modified with MPIOs in the brain. Successfully detected signals from MPIOs-MSCs migrating towards the tumour in both control and irradiated animals following IN administration. This study is the first instance of using MRI to confirm the migration of MSCs delivered intranasally towards brain tumours [156]. MRI images showed the migration path of MPIOs-MSCs toward brain tumours (Figure 11). The movement was monitored through sequential T1-weighted images for both non-irradiated and irradiated mice. The presence of tumours was identified on T2-weighted MRI images in irradiated mice. The MPIO signals,

indicating the cells' location, were clearer in irradiated animals and could be seen moving towards the tumour from the basal region of the brain. Similar observations were made in axial MRI images, where the MPIO signals appeared as hypointensity spots. The authors observed an accumulation of MPIOs-MSCs in the brain and extending towards the tumour 48 h after IN administration. This was evidenced by signal reduction on T2-weighted images and hyperintensity on T1-weighted ones. These findings were further supported by the post-mortem analysis of brain tissue stained with Prussian blue, which confirmed the MSCs' presence at the tumour sites in both control and irradiated animals. This confirms the targeted movement of MSCs delivered intranasally towards tumours within the brain.

An SPIO used to monitor and track IN administration of NSCs through the olfactory epithelium towards inserted glioma xenografts [155]. The delivery device was a micropipette. To optimise the delivery and migration of NSCs to the brain, the study employed methimazole to temporarily disrupt the olfactory epithelium. This pharmacological intervention delayed the clearance of NSCs from the nasal cavity, allowing for prolonged persistence and improved migration into the brain. BLI showed that NSCs were detectable in the nasal cavity within 30 min post-administration, with significant persistence



Figure 11. MRI images of MPIOs-MSCs migration in mice post-IN administration. Panel (A) presents coronal MRI images where images a and b serve as control showing absence of MSCs and tumour, thus no detectable signal. Images c and d show the migration pathway of MSCs in a nonirradiated, tumour-bearing mouse, while e through g display the migration in an irradiated mouse, all through T1-weighted imaging. T2-weighted images h, i, and j from an irradiated mouse highlight both the migrating MSCs, indicated by red arrows, and the tumour, pinpointed by a yellow arrow in j. Panel (B) illustrates axial T1-weighted brain images of an irradiated, tumour-bearing mouse from a to e, with red arrows denoting the direction of MSC movement. These images were captured on the second day following IN administration of MPIOs-MSCs, providing insight into the MSCs' behaviour and their potential therapeutic trajectory towards the tumour site. Adapted with permission from [156], copyright 2014 Elsevier.

observed at 6 and 24 h in methimazole-treated groups. MRI further confirmed enhanced accumulation of NSCs in tumour regions at 24, 48, and 120 h post-administration, demonstrating the effective penetration and retention facilitated by methimazole treatment. The team reported hypointensity and reduced T2 values, confirming the migration of SPIO to the brain. The most significant reduction in T2 values peaked at 120 h post-administration of the SPIO. These findings were validated against histology, confirming the presence of SPIO-NSCs within the tumour and its surroundings. The study identified the olfactory bulb and peri-tumoral brain regions as key uptake areas for the NSCs. Histological analyses using Prussian blue staining confirmed the presence of SPIO-NSCs in these regions, while confocal microscopy revealed increased GFP signals in the tumour areas, indicating successful migration and retention of NSCs in methimazole-treated animals. Additionally, hexon protein staining showed a significantly higher viral load in glioma tissues after NSCs-oncolytic virus treatment in methimazole-treated animals, highlighting the improved delivery of the therapeutic payload. Overall, the study demonstrated that methimazole pretreatment effectively enhances the IN delivery of NSCs to brain tumours by disrupting the olfactory epithelium, thereby delaying NSC clearance and facilitating their migration and retention in glioma tissues. This approach holds promise for improving the therapeutic efficacy of non-invasive treatments for malignant brain tumours, offering a potential platform for future clinical applications.

Chastkofsky *et al.* demonstrated that upon IN administration, these SPIOs transport through the trigeminal pathway to the xenografts in the brainstem [161]. The study used a patient-derived xenograft model of diffuse intrinsic pontine glioma in nude mice. MRI was employed to trace and quantify SPIO-MSCs following IN delivery. The MSCs were administered intranasally and observed to migrate along the trigeminal nerve towards the diffuse intrinsic pontine glioma xenografts located in the brainstem. MRI imaging, which tracked the MSC's movement, confirmed this pathway. The MSCs were found to migrate to the tumour within 48 h post-IN, with the most concentrated hypointense signals detected in the tumour region at this time point, indicating that MSCs reach the tumour site within 2 days. At later time points (5 days post-IN), a more diffuse hypointense signal was observed in the brainstem. The MSC's presence around the tumour was further confirmed by Prussian Blue staining and immunofluorescence. Immunofluorescence with

antibodies specific for CD105 (a marker for MSCs) and H3-K27M (a mutation-specific for the diffuse intrinsic pontine glioma xenograft) confirmed the presence of MSCs around the tumour 5 days post-IN. These findings supported the MRI results, confirming the localisation and migration of MSCs to the tumour site.

In a different study, SPIO was used to monitor IN administration of MSCs to asses stem cell therapies for brain injuries and diffuse intrinsic pontine glioma [162]. The objective was to evaluate the feasibility and efficacy of this method in a traumatic brain injury mouse model using MRI. The MSCs-SPIO NPs tagged with FITC and delivered intranasally using a pipette. The total volume of 18 µL was administered. The SPIO NPs served as the imaging agents, enabling the non-invasive tracking of the labelled MSCs via MRI. T2\*-weighted MRI was employed for its sensitivity in detecting the hypointense areas indicative of SPIO presence. This method allowed real-time visualisation and longitudinal tracking of the MSCs as they migrated toward the site of cortical injury. The study observed that SPIO-MSCs were detectable as strong hypointense areas on T2\*-weighted images as early as 24 h post-delivery. These areas were predominantly located medial to the cortical injury, suggesting targeted migration of the MSCs. Importantly, these hypointense signals persisted up to 14 days post-delivery, confirming prolonged retention of the labelled cells at the injury site. While the study successfully visualised the presence of SPIO-MSCs at the injury site via MRI, it did not provide specific quantitative data on the exact percentage of the administered cells that reached the brain. The focus was more on demonstrating the feasibility and tracking capability rather than precise quantification. To corroborate the MRI findings, high-resolution histochemical analysis was performed. Prussian blue staining confirmed the presence of SPIO-MSCs in brain tissue sections, revealing blue-coloured iron deposits in the labelled cells. Additionally, FITC fluorescence microscopy further validated these findings by showing FITC-tagged SPIO-positive cells in the injured cortex, correlating well with the MRI results. These histochemical techniques provided robust evidence supporting the non-invasive imaging findings. The study highlighted the effectiveness of using T2\*-weighted MRI for tracking SPIO-MSCs, noting its sensitivity and accurate association with histological findings. This method provided clear and reliable data on the biodistribution and migration of the MSCs within the brain.

Simorgh *et al.* conjugated olfactory ecto-MSCs with Alginate (Alg)-coated SPIOs to target the striatum in the brain [163]. The cells utilised both the olfactory and trigeminal nerve pathways to reach the CNS. The magnetic targeting cell delivery method significantly enhanced this process. By using magnetic forces to direct and concentrate the cells to the desired brain regions, the approach minimised cell loss during transit and increased the efficiency of cell delivery. The MRI scans were pivotal in tracing the perfusion of Alg-SPIO olfactory ecto-MSCs. Images captured before and two days after cell infusion revealed significant findings, indicating that the initial uptake and detectable presence of the cells in the brain occurred within this timeframe. The primary uptake region was the striatum, a critical area affected by Parkinson's disease. This was confirmed through MRI imaging, which showed a reduction in signal intensity in the striatum region, indicating the presence and concentration of the labelled stem cells. The study also demonstrated long-term retention of the stem cells. MRI evaluations showed the presence of labelled cells in the brain up to eight weeks post-administration. Histological analyses, including Nissl staining, confirmed the survival of dopaminergic neurons in the substantia nigra for this duration, showcasing the effectiveness of the stem cell integration. The use of magnetic forces significantly increased the number of cells that reached the lesion site. As such, this study highlights the potential of combining IN administration with magnetic targeting for delivering stem cells to the brain. The approach shows great promise for treating Parkinson's disease by ensuring efficient delivery, substantial retention, and functional integration of stem cells in the affected brain regions. The findings, confirmed by MRI and histochemical analyses, demonstrate the effectiveness of this novel therapeutic strategy.

In Kou *et al.*'s study on IN administration of NPs for CNS delivery, PEGylated ultrasmall Fe<sub>3</sub>O<sub>4</sub> NPs served as a dual-modality tracer, utilising both MRI and fluorescence imaging [164]. These NPs, designed to enhance MRI contrast and tagged with the fluorescent dye (Cy5.5), were monitored to understand their distribution and eventual elimination in the rat brain. The MRI data highlighted a dynamic distribution of the NPs, particularly in the olfactory bulb, pons, and cerebellum, revealing temporal variations in their uptake. Notably, the olfactory bulb showed increased MRI signals as early as 5 min post-administration, while the pons and cerebellum exhibited similar increases around 15 min post-administration. This variation in timing suggests different pathways and rates of NP penetration into various brain regions. Complementing the MRI data, fluorescence imaging provided a more detailed view

at the tissue level, confirming that the trigeminal and olfactory nerves are significant pathways for the NPs to reach the CNS. The study showed that the NPs primarily travelled along these nerves to enter the brain, reaching the subarachnoid space and CSF. Once in the CSF, the NPs circulated throughout the cerebral cistern, allowing them to spread to other brain regions. The elimination of the NPs from the brain followed the CSF circulation, eventually being excreted via the arachnoid villus and superior sagittal sinus into the venous system. The maximum accumulation of Fe<sub>3</sub>O<sub>4</sub> NPs in the brain was 5.97 μg/g at 1 h post-administration, indicating a significant initial uptake followed by clearance within a few hours. High-resolution histochemical findings were confirmed through fluorescence microscopy and Prussian blue staining. Fluorescence microscopy of brain slices showed that the NPs were mainly accumulated in the meninges. Prussian blue staining validated the presence of Fe3O4 NPs in the connective tissues of the trigeminal nerve and brain. No intracellular transport within the axons of olfactory neurons was observed, indicating paracellular transport through the nerve sheaths. Notably, the NPs traverse through the perineural space, ultimately infiltrating the brain parenchyma, and provide valuable insights into effective CNS drug delivery mechanisms, potentially enhancing treatment strategies for CNS diseases.

Law and colleagues (2023) developed a CT contrast agent using iohexol-loaded liposomes with >10% PEG to monitor the nose-to-brain tract using chemical exchange saturation transfer (CEST) MRI [165]. The choice of iohexol was due to its ability to produce a distinct CEST contrast at 4.3 ppm on the Z-spectrum and its single CEST property, preventing readout overlaps when tracking multiple components of the *in vivo* delivery system. Which was detectable by CEST MRI due to specific amide protons. *In vitro*, these liposomes generated significant CEST contrasts at 4.3 ppm, 3.4 ppm, and 1.2 ppm. After IN administration, signal increases in CEST contrast were observed in the olfactory bulb and frontal lobe with the 10% PEG formulation, especially prominent at 4.3 ppm. Contrarily, the 1% PEG formulation showed no notable increase. The brain showed a 62% increase in CEST at 4.3 ppm in the olfactory bulb and a 40% increase in the frontal lobe. Additionally, CEST contrast at -3.4 ppm indicated a 11% increase in the olfactory bulb and 9% in the frontal lobe. The estimated dose of iohexol delivered from the olfactory bulb to the frontal lobe was 57%, while 87% of the liposomes were delivered, indicating partial release of iohexol from the liposomes. Elimination of the tracer

involved partial release of iohexol, with approximately 36% released within the first hour. It was suggested that elimination likely occurs through the CSF and systemic absorption. Histochemical findings validated the MRI results, with a high rhodamine signal observed in the olfactory bulb and frontal lobe of mice treated with 10% PEG iohexol liposomes. This signal was significantly higher than in mice treated with 1% PEG iohexol liposomes, supporting the MRI findings of higher uptake and retention with the 10% PEG formulation. The histological analysis confirmed the distribution of liposomes in specific brain regions, particularly in CSF-rich areas like the external plexiform layer and the region between the left and right olfactory bulbs. The study concluded that the multiple CEST contrast approach is effective for non-invasively detecting and monitoring the delivery of liposome-based drugs from the nose to the brain. The findings emphasise the importance of PEG coating and particle size in overcoming nasal mucus barriers and achieving significant brain uptake. This approach provides valuable insights for optimising the INDD, highlighting the potential of CEST MRI for guiding the design and monitoring of such systems. The distinctive CEST MRI contrasts can be used to aid in non-invasively tracking the nose-to-brain delivery of liposomes.

A study by Seino *et al.* examined the efficacy of IN administration of SPIO as a contrast agent for magnetic particle imaging [166]. They focused on enhancing the delivery and targeting of these NPs to specific brain regions. SPIOs were modified by compositing them with AuNPs, facilitating additional surface modification via gold-sulfur bonds without compromising their magnetic properties. PEG was used to coat the NPs to prevent aggregation and nonspecific adsorption. The NPs were further functionalised with tracer molecules called ABC 595 designed to target Aβ. The resulting tracer, ABC-PEG-Au/FcM, consisted of Ferucarbotran Magnetic (FcM), AuNPs, PEG coating, and the ABC 595 probe molecule. *In vivo* experiments involved administering these PEGylated magnetic NPs intranasally to mouse models. The study employed superconducting quantum interference device measurements to track the migration and accumulation of NPs in the brain. Results showed that approximately 0.3-0.6% of ID successfully reached the brain. The NPs migrate through the nasal mucosa to the olfactory bulb, subsequently accumulating at the target site Aβ plaques, confirmed through fluorescence microscopy. This selective targeting is significant for imaging applications in Alzheimer's

disease. Additionally, these NPs maintained their magnetic properties and did not ionise during migration. Over time, the NPs detached from the target sites, suggesting a mechanism for their clearance from the brain, which is beneficial from a safety perspective. The study concluded that IN administration is an effective method for delivering magnetic NPs into the brain, providing a non-invasive alternative to bypass the BBB. These NPs also retain their magnetic properties upon delivery and accumulate at target sites, offering promise for early diagnosis and monitoring of neurological conditions.

# 2.4.4. Remarks on MRI

MRI has become a crucial imaging modality in neuroscience, particularly in monitoring INDD. Its ability to differentiate between various soft tissues with high contrast makes it an invaluable tool for detecting diseases and evaluating therapeutic outcomes. In the context of nose-to-brain pathways, MRI provides real-time, non-invasive visualisation and tracking of drug movement, typically achieved using contrast agents involving metals.

The contrast agents, such as Gd-based chelates, USPIO, MPIOs, SPIONs, and Mn-based agents, significantly affect the relaxation times of tissues, altering T1 and T2 relaxation times for enhanced imaging. For instance, studies have shown that Gd agents can access the brain rapidly post-IN administration, with changes in T1 relaxation times observable within 30 min. A notable study by Yadav *et al.* compared the uptake of Gd encapsulated NE (Gd-NE) and aqueous solution of Gd-DTPA in the rat brain, revealing significant uptake in various brain regions and suggesting transport through both trigeminal and olfactory pathways.

MRI has also been instrumental in tracking Mn2+-based agents, typically taken up by active neurons, allowing for studies on brain function and neuronal connections. However, neurotoxicity concerns around the use of Mn2+ have led to the investigation of Mn2+-based NPs and chelates. These agents, when given intranasally, show dynamic and quantitative data on trans-synaptic translocation and nose-to-brain migration, with visual stimulation enhancing the transport of  $Mn^{2+}$  from the olfactory bulb to the visual cortex.

In addition to what may be considered as traditional MRI contrast agents, studies have also utilised USPIO, SPIO, and MPIO. For example, USPIO conjugated with astrocyte-derived extracellular vesicles demonstrated the ability to bypass the BBB through the olfactory and trigeminal pathways, leading to efficient CNS delivery. Similarly, MPIOs

have been used to track the IN administration of MSCs to the brain, highlighting the potential of this route for drug delivery to specific brain regions.

These MRI studies offer clear insights into the dynamics of drug transport and distribution in the brain following IN administration. The ability to visualise and track these processes in real time has significant implications for the development of targeted therapies for CNS disorders, providing a better understanding of the potential and limitations of INDD.

# **2.5. Focused ultrasound**

Research using FUS in combination with IN administration has made significant strides over the last decade. FUS is a non-invasive therapeutic technology that uses frequencies above human hearing and up to 18 Mhz to target tissue deep in the body without incisions or radiation. Its technique has been gaining traction as a potential tool for enhancing INDD. FUS leverages the mechanical and thermal effects produced by US waves to transiently open the BBB, enabling targeted delivery of therapeutic agents [167-169]. To ensure the effective and safe translation of US technologies in drug delivery to clinical applications, Isaac's theranostics review outlines crucial guidelines for clinical translation [170].

#### 2.5.1. FUS and microbubbles

FUS was primarily spearheaded by Chen *et al.* and Ye *et al.* conducted studies [171-173]. The method entails the IV delivery of microbubbles succeeded by the utilisation of FUS to induce an opening of the BBB [168, 169]. The process of FUS involves the external transmission of US pulses through the skull, which are then focused onto a small area, usually measuring a few millimetres, located in the subcortical structures [174]. In their seminal 2014 and 2016 studies, Chen H *et al.* developed an innovative strategy that leverages FUS to enhance the delivery of IN administration drugs to specific brain regions. They provided proof-of-concept for delivering a model drug (Dex) and brain-derived neurotrophic factor (BNDF) to the caudate putamen of mouse brains. Their approach achieved drug delivery efficiency comparable to the nose-to-brain alone, indicating that FUS could potentially enhance the nose-to-brain delivery through a process of active pumping [171, 172].

# 2.5.2. Gold nanoclusters

In 2018, a study by Ye *et al.* used FUS combined with microbubble-mediated IN delivery (FUS+IN) to administer gold nanoclusters (AuNCs) to the brain [173], as shown in Figure 12. The IN-delivery device involved administering 24 µL of AuNCs. The AuNCs

used in the study had a hydrodynamic size of  $5.60 \pm$ 1.50 nm. The delivery pathway for the AuNCs was through the nasal route, specifically targeting the olfactory and trigeminal nerve pathways. This approach led to the NPs bypassing the BBB and directly entering the brain. FUS-induced microbubble cavitation further enhanced the transport of these NPs to the targeted brain regions. This mechanism, referred to as the "microbubble pump effect," is similar to the perivascular pump effect and enhances the distribution of intranasally administered agents to the FUS-targeted brain location. The uptake time of the AuNCs was evaluated using various imaging techniques. PET/CT imaging and *ex vivo* gamma counting were performed 1 h post-administration to assess the biodistribution of [64Cu]AuNCs. The data indicated that the amount of radioactive tracer was >10-fold higher in the blood, lungs, liver, spleen, kidneys, and heart when IV injection was used, compared to IN administration. The study found significant fluorescence in the brainstem at 1 h post-administration, indicating successful uptake of the NPs. The primary uptake region identified was the brainstem, specifically the left side targeted by FUS. Comparisons were made between the FUS-treated side and the contralateral side to evaluate the effectiveness of the technique. The retention time of the NPs was studied as well. Enhanced fluorescence signals in the trigeminal nerve were observed 1 h post-administration, which significantly diminished by 24 h, suggesting the clearance of the NPs from the nerve. This rapid clearance indicated that the NPs did not remain in the nerve tissue for extended periods, reducing potential toxicity risks. The study used two types of imaging agents: [64Cu]AuNCs and TR-labelled AuNCs (TR-AuNCs). The distribution of these agents was evaluated using multiple imaging modalities. PET/CT imaging provided whole-body biodistribution data, autoradiography assessed the spatial distribution of [64Cu]AuNCs in brain slices, and fluorescence microscopy examined the distribution of TR-AuNCs in the brainstem. The findings revealed a significantly higher accumulation of both [<sup>64</sup>Cu]AuNCs and TR-AuNCs in the FUS-targeted brain regions compared to the non-targeted regions, demonstrating the effectiveness of FUS in enhancing localised delivery. Quantification of the drug/tracer reaching the brain showed that FUS+IN significantly increased the delivery efficiency. The radioactivity of [64Cu]AuNCs in the FUS-targeted brainstem was 2.72 times higher than IN delivery alone. Similarly, the fluorescence intensity of TR-AuNCs in the FUS-targeted region was 2.32 times higher than in the

non-targeted side. These results indicated that FUSIN effectively enhanced the localised delivery of NPs to the brain. The elimination of [64Cu]-AuNCs was characterised by minimal presence in the blood and major organs after IN administration. That is, the NPs were predominantly cleared via mucosal removal to the nasopharynx and subsequent accumulation in the stomach and intestines for fecal excretion. Fluorescence signals from TR-AuNCs in the trigeminal nerve diminished by 24 h, indicating effective clearance from the nerve tissue. Histochemical analysis was conducted to assess any potential tissue damage resulting from the FUS+IN treatment. The study found no histological-level tissue damage in the nose, trigeminal nerve, or brain. Hematoxylin and eosin staining of nasal tissue and LFB-CV staining of the trigeminal nerve showed no changes compared to the control groups. These findings highlight the short-term safety of FUS+IN, with no observed histological damage following treatment, and the study supports the potential of FUS+IN as a promising technique for non-invasive, spatially targeted, and safe delivery of NPs to the brain with minimal systemic exposure. The combination of PET/CT, autoradiography, and fluorescence microscopy provided comprehensive insights into the distribution, retention, and clearance of the NPs in the brain. Furthermore, the study aligns with previous research findings that IN administration delivery of larger NPs, such as micelles with a diameter of approximately 600 nm, is associated with minimised systemic exposure [175]. This study extends understanding on NP biodistribution by demonstrating that small-size AuNCs share with larger size NPs the advantage of minimising systemic exposure. This comparison on the versatility of NP size in achieving reduced systemic toxicity through targeted delivery methods offers valuable insights into the design of safer nanotherapeutics. Notably, this appears to be the only study employing [64Cu] in the context of nose-to-brain. Future research should continue to validate these findings through direct quantification methods and long-term safety evaluations, including histochemical analyses to confirm imaging results. This group further expanded the potential applications of FUS+IN administration in two separate studies. They explored the effectiveness of the method in concentrating the delivery of anti-programmed cell death ligand 1 (aPD-L1) antibody to the brain labelled with near-infrared fluorescent dye (IR800), and in facilitating adeno-associated viral delivery (AAV5) labelled with EGFP to the brain. Both studies yielded highly encouraging results, reaffirming the potential of FUS-assisted nose-to-brain delivery as a versatile, efficient method for targeted drug delivery within the CNS [176, 177].

#### 2.5.3. Remarks on FUS

The mechanism of FUS+IN administration is indeed an active area of research. The nose-to-brain delivery method utilises the olfactory and trigeminal nerves ending in the nasal cavity as direct routes for medication delivery into the brain. The most likely transport mechanism from the nose-to-brain is bulk transport along the channels surrounding these nerves, a conclusion drawn from the rapid speed of transport observed. After reaching brain entry points, the IN administration agents are then distributed throughout the brain along the cerebral perivascular spaces. This distribution is aided by the perivascular pump effect, where the heartbeat-driven pulsations of the blood vessel wall drive the agents through these spaces. This innovative approach combines the precision of FUS in opening the BBB and the direct drug delivery advantages of the IN administration method. The existing series of studies have opened new frontiers in non-invasive therapeutics and hold value for the treatment of various CNS disorders having localised distributional characteristics.



**Figure 12.** Demonstrating FUS+IN administration of NP and the potential pathways from nose-to-brain. "Reproduced with permission from [173], copyright 2018 ELSEVIER".

### **2.6. Computed tomography**

CT provides detailed anatomical pictures by combining X-ray technology and tomographic image reconstruction. Despite its high-resolution imaging capability, CT's role in the nose-to-brain pathway is relatively limited due to the scarce contrast agents available and its low sensitivity and lack of soft tissue contrast [178]. However, its value significantly increases when combined with modalities such as PET or SPECT, offering a more comprehensive understanding of nose-to-brain applications. The CT scan supplies detailed anatomical images for reference, while the PET or SPECT scan provides dynamic information. This combination can offer complementary insights into the contrast agent's delivery process and its distribution profile [178].

### 2.6.1. Gold nanoparticles

The application of AuNPs as contrast agents in CT imaging has been explored in the INDD pathway primarily because AuNPs enhance the visibility of the administered drugs and their biodistribution. In 2018, Betzer *et al.* addressed a key challenge in exosome research tracking these vesicles *in vivo* [179]. They labelled exosomes derived from MSCs with AuNPs and successfully used *in vivo* micro-CT imaging to track the labelled exosomes in an animal model of acute striatal stroke. They found an accumulation of the exosomes in the murine model's brain within 1 h of administration, with a concentration at the stroke site apparent after 3 h. These exosomes remained at the lesion site for up to 24 h, while in contrast, the control group showed no specific exosome accumulation and clearance from the brain began after 24 h. These findings suggest AuNPs could serve as effective markers for tracking drug distribution and have potential for use in brain theranostics.

In 2019, Salem *et al.* expanded on this work by investigating brain-targeted resveratrol delivery via nose-to-brain pathway lipid vesicles labelled with AuNPs [180]. The AuNPs enabled validation of the cellular uptake of the formulated nanomaterials in the brain. Particularly, transferosomes displayed superior permeation rates and a higher fluorescence intensity compared to NEs, thereby demonstrating a significant improvement in behaviour acquisition and spatial memory function in amnesic rats. The authors successfully demonstrated AuNP accumulation in the brains of all CT-treated rats (see Figure 13). By comparing the CT images, they could distinctly identify bright regions in the brain, reflecting the accumulation of AuNPs (indicated by red arrows). These brightly illuminated areas are distinguishable from the surrounding tissues due to the X-ray

attenuation induced by the AuNPs. CT images could distinguish specific bright cerebral areas that represent the accumulation of AuNPs.

Later, Bekhet *et al.* formulated citicoline-loaded niosomes that were labelled with AuNPs with the goal of improving the management of epilepsy disorders [181]. They used CT imaging to examine brain uptake and cellular translocation of this formulation. The results of the study indicated that a low dosage of IN administration citicoline-loaded niosomes in a soluble gel had a protective effect, significantly extending the latency period before the onset of convulsions. Importantly, the CT images displayed high attenuation of the X-ray beam in different brain regions to indicate AuNP accumulation. The bright patches on the CT scans suggest that IN administration of the AuNP-labelled citicoline-loaded niosomes effectively bypassed the BBB and reached various regions of the brain.

### 2.6.2. Remarks on CT

Collectively, studies using CT underscore the potential of AuNPs as contrast agents in the INDD pathway. By integrating AuNPs with various delivery vehicles such as exosomes, transferosomes, and niosomes, the targeted delivery of therapeutics can be traced and validated. AuNPs can, by bypassing the BBB, reach various regions of the brain effectively. This approach not only enhances the efficacy of therapeutic agents, but also provides a powerful tool for real-time tracking and verification of drug delivery through CT imaging. As such, AuNPs promises to be an indispensable asset in optimising the nose-to-brain pathway, boosting the potential of IN administration for improved disease diagnosis, monitoring, and therapy. However, the exact time it takes for AuNPs to reach these regions, their retention time in the brain, and their distribution across the brain can vary significantly and depend on multiple factors. These factors include the specific formulation used (e.g., exosomes, transferosomes, or niosomes), the disease model, and the specific imaging technique used. As for whether AuNPs can provide full brain coverage in nose-to-brain delivery, more research is needed to form a conclusive view. Given the complex and heterogeneous nature of the brain, it may be challenging for any single delivery system to achieve complete brain coverage. However, AuNPs certainly show promise in enhancing the reach and efficacy of therapeutically active agents in various brain regions. Further research is necessary to fully understand the kinetics, biodistribution, and clearance of AuNPs in nose-to-brain delivery and to optimise their use for different therapeutic agents and disease states.



**Figure 13.** Demonstrates CT images depicting the uptake of AuNPs in the brain. The images compare three scenarios: (A) an untreated rat, (B) a resveratrol transferosome<br>capped with AuNPs treated rat, and (C) a resveratrol [180], copyright 2019 Taylor & Francis.









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### **3. Methods**

#### **3.1. Literature search methods**

Our literature search adhered to the PRISMA-S guidelines [200]. We conducted a comprehensive search of PubMed, Scopus, Embase, and Web of Science, covering publications from 1989 to 2024 (see Figure 14). The initial phase involved the removal of duplicate studies from the database search results. Subsequently, we screened the remaining articles by examining titles and abstracts. The final stage entailed a thorough review of the full texts of these articles to determine their relevance and suitability for inclusion in our review.

#### **3.2. Literature search strategy**

The search strategy was designed to encompass a broad range of studies pertinent to our review. Utilising the resources of the University of Queensland library, we searched the PubMed, Scopus, Embase, and Web of Science databases using a combination of specific and alternative terms. Our search query was as follows:

("nose to brain" OR nose-to-brain OR intranasal OR intranasally OR "intranasal drug delivery" OR intramucosal) AND (Imaging OR Image OR "Magnetic resonance Imaging" OR MRI OR "Positron emission tomography" OR PET OR "Single photon emission computerised tomography" OR SPECT OR "Computed tomography" OR CT OR "Optical Imaging" OR OI OR "Fluorescence imaging" OR FLI OR "Bioluminescence imaging" OR BLI OR PET OR PET-CT OR PET-MRI OR PET) NOT (sinuses OR sinusitis OR facial OR respiratory OR sedation OR lung).

The literature search included studies published in English employing one or more imaging techniques, such as MRI, PET, SPECT, CT, OI, FLI, BLI, or combinations like PET-CT or PET-MRI, where the imaging agent was administered intranasally. Studies were excluded if they were review articles, brief communications, or abstracts lacking detailed methodology. Additionally, studies published in languages other than English or those focusing on IN administration for purposes other than brain imaging were not considered.

#### **3.3. Literature search results**

The results of our literature search are summarised in the figure below and structured according to the PRISMA-S guidelines [200].



**Figure 14.** Diagram of literature search based on PRISMA-S guidelines.

# **4. Conclusions**

The exploration of IN administration as a route for delivering imaging agents to the brain has been significantly advanced by diverse imaging modalities, each offering unique insights into the dynamics of agent delivery, distribution, and nasal cavity-to-brain interactions. The choice of imaging modality, contrast agent, and formulation like NEs, MEs, and NPs play a pivotal role in determining the efficiency, specificity, and safety of the IN administration process. PET Imaging is a cornerstone in understanding IN administration's molecular mechanisms. Utilising radioisotopes such as 18F and 11C, PET imaging is particularly valuable, as it offers a detailed view of the metabolic processes and direct transport of imaging agents within brain region parenchyma.

In examining the formulations used in these studies, the role of NE, ME, and NPs is particularly noteworthy. These engineered formulations enhance the bioavailability and brain targeting of imaging agents. NEs, for instance, demonstrate superior permeation rates and have been effectively employed in tracking agents like Gd and various drugs. MEs, with their unique composition, aid in stabilising and delivering imaging agents, as seen in studies involving citicoline-loaded niosomes. NPs, particularly AuNPs, have revolutionised CT imaging, offering enhanced visibility and precise tracking of drug distribution.

Each imaging modality and formulation offers distinct advantages and challenges. PET and SPECT are highly sensitive and provide rapid detection, making them suitable for real-time monitoring. OI, with its cellular-level resolution, is invaluable in toxicity studies and detailed tissue distribution analysis. MRI, with its superior resolution and non-invasive nature, is ideal for detailed anatomical and functional studies, while CT's value lies in its anatomical clarity, especially when combined with PET or SPECT.

The exploration of IN administration for imaging agents is a multidimensional field that benefits significantly from the diverse capabilities of these imaging modalities. The synergy between these techniques and the careful selection of appropriate formulations like NE, ME, and NPs are crucial in advancing our understanding of IN administration's potential in brain imaging and drug delivery. As this field evolves, further studies are needed to optimise the various imaging modalities and formulations, with the positive outcome of enhancing their efficiency, specificity, and safety in clinical applications.

The exploration of IN administration for delivering imaging agents to the brain is a growing field that has significantly advanced our understanding of drug delivery mechanisms, particularly where bypassing of the BBB is important. This review considered the SPECT, PET, OI, MRI, FUS and CT imaging modalities. The integration of these imaging techniques has offered unprecedented insights into the pharmacokinetics, biodistribution, and efficacy of IN administration. Advancements to date are pivotal not only in tracking and validating the efficacy of drug delivery but also in understanding the interaction of the agents used in accessing the brain tissues. Moreover, the review underscores the potential of IN administration in offering a non-invasive, efficient, and potentially safer alternative for drug delivery to the brain.

The ability of IN administration to bypass the BBB opens new avenues for treating neurological disorders, enhancing the efficacy of therapeutics, and reducing systemic side effects. It also paves the way for innovative diagnostic and therapeutic strategies, particularly in the context of neurodegenerative diseases, brain injuries, and brain tumours.

The field of IN administration in neuroimaging and drug delivery is on the brink of transformative advancements. Central to these developments is the need for new tracers and contrast agents optimised for the nose-to-brain pathway, focusing on enhanced targeting capabilities to improve efficacy. The integration of multiple imaging modalities promises a more comprehensive analysis, combining the strengths of each technique for a holistic understanding of drug dynamics in the brain.

Advancements in NP technology is poised to play a pivotal role, with multifunctional NPs offering exciting possibilities in improving the delivery and visualisation of imaging agents. These NPs, trackable by various imaging modalities and capable of carrying therapeutic payloads, are ripe for exploration. Safety and efficacy remain paramount, with long-term studies crucial for understanding the systemic effects and clearance mechanisms of IN administration agents. Innovations in delivery systems that can precisely regulate dosage, penetration, and release of imaging agents will further enhance the precision of IN administration.

In conclusion, the integration of advanced imaging modalities and innovative formulations in IN administration research has opened new frontiers in neurophysics and pharmacology. Figure 15 visually summarises the contributions of each imaging modality, elucidating the pathways and tracers used in the exploration of INDD.



**Figure 15.** Comparative analysis of the imaging modalities in INDD to the Brain. This figure presents six pie charts, each representing a different imaging modality utilised to track<br>the migration of specific tracers from routes. These visualisations underscore the diverse mechanisms and efficiency of nose-to-brain delivery pathways across different imaging agents. [created with BioRender.com].

This representation not only complements the detailed remarks within this review but also highlights the practical application and comparative benefits of these technologies in enhancing our understanding of drug dynamics in the brain. Table 2 provides a consolidated overview of the extensive work conducted in this field, underscoring the multi-dimensional nature of this research and its potential to revolutionise CNS diagnostics and therapeutics. This approach offers hope for more effective delivery of treatments for psychiatric and neurological disorders, with non-invasive techniques that may better target and mitigate these complex conditions. Future research is essential to address the highlighted challenges and to fully realise the potential of nose-to-brain delivery of imaging agents. Continued advancements in this area are crucial for improving the precision, efficiency, and safety of these delivery systems, ultimately enhancing patient outcomes in clinical settings. While the current review provides a comprehensive overview of the strengths and weaknesses of various imaging modalities and their applications in INDD, future research should also consider the development and optimisation of delivery devices and methods. These advancements are essential for maximising the efficacy of nose-to-brain delivery systems. Innovative delivery devices, such as micro-needles, nebulisers, and advanced nasal sprays, can significantly enhance the precision and efficiency of intranasal administration. Furthermore, the integration of real-time monitoring systems within these devices could provide immediate feedback on the delivery process, ensuring optimal dosing and minimising potential side effects. By focusing on both the imaging techniques and the delivery mechanisms, future studies can provide a more holistic approach to INDD, ultimately improving therapeutic outcomes and patient safety.

**Table 2.** Summary of key research findings on INDD systems for enhanced brain targeting, monitored by medical imaging: a systematic review of imaging agents, modalities, objectives, and achievements.

	Section Imaging agents	<b>Modalities</b>	time/ scan window time	Brain retention Validation method	Objectives	Achievements	Study
2.1.1	[99mTc]	SPECT/Gam 8 h ma		Biodistribution study.	To prepare and evaluate MEs containing zolmitriptan for rapid drug delivery to the brain. It also aimed to characterise these ME and assess their biodistribution in rats through different administration routes.	The result demonstrated substantial uptake of the drug into the brain following IN administration of the zolmitriptan mucoadhesive ME. This imaging evidence supports the effectiveness of the IN administration of zolmitriptan mucoadhesive ME for targeted drug delivery to the brain.	Vyas et al. (2005) $[24]$
2.1.1	$[^{99m}Tc]$	SPECT/Gam 8h ma		Biodistribution study & transmission electron microscopy.	To develop and assess clonazepam MEs for INDD, specifically targeting the treatment of acute status epileptic patients.	The study indicated that the IN administration of clonazepam mucoadhesive ME resulted in superior brain uptake of the drug, as evidenced by higher brain/blood uptake ratios and clear visualisation in rabbit brain scintigraphy imaging.	Vyas et al. (2006) [26]
2.1.1	$[^{99m}Tc]$	SPECT/Gam 8 h ma		Biodistribution study & electron microscopy.	To formulate ME of sumatriptan and sumatriptan succinate and compare their effectiveness in delivering the drug rapidly to the brain during acute migraine attacks.	The imaging results confirmed that sumatriptan succinate was uptaken in the brain more efficiently when administered through IN administration of both formulated MEs than in other forms and delivery methods.	Vyas et al. (2006) $[27]$
2.1.1	$[^{99m}Tc]$	SPECT/Gam 8 h ma		Biodistribution study.	To formulate an NE containing risperidone for targeted INDD drug delivery. Aimed to compare the biodistribution and brain localisation of NE-containing risperidone in different formulations for IV injection and IN administration in Swiss albino rats.	The result demonstrated rapid and extensive transport of the drug to the brain, and bypassing the BBB. This was evident from higher brain/blood uptake ratios and drug transport efficiency when compared to the other formulations and administration routes.	Kumar et al. (2008) $[28]$
2.1.1	[99mTc]	SPECT/Gam 4 h ma		Biodistribution study.	To prepare and characterise cabergoline IN administration ME formulations for effective brain drug delivery and assess their performance in weight control.	Imaging confirmed the localisation of the cabergoline in the brain following IN, IV, and oral administrations. The [99mTc]cabergoline formulations showed higher brain/blood uptake ratios, drug targeting efficiency, and direct drug transport to the brain via IN administration compared to IV injection or oral administration.	Sharma et al. (2009) $[29]$
2.1.1	[99mTc]	SPECT/Gam 8h ma		Biodistribution & pharmacokinetic studies.	To evaluate the drug biodistribution of thiolated chitosan NPs of tizanidine hydrochloride after IN administration.	Results showed that the brain uptake of the drug was significantly enhanced after the thiolation of chitosan, indicating improved transnasal permeation.	Patel et al. (2012) $[30]$
2.1.1	[99mTc]	SPECT/Gam 8h ma		Biodistribution study.	To evaluate the effectiveness of direct nose-to-brain delivery of tacrine, a cholinesterase	The study demonstrated that IN administration of tacrine when compared to IV injection, resulted in faster delivery to the brain, higher	Jogani et al. (2007) $[32]$


















Abbreviations: [<sup>11</sup>C]: carbon-11; [<sup>18</sup>F]: fluorine-18; [<sup>3</sup>H]: tritium; [<sup>68</sup>Ga]: gallium-68 [Mn]: manganese; [<sup>64</sup>Cu]: copper-64; [<sup>86</sup>Rb]: rubidium-86; [<sup>89</sup>Zr]: zirconium-89; [<sup>99mTc</sup>]: technetium-99m; [<sup>111</sup>In]: Indium-111; [<sup>123</sup>I]: Iodine-123; [<sup>125</sup>I]: Iodine-125; [<sup>131</sup>I]: Iodine-125; [<sup>201</sup>I]: thallium-201; AAV5: adeno-associated virus serotype 5; ADEVs: astrocyte-derived extracellular vesicles; Aβ: amyloid β; Alg: Alginate; AuNCs: gold nanoclusters; AuNRs: gold nanorods; BBB: blood-brain barrier; BLI: bioluminescence imaging; BODIPY: boron-dipyrromethene; BSA: bovine serum albumin; CB: cholera toxin B subunit; CEST: chemical exchange saturation transfer; CNS: central nervous system; CSF: cerebrospinal fluid; CSA: cyclosporine A; CT: computed tomography; CTG: CellTracker Green; Cy: cyanine dye; Ct: cholera toxin; Cou-6: coumarin-6; DIPG: diffuse intrinsic pontine glioma; DTPA: diethylenetriamine pentaacetate; Dex: dextran; DiR: 1,1'-dioctadecyl-3,3,3',3'-tetramethylindotricarbocyanine iodide; EGFP: enhanced green fluorescent protein; ENT1: equilibrative nucleoside transporter 1; FBP: Fas-signaling blocking peptides; FDG: fluorodeoxyglucose; FDS: 2-deoxy-2-fluorosorbitol; FITC: fluorescein isothiocyanate; FLI: fluorescence imaging; FLUC: firefly luciferase; FLT: fluorothymidine; Fe3O4: iron oxide; FUS: focused ultrasound; AuNP: gold nanoparticle; Gd<sup>+3</sup>: gadolinium ion; GFP: green fluorescent protein; GM1: monosialoganglioside; IGF-1: Insulin-like growth factor-1; IFN-β1b: interferon-β1b; INDD: intranasal drug delivery; IN: intranasal; IR780: near-infrared dye 780; IV: intravenous; LDH: layered double hydroxide; LPNPs: lipid-polymer nanoparticles; MEs: microemulsions; MRI: magnetic resonance imaging; MIL-100: a type of metal-organic framework; MSCs: mesenchymal stem cells; MTX: methotrexate; Mn2+: manganese ion; miRNA: micro RNA; MPIOs: micron-size paramagnetic iron oxide; NOTA: ((1,4,7-triazanonane-1,4,7-triyl) triacetic acid); NE: nanoemulsion; NAC: N-acetyl-L-cysteine; NPs: nanoparticles; NSCs: neural stem cells; OI: optical imaging; PET: positron emission tomography; PHT: phenytoin; PLGA: polylactic-co-glycolic acid; PS-YG: polystyrene nanoparticles labelled with yellow green; QDs: quantum dots; SPIONs: superparamagnetic iron oxide nanoparticles; SV2A: synaptic vesicle glycoprotein 2A; siRNA: small interfering RNA; SPECT: single photon emission computed tomography; TR-Dex: Texas Red dextran; USPIO: ultrasmall superparamagnetic iron oxide; VEGF: vascular endothelial growth factor; WGA: wheat germ agglutinin; bFGF: basic fibroblast growth factor; hNSCs: human neural stem cells; polyGIONs: polyfunctional gold-iron oxide NPs.

## **Abbreviations**

[11C]: carbon-11; [18F]: fluorine-18; 2D: two dimensional; 3D: three dimensional; [3H]: tritium; 5-HT<sub>1B/1D</sub>: subtypes of serotonin receptors; [<sup>68</sup>Ga]: gallium-68; [51Mn]: manganese-51; [52Mn]: manganese-52;  $[54Mn]$ : manganese-54;  $54MnCl<sub>2</sub>$ : manganese-54 dichloride; [64Cu]: copper-64; [86Rb]: rubidium-86; [89Zr]: zirconium-89; [99mTc]: technetium-99m; [111In]: Indium-111; [123I]: Iodine-123; [125]]: Iodine-125; [131]: Iodine-131; [201Tl]: thallium-201; 201TlCl: thallium-201 chloride; AAV5: adeno-associated virus serotype 5; ADEVs: astrocytederived extracellular vesicles; AlF-NODA-dLVT: aluminum fluoride-NODA-oxytocin derivative; Aβ: amyloid β; Alg: alginate; AuNCs: gold nanoclusters; AuNRs: gold nanorods; AUC: area under the curve; BBB: blood-brain barrier; BLI: bioluminescence imaging; BNDF: brain-derived neurotrophic factor; BODIPY: boron-dipyrromethene; BSA: bovine serum albumin; CB: cholera toxin B subunit; CEST: chemical exchange saturation transfer; CHC: hydroxycinnamic acid; CNS: central nervous system; CNT: concentrative nucleoside transporters; CSF: cerebrospinal fluid; CSA: cyclosporine A; CF: carbonised framework; CT: computed tomography; CTG: CellTracker Green; Cy: cyanine dye; CdSe-ZnS: cadmium selenide-zinc sulfide; Ct: cholera toxin; Cou-6: coumarin-6; C<sub>max</sub>: maximum concentration; DIPG: diffuse intrinsic pontine glioma; DSF-INEG: disulfiram-loaded ion-sensitive nanoemulsion gel; DTP%: direct transport percentage; DTPA: diethylenetriamine pentaacetate; DTE%: drug targeting efficiency percentage; Dex: dextran; DiR: 1,1'-dioctadecyl-3,3,3',3'-tetramethylindotricarbocyani ne iodide; EPO: erythropoietin; EGFP: enhanced green fluorescent protein; ENT1: equilibrative nucleoside transporter 1; FA-TEPA: folic acid tetraethylenepentamine conjugate; FDG: fluorodeoxyglucose; FDS: 2-deoxy-2-fluorosorbitol; FBP: Fas-signaling blocking peptides; FcM: Ferucarbotran Magnetic; FITC: fluorescein isothiocyanate; FLI: fluorescence imaging;

FLUC: firefly luciferase; FLT: fluorothymidine; Fe: iron; Fe3O4: iron oxide; FUS: focused ultrasound; AuNP: gold nanoparticle; Gd+3: gadolinium ion; GFP: green fluorescent protein; GM1: monosialoganglioside; IGF-1: Insulin-like growth factor-1; IFN-β1b: interferon-β1b; ID/g: injected dose per gram; INDD: intranasal drug delivery; IN: intranasal; IR780: near-infrared dye 780; IV: intravenous; LDH: layered double hydroxide; LPNPs: lipid-polymer nanoparticles; MEs: microemulsions; MRI: magnetic resonance imaging; MIL-100: a type of metal-organic framework; MSCs: mesenchymal stem cells; MTX: methotrexate; Mn<sup>2+</sup>: manganese ion; miRNA: micro RNA; MPIOs: micron-size paramagnetic iron oxide; NOTA: ((1,4,7-triazanonane-1,4,7-triyl)triacetic acid); NBMPR: nitrobenzylmercaptopurine riboside; NE: nanoemulsion; NMDA: N-methyl-D-aspartate; NAC: N-acetyl-L-cysteine; NPs: nanoparticles; NSCs: neural stem cells; OI: optical imaging; PET: positron emission tomography; PHT: phenytoin; PLA–PEG–DSPE: polylactic acid –polyethylene glycol–distearoylphosphatidylethanolamine; PLGA: polylactic-co-glycolic acid; PRISMA-S: Preferred Reporting Items for Systematic Reviews and Meta-Analyses extension for Scoping Review; PS-YG: polystyrene nanoparticles labelled with yellow green; QDs: quantum dots; R8-YAβ(25–35)-PEI: polyarginine-amyloid beta peptide-polyethyleneimine; SPIONs: superparamagnetic iron oxide nanoparticles; SV2A: synaptic vesicle glycoprotein 2A; siRNA: small interfering RNA; SUV: standard uptake value; SPECT: single photon emission computed tomography; TACs: time-activity curves;  $T_{\text{max}}$ : time to reach maximum; TR: Texas Red; TT: tetanus toxoid; USPIO: ultrasmall superparamagnetic iron oxide; VEGF: vascular endothelial growth factor; WGA: wheat germ agglutinin; bFGF: basic fibroblast growth factor; cpm: counts per minute; hNSCs: human neural stem cells; polyGIONs: polyfunctional gold-iron oxide NPs.

## **Authorship contributions**

AA, HP, BP and VV contributed to the

conception and design of the work, the acquisition, analysis, or interpretation of data, and are accountable for all aspects of the work, ensuring that questions related to the accuracy or integrity of the work are appropriately investigated and resolved. KT helped draft and review the manuscript for its criticality. All authors provided approval for publication of the final version.

## **Competing Interests**

The authors have declared that no competing interest exists.

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