Supplementary figures



Figure S1 Molecular and morphological identification of GSC and DGCs. A Representative images of immunofluorescence staining showing the stemness markers expression of GSCs. Scale bars = 100 μ m. B Representative images of the cellular morphology of DGCs. Scale bars = 50 μ m. C Representative images of immunofluorescence staining showing the differentiational markers expression of DGCs. Scale bars = 50 μ m. D Western blotting analysis of stemness markers on GSCs and DGCs.



Figure S2 Exosomal circCMTM3 promotes proliferation and migration of DGCs and is corelated with angiogenesis pathway. A, B RT-qPCR assays displaying the expression of circCMTM3 in GSCs (A) and GDEs (B) after overexpression or knockdown. C, D MTS assays reveling the cell viabilities of DGC01 (C) and DGC03 (D) by incubating with different groups of GDEs. E, F Representative images (E) visualized in Hstudio and quantification (F) of the DGC03 migration ability (n = 5). Scale bars = 50 μ m. G Correlation of exosomal circCMTM3 expression with angiogenesis pathway genes. Data are presented as means \pm SD (three independent

experiments). *p < 0.05; **p < 0.01; ***p < 0.001; ns, no significance.



Figure S3 Exosomal circCMTM3 upregulates STAT5A/B expressional level via inhibiting the ubiquitin- proteasomal pathway. A-D RT-qPCR assays showing the mRNA expression of STAT5A and STAT5B in CNOT4 -overexpressed or silencing DGCs. E, F CircCMTM3 can bind to STAT5A and STAT5B proteins via CatRAPID prediction. G-J RT-qPCR assays displaying the mRNA expression of STAT5A and STAT5B in DGCs with treatment of different groups of GDEs. K-N Co-IP assays illustrating interaction efficiency of CNOT4 and STAT5A (K, L) as well as CNOT4 and STAT5B (M, N) in DGCs under the condition of exosomal circCMTM3 knockdown. O-R Western blotting analysis showing STAT5A and STAT5B expression in CNOT4-silenced DGCs with treatment by GDEs containing downregulated circCMTM3 and CHX (50 µg/ml) (O, Q), meanwhile, quantitative analysis on STAT5A and STAT5B expression half-life time (t1/2) reflecting degradation rates (P, R). S, T Western blotting assays showing STAT5A (S) and STAT5B (T) expression in CNOT4silenced DGC01 treated with or without MG-132(50 µM) after incubating with circCMTM3-downregulated GDEs. Data are presented as means ± SD (three independent experiments). *p < 0.05; **p < 0.01; ***p < 0.001; ns, no significance.



Figure S4 Exosomal circCMTM3 interacts with STAT5A, STAT5B and JAK2 respectively in DGCs. A-C LC-MS/MS spectrum showing the STAT5A, STAT5B and JAK2 peptides pulled down by circCMTM3 probe respectively. D-F RNA pull-down assay to validate the interaction between exosomal circCMTM3 and JAK2, STAT5A, and STAT5B respectively in DGC03. G-L RIP assays showing anti-STAT5A (G, J), anti-STAT5B (H, K) and anti-JAK2 (I, L) treatment caused exosomal circCMTM3 enrichment in DGC03 by treatment with different groups of GDEs. Data are presented as means \pm SD (three independent experiments). *p < 0.05; **p < 0.01; ***p < 0.001; ns, no significance.



Figure S5 Exosomal circCMTM3 induces VM formation in vitro via upregulating p-STAT5A expression. A, B MTS assays assessing the cell viabilities of DGC01 (A) and DGC03 (B) incubated with circCMTM3-EV GDEs and circCMTM3-OE GDEs with DMSO or Stafia-1. C, D EdU assays (C) and quantified analysis (D) showing the proliferation of DGC01 and DGC03 with the indicated treatments. Scale bars = 100 μ m. E, F Representative photographs (E) and quantification (F) of the Transwell assay of DGCs from each group. Scale bars = 50 μ m. G, H Representative photographs (G) and quantification of relative migration distance (H) exhibiting the migration ability of DGC01 and DGC03 detected by HoloMonitor and visualized in Hstudio (n = 5) in each group. Scale bars = 50 μ m. I-K Representative images (I) and quantification (J, K) showing tube formation of DGCs. Scale bars=100 μ m. Data are presented as means ± SD (three independent experiments). *p < 0.05; **p < 0.01; ***p < 0.001; ns, no significance.



Figure S6 Exosomal circCMTM3 activates expression of CHI3L2 in DGCs. A Representative images of immunofluorescence staining showing the expression and subcellular distribution in DGCs with knockdown or overexpression exosomal circCMTM3 treatment. Scale bars = 50 μ m. B Correlation analysis of STAT5A and CHI3L2 based on TCGA-glioma, CGGA325 and CGGA693 datasets. C ChIP analyses of the STAT5A-bound promoter regions of CHI3L2. The corresponding isotype IgG was used as a negative control. D Quantification of ChIP assay showing the STAT5A binding sites in the CHI3L2 promoters in DGC03 treated with circCMTM3-OE GDEs with or without Stafia-1intervention. E, F ChIP-qPCR detecting H3K4me (E) and H3K27ac (F) enrichment in the CHI3L2 promoters in DGC03 with indicated intervention. G The Dual-luciferase reporter assays revealing the luciferase promoter activities of CHI3L2 with treatment by different group GDEs and Stafia-1 or not in DGC03. Data are presented as means ± SD (three independent experiments). *p < 0.05; **p < 0.01; ***p < 0.001; ns, no significance.



Figure S7 Exosomal circCMTM3 mediates resistance to anti-angiogenic therapy by promoting VM formation in GBM. A, B Representative H&E staining (A) and correspond quantification (B) displaying tumor size in brain slices from different experimental groups (n = 5), Scale bars = 1 mm. C Kaplan–Meier analysis of mice from the indicated groups (n = 5). D Representative H&E staining images demonstrating the morphological characteristics of GBM. Double staining showing the VM formation assessed by PAS and anti-CD31 immunohistochemical staining in tumor tissue. Red arrows indicating the VM tubular structures with PAS⁺/CD31⁻. Scale bars = 20 µm. E Assessment of VM scores in each group. Data are presented as means \pm SD (three

independent experiments). *p < 0.05; **p < 0.01; ***p < 0.001; ns, no significance.