Supporting Information

Mesenchymal stem cell-derived extracellular vesicles targeting irradiated intestine exert therapeutic effects

Ningning He^{1#}, Mingxin Dong^{2#}, Yuxiao Sun^{1#}, Mengmeng Yang¹, Yan Wang¹, Liqing Du¹,

Kaihua Ji¹, Jinhan Wang¹, Manman Zhang¹, Yeqing Gu¹, Xinran Lu¹, Yang Liu¹, Qin Wang¹,

Zongjin Li³, Huijuan Song^{1*}, Chang Xu^{1*} and Qiang Liu^{1*}

¹Tianjin Key Laboratory of Radiation Medicine and Molecular Nuclear Medicine, Institute of Radiation Medicine, State Key Laboratory of Advanced Medical Materials and Devices, Chinese Academy of Medical Science and Peking Union Medical College, Tianjin 300192, China.

²State Key Laboratory of Experimental Hematology, National Clinical Research Center for Blood Diseases, Haihe Laboratory of Cell Ecosystem, Institute of Hematology & Blood Diseases Hospital, Chinese Academy of Medical Sciences & Peking Union Medical College, Tianjin, 300020, China.

³School of Medicine, Nankai University, Tianjin, China.

[#]These authors contributed equally to this work.

*Correspondence: Qiang Liu, Institute of Radiation Medicine, Chinese Academy of Medical Sciences & Peking Union Medical College, Tianjin Key Laboratory of Molecular Nuclear Medicine, Tianjin, 300192, China. Ph:+86-22-8568-3008, Fax: +86-22-8568-3033, E-mail: <u>liuqiang@irm-cams.ac.cn</u>; Chang Xu, Institute of Radiation Medicine, Chinese Academy of Medical Sciences & Peking Union Medical College, Tianjin Key Laboratory of Molecular Nuclear Medicine, Tianjin, 300192, China. E-mail: <u>xuchang@irm-cams.ac.cn</u>; Huijuan Song, Institute of Radiation Medicine, Chinese Academy of Medical Sciences & Peking Union Medical College, Tianjin Key Laboratory of Molecular Nuclear Medicine, Tianjin, 300192, China. E-mail: <u>songhuijuan@irm-cams.ac.cn</u>.

Supporting Information Contents:

Table S1

Figures S1-S10

Table S1. Primer sequences	used for real-time RT-PCR.
----------------------------	----------------------------

Gene	Forward primer $(5^{\circ} \rightarrow 3^{\circ})$	Reverse primer $(5' \rightarrow 3')$	
GADPH	AGGTCGGTGTGAACGGATTTG	TGTAGACCATGTAGTTGAGGTCA	
IL-1β	GCAACTGTTCCTGAACTCAACT	ATCTTTTGGGGTCCGTCAACT	
IL-6	TAGTCCTTCCTACCCCAATTTCC	TTGGTCCTTAGCCACTCCTTC	
TNF-α	CCTGTAGCCCACGTCGTAG	GGGAGTAGACAAGGTACAACCC	
IL-10	GCTCTTACTGACTGGCATGAG	CGCAGCTCTAGGAGCATGTG	
IL-22	AATGAAAAGGCCCCCAAGGTAGT	GTCGTTTCCGCAACAAGTCCTCT	
	TATCC	тс	
Stat3	GCCATCCTAAGCACAAAGCC	GGGAATGTCGGGGTAGAGGT	
c-Myc	GACTGTATGTGGAGCGGTTTCT	TGCTGTCGTTGAGCGGGTA	
Survivin	ATCGCCACCTTCAAGAACTG	AATCAGGCTCGTTCTCGGTA	
VEGF	AACGATGAAGCCCTGGAGTG	TGAGAGGTCTGGTTCCCGA	
Bcl-2	GATGACTGAGTACCTGAACCG	CAGAGACAGCCAGGAGAAATC	
SOCS3	GGAGAGCGGATTCTACTGGA	TGACGCTCAACGTGAAGAAG	
Reg3b	ACTCCCTGAAGAATATACCCTCC	CGCTATTGAGCACAGATACGAG	

Reg3g	ATGCTTCCCCGTATAACCATCA	
Reg3g	ATGCTTCCCCGTATAACCATCA	



Figure S1. Ex vivo imaging of NIR dyes in radiation-induced intestinal injury mice. There

showed no radiation dose-dependent of NIR dyes distribution in intestines.



Figure S2. *Ex vivo* **imaging of MSC-EVs in radiation-induced intestinal injury mice.** (A) Distributions of NIR-labeled EVs at 48 h after NIR-EVs administration. (B) Distributions of NIR-labeled EVs at 72 h after NIR-EVs administration.



Figure S3. Radiation increase cellular uptake of MSC-EVs. (A) Representative immunofluorescence images showing uptake of PKH26-labeled MSC-EVs by MODE-K cells under different irradiation doses. Scale bar, 25 μ m. (B) Graph showing the percentage uptake of fluorescent MSC-EVs (red) by MODE-K cells. The results showing radiation increases the binding of MSC-EVs to irradiated MODE-K cells. ***P* < 0.01 *vs*. 0 Gy, ^{##}*P* < 0.01 *vs*. 2 Gy, n=3.



Figure S4. Effects of MSC-EVs on DNA-damage Responses. (A and B) Representative images of γ H2AX (green) and quantitative data. Scale bar, 100 µm. (C and D) Representative images of comet assay and quantitative data. MSC-EVs ameliorated DNA damage after radiation. Scale bar, 100 µm. ***P* < 0.01 *vs*. EV, ****P* < 0.001 *vs*. EV, #*P* < 0.05 *vs*. IR+PBS, ###*P* < 0.001 *vs*. IR+PBS, n=3.







Figure S6. The effect of MSC-EVs on the protection of intestinal cells in irradiated mice. (A) Immunohistochemistry of Lysozyme in intestine tissues. Scale bar, 100 μ m. (B) qRT-PCR of Lysozyme, and the expressions of regeneration-associated genes (Reg3b and Reg3g) were increased in the MSC-EV group. ***P* < 0.01 *vs*. Ctrl, ****P* < 0.001 *vs*. Ctrl, #*P* < 0.05 *vs*. IR+PBS, ##*P* < 0.01 *vs*. IR+PBS, n=3.



Figure S7. Immunohistochemistry of TUNEL in intestine tissues. MSC-EVs significantly reduced the apoptosis level in the intestine of mice after radiation.



Figure S8. (A) Statistical histogram of differential genes. Red represents up-regulated genes, blue represents down-regulated genes, and the ordinate represents the number of differential genes. (B) Western blot of Akt and ERK pathway related protein expression. There was no obvious difference among three groups. (C) Western blot of Stat3 pathway related protein expression. The Stat3 pathway was inactivated after blocked the PS on cell membrane and neutralized the MFGE8 protein on the MSC-EVs.



Figure S9. (A) qRT-PCR analysis of the intestinal tissue showed that MSC-EVs treatment significantly increased the level of miRNA-455-5p in the intestinal tissue. ^{**}P < 0.01 vs. Ctrl, n=3. (B) Quantification analysis of the SOCS3 protein in Figure 8F. ^{##}P < 0.01 vs. EV-inhibitor^{NC}, n=3.



Figure S10. (A) Western blot of Stat3 pathway related protein expression. p-Stat3 and VEGFR1 protein level were inhibited after treatment with EV-miR-455-5p inhibitor. ${}^{\#}P < 0.05$ *vs.* EV- inhibitor^{NC}, ${}^{\#}P < 0.01$ *vs.* EV-inhibitor^{NC}, n=3.