- **Supplementary information** 1
- Targeted degradation of VEGF with bispecific aptamer-based LYTACs 3 ameliorates pathological retinal angiogenesis 4
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Table S1

Name	Sequence (5'-3')
M6PR aptamer	GGCGCGTAGATGACGAGCAGTCCTAACATCGTTTAG
	GAC
VEGE antamer-1	TGTGGGGGTGGACTGGGTGGGTACC
VEGF aptamer-2	TGTGGGGGTGGACGGGCCGGGTAGA
VEGF aptamer-3	GCCCGTCTTCCAGACAAGAGTGCAGGGC
M6PR-A for V1, V2	CGTAAATCAGTCATAGGGCGCGTAGATGACGAGCA
and V3	GTCCTAACATCGTTTAGGAC
M6PR-A for V4, V5	<i>P</i> -
and V6	CCCCCACACGTAAATCAGTCATAGGGCGCGTAGATG
	ACGAGCAGTCCTAACATCGTTTAGGAC
VEGF-A1 for V1	TATGACTGATTTACGTGTGGGGGGGGGGGACTGGGTGGG
	TACCC
VEGF-A2 for V2	TATGACTGATTTACG <u>TGTGGGGGGTGGACGGGCCGG</u>
	GTAGA
VEGF-A3 for V3	TATGACTGATTTACGGCCCGTCTTCCAGACAAGAGT
	GCAGGGC
VEGF-A1 for V4	TATGACTGATTTACG <u>TGTGGGGGG</u> TGGACTGGGTGGG
	TACCC
VEGF-A2 for V5	TATGACTGATTTACGTGTGGGGGG <u>TGTGGGGGGTGGAC</u>
	GGGCCGGGTAGA
VEGF-A3 for V6	TATGACTGATTTACGTGTGGGGGGGGCCCGTCTTCCAG

ACAAGAGTGCAGGGC The sequences of aptamers (M6PR aptamer and VEGF aptamer) are underlined. The linkers (15-bp and 23-bp) of the corresponding VED-LYTACs (V1-V6) are labeled with

red color.



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Figure S1. Construction of VED-LYTACs and evaluation of its ability to degrade 22 VEGF. (A) Denatured polyacrylamide gel analysis of the V4, V5 and V6. (B) ELISA 23 assay for analysis the VEGF level from bEND.3 cells treated with different 24 concentrations of V5 for 12 h (n = 3 independent experiments). (C) ELISA assay for 25 analysis the VEGF level from bEND.3 cells treated with V5 for 6 h or 12 h (n = 326 independent experiments). (D and E) Quantifications of fluorescence intensity for 27 lysosome colocalization analysis in HUVECs treated with Cy5-labeled individual 28 29 aptamers or V5. (F and G) Quantification of fluorescence intensity for lysosome colocalization analysis in HUVECs treated with Cy5-labeled V5, along with GFP-30 M6PR (F) or FAM-A1 aptamer (G) to label M6PR or VEGF proteins. 31

- 32 Data are presented as mean \pm SEM. **p < 0.01, ***p < 0.001, ****p < 0.0001.
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Figure S2. VED-LYTACs inhibit cell migration in bEND.3 cells. (A-C) Representative images (A) and quantifications of the relative migration rate (B, n = 30areas from 3 independent experiments) and percentage of wound closure (C, n = 30areas from 3 independent experiments) for the scratch migration assay in bEND.3 cells treated with different concentrations of V5. Scale bar, 80 µm.

41 Data are presented as mean \pm SEM. ****p < 0.0001.







Figure S3. A2 aptamer exhibits less efficacy in inhibiting angiogenesis compared 45 to V5. (A-C) Representative images (A) and quantifications of the relative migration 46 rate (B, n = 30 areas from 3 independent experiments) and percentage of wound closure 47 48 (C, n = 30 areas from 3 independent experiments) for the scratch migration assay in HUVECs treated with different concentrations of A2. Scale bar, 80 µm. (D-F) 49 Representative images (D) and quantifications of the branch points (E, n = 3) 50 independent experiments) and the tube length (F, n = 3 independent experiments) for 51 52 tube formation assay in HUVECs treated with different concentrations of A2. Scale bar, 53 200 µm. (G-I) Representative images (G) and quantifications of the total sprout length (H, n = 50-80 sprouts from 3 independent experiments) and numbers (I, n = 12 spheroids 54 55 from 3 independent experiments) for sprouting assay from HUVECs treated with

56 different concentrations of A2. Scale bar, 30 μ m. (J-L) Representative images (J) and 57 quantifications of the neovascular length (K, n = 30 fields from 3 independent 58 experiments) and the sprout numbers per field (L, n = 30 fields from 3 independent 59 experiments) for sprouting assay from mice aortic ring treated with different 60 concentrations of A2. Scale bar, 300 μ m.

Data are presented as mean \pm SEM. *p < 0.05, **p < 0.01, ***p < 0.001; ns, not significant.

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Figure S4. The biosafety of VED-LYTACs in an ocular application. (A and B) 65 Photomicrographs (A) and quantification (B) of the retinal histology assessed by 66 hematoxylin and eosin (H&E) staining in control and V5-injected mice (6 μ M, n = 1267 mice from three independent experiments). ONL, outer nuclear layer. (C-E) ERG 68 recordings (C) and measurement of retinal a-wave (D, n = 24 eyes from 3 independent 69 experiments) and b-wave (E, n = 24 eyes from 3 independent experiments) amplitudes 70 for control and V5 (6 μ M)-injected mice under scotopic conditions at 3 cd s m⁻² flash 71 intensity. 72

73 Data are presented as mean \pm SEM. # and ns, not significant.



75 76 Figure S5. Anti-angiogenic effect of V5 in vivo. (A) Images of mice injected with 2 µM of Cy5-V5 for the indicated times (left) or different concentrations of Cy5-V5 for 77 48 h (right) using the in vivo imaging system. (B) ELISA assay for analysis the VEGF 78 level from retinas treated with 6 μ M of V5 for indicated times (n = 3 independent 79 experiments). (C) Quantification of the percentage of the neovascular area in retinas 80 from control or VEGF-injected mice treated with different concentrations of V5 (n =81 30 fields from 3 independent experiments). (D) Quantification of the percentage of 82 RBC leakage in retinas from control or VEGF-injected mice treated with different 83 concentrations of V5 (n = 3 independent experiments). (E) Quantification of the relative 84 claudin-5 intensity in retinas from control or VEGF-injected mice treated with different 85 concentrations of V5 (n = 3 independent experiments). 86

B7 Data are presented as mean \pm SEM. *p < 0.05, **p < 0.01, ***p < 0.001, ****p < 0.001, ****p < 0.0001.



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Figure S6. Anti-angiogenic effect of V5 in OIR mouse models. (A) 91 Immunofluorescence images of the retinal vasculature stained with anti-CD31 in retinas 92 from control or OIR mice. Scale bar, 30 µm. (B) ELISA assay for analysis the VEGF 93 94 level in retinas from control or OIR mice treated with different concentrations of V5 (n = 3 independent experiments). (C) Quantification of the percentage of the neovascular 95 area in retinas from control or OIR mice treated with different concentration of V5 (n 96 97 = 30 fields from 3 independent experiments). (D) ELISA assay for analysis the VEGF 98 level in retinas from control or STZ-induced diabetic mice treated with different concentrations of V5 (n = 3 independent experiments). 99

100 Data are presented as mean \pm SEM. ***p < 0.001, ****p < 0.0001.

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