Figure S1. Fibroblast subtype identification. (A) Dot plot showing marker gene expression of major cell types. (B) UMAP shows the sample type and individual sample distribution of all cells. (C) Stacking plot shows the proportion of major cell types in peripheral blood and five tissue samples. (D) UMAP shows the sample type and individual sample distribution of fibroblasts. (E) Feature plot shows the marker gene expression of selected fibroblast subtypes. (F) Heatmap showing sample preference of fibroblast subtypes, where OR > 1.5 was considered significantly enriched for that cell in that type of sample, and OR < 0.5 was considered significantly not enriched. (G) Volcano plot showing the difference in the proportion of major cell types in ICC (n = 31) versus AL (n = 14). (H) Volcano plot comparing the relative abundance of fibroblast subtypes in ICC versus HCC. (I) Heatmap showing the Ucell enrichment scores of key biological entries in the Fb 03 FAP subtype in different tissue types. (J) Bar plot shows the proportion of FAP + CAF from TCGA samples after deconvolution by CIBERSORTx. Paired point plot shows the high proportion of FAP + CAF in the paired tumor samples from the single-cell discovery cohort. *, P < 0.05; **, P < 0.01; ***, P < 0.001; ****, P < 0.0001.

Figure S2. Functional analysis of fibroblast subtypes. (A) Dot plot showing selected biological terms or pathways significantly enriched for each fibroblast subtype. (B) Heatmap shows the enrichment scores of relevant metabolic pathways for fibroblast subtypes calculated by R package scMetabolism; bar plot shows the overall metabolic score of each fibroblast subtype, with the vertical coordinate being the value after scaling of metabolic score. (C) Heatmap showing the enrichment of 50 key cancer hallmarks in different fibroblast subtypes.

Figure S3. Comparison of Scissor cell-related genes and fibroblast subtypes. (A) Volcano plot showing genes differentially expressed between Scissor ⁺ and Scissor ⁻ cells. Genes with $|\log FC| > 2$, adj. P < 0.05 were considered significantly different. (B) Dot plot showing the expression preference of genes highly expressed in Scissor ⁺ or Scissor ⁻ cells in HCC and ICC.

Figure S4. Fluorescent expression of FAP in tumor and paracancer. Immunofluorescence images show that FAP is more abundantly expressed in tumor samples compared to paracancer. Viewed by SlideViewer with field of view sizes of 2000 μ m and 400 μ m.

Figure S5. Correlation and co-localization analysis of fibroblasts with macrophages. (A) Heatmap showing Spearman's correlation of the proportion of fibroblasts infiltrating with other major cell types in five independent bulk transcriptome cohorts based on CIBERSORTx deconvolution imputation. Scatterplots show the correlation between the proportion of fibroblasts infiltrated with macrophages. (B) Heatmap showing the correlation of spatial localization between cells based on R package Cards imputed in HCC and ICC ST slides, high correlation indicates high spatial localization between cells. (C) Cell score ST plot showing the presence of spatial co-localization of imputed fibroblasts and macrophages.

Figure S6. Identification of key TAMs. (A) UMAP shows the distribution of sample type and individual sample in macrophage subtypes. (B) Dot plot shows the expression of macrophage subtype marker genes in different subtypes (left); tissue enrichment preference shows the heatmap of macrophage subtypes (middle); stacking plot shows the relative proportion of macrophage subtypes in different tissues (right). (C) RNA velocity analysis indicates that Mph DAB2 and Mph SPP1 are terminally differentiated cell types. (D) Heatmap showing the Ucell enrichment score of macrophageassociated pathways in different macrophage subtypes. (E) Scatter plot showing the differentially expressed genes of DAB2 + TAM (Mph 03) versus SPP1 + TAM (Mph 04). (F) KM curves show that tumor patients with high Mph 03 or Mph 04 score calculated by ssGSEA had worse overall survival. (G) Bar plot showing the proportions of DAB2 + /SPP1 + TAMs after deconvolution by CIBERSORTx based on TCGA samples. (H) Immunofluorescence images confirmed that DAB2 + TAM and SPP1 + TAM were enriched in HCC and ICC tumor samples, respectively. Scale bars are 20 µm and 200 µm.

Figure S7. Gene expression and spot annotation of ST boundary slides. (A) Spatial feature plot showing spatial expression of FAP + CAF and selected macrophage subtype marker genes. (B) Unbiased clustering of ST spots in HCC3 and HCC4 slides and cell type annotation for each cluster. Dot plots showing the expression of specific marker genes for each cluster in HCC3 and HCC4 slides. (C) Dot plot showing the results of GO enrichment analysis of the co-localized regions of FAP + CAF and TAM.

Figure S8. Spatial distribution of immune cells. (A and B) Malignant spots (Mal, red), boundary spots (Bdy, blue), and

non-malignant spots (nMal, orange) were annotated on the tissue slides by R package Cottrazm. (C and D) Feature plots showing the expression of selected immune cell-related genes. (E and F) Box plots showing the proportion of immune cells in the annotated regions.

Figure S9. FAP + CAF recruit macrophages and promotes M2 polarization. (A) Heatmap showing the prior interaction potential of ligand and receptor from TAM to FAP + CAF. The dots represent genes significantly associated with survival of TCGA-LIHC patients (Cox P < 0.05), red represents better prognosis (HR > 1) and blue color represents worser prognosis (HR < 1). (B) Spatial dot plot showing the spatial expression of *PDGFB* in *DAB2* ⁺TAM and corresponding receptor genes in FAP + CAF in HCC1 L and HCC2 L slides. (C) Circos plot shows the weights of signal sent by FAP + CAF to other cell types in HCC or ICC; heatmap shows the weights of signaling exchange between all cell types. (D) Signal enrichment analysis based on cellular communication presents the strength of efferent signaling enrichment pathways of different cell types in HCC or ICC. (E) Heatmap shows the prior interaction potential of ligand and receptor from FAP + CAF to TAM; dot plot shows the expression of ligand and receptor genes in fibroblast subtypes and macrophage subtypes. (F) Dot plot showing the pathways to which FAP + CAF ligand genes are significantly enriched.

Figure S10. Cell communication of FAP + CAF in ICC. (A) Black heatmap shows significant top ligands and receptors between FAP + CAF and tumor or endothelial cells; red heatmap shows growth factor-associated ligands and receptors. (B) UMAP and dot plot showing high expression of *VEGFB* in fibroblasts of ICC samples. (C) KM curve showing that ICC patients with high *VEGFB* expression have shorter OS. (D) Scatter plot showing *VEGFB* is significantly positively correlated with *ADM* receptor *RAMP1* and *CALCRL*. (E) Spatial dot plot showing the spatial expression of *ADM* in *SPP1* ⁺TAM and *RAMP1* and *CALCRL* in *FAP* ⁺CAF. (F) Conjecture of communication between *SPP1* ⁺TAM, *FAP* ⁺CAF, endothelial cells, and tumor cells in ICC.

Figure S11. Drugs predicted to block TAM-*FAP* + CAF interaction. (A) Dot plot showing the targeted drugs with the high correlation with the LRscore based on TCGA-LIHC samples. A higher positive correlation means that the drug is more sensitive for patients with a low LRscore, and a negative correlation means that the drug is more sensitive for patients with a high LRscore. (B and C) Bar plot showing the small molecule drugs predicted by the R package sc2MeNetDrug to block TAM-*FAP* + CAF communication, where a high negative enrichment score represents a higher likelihood that the drug will work; the network plot shows clustering based on drug structure.

Figure S12. Single-cell and bulk pan-cancer analysis of FAP + CAF and DAB2 + TAM. (A and B) UMAP show the identification of FAP + CAF and DAB2 + TAM and associated gene expression from the integrated pan-cancer single-cell cohort. (C) Bar plot showing the proportion of FAP + CAF and DAB2 + TAM infiltration based on gene FAP and DAB2 expression grouping. (D) KM curves showing that high FAP

or *DAB2* gene expression is associated with worse OS in pancancer patients. (E) KM curve showing that patients with high *FAP* and *DAB2* gene expression had the shortest OS. (F) KM curves for 39 cancers show that patients with high *FAP* and *DAB2* gene expression usually predict shorter OS. (G and H) UMAP showing identification of *FAP* ⁺ CAF and *DAB2* ⁺ TAM from an integrated pan-cancer immunotherapy singlecell cohort; dot plot showing selected subtype-specific gene expression.

Figure S13. Spatial distribution of FAP and DAB2 at the boundaries of multiple cancers. Tissue slides were annotated by malignant spots (Mal, red), boundary spots (Bdy, blue), and non-malignant spots (nMal, orange), including lung (LUAD, GSM5420751), adenocarcinoma renal cell carcinoma (RCC, GSM5924036), medulloblastoma (MB, EGAS00001006124), pancreatic ductal adenocarcinoma (PDAC, GSM6505134), squamous cell carcinoma (SCC, V10F24 015 A1, doi.org/10.17632/2bh5fchcv6.1), ependymoma (EPN, GSM5844724), colorectal cancer (CRC, P6, HRA000979), head and neck squamous cell carcinoma (HNSC, GSM5494476), ovarian cancer (OV, Human Ovarian 11 Capture Area from 10x), Cancer. prostate mm adenocarcinoma (PRAD, EGAS00001006124), gastrointestinal stromal tumor (GIST, GSM6177607).

Figure S1.

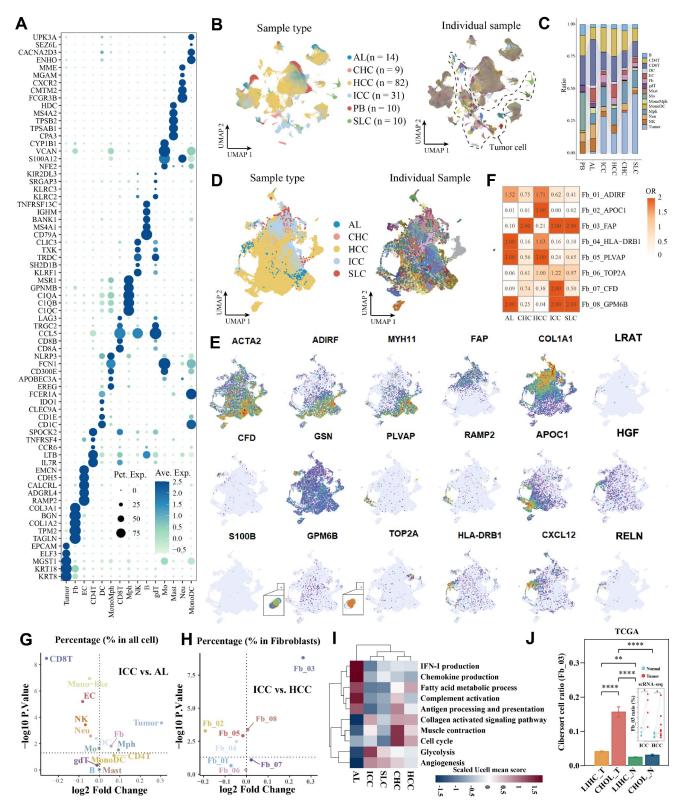


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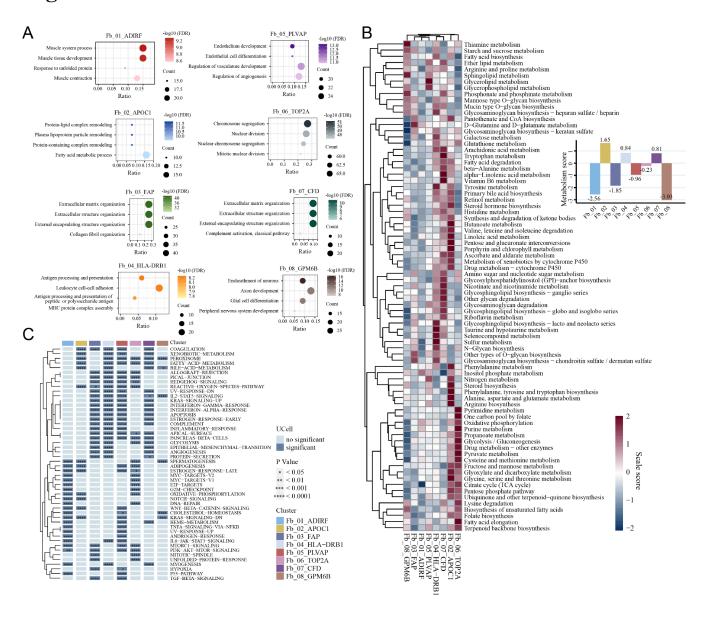


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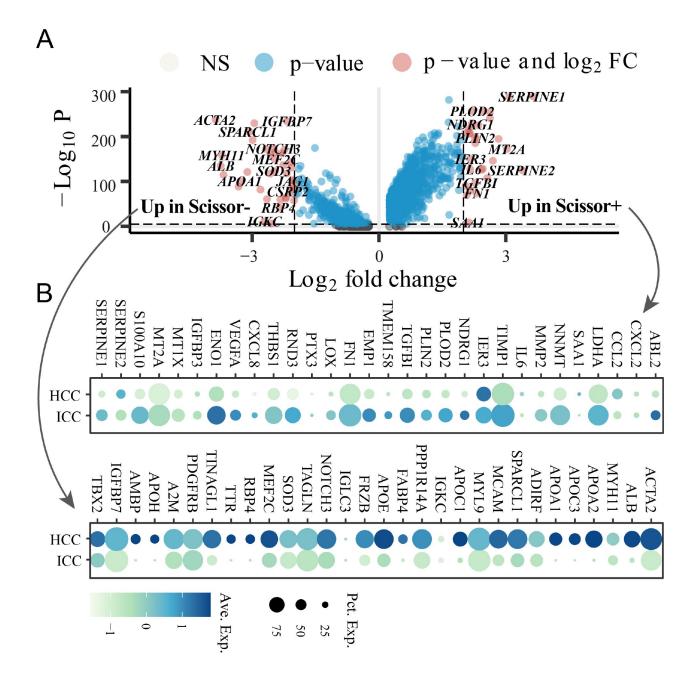
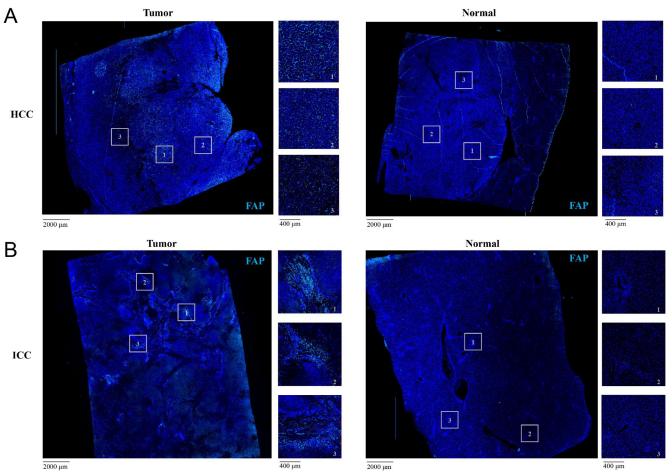


Figure S4.



2000 µm

Figure S5.

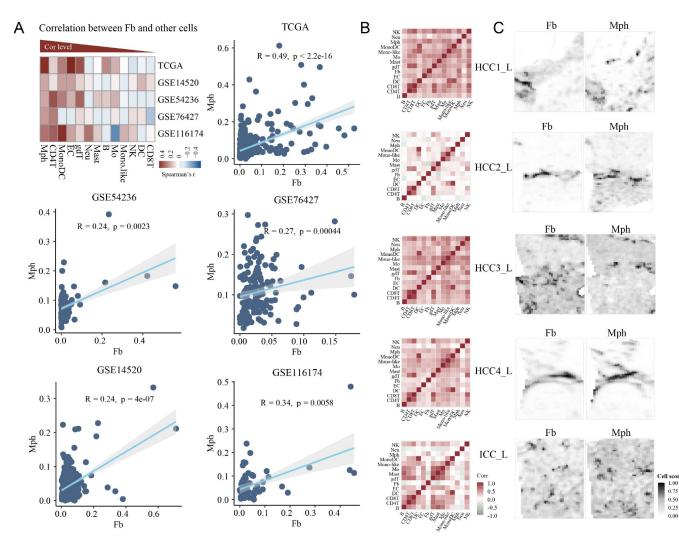


Figure S6.

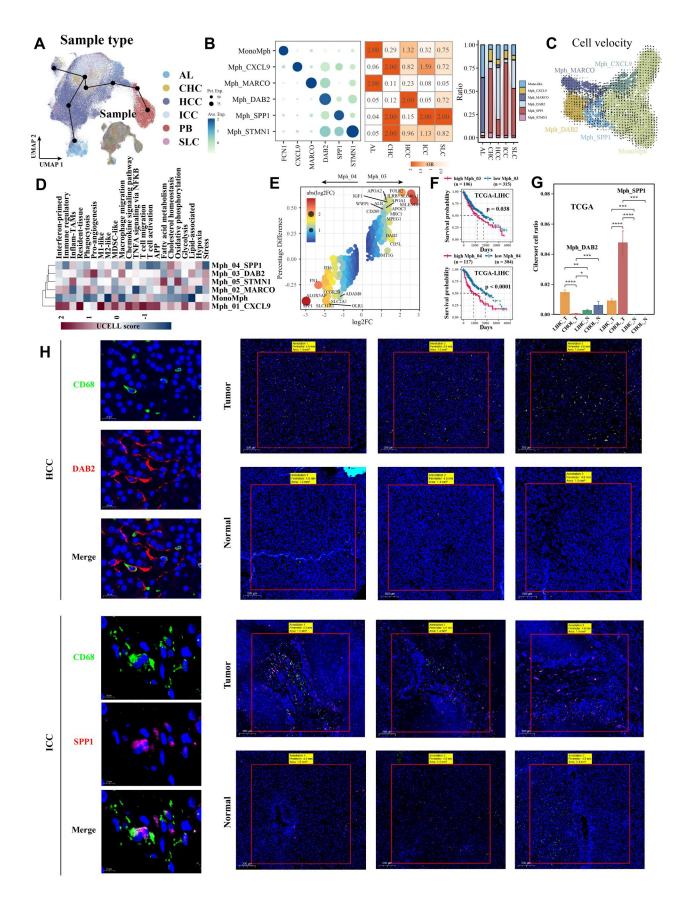
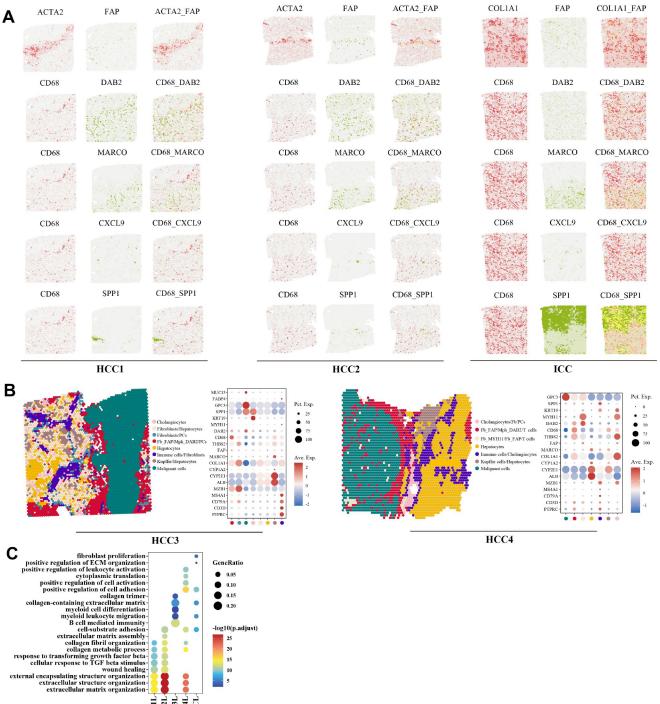


Figure S7.



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HCCIL-HCC2L-HCC3L-HCC4L-ICCL-

Figure S8.

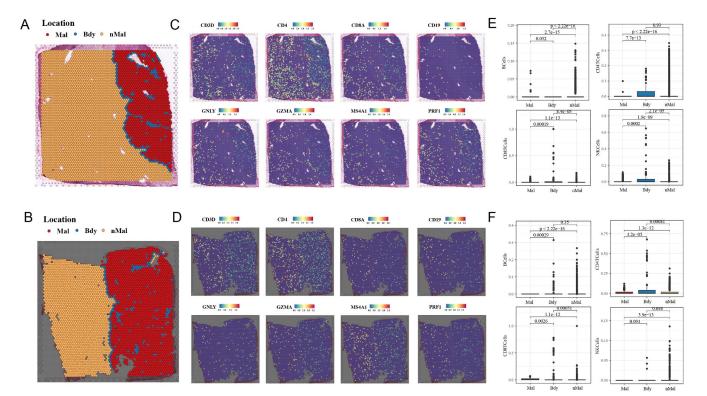


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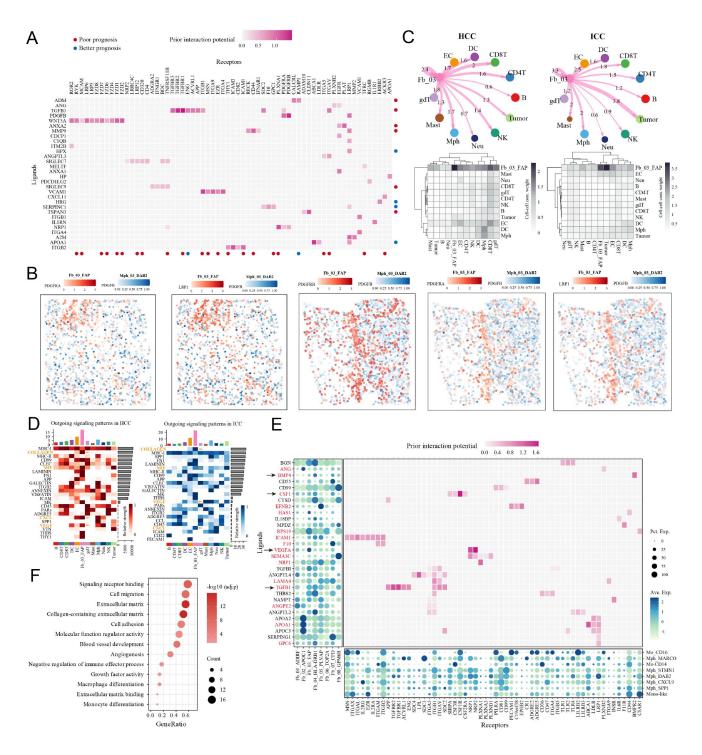


Figure S10.

high VEGFB (n = 92) А С ---- low VEGFB (n = 23) В VEGFB Ext Exp COL3A1_ADGRG1 COL3A1_ADGRG1 15 SLC . p = 0.0093 45 COL4A2_ADGRG6 Means PPIA_BSG ē 10 ICC Survival probability COL4A1_ADGRG6 APP_CD74 2 5 HCC 1.0 0.5 0.0 -0.5 -1.0 UMAP 0 THY1 ADGRE5 VSIR_HLA-E 0.5 0 CHC JAG1_CD46 VEGFB_FLT1 -5 Secreted: TRUE TNFSF12 TNFRSF12A THBS2_CD36 -10 AL IGFBP3 TMEM219 FALSE GSE89749 VEGFA_FLT1 -10 0 10 5 UMAP 1 GAS6_TYRO3 Means THY1_ADGRE5 Day D PLAU_PLAUR VEGFB_NRP1 JAG1_NOTCH2 13 13 0.0 0.5 1.0 VEGFA_NRP2 SEMA4C_PLXNB2 PGF_FLT1 HBEGF EGFR VEGFB VEGFB 12 VEGFA_NRP1 TYROBP CD44 VEGFB NRP1 VEGFA KDR PRNP ADGRG6 FGF7_FGFR2 11 11 SEMA3C_NRP2 TGFB1_TGFBR3 0.21, p = 0.019HGF MET 0.35, p = 0.00011 THBS1_CD36 SEMA3C NRP2 GSE89749 GSE89749 FGF1_FGFR3 • GJA1_GJA1 PTPRF_LRRC4C FGF1_FGFR4 10 11 9 SEMA3F_NRP2 CALCRL RAMP1 FGF2_FGFR3 PDGFC_PDGFRA PGF_NRP1 PDGFB_PDGFRA NTN4_UNC5B Fb_03_FAP Fb_03_FAP FGF1_FGFR2 Е JAG1_VASN RAMPI CALCRL FGFR4_SDC2 FGF2_FGFR4 IGFBP3_TMEM219 JAG1_NOTCH3 FGF1_FGFR1 JAG2_NOTCH3 PDGFA_PDGFRA IGF1 IGF1R JAG1 CD46 VSIR HLA-E FGF2_FGFR2 GAS6 AXL SDC1_ADGRA2 FGF2_FGFR1 EFNA1_EPHA3 PDGFB_PDGFRA DLL4_NOTCH3 PDGFC PDGFRA VEGFA NRP2 JAG1_NOTCH3 PDGFA_PDGFRA PDGFD_PDGFRB PDGFB_PDGFRB VEGFA_NRP2 CD93_IFNGR1 PDGFD_PDGFRB PDGFB_PDGFRB VEGFA_NRP1 LGALS9 P4HB to 13 Turnet 19 - georged PDGFD_PDGFRB to 10 to 10 to 10 to 10 Endothelial/Cancer cell F SPP1+ Mph FAP⁺ CAF PPIA_BSG VEGFA NRP1 PDGFB_PDGFRB CALCRL NRP1 0 VEGEB ADM RAMPI

Figure S11.

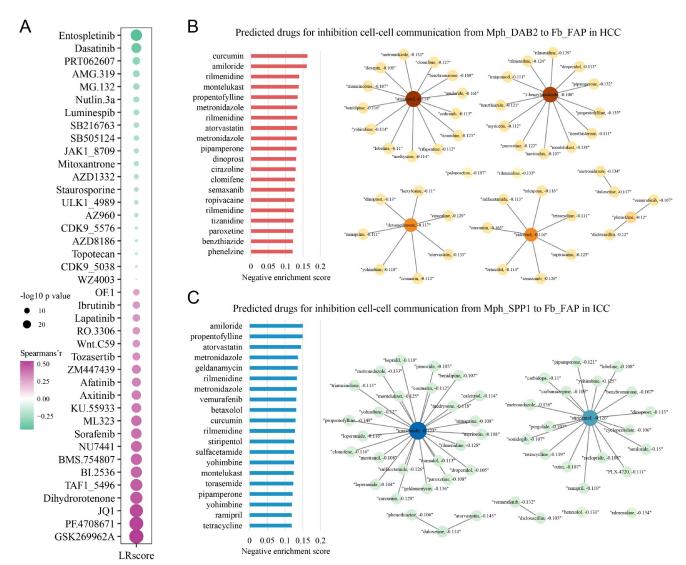


Figure S12.

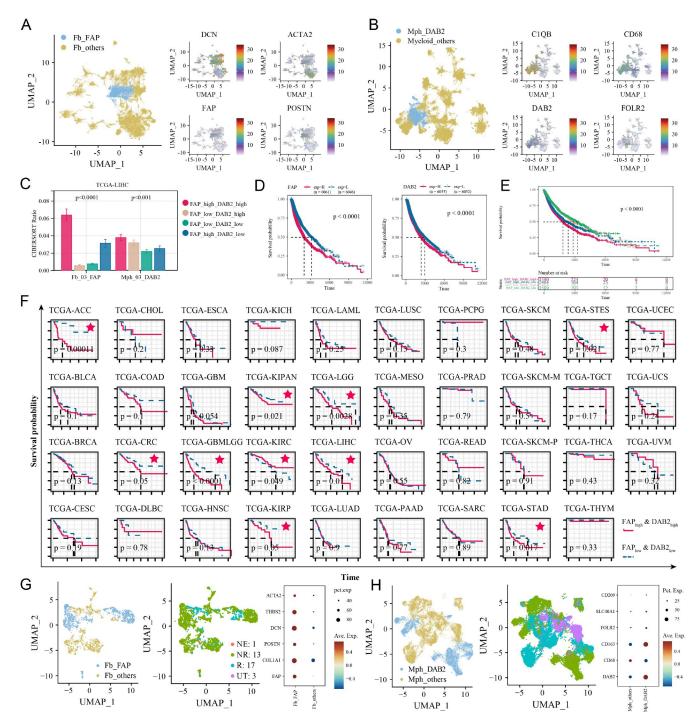
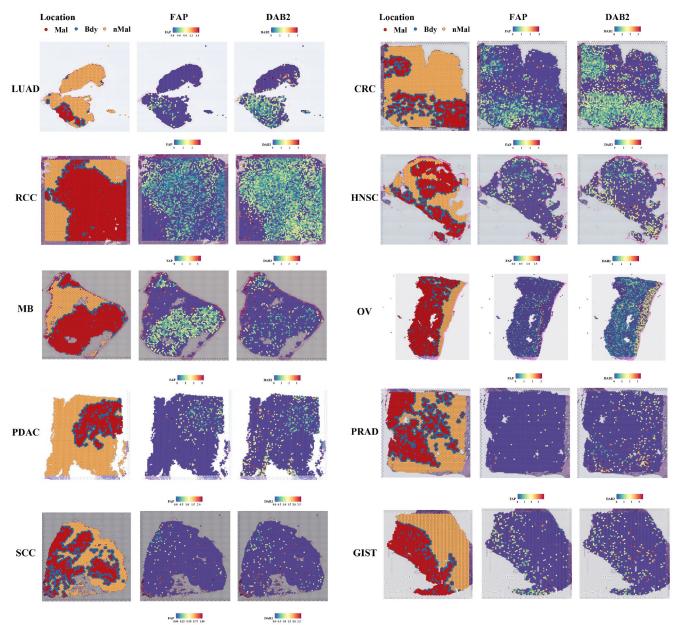


Figure S13.



EPN





