Supplemental Information

Transfer of cardiomyocyte-derived extracellular vesicles to neighboring cardiac cells requires tunneling nanotubes during heart development

Ting Chen et al.



Figure S1. Direct fluorescence of different organs sections from P0, P8, and P28 *Tnnt2-Cre*; doublefloxed inverted *CD9/EGFP* - mice and double staining with CD9 and EGFP both *in vitro* and *in vivo*. No EGFP signal was detected in different organs sections from P0, P8, and P28 *Tnnt2-Cre*; double-floxed inverted *CD9/EGFP* - mice. Double staining with CD9 and EGPF show co-localization both *in vitro* and in *vivo* in *CD9/EGFP*⁺ mice.



Figure S2. GFP antibody was used to amplify EGFP signals.

Immunofluorescence staining of heart sections from P0, P8, and P28 *Tnnt2-Cre*; double-floxed inverted *CD9/EGFP* $^+$ and $^-$ mice The GFP antibody signals (red) were co-localized with the EGFP signals (green). Positive: *Tnnt2-Cre*; double-floxed inverted *CD9/EGFP* $^+$ mice, Negative: *Tnnt2-Cre*; double-floxed inverted *CD9/EGFP* $^-$ mice.



Figure S3. EGFP-labeled EVs were present in a fraction of cardiomyocytes.

Immunofluorescence staining of EGFP⁺ cardiac cultures for EGFP (green), MYH1 (red), and DAPI (blue) (n = 4).

Table S1. The pregnant αMHC -MerCreMer mouse exhibited poor outcomes after tamoxifen administration.

Mouse ID	Methods	Outcome
1	Injecting tamoxifen 50 mg/kg body weight/day to pregnant	Stillbirth
2	mice for 2 consecutive days.	Dystocia
3		Dystocia
4	injecting tamoxinen 20 ing/kg body weight/day to pregnant	Dystocia
5	mice for 4 consecutive days from E14-E17.	Dystocia

Gene	Primer	Sequence
Cre	Cre Trangene F	GCGGTCTGGCAGTAAAAACTATC
	Cre Trangene R	GTGAAACAGCATTGCTGTCACTT
CD9-EGFP	EGFP F	AAGCTGACCCTGAAGTTCATCTGC
	EGFP R	CTTGTAGTTGCCGTCGTCCTTGAA
Interleukin-2	Internal Ctrl F	CTAGGCCACAGAATTGAAAGATCT
	Internal Ctrl R	GTAGGTGGAAATTCTAGCATCATCC

 Table S2. List of genotyping primers used in qPCR-HRM curve analysis.

Gene	Primer	Sequence
CD9-EGFP	PSV 0504	GCAACGTGCTGGTTATTGTG
	PSV 0506	GTCCAATAGCAAGCACTGCA
Beta-actin	Beta-actin F	GCTGTATTCCCCTCCATCGTG
	Beta-actin R	CACGGTTGGCCTTAGGGTTCAG
RPL13A	RPL13A F	CGAAGATGGCGGAGGGGGCAG
	RPL13A R	ACCACCACCTTCCGGCCCAG

 Table S3. List of qRT-PCR primers used in this study.

Antibody	Manufacturer, catalog #	Host species	Dilution
GFP	Abcam (Cat. no.: ab290)	Rabbit	1:500 (IHC, ICC, Flow)
GFP	Abcam (Cat. no.:	Chicken	1:500 (IHC)
	ab13970)		
MYH1	DSHB (Clone MF20)	Mouse	1:200 (IHC, ICC), 1:300
			(Flow)
GFP	GeneTex(Cat. no.:	Rabbit	1:1000 (WB)
	GTX113617)		
CD31	BD Pharmingen (Cat. no.:	Rat	1:50 (IHC)
	553370)		
Vimentin	Abcam (Cat. no.:	Rabbit	1:500 (IHC)
	ab45939)		
Phalloidin	Invitrogen (Cat. no.:	Amanita	1:800 (IHC)
	A30107)	phalloides	
Connexin 43	Sigma-Aldrich (Cat. no.:	Mouse	1:250 (IHC)
	SAB4200730)		
Donkey anti-Rabbit	Invitrogen (Cat. no.: A-	Donkey	1:200 (IHC, ICC, Flow)
IgG, Alexa Fluor 488	21206)		
Goat anti-Chicken IgY,	Invitrogen (Cat. no.: A-	Goat	1:200 (IHC)
Alexa Fluor 488	11039)		
Donkey anti-Rabbit	Invitrogen (Cat. no.: A-	Donkey	1:200 (IHC)
IgG, Alexa Fluor 555	31572)		
Donkey anti-Mouse	Invitrogen (Cat. no.: A-	Donkey	1:200 (IHC, ICC, Flow)
IgG, Alexa Fluor 555	31570)		
Goat anti-Rat IgG,	Invitrogen (Cat. no.: A-	Goat	1:200 (IHC)
Alexa Fluor 555	21434)		
Donkey anti-Mouse	Invitrogen (Cat. no.: A-	Donkey	1:200 (Flow)
IgG, Alexa Fluor 647	31571)		
Goat Anti-Rabbit	Dako (Cat. no.: P0448)	Goat	1:2000 (WB)
Immunoglobulins/HRP			

Table S4. List of antibodies used in this study.

Movie S1. Time-lapse microscopic imaging of tunneling nanotubes linking EGFP⁺ cardiomyocyte with a recipient cell of EGFP⁻ origin.

EGFP signals appeared along the nanotube and at the junction of the recipient cell.