Supplementary materials



Figure S1, Related to Figure 1. CREPT is upregulated in the cancerized field adjacent to the tumor. IHC staining of CREPT in human cervical carcinoma (A) and colorectal carcinoma (B). CREPT is highly expressed in the primary tumor (a), dysplasia (b) and the proximal normal glands (c), but not expressed in the distal normal glands (d). (C) IHC staining of CREPT in human hyperplastic mammary tissue.



Figure S2, Related to Figure 2. CDEs induce CREPT expression in non-malignant epithelial cells. (A) Western blot of CREPT in protein extracts of NMuMG cells (in the lower chamber) cultured with NMuMG/ 4T1 cells (in the upper chamber) treated with DMSO/ GW4869 (5 μ M). (B) Representative TEM image of small extracellular vesicles from MDA-MB-231 cells confirmed the expected teacup-shaped morphology of vesicles. (C) Nanoparticle Tracking Analysis of small extracellular vesicles derived from MDA-MB-231, 4T1, MCF10A and NMuMG cells showed the size distributed between 30-200 nm. (D) Western blot of CD9, CD63, CD81 and Calnexin in protein extracts of small extracellular vesicles from NMuMG, MCF10A, 4T1 and MDA-MB-231 cells. (E) Western blot of CREPT in protein extracts of NCM460 cells treated for 3 days with/ without NCM460EXO or SW620EXO in the concentration of 1 \times 10⁹ particles/mL.



Figure S3, Related to Figure 3. CDEs induce CREPT expression through ERK activation. (A) Western blot of Alix, Flotillin 1 (two common exosomal markers) and CREPT in protein extracts of cells and small extracellular vesicles from 4T1 and MDA-MB-231 cells. (B) Western blot of Alix and CREPT in protein extracts of cells and small extracellular vesicles from B16 cells. (C) The luciferase reporter activity in CHO cells transfected with a luciferase gene driven by CREPT-promoter and treated for 24 h with 0, 2, 4 and 8 \times 10⁹ particles/mL B16EXO. (D) The volcano plot of differentially enriched proteins in SW620EXO compared with NCM460EXO. Red dots indicate 631enriched proteins (fold change > 1.5, p-value < 0.05), blue dots indicate 439 reduced proteins (fold change < 0.67, p-value < 0.05). (E) KEGG analysis of the 631 differentially enriched proteins in SW620EXO. The -Log10(p-value) is indicated. (F) The heatmap of relative levels of 19 protein extracts of 4T1 cells treated with ERK inhibitor (SCH772984) in the concentration of 0, 2, 4 and 6 μ M. (H) The ChIP-Seq results from Cistrome DB showed ELK1 occupied the CREPT promoter region of Hela-S3, K562, MCF7 and A549 cells.



Figure S4, Related to Figure 4. Over-expression of CREPT enhances the proliferation of non-malignant epithelial cells. Western blot results showing CREPT over-expression (OE) and knockout (KO) in CHO (A), NMuMG (B), and MCF10A (C) cell lines. Actin was used as a control. WT, wild-type. (D) The volcano plot of DEGs after CREPT knockout in CHO. (E) The bubble plot of KEGG enrichment analysis of down-regulated genes after CREPT knockout in CHO.



Figure S5, Related to Figure 5. CREPT is required for cancerization induced by CDEs. (A) CCK-8 assay during 6 days of NMuMG cells treated with 4T1EXO for 0, 1, 3 or 6 days. The cell number indicated by OD450 is normalized by the value of day 0. (B) Representative images of cells stained with crystal violet showing colony formation from MCF10A CREPT-WT and CREPT-KO treated with PBS or 231EXO for two weeks. (C) Quantification of colony formation assay (B), the data shows the colony number of each group. (D) The HE staining of the skin in wild-type mice (n=5) after the injection of PBS, CHOEXO or B16EXO in the skin for 14 days and the skin in the mice with systemic knockout of CREPT (n = 5) after the injection of B16EXO in the skin for 14 days. (E) The statistical results of Figure S5D, n = 5. (F) IHC staining of Ki67 in the corresponding skin tissue in Figure S5D. (G) The statistical results of Figure S5F, n = 20.



Figure S6, Related to Figure 6. CREPT-mediated TNFR2 signaling promotes field cancerization induced by CDEs. (A) The volcano plot of the differentially expressed genes in NMuMG cells treated with 4T1EXO compared with untreated NMuMG cells. The blue dots show the significantly (fold change < 0.5, p-value < 0.001) down-regulated genes and the red dots show the significantly (fold change > 2, p-value < 0.001) up-regulated genes. (B) KEGG analysis of the differentially expressed genes between untreated NMuMG cells and NMuMG cells treated with 4T1EXO for 2 weeks. The input gene number, p-value and rich factor of each signaling pathway are indicated. (C) The verification of the inflammationrelated-mRNA levels of TNF, NOD2, CXCL1 and CSF1 by qPCR in NMuMG cells treated with 4T1EXO on day 0, 3, 7 and 16. (D) The Venn diagram shows the differentially expressed gene number in NMuMG WT/ KO cells treated with/ without 4T1EXO. (E) KEGG analysis of genes that were significantly upregulated in NMuMG CREPT-WT cells but not significantly upregulated in NMuMG CREPT-KO cells. The input gene number, p-value and rich factor of each signaling pathway are indicated. (F) Heatmap of the mRNA levels of genes related to TNF signaling pathway in NMuMG CREPT-WT and CREPT-KO cells treated with PBS or 4T1EXO. The mRNA levels of Tnfrsf1b (G) and Pik3cd (H) was quantified by qPCR of NMuMG WT/ KO cells treated with/without 4T1EXO. (I) Western blot of CREPT, p-AKT, AKT, p-p65 and p65 in protein extracts of NMuMG CREPT-WT and CREPT-KO cells treated with 4T1EXO for 0, 1, 3, 6, 9 and 11 days.

Table S1. The clinical characteristics of tumors used in PDX model				
	Age	type	TNM	ER/PR/ HER-2
Patient 1	61	invasive ductal carcinoma	T2N0M0	-/-/-
Patient 2	60	invasive ductal carcinoma	T2N0M0	_/_/-
Patient 3	63	invasive ductal carcinoma	T3N0M0	-/-/-