

Supporting Information

Dietary cobalt oxide nanoparticles alleviate aging through activation of mitochondrial UPR in *Caenorhabditis elegans*

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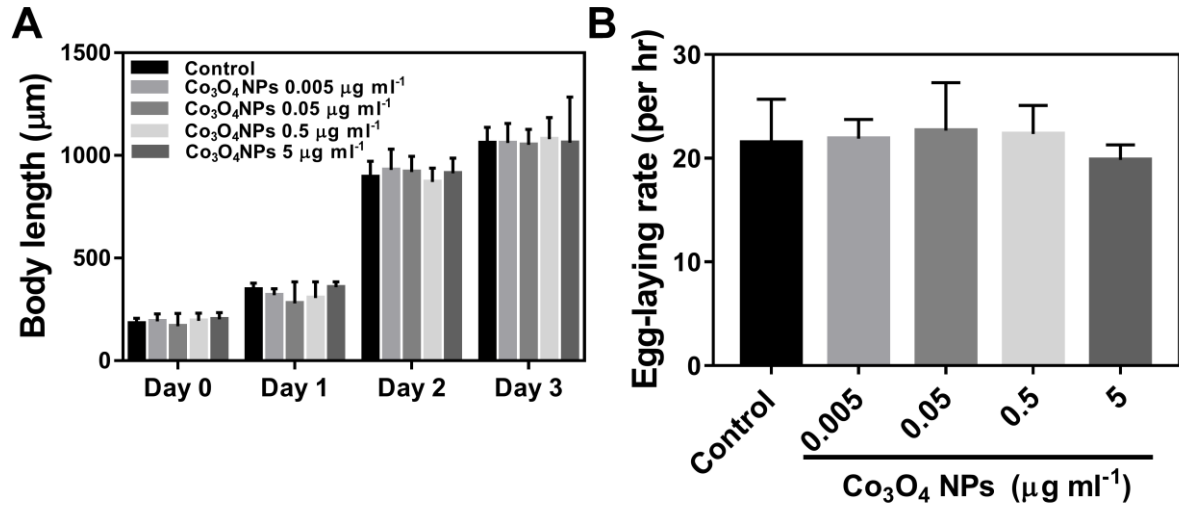


Figure S1. Biosafety assessment of Co₃O₄ NPs on *C. elegans*. Effects of Co₃O₄ NPs on worms were shown in body length (A) and egg-laying rate (B). Wild-type worms at L1 larva stage were treated with Co₃O₄ NPs with gradient doses (0.005-5 µg ml⁻¹) for three days. Bars represent means ± SD.

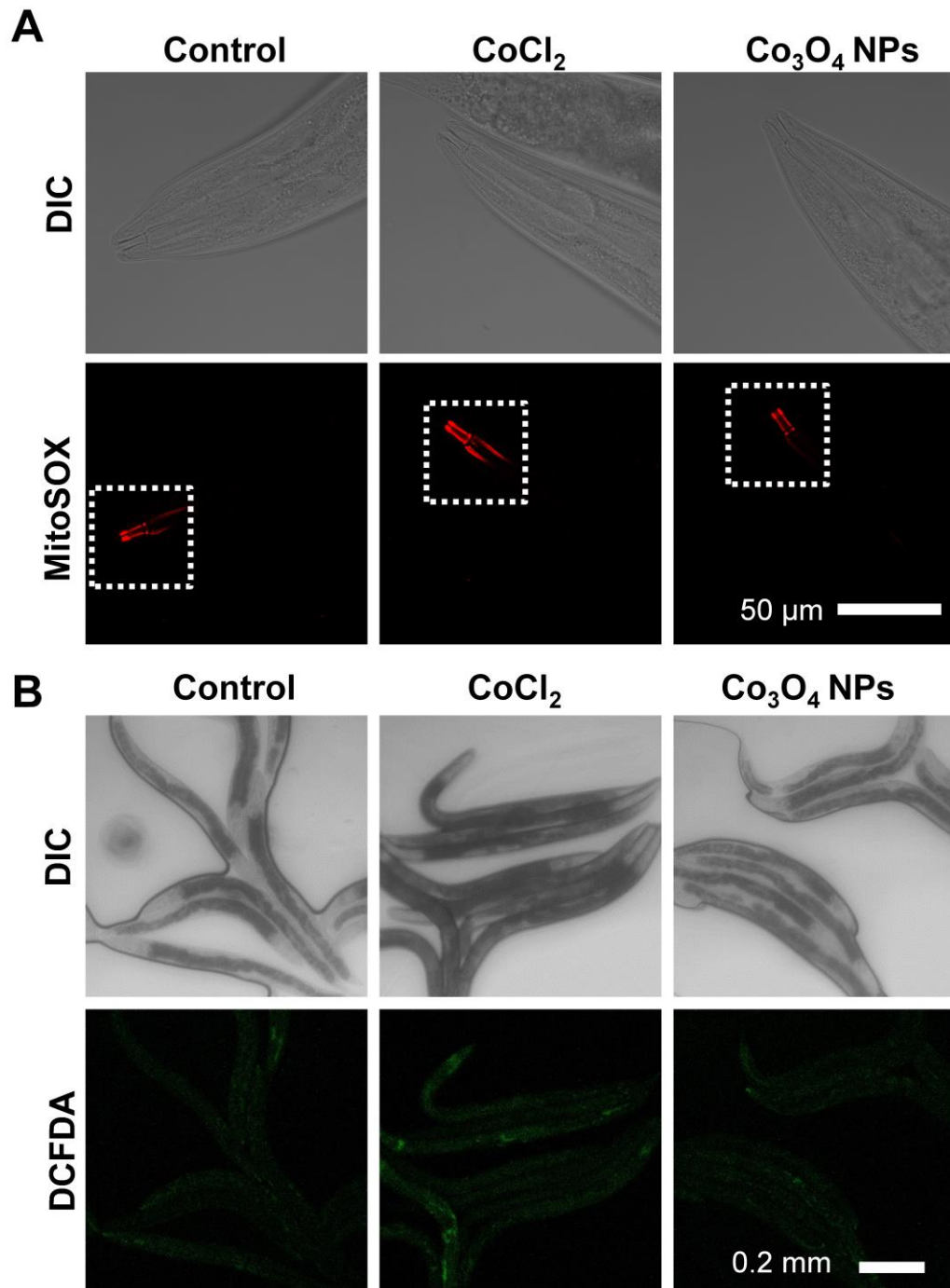


Figure S2. Representative pictures of mitochondrial ROS on nematode head (A) and cellular ROS (B) in Co₃O₄ NPs, CoCl₂ or mock-treated worms on adult day 1.

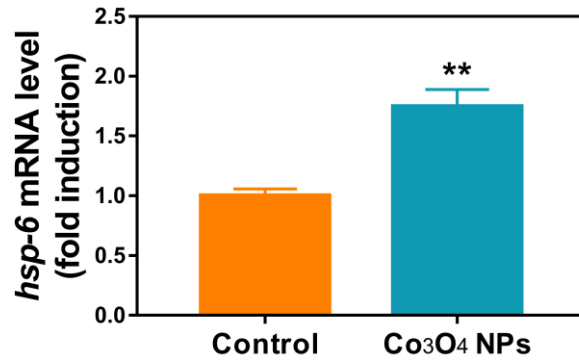


Figure S3. Influence of Co₃O₄ NPs on *hsp-6* (homologous gene of *hsp60*) expression. *mRNA* expression of *hsp-6* of worms on adult day 1 pretreated with Co₃O₄ NPs.

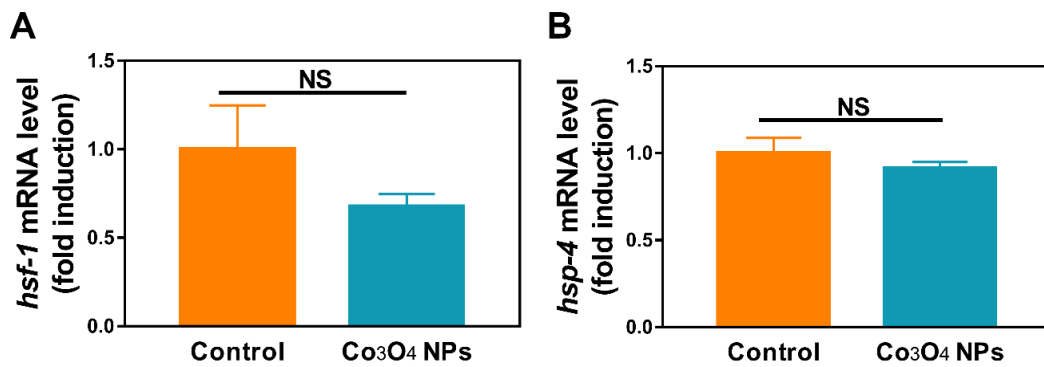


Figure S4. Influence of Co₃O₄ NPs on *hsf-1* and *hsp-4* expression. (A) Expression of *hsf-1* in worms on adult day 1 pretreated with Co₃O₄ NPs. (B) Expression of *hsp-4* in worms on adult day 1 pretreated with Co₃O₄ NPs..

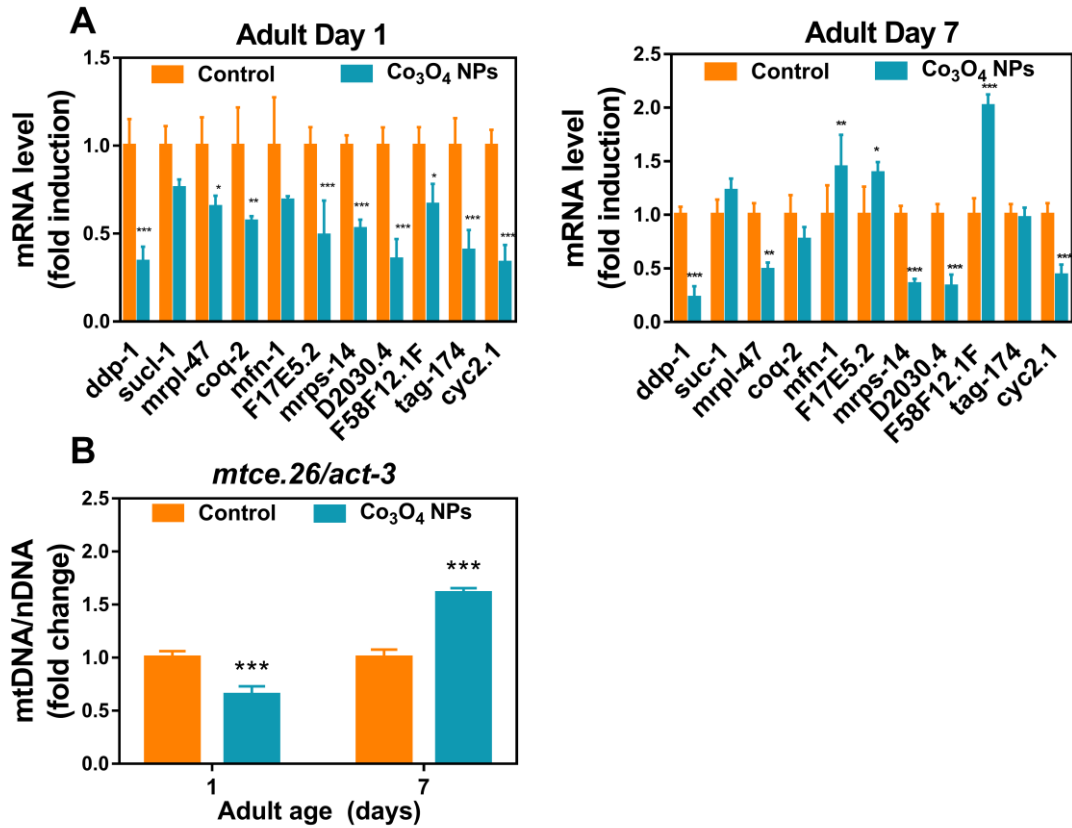


Figure S5. The effect of Co₃O₄ NPs on mitochondrial abundance. (A) RT-qPCR analysis of changes in the expression of nuclear genes encoding mitochondrial proteins in wild-type worms on adulthood days 1 and 7 pretreated with Co₃O₄ NPs. (B) RT-qPCR analysis of mtDNA/nDNA ratio (*mtce.26/act-3*) in wild-type worms on adulthood days 1 and 7 pretreated with Co₃O₄ NPs.

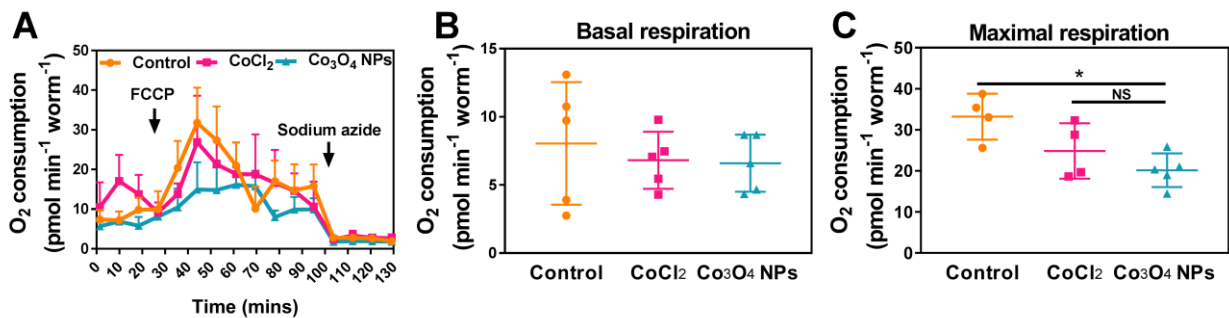


Figure S6. Influence of Co₃O₄ NPs on oxygen consumption rate (OCR) compared to CoCl₂ and negative control. (A) Oxygen consumption curves of Co₃O₄ NPs, CoCl₂ treated worms or blank control in the presence FCCP and NaN₃ on adult day 4. Comparison of the basal oxygen consumption rate (B) and maximum oxygen consumption rate (C) in N₂ wildtype worms treated with Co₃O₄ NPs or CoCl₂ on adult day 4.

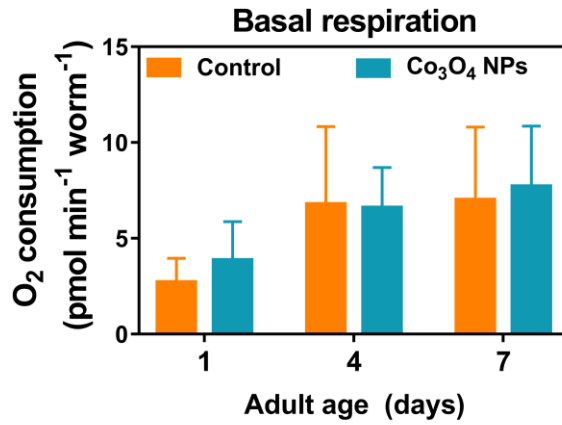


Figure S7. Basal oxygen consumption of N₂ wildtype worms treated with Co₃O₄ NPs or not on adult days 1, 4 and 7.

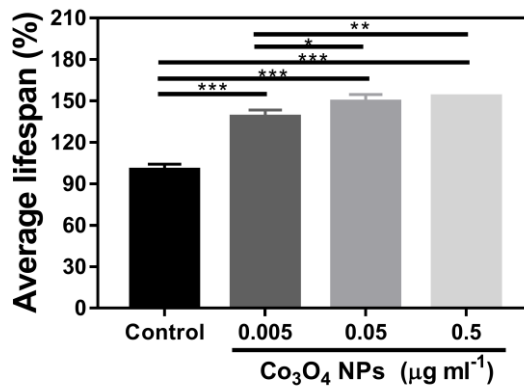


Figure S8. The average lifespan (%) of worms treated with Co₃O₄ NPs (from 0 to 0.5 μg ml⁻¹).

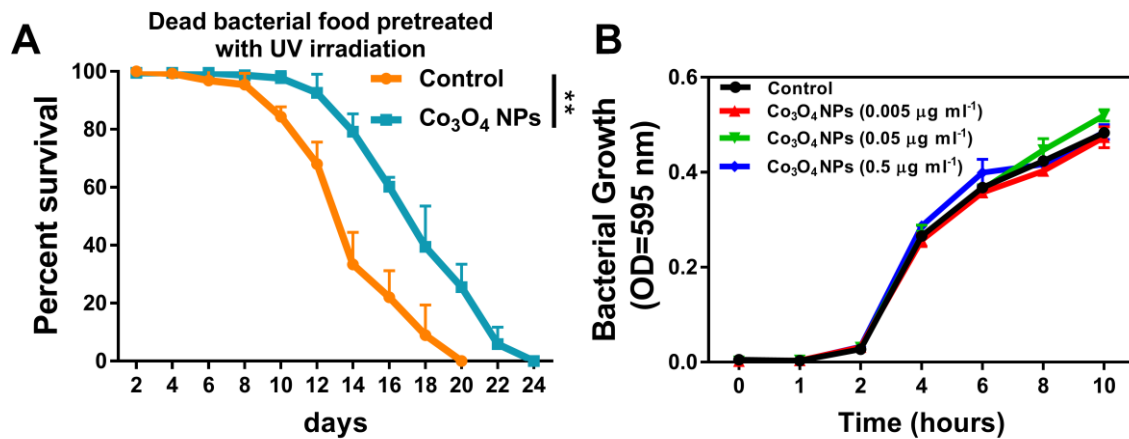


Figure S9. (A) Lifespan curves of *C. elegans* treated with Co_3O_4 NPs in dead bacterial food pretreated with UV irradiation for 30 minutes. (B) OP50 bacteria in liquid LB treated with different concentrations of Co_3O_4 NPs did not show growth retardation.

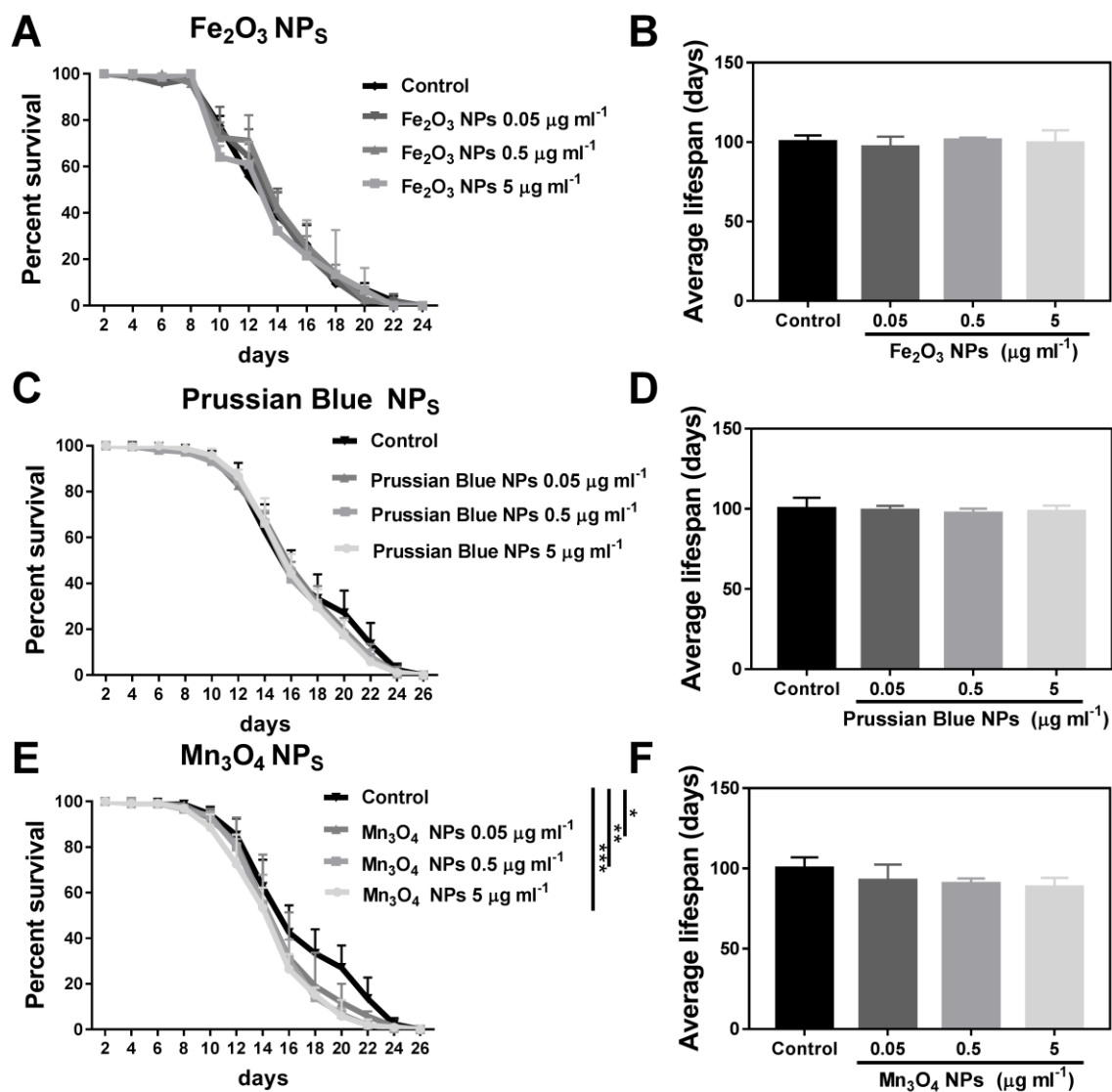


Figure S10. Influence of Fe_2O_3 NPs, Prussian Blue NPs and Mn_3O_4 NPs on lifespan of *C. elegans*. (A) Lifespan curves of *C. elegans* treated with Fe_2O_3 NPs. (B) The average lifespan (%) of worms treated with Fe_2O_3 NPs (from 0 to $5 \mu\text{g ml}^{-1}$). (C) Lifespan curves of *C. elegans* treated with Prussian Blue NPs. (D) The average lifespan (%) of worms treated with Prussian Blue NPs (from 0 to $5 \mu\text{g ml}^{-1}$). (E) Lifespan curves of *C. elegans* treated with Mn_3O_4 NPs. (F) The average lifespan (%) of worms treated with Mn_3O_4 NPs (from 0 to $5 \mu\text{g ml}^{-1}$).

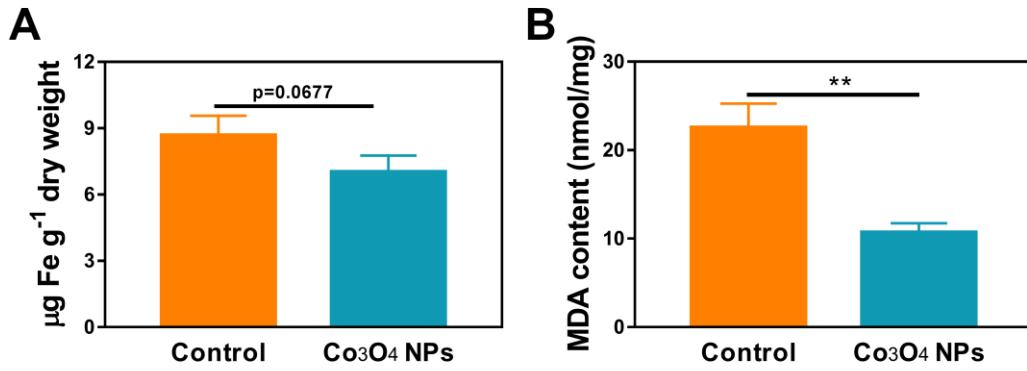


Figure S11. Influence of Co_3O_4 NPs on Fe accumulation and MDA content in *C. elegans*
 A. *In vivo* Fe levels ($\mu\text{g Fe}$ per dry weight measured via ICP-MS) in worms treated with or without Co_3O_4 NPs on adult day 7. B. Malondialdehyde (MDA) content in worms treated with or without Co_3O_4 NPs on adult day 7.

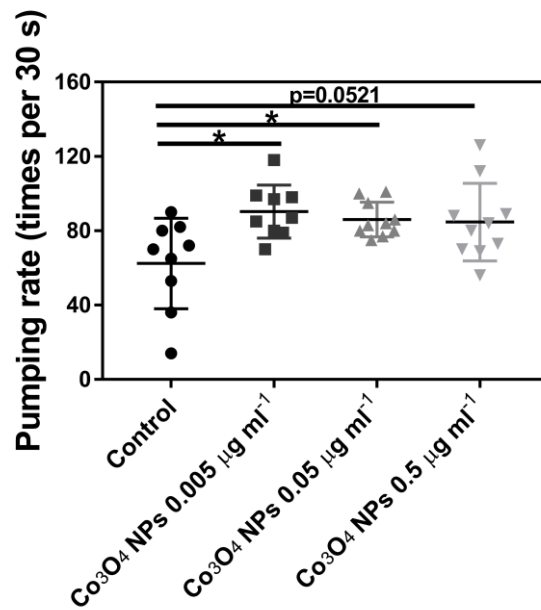


Figure S12. Pharyngeal pumping in worms treated with Co_3O_4 NPs at different doses on adult day 6.

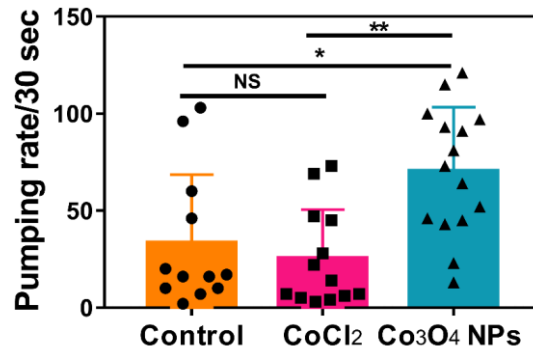


Figure S13. Pharyngeal pumping in worms treated with Co_3O_4 NPs or CoCl_2 at $0.05 \mu\text{g ml}^{-1}$ on adult day 9.

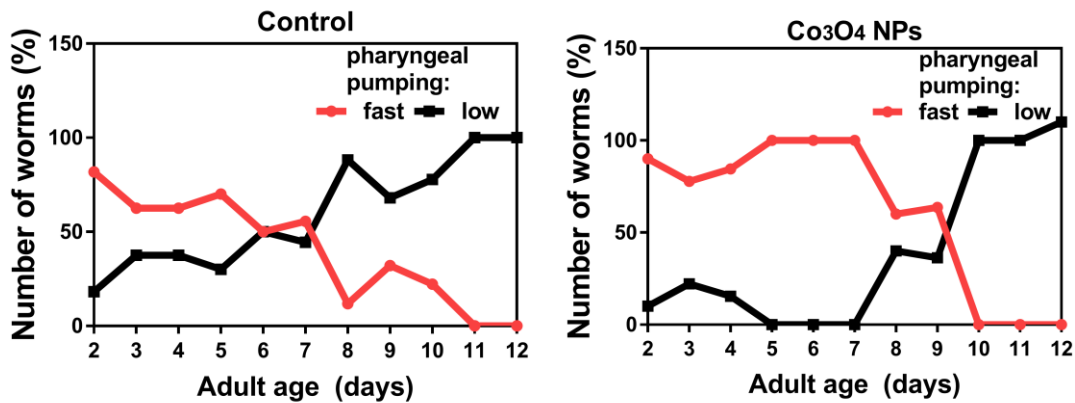


Figure S14. Co_3O_4 NPs' effect on pumping rate by quantifying the ratio of worms that are fast-, slow-pumpers. Typically, pharyngeal contractions were divided the following subgroups: <6 per minute (not pumping), 6-147 per minute (slow pumping), and >147 (fast pumping), respectively.

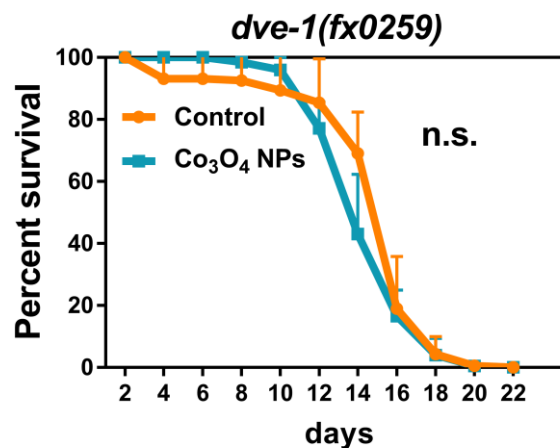


Figure S15. Supplementation of Co_3O_4 NPs does not extend lifespan in the *dev-1(fx0259)*

mutant (n.s. = not significant).

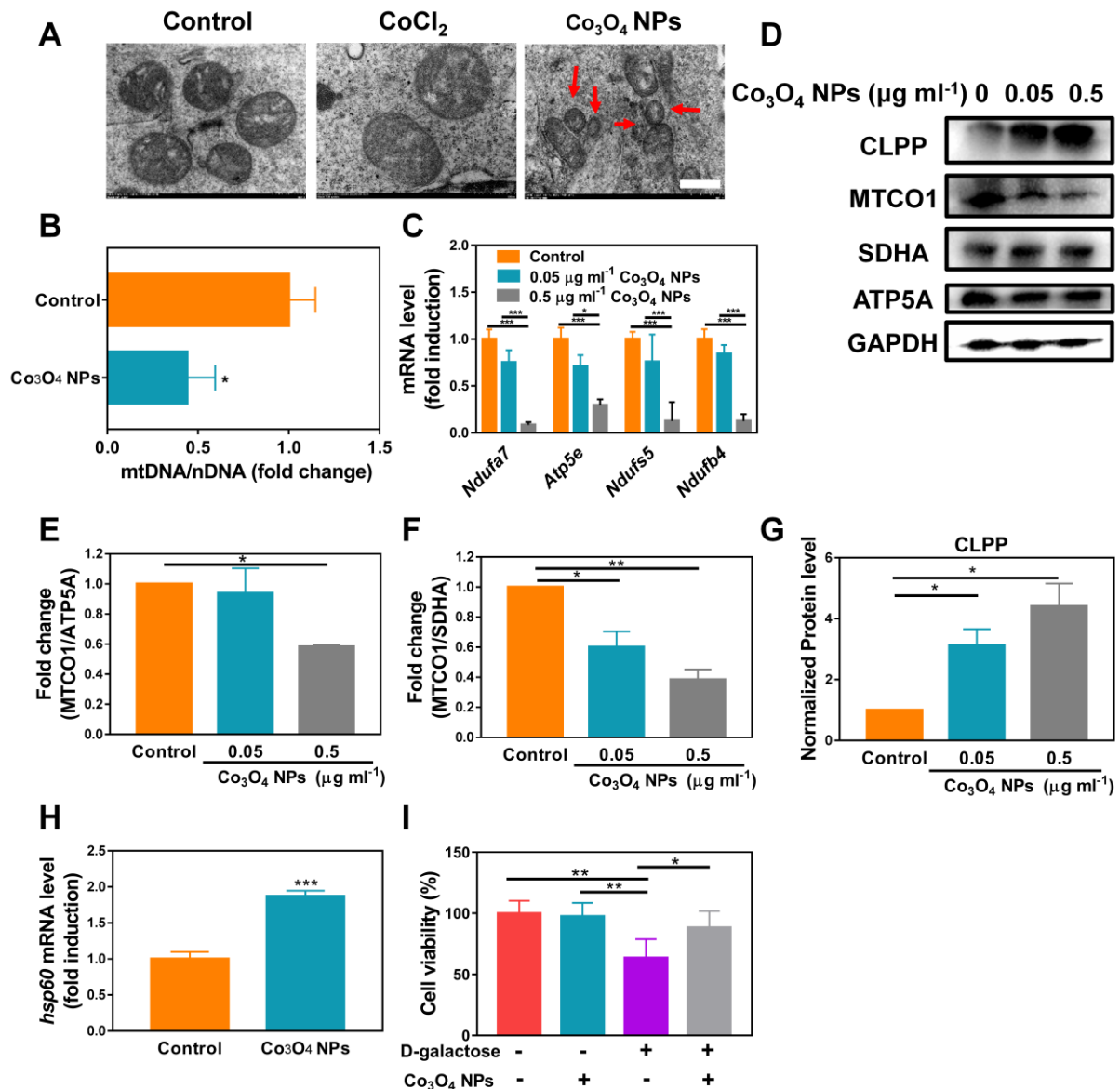


Figure S16. Effect of Co₃O₄ NPs on mitochondrial homeostasis in mammalian cells. (A) Mitochondrial morphology of cells treated with Co₃O₄ NPs, CoCl₂ or blank control by transmission electron microscopy. Mitochondrial in smaller size versus average was indicated by red arrows (B) Treatment with Co₃O₄ NPs decreases the ratio between mitochondrial DNA (mtDNA) and nuclear DNA (nDNA), a common marker for mitochondrial abundance in HEK293t cells. (C) Effects of Co₃O₄ NPs on the transcription of mitochondrial-related genes in HEK293t cells. (D) Western blots pics of CLPP, MTCO1, SDHA and ATP5A in HEK293t cells treated with or without Co₃O₄ NPs. GAPDH served as a loading control. (E) Quantitative analysis on the ratio of mtDNA-encoded MTCO1 and nDNA-encoded ATP5A in HEK293t cells treated with Co₃O₄ NPs. (F) Quantitative analysis on the ratio of mtDNA-encoded MTCO1 proteins and nDNA-encoded SDHA in HEK293t cells treated with Co₃O₄ NPs. (G) Quantitative

analysis on the UPR^{mt} protease CLPP expression in HEK293t cells treated with Co₃O₄ NPs. (H) The mRNA expression level of *hsp60* in HEK293t cells treated with Co₃O₄ NPs. (I) Effect of Co₃O₄ NPs on D-galactose-induced cellular viability decline. HEK293t cells were pretreated with Co₃O₄ NPs at 0.05 µg ml⁻¹ in DMEM medium, and then exposed to D-galactose (20 g L⁻¹) for 48 h. CCK8 assay was used to measure cellular viability. Bar graphs are expressed as mean±SEM. *P<0.05, **P<0.01, ***P<0.001. Scale bar=5 µm.

Table S1. Primers used in RT-qPCR

Gene name	Sequence (5'-3')
<i>cts-1-rv</i>	GGTACAGGTTGCGATAGATGATAGC
<i>cts-1-fw</i>	CTCGACAACCTCCCAGATAACC
<i>cox-4-rv</i>	AGGTTGGCGGCAGTTCTGGG
<i>cox-4-fw</i>	GCCCCAATTCGCGCCAAGGA
<i>hvk-1-rv</i>	CTAGAGATGACGTCACACACTTCTC
<i>hvk-1-fw</i>	GTGCGACGAGTACTTTCTCAACTG
<i>fzo-1-rv</i>	AGTCGGCATTCCCCTGATTCCG
<i>fzo-1-fw</i>	TCTGCAGGTTGAAGGTTCAGAAGGC
<i>opa-1-rv</i>	CTTGCCATCCATTCTGCCCA
<i>opa-1-fw</i>	TCGCGGCTAGAACGTGGTATGA
<i>hsp-6-rv</i>	AGCGATGATCTTATCTCCAGCGTCC
<i>hsp-6-fw</i>	AACCACCGTCAACAACGCCG
<i>pyc-1-rv</i>	GTGATCATAATCCTGGTCTACTGC
<i>pyc-1-fw</i>	TCCAATACTCCTCTTGCTACTGAC
<i>ddp-1-rv</i>	AGTGCTCGACCATGAAGTTG
<i>ddp-1-fw</i>	AACAAGTGCACACGCTCA
<i>sucl-1-rv</i>	CAGCTGATCCTCCGATTCT
<i>sucl-1-fw</i>	GTCGGATTCGGACAGACTTT
<i>mrpl-47-rv</i>	ACTCGTGGAGCTCCTCTCTTGA
<i>mrpl-47-fw</i>	CGACGACGATGCCTACGTGA
<i>mrps-14-rv</i>	TTGGACTCCACTGAGAGCTG

<i>mrps-14-fw</i>	CGATCATCCTCGTCTGATTC
<i>coq-2-rv</i>	CCGATGTTCTTCCTCGATTC
<i>coq-2-fw</i>	AGTCAAACCTCATGGATGGGAA
<i>mfn-1-rv</i>	TGCAGGAACCTGGAATATGA
<i>mfn-1-fw</i>	GATCCAGCAAATAGGCGAAT
<i>F17E5.2-rv</i>	CCTTCATGAAATTCGGTGTG
<i>F17E5.2-fw</i>	TCGAACAAGGCTTCAAGCTA
<i>F58F12.1-rv</i>	TGATCAATGCTTCAGCAACTT
<i>F58F12.1-fw</i>	CTTGATGCTGCCCAAAGAG
<i>D2030.4-rv</i>	TTCAAGCAGTGGCTTCTTTG
<i>D2030.4-fw</i>	TGGAGCTTGGGATAAATGTG
<i>tag-174-rv</i>	TAGTGCTTCTCACGGTCAGC
<i>tag-174-fw</i>	TCGAGTACGCCTTCTTGAAC
<i>cyc-2.1-rv</i>	TTCTTGAGTCCAGCGAACAC
<i>cyc-2.1-fw</i>	CGCTGCTAACAAGAACAAGG
<i>act-1-rv</i>	GTAGCAGAGCTTCTCCTTGATGTC
<i>act-1-fw</i>	GCTGGACGTGATCTTACTGATTACC
<i>act-3-rv</i>	GGTGGTTCTCCGGAAAGAA
<i>act-3-fw</i>	TGCGACATTGATATCCGTAAGG
<i>nd-1-rv</i>	AAGCTTGTGCTAATCCCATAAATGT
<i>nd-1-fw</i>	AGCGTCATTTATTGGGAAGAAGAC
<i>mtce.26-rv</i>	CAGGGTGCCCCATTGTTCTT
<i>mtce.26-fw</i>	GGTTGTGGGACTAGGTGAACA
<i>HK2-rv</i>	GGGAACACAAAAGACCTCTTCTGG
<i>HK2-fw</i>	GCCAGCCTCTCCTGATTTTAGTGT
<i>16s-rv</i>	TCGTTTGGTTTCGGGGTTTC
<i>16s-fw</i>	CCGCAAGGGAAAGATGAAAGAC
<i>Ndufa7-rv</i>	GAGCTTGTGGCTAGGACCC
<i>Ndufa7-fw</i>	TGCAGCTACGCTACCAGGA
<i>Ndufs5-rv</i>	CCGAAGCAAACACTCTACGAAAT

<i>Ndufs5-fw</i>	TGCACATGGAATCGGTTATACTC
<i>Ndufb4-rv</i>	GAAGCAGGTACTCTCGTTTCAG
<i>Ndufb4-fw</i>	ATGTCGTTCCCAAAGTATAAGCC
<i>Atp5e-rv</i>	GGAGTATCGGATGTAGCTGAGT
<i>Atp5e-fw</i>	GTGGCCTACTGGAGACAGG
<i>hsp60-rv</i>	AGCCCGAGTGAGATGAGGAG
<i>hsp60-fw</i>	ATGCTTCGGTTACCCACAGTC