

1 **Supplementary Materials and Methods**

2 **Mice and Cell lines**

3 The female C57BL/6 mice at 6-8 weeks of age were purchased from Joint Ventures
4 Sipper BK Experimental Animal (Shanghai, China). *CD4^{Cre}* and *Foxp3^{YFP-Cre}* mice as
5 described previously were purchased from Cyagen Biosciences. *Ragl^{-/-}* mice were
6 purchased from Model Animal Research Center. All strains of mice were housed in a
7 specific pathogen-free facility. The experimental protocols were approved by the
8 Animal Care and Use Committee of Medical School of Zhejiang University (Hangzhou,
9 China). The *CD4⁺Foxp3/CD25⁺CD69⁺* and *CD4⁺Foxp3/CD25⁺CD69⁻* Tregs were
10 sorted using the FACSDiVa system (Becton Dickinson), respectively.

11 **Tregs suppression assay**

12 The suppression of *CD4⁺* T cell proliferation in each group was determined by the CFSE
13 incorporation assay. The murine splenic *CD4⁺* T cells were isolated using a *CD4⁺* T-
14 cell isolation kit II (Miltenyi Biotec, Germany) and were labeled with CFSE (Invitrogen)
15 according to the manufacturer's instruction. CFSE labeled *CD4⁺* T cells (1×10^6 /ml)
16 were stimulated with 1 μ L of anti-*CD3/CD28*-coated beads (Invitrogen). Purified
17 *CD69^{fl/fl}-iTregs*, *CD4^{Cre}CD69^{fl/fl}-iTregs* at a ratio of 1: 1, 1: 2, or 1:4 were added, the
18 effector *CD4⁺* T cells served as the control. Three days later, the cells were harvested,
19 and the proliferation of effector *CD4⁺* T cells was analyzed using flow cytometry.

20 **ELISA**

21 *IL-10* and *TGF- β 1* levels in the supernatant of cultured cells and the *IL-6*, *IFN- γ* , and
22 *IL-17* levels in the colon of mice were measured using specific kits (eBioscience, USA)
23 for ELISA. For analysis of colon explant cultures, the colons of mice were flushed with
24 PBS containing 30% antibiotics, and open along a longitudinal axis. Then, pieces of
25 tissue ($\sim 3\text{-mm}^2$) were obtained from the distal colon and incubated for 24 hours in
26 RPMI supplemented with 10% FCS and 20% antibiotics (1 punch biopsy/100 ml
27 medium). Supernatants were collected and kept frozen until assessment. For the
28 detection of *TGF- β 1*, 100 μ L of supernatants were acidified with 20 μ L of 1N HCl at
29 room temperature for 10 min and then neutralized with 20 μ L of 1N NaOH to activate
30 latent *TGF- β 1* into its immunoreactive form.

31 **RNA isolation, cDNA synthesis, and quantitative real-time PCR**

32 Total RNA from the indicated types of cells was extracted using Trizol reagents, and
33 the RNA samples were reverse transcribed into cDNA using a high-capacity cDNA
34 Reverse Transcription Kit (Thermo Fisher, USA). The relative levels of target mRNAs
35 were determined by qRT-PCR. The sequences of primers are listed in Table S1. Data
36 were analyzed using the comparative Ct method using β -actin as the normalization
37 control.

38 **Confocal microscopy**

39 CD4⁺ cells seeded on chamber slides were cultured overnight. After treatment with
40 MG132 for 2h, cells were fixed with 4% paraformaldehyde and permeabilized in 0.1%
41 Triton X-100/PBS for 10 min. After blocking with 5% bovine serum albumin, cells
42 were incubated with rat anti-HSF1 monoclonal antibody followed by decoration with
43 fluorescein-conjugated anti-rabbit IgG. The images were acquired on confocal
44 microscope.

45 **FACS**

46 To analyze the differentiation of Th1 or Th17 cells, CD4⁺ T cells were incubated with
47 the cell stimulation cocktail (eBioscience, USA) for 5 h at 37 °C and stained with PE-
48 Cy7-anti-CD4 antibodies. Cells were fixed and stained with PE-anti-IFN- γ (Invitrogen,
49 Clone: XMG1.2; Catalog: 12-7311-82), APC-anti-IL-17 (Invitrogen, Clone: eBio17B7;
50 Catalog: 17-7177-81) antibodies, or isotype controls after permeabilization. The
51 percentages of CD4⁺IFN- γ ⁺ and CD4⁺IL-17⁺ cells were determined by flow cytometry.

52 **Protein half-life assay**

53 CD4⁺ T cells were pretreated with or without MG132 (Sigma Aldrich) for 2 hours then
54 incubated with 50 μ g/ml CHX (Sigma Aldrich) for 0,6, 12, 18 hours or 25 before
55 Western blot analysis.

56 **Colitis induction**

57 *CD69^{fl/fl}* or *Foxp3^{YFP-Cre}CD69^{fl/fl}* Mice were orally given 2% DSS (MW 36,000-
58 50,000, MP Biomedicals, USA) in drinking water for 5 days, followed by regular
59 drinking water without DSS. Body weight was measured every 24 h during the

60 experiment and mice were sacrificed at the indicated day. For T-cell transfer-induced
 61 colitis, CD4⁺CD45RB^{hi} T-cells from C57BL/6 mice were enriched (CD4⁺ T-Cell
 62 Isolation Kit; Miltenyi Biotec) and single-cell suspensions were stained with APC-
 63 anti-CD4 (GK1.5), and PE-anti-CD45RB (C363.16A), all from eBioscience, followed
 64 by cell sorting (FACS AriaII) (purification > 99%). Rag1^{-/-} recipient mice received 5
 65 × 10⁵ CD4⁺CD45RB^{hi} T-cells by intravenous (i.v.) injection, 1 × 10⁶ Control-
 66 iTregs or PSI-iTregs were injected i.v. 21 days later. At the end of the experiment, the
 67 large intestines of individual mice were dissected out and fixed in 10% phosphate-
 68 buffered formalin.

69 **Statistical analyses**

70 Data are expressed as the mean ± SD. The difference among the groups was were
 71 analyzed by one-way ANOVA and Student *t* test where applicable using GraphPad
 72 Prism 8. *P* < 0.05 was considered statistically significant.

73

74 **Table S1 The sequences of primers**

Real-Time PCR primers		
Gene	Forward primer (5'-3')	Reverse primer (5'-3')
IL-10	CCA AGC CTT GGA AA GA	TTT TCA CAG GGG AGA AAT CG
TGF-β1	AACTGCACCCACTTCCCAGTC	CATTAAGGAGTCGGTTAGCAG
HSPA1A	TGGTGCAGTCCGACATGAAG	GCTGAGAGTCGTTGAAGTAGGC
DNAJB1	TTCGACCGCTATGGAGAGGAA	CACCGAAGAACTCAGCAAACA
HSP90AB1	GTCCGCCGTGTGTTTCATCAT	GCACTTCTTGACGATGTTCTTGC
Actin	AACAGTCCGCCTAGAAGCAC	CGTTGACATCCGTAAAGACC

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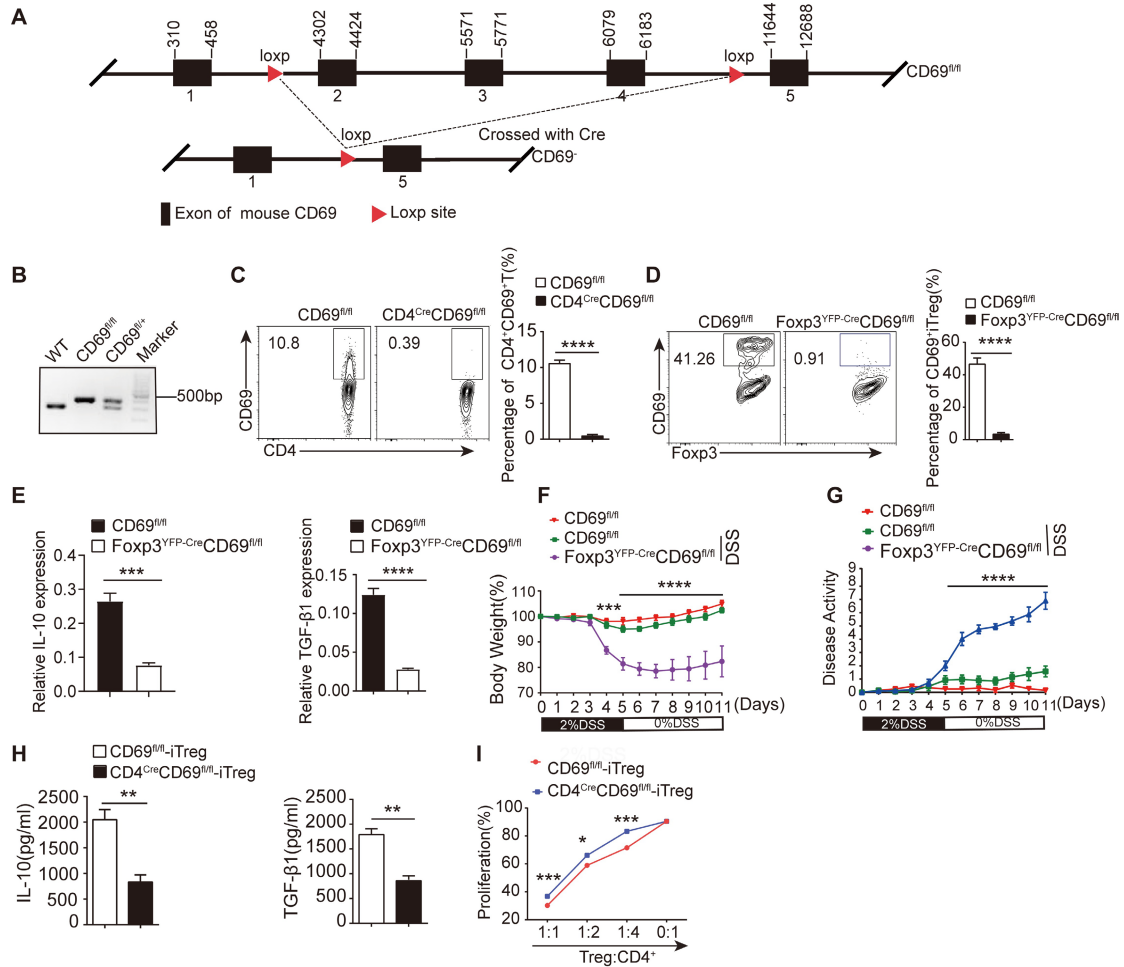
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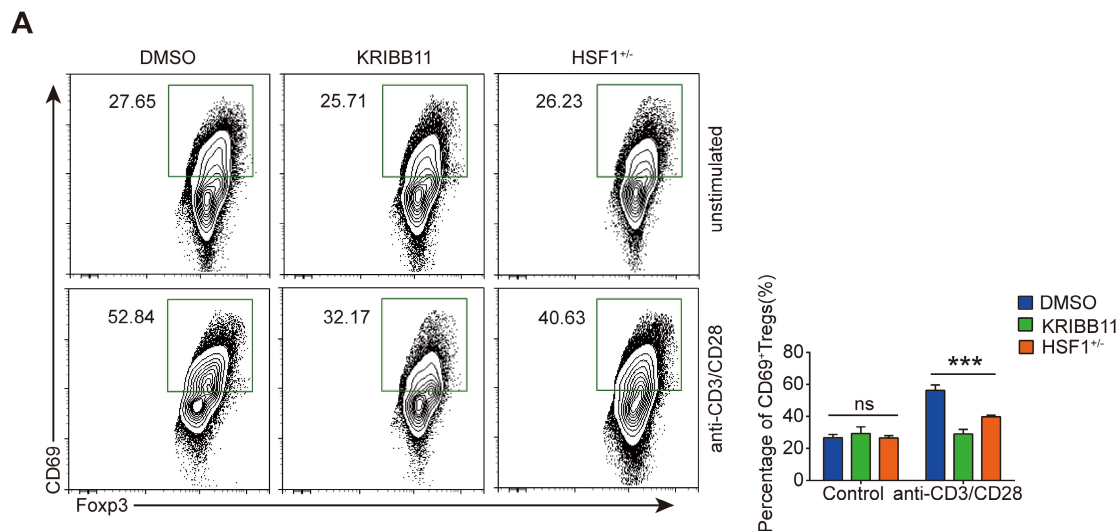
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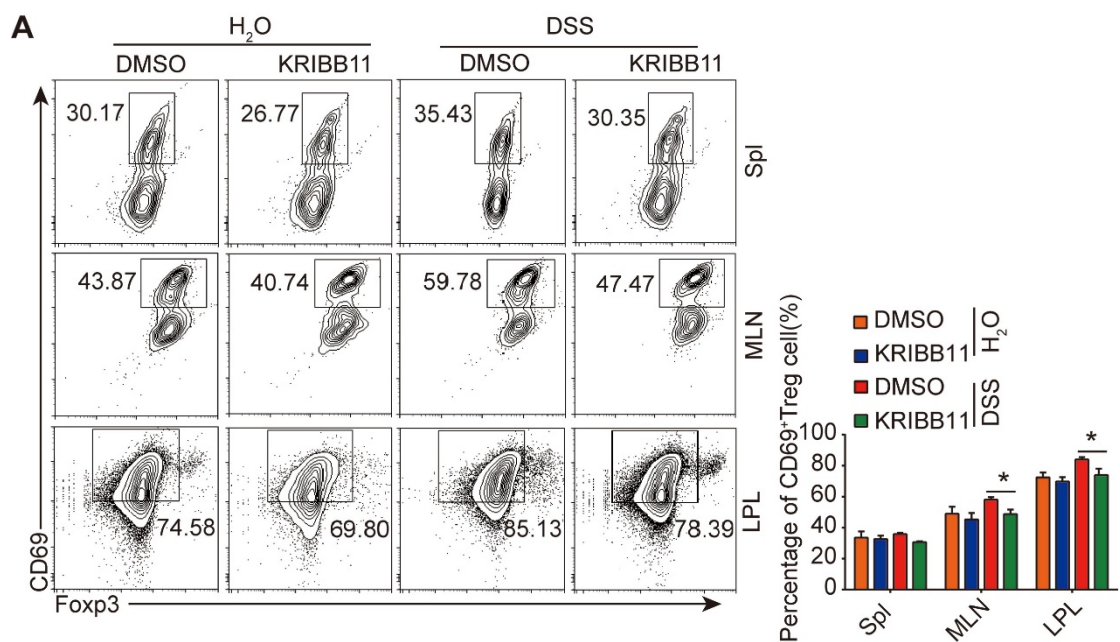
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85 **Figure S1. Conditional targeting of the mouse *CD69* gene in Treg exacerbated**
 86 **DSS-induced colitis. (A)** Strategy of *CD69* conditional knockout mice. **(B)**, Genomic
 87 DNA was isolated from tail of indicated mice and then used for PCR analysis of
 88 *CD69* deletion and the *LoxP*-flanked allele. **(C and D)** Phenotype of *CD69*^{fl/fl} ×
 89 *CD4*^{Cre} or *Foxp3*^{YFP-Cre} offspring, MLN from conditional knock-out mice were used
 90 for analysis the expression of CD69⁺ Tregs. **(E)** The expression of IL-10 and TGF-β1
 91 in Treg from *CD69*^{fl/fl} or *Foxp3*^{YFP-Cre}*CD69*^{fl/fl} mice were analysis by qRT-PCR. **(F and**
 92 **G)** *CD69*^{fl/fl} and *Foxp3*^{YFP-Cre}*CD69*^{fl/fl} mice were administrated with 2.0% DSS in
 93 drinking water for 5 days, and then they were given by normal water in the following
 94 days. Average body weight is shown as percentage relative to initial value. Results are
 95 means ± SD (n = 7). **(H)** The IL-10 and TGF-β1 in supernatant of induced
 96 Foxp3⁺CD4⁺Tregs from *CD69*^{fl/fl} and *CD4*^{Cre}*CD69*^{fl/fl} mice were analysis by ELISA.
 97 **(I)** The proliferation of CD4⁺ T cells was analyzed using flow cytometry. The cells

98 were first gated on living lymphocytes and then on CFSE⁺ T cells (n = 3). Data are
 99 representative of three independent experiments. Comparisons were made using one-
 100 way ANOVA or Student's unpaired *t* test. **P* < 0.05; ***P* < 0.01; ****P* < 0.001;
 101 *****P* < 0.0001.

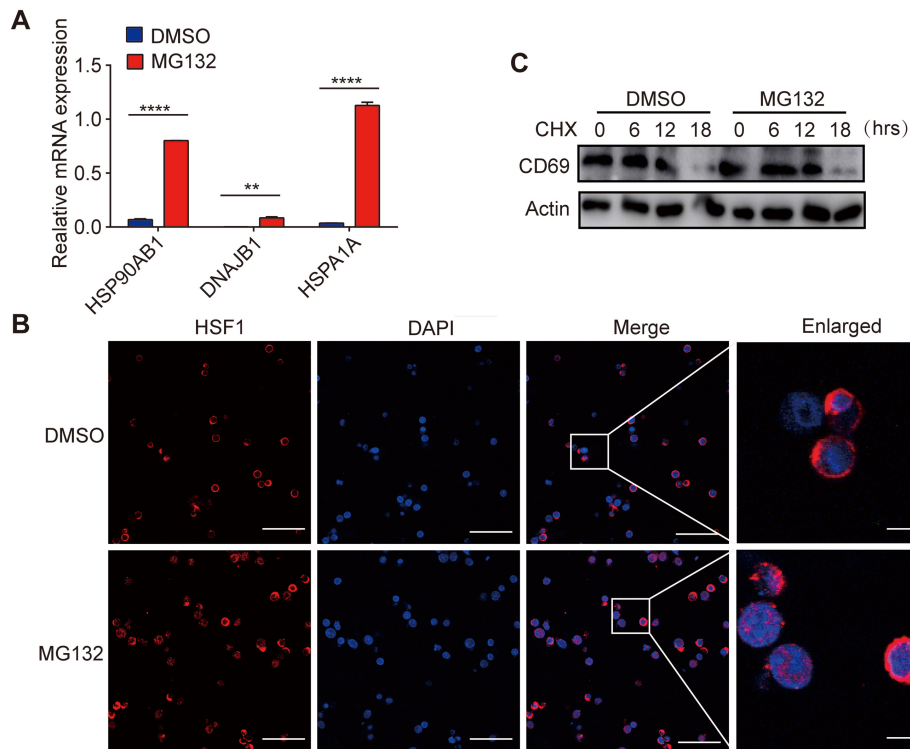


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 103 **Figure S2. Inhibition of HSF1 reduce CD69 expression of Tregs stimulation with**
 104 **anti-CD3/CD28. (A)** 1 × 10⁶/ml Tregs were stimulated with or without anti-
 105 CD3/CD28 antibodies and treated with KRIBB11 for the indicated time. Relative
 106 levels of CD69 expression in different groups of Tregs were determined by FACS.
 107 Results are means ± SD. *** *p* < 0.001, as analyzed by Student's unpaired *t* test.



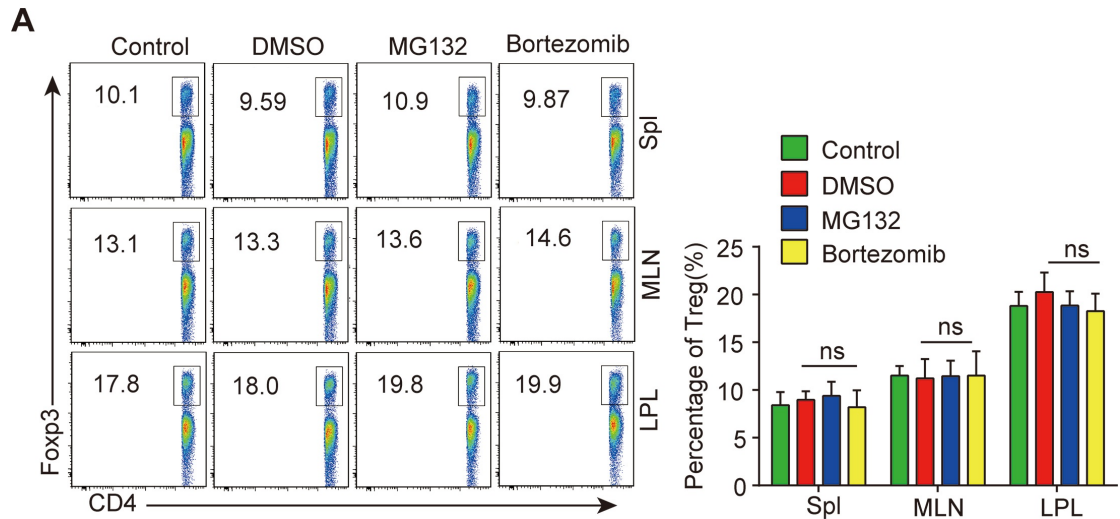
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109 **Figure S3. The differentiation of CD69⁺ Treg cells is inhibited in DSS induced**
 110 **colitis mice treated with KRIBB11.** (A) Spleen, MLN and colonic lamina propria cells
 111 were isolated from control or mice injected with HSF1 inhibitor KRIBB11 after
 112 administration of DSS for 5 days. The frequency of Tregs and CD69⁺ Tregs was
 113 detected by flow cytometry. The graph shows the average percentage of Foxp3⁺ cells
 114 among CD4⁺ T cells and CD69⁺ cells among Tregs. Results are means ± SD (n = 5 mice
 115 per each group). **p* < 0.05, as analyzed by Student's unpaired *t* test.



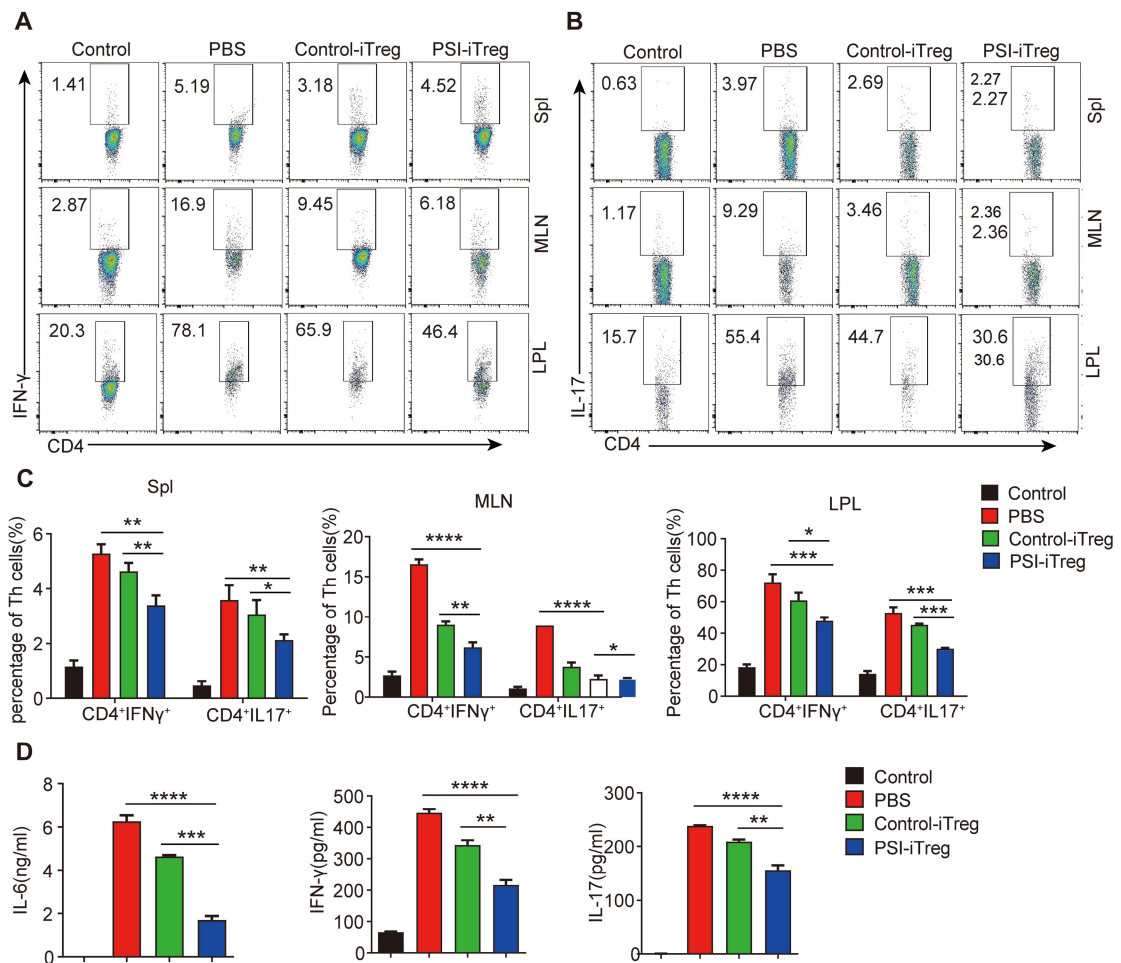
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 117 **Figure S4. MG132 stabilized HSF1 protein to activate the transcription of its**
 118 **target genes.** (A) *HSP90AB1*, *HSP90A1A* and *DNAJB1* normalized to β-actin were
 119 analyzed by qRT-PCR; (B) CD4⁺ T cells were treated with MG132 for 2 h, HSF1
 120 immunofluorescence staining was performed. (C) CD4⁺ T cells were treated with CHX
 121 alone or pretreatment with MG132 for time periods as indicated and CD69 protein
 122 level was measured by western blotting. The data are presented as the means ± SD of
 123 three repeated experiments. ***p* < 0.01, *****p* < 0.0001 as analyzed by Student's
 124 unpaired *t* test.

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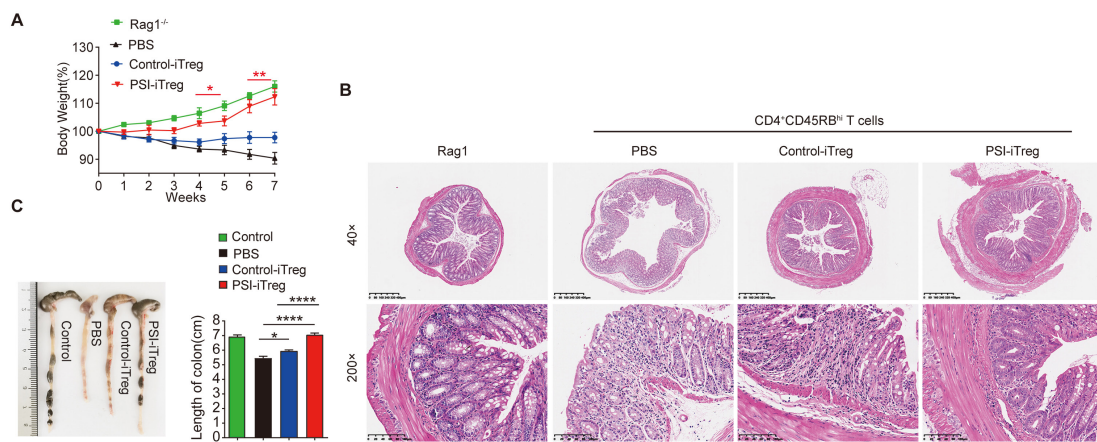
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127 **Figure S5. (A)** Density plots showing total CD4⁺Foxp3⁺ cells from freshly isolated
 128 spleen, MLN and colonic LPL in mice treated with MG132 and Bortezomib.
 129 Representative images of the data expressed as mean \pm SD of three independent
 130 experiments. ns, not significant, as analyzed by ANOVA or Student's *t*-test.



131

132 **Figure S6. PSI-iTreg prevents mice from exacerbation of DSS-induced colitis. (A-**
 133 **C)** Each group of T cells was stimulated with the cell stimulation cocktail for 6 h and
 134 then stained with anti-mouse CD4 and anti-IFN- γ or anti-IL-17 antibodies, followed
 135 by flow cytometry. **(D)** Colon tissue was cultured overnight, and cytokines in the
 136 supernatant were measured by ELISA (n = 5). Representative images of the data
 137 expressed as the mean \pm SD of three independent experiments. * p < 0.05, ** p < 0.01,
 138 *** p < 0.001, **** p < 0.0001, as analyzed by One-way ANOVA or Student's
 139 unpaired t test.



140
 141 **Figure S7. Adoptive transfer of PSI-iTregs inhibits the disease progression in a**
 142 **mouse model of T-cell transfer-induced colitis.** Spleen cells from C57BL/6 mice
 143 were enriched for CD4⁺ T-cells and then stained with anti-CD4 and anti-CD45RB
 144 monoclonal antibodies. Then CD4⁺Foxp3⁻CD45RB^{hi} T-cells were sorted and injected
 145 i.v into immunodeficient *Rag1*^{-/-} mice. Groups of mice were injected i.v with Control-
 146 iTregs and PSI-iTregs (1 \times 10⁶/mouse/injection) on days 21 (n = 7). **(A)** The body
 147 weights were measured for 7 weeks. Each point represents average weight data
 148 pooled from 7 mice \pm SD. **(B)** Histological appearance 7 weeks after colitis induction.
 149 **(C)** Appearance and statistical analysis of colon length. Representative colonic
 150 sections stained with H&E (Magnification: 40 \times and 200 \times). Data are representative
 151 images or expressed as the mean \pm SD of three independent experiments (n = 7 per
 152 group). * P < 0.05, ** P < 0.01, analyzed by ANOVA and Student's t -test.