Supporting Information

Controlled intracellular aggregation of magnetic particles improves permeation and retention for magnetic hyperthermia promotion and immune activation

Ao Hu^{1, 2}, Yiyao Pu^{1, 2}, Na Xu^{1, 2}, Zhongyuan Cai^{1, 2}, Ran Sun³, Shengxiang Fu^{1, 2}, Rongrong Jin^{*, 1, 2}, Yingkun Guo³, Hua Ai^{1, 2}, Yu Nie^{*, 1, 2}, and Xintao Shuai⁴

¹National Engineering Research Center for Biomaterials, Sichuan University, Chengdu, 610064, P. R. China

²College of Biomedical Engineering, Sichuan University, Chengdu, 610065, P. R. China
³Development and Related Diseases of Women and Children Key Laboratory of Sichuan
Province, West China Second University Hospital, Sichuan University, Chengdu 610041,
P. R. China
⁴Nanomedicine Research Center, The Third Affiliated Hospital of Sun Yat-sen University,

Guangzhou 510630, China

Table 1. Primer nucleic acid sequence for detection giant cells formation and macrophages

| Gene Name | Primer sequence (5'-3') |
|----------------|-------------------------|
| Mrc1-Forward | ACGAGCAGGTGCAGTTTACA |
| Mrc1-Reverse | TCAGGAGTTGTTGTGGGGCTC |
| SR-A-Forward | CCAAACGCACTCCCCTTACT |
| SR-A-Reverse | CCACACCAGTAGCAGGACAG |
| CXCL11-Forward | GAACAGGAAGGTCACAGCCATA |
| CXCL11-Reverse | CTCTGCCATTTTGACGGCTTT |
| CD68-Forward | GGGGCTCTTGGGAACTACAC |
| CD68-Reverse | GTACCGTCACAACCTCCCTG |
| CD80-Forward | TTCACCTGGGAAAAACCCCC |
| CD80-Reverse | CCCGAAGGTAAGGCTGTTGT |
| H2-Eb1-Forward | ATAAATTCCTTGTGCGGCGG |
| H2-Eb1-Reverse | CCAGTCTCCATTTCGGACCA |
| TNF-α-Forward | CTGAACTTCGGGGTGATC |
| TNF-α-Reverse | TCCTCCACTTGGTGGTTT |
| iNOS-Forward | CACGGACGAGACGGATAG |
| iNOS -Reverse | CACTGACACTTCGCACAAA |
| IL-1β-Forward | AGCACCTTCTTTTCCTTC |
| IL-1β-Reverse | TGCCGTCTTTCATTACAC |

polarization related gene.

Table S2. Calculated specific absorption rate (SAR) (W/g) and T_2 relaxivity (r_2) (mM⁻¹s⁻¹)

| | SAR (W/g) | $r_2 (\mathrm{mM}^{-1}\mathrm{s}^{-1})$ |
|------------|-----------|---|
| M5 | 41.8 | 48.3 |
| M20 | 560.1 | 279.0 |
| M20&DPA/HA | 547.6 | 232.1 |
| M5&20 | 413.8 | 579.6 |
| M20&20 | 844.4 | 465.1 |

values of various MNPs.

| | White blood cell | Lymphocytes | Red blood cell | Hemoglobin |
|---------|---------------------|----------------|----------------------|---------------------------------|
| | $(\times 10^{9}/L)$ | (70) | $(\times 10^{12}/L)$ | $(\underline{S},\underline{L})$ |
| Control | 1.6 ± 0.2 | 76.8 ± 2.8 | 7.32 ± 0.4 | 143 ± 13 |
| M5&20 | 1.0 ± 0.4 | 71.6 ± 2.7 | 7.03 ± 0.3 | 114 ± 15 |
| M20&20 | 1.8 ± 0.5 | 86.0 ± 2.3 | 7.48 ± 0.3 | 140 ± 10 |

Table S3. Blood routine and biochemical indicators of mice treated with different MNPs.

Continued Table S3. Blood routine and biochemical indicators of mice treated with different MNPs.

| | Hematocrit (%) | Mean corpuscular hemoglobin (pg) | Platelet count (10 ⁹ /L) |
|---------|-------------------|-------------------------------------|-------------------------------------|
| Control | 34.8 ± 3.1 | 18.5 ± 1.2 | 590 ± 35 |
| M5&20 | 44.4 ± 4.2 | 16.9 ± 1.5 | 1061 ± 54 |
| M20&20 | 35.9 ± 3.2 | 18.9 ± 1.4 | 499 ± 42 |



Figure S1. Characterization of various MNPs. (A) XRD spectra of MNPs with 5 nm and 20 nm. (B) FTIR spectra of various MNPs (MNPs, M5, M20, M20@DPA and M20@DPA/HA). (C) Hysteresis loops of M5, M20 and M20@DPA/HA at 20000 Oe.



Figure S2. Characterization of BOC-DPAA and DPA. (A) Synthesis process of BOC-DPAA and DPA. (B) ¹H NMR and (C) mass spectrum of BOC-DPAA. (D) ¹H NMR and (E) mass spectrum of DPA.



Figure S3. Characterization of HA and HA-CHO. (A) Synthesis process of HA-CHO. (B)

¹H NMR and (C) FTIR spectra of HA and HA-CHO.



Figure S4. Characterization of different individual MNPs and pH-responsive aggregation. Size distribution of (A) M5&20 and (B) M20&20 incubated in different pH conditions for 5 h. TEM images of (C) M5&20 and (D) M20&20 incubated in pH 7.4 for 5 h. Scale bars: 50 nm.



Figure S5. Magneto-thermal conversion efficiency and MRI performance of varied MNPs. Temperature change curves over time of individual MNPs and its aggregation under AMF (15 KA/m, 300 kHz) with (A) 0.5 mg Fe/mL and (B) 1.0 mg Fe/mL. (C) T_2 relaxation rate ($1/T_2$, s⁻¹) as a function of Fe concentration (mM) for different MNPs and aggregations

under a 7.0 T magnetic field. Infrared thermal imaging photos of MNPs and aggregations under AMF for 120 s with (D) 0.5 mg Fe/mL and (E) 1.0 mg Fe/mL, respectively.



Figure S6. Infrared thermography of PBS and DMEM with AMF (15 KA/m, 300 kHz) for

20 min.



Figure S7. Killing effect of tumor cells on 4T1 by different MNPs. (A) Generation of ROS after 24 h co-incubation with different MNPs without AMF by fluorescent probe (DCFH-DA assay) staining. (B) Semi-quantitative analysis of ROS generation by MFI of fluorescent probe. (C) Live/dead staining with calcein-AM and PI after co-incubation with different MNPs under AMF. (D) Live/dead staining with calcein-AM and PI after co-incubation with different MNPs in the absence of AMF. Scale bars: 100 μ m. *** P < 0.001.



Figure S8. Immune activation in giant cells formation after treated with different MNPs. (A) Cellular uptake of different MNPs (M20@DPA, A-M5&20, A-M20&20) after 24 h incubation on RAW264.7 by Prussian blue staining. Scale bars: 250 and 50 μ m, respectively. (B) Intracellular Fe concentrations of different MNPs, detected by ICP-OES analysis after 24 h incubation. (C) Detection of giant cells formation in RAW264.7 by

cytoskeleton fluorescence staining with phalloidin (red) and DAPI (blue) after different treatments. Scale bars: 10 μ m. Average number (D) and percentage acreage (E) of the giant cells, calculated from the Prussian blue staining in Figure S8 A. *** P < 0.001.



Figure S9. M1 polarization-related gene expression on RAW264.7 cells after treated with

different MNPs for 24 h. ** P < 0.01, *** P < 0.001.



Figure S10. M1 polarization-related gene expression on RAW264.7 cells treated with AMF for 20 min after treated by different MNPs for 24 h. *** P < 0.001.



Figure S11. (A) Immunofluorescence (IF) staining images of F4/80 and CD86 on tumor tissue sections in control and M20&20 treated group, respectively. (B) MFI of F4/80 and CD86 calculated from the left CLSM images. *** P < 0.001.



Figure S12. (A) Immunofluorescence (IF) staining images of CD3, CD4 and CD25 on tumor tissue sections in control and M20&20 treated group, respectively. (B) MFI of CD3, CD4 and CD25 calculated from the left IF images. *** P < 0.001.



Figure S13. Therapeutic effects of intracellular aggregation of MNPs. (A) Body weight curves of mice treated with different MNPs during the treatment. (B) Tumor weight excised from mice post 15-day treatment. (C) The percentage of necrosis area (%) calculated from each tumor (n = 5). *** P < 0.001.



Figure S14. The blood biochemical indicators related to liver and kidney function after indicated treatment.



Figure S15. Biodistribution of M5&20 and M20&20 in 4T1 mouse mammary tumor model

via 24 h-intravenous injection. *** P < 0.001.



Figure S16. Fe concentration of tumor treated by M5&20 and M20&20 for 24 h. ** P <

0.01.



Figure S17. H&E analysis on sections of main organs at day 15 post treatment. Scale bars:

200 $\mu m.$ Data are presented as mean \pm SD (n = 5).



Figure S18. Number of lung metastatic nodules on day 15 after treated with different groups. *** P < 0.001.



Figure S19. (A) In vivo *T*₂-weighted images of various MNPs at the predetermined time.(B) SNR values of various MNPs based on MRI signal over time.