Essential role of IL-17 in acute exacerbation of pulmonary fibrosis induced by non-typeable *Haemophilus influenzae*

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Figure S1. Establishment of pulmonary fibrosis mouse model. (A) The diagram illustrating the establishment of the mouse model of bleomycin (BLM)-induced pulmonary fibrosis. **(B)** Body weight changes of the mice intranasally administrated with BLM in dosages of 60 μ g, 30 μ g and 15 μ g per mouse severally. **(C)** Survival rate comparison among the mice intranasally administrated with BLM in dosage of 60 μ g, 30 μ g and 15 μ g per mouse severally. **(D)** Representative images of lung sections stained with H&E on day 0, 3, 7, 14 and 21 after BLM administration in the dosages of 15, 30 and 60 μ g per mouse severally (scale bar: 500 μ m). Data are expressed as the mean±SEM of 5-10 mice/group. *P < 0.05; **P < 0.01; ***P < 0.001.



Figure S2. The airway damage and fibrosis induced by instillation of BLM. Lung tissues were collected on day 0, 3, 7, 14, and 21 after 30 μ g BLM administration. (A and B) Immunostaining with an antibody to the alveolar epithelial type I cells (AECI, RAGE⁺) and alveolar epithelial type I cells (AECI, SPC⁺). (C) Histological analysis of lung sections showing proliferating basal cells (Krt5⁺, area within the yellow dotted line) on day 7 and 14 after BLM administration. (D and E) Pulmonary fibrosis was evaluated by the immunostaining with an antibody to myofibroblasts (α -SMA⁺) and Masson's trichrome staining specific for collagen fibers. DAPI: represents the nucleus. Scale bars: A, B, D and E: 50 µm; C: 100 µm.



Figure S3. Kinetics of T cell response to NT127 infection. (A) Kinetics of NT127-specific T cells producing IL-17 (IL-17⁺T cell) and IFN- γ (IFN- γ^+ T cell) were determined in the lung and spleen of mice. **(B)** Statistical analyses for the results of (A). Data are expressed as the mean±SEM (n=5 mice/group). *P < 0.05; **P < 0.01; ***P < 0.001.



Figure S4. Gating strategy used for the identification of cytokines-producing T cell subsets in murine lung.



Figure S5. The characteristics of splenic T cells response in AE-IPF mice. IL-17 and IFN- γ production by total T cells (CD3⁺) after stimulation with PMA/Ionomcyin as visualized by flow cytometry and calculated as the percentage and number of IL-17⁺CD3⁺ and IFN- γ ⁺CD3⁺ in the speen of naive, BLM-instilled, NT127-infected and BLM/NT127-instilled mice on day 7 after NT127 infection (day 14 after BLM administration). Data are expressed as the mean±SEM(n=5 per group), *P < 0.05; **P < 0.01; ***P < 0.001.



Figure S6. Weak IL-22-producing CD4⁺ and $\gamma\delta$ **T cells response in the lung.** IL-22 secretion by pulmonary CD4⁺ and $\gamma\delta$ T cells after stimulation with PMA/Ionomcyin as visualized by flow cytometry and calculated as the percentage and number of IL-17⁺CD4⁺, IFN- γ ⁺CD4⁺, IL-17⁺ $\gamma\delta$ in naive, BLM, NT127 and BLM/NT127 mice on day 7 after NT127 infection (day 14 after BLM administration). Data are expressed as the mean ±SEM (n=5 per group), *P < 0.05; **P < 0.01; ***P < 0.001.



Figure S7. Impact of IL-17 on TGF- β production by pulmonary CD4⁺ and $\gamma\delta$ T cells in AE-IPF mice. TGF- β expression by pulmonary CD4⁺ and $\gamma\delta$ T cells after stimulation with PMA/Ionomcyin in WT and IL-17 KO mice were analysed by flow cytometry and calculated as the percentage and number of TGF- β ⁺CD4⁺ (A) and TGF- β ⁺ $\gamma\delta$ T cells (B) after intranasally instilling with BLM, NT127 and BLM/NT127. Data are expressed as the mean±SEM (n=5 per group), *P < 0.05; **P < 0.01; ***P < 0.001.



Figure S8. Gating strategy used for the identification of major inflammatory cell populations in murine lung. Gates containing multiple cell populations are numbered (R1-R7). Gates containing a single cell population are labeled with neutrophil, eosinophil and macrophage.