

Supplementary information

Supplementary Figures

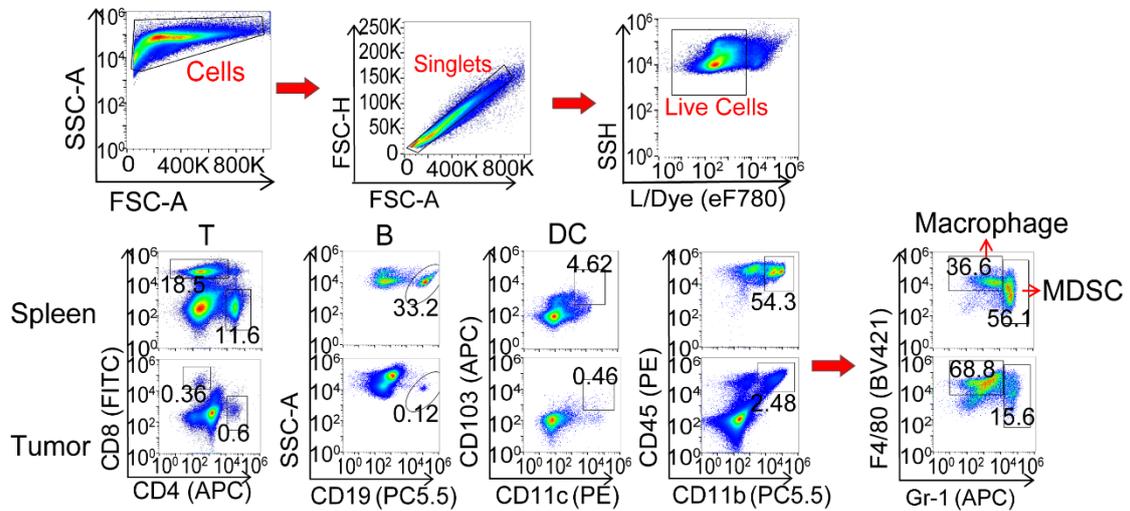


Figure S1. Gating strategy of immune cells in MC38 tumor bearing mice. Gating strategy for T cells (CD3⁺CD8⁺ or CD3⁺CD4⁺), B cells (CD19⁺), DCs (CD11c⁺CD103⁺), MDSCs (CD11b⁺CD45⁺Gr1⁺) and macrophages (CD11b⁺CD45⁺F4/80⁺) in mouse spleen and tumor.

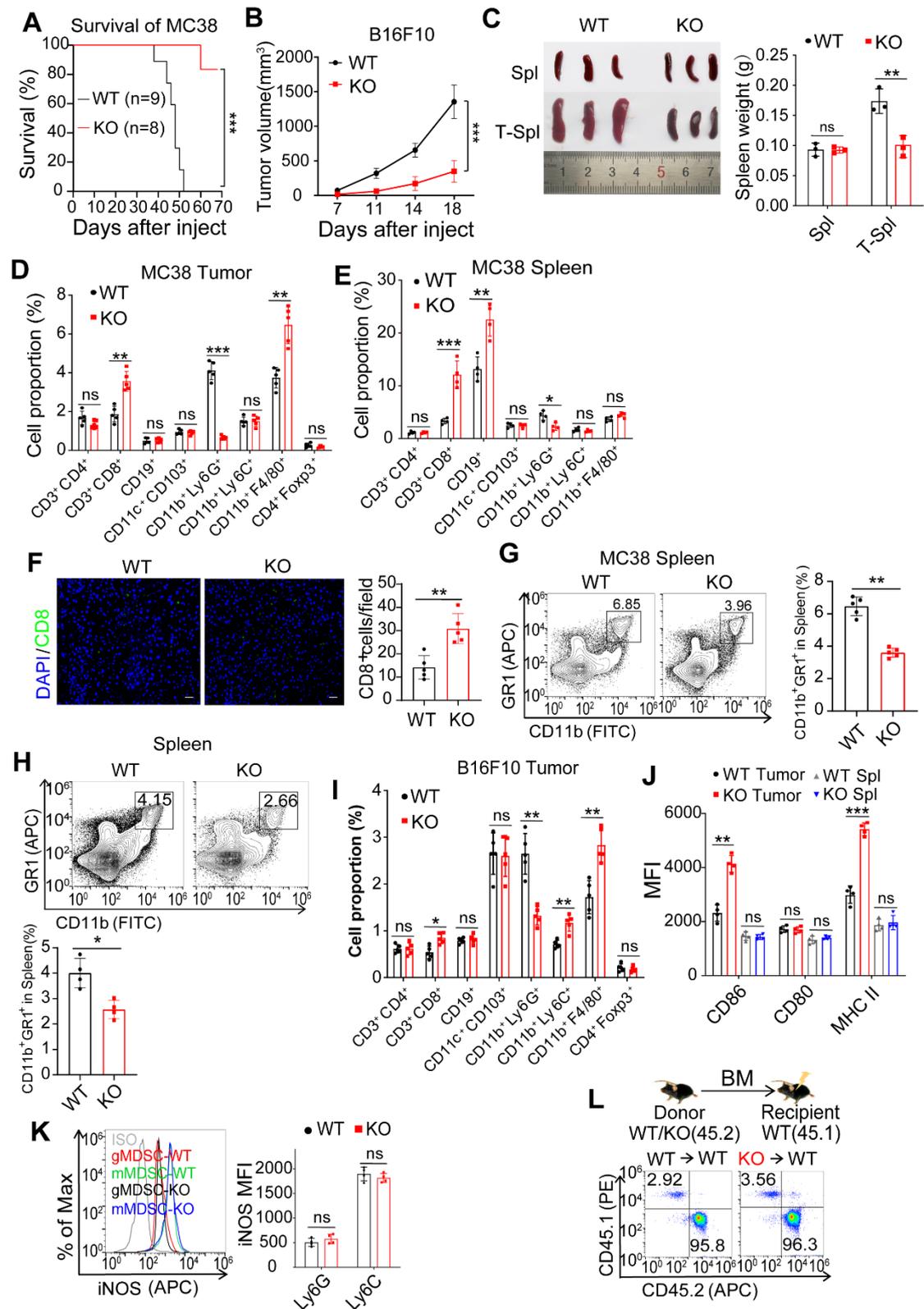


Figure S2. FcγRIIB deficiency reduces MDSCs in the TME. (A) Survival of MC38 tumor-bearing WT or FcγRIIB^{-/-} (KO) mice, one of three representative experiments is shown, with $n = 8-9$ mice each group (log-rank [Mantel-Cox] test, $p < 0.001$). (B) A total of 5×10^5 B16F10 melanoma cells were subcutaneously injected into WT or KO mice, tumor growth was monitored every 3 or 4 d. (C) WT or KO mice were injected

subcutaneously with 10^6 MC38 cells, mice were sacrificed at day 21 post-grafting, and spleen from tumor free mice (Spl) and tumor bearing mice (T-Spl) were collected and weighed, representative spleen images are shown, $n = 3$. **(D and E)** WT or KO mice were injected subcutaneously with 10^6 MC38 cells, Mice were euthanized on day 14, the frequency of T cells, B cells, DCs, MDSCs, macrophages and Tregs in tumor tissues or spleen were determined by FCM. **(F)** The frequency of $CD11b^+Gr1^+$ MDSCs in spleen from WT or KO tumor bearing mice was assessed 14 days after MC38 tumor inoculation, $n = 5$. **(F)** The $CD8^+$ cells in MC38 tumor tissue sections from WT or KO mice was detected by immunofluorescence. Scale bars: 50 μm . **(G)** The frequency of $CD11b^+Gr1^+$ MDSCs in spleen from WT or KO tumor bearing mice was assessed 14 days after MC38 tumor inoculation, $n = 5$. **(H)** The frequency of $CD11b^+Gr1^+$ cells in spleen from WT or KO naïve mice was determined by FCM, $n = 4$. **(I)** WT or KO mice were injected subcutaneously with 5×10^5 B16F10 melanoma cells for 14 d, the frequency of T cells, B cells, DCs, MDSCs, macrophages and Tregs in tumor tissues were determined by FCM, $n = 4$. **(J)** CD86, CD80 and MHC II expression for $CD11b^+F4/80^+$ cells from tumor and spleen were determined by FCM, $n = 4$. **(K)** iNOS expression in gMDSCs and mMDSCs from WT or KO MC38 tumor tissues were analyzed by FCM; $n = 4$. **(L)** Recipient mice were irradiated and then received WT or $Fc\gamma RIIb^{-/-}$ mouse-derived BM cells. CD45.1 and CD45.2 expressions on peripheral blood MDSCs were detected after BM reconstitution. Data are expressed as means \pm SD. $*P < 0.05$, $**P < 0.01$, $***P < 0.001$, by Mann-Whitney test. *ns*, no significant difference.

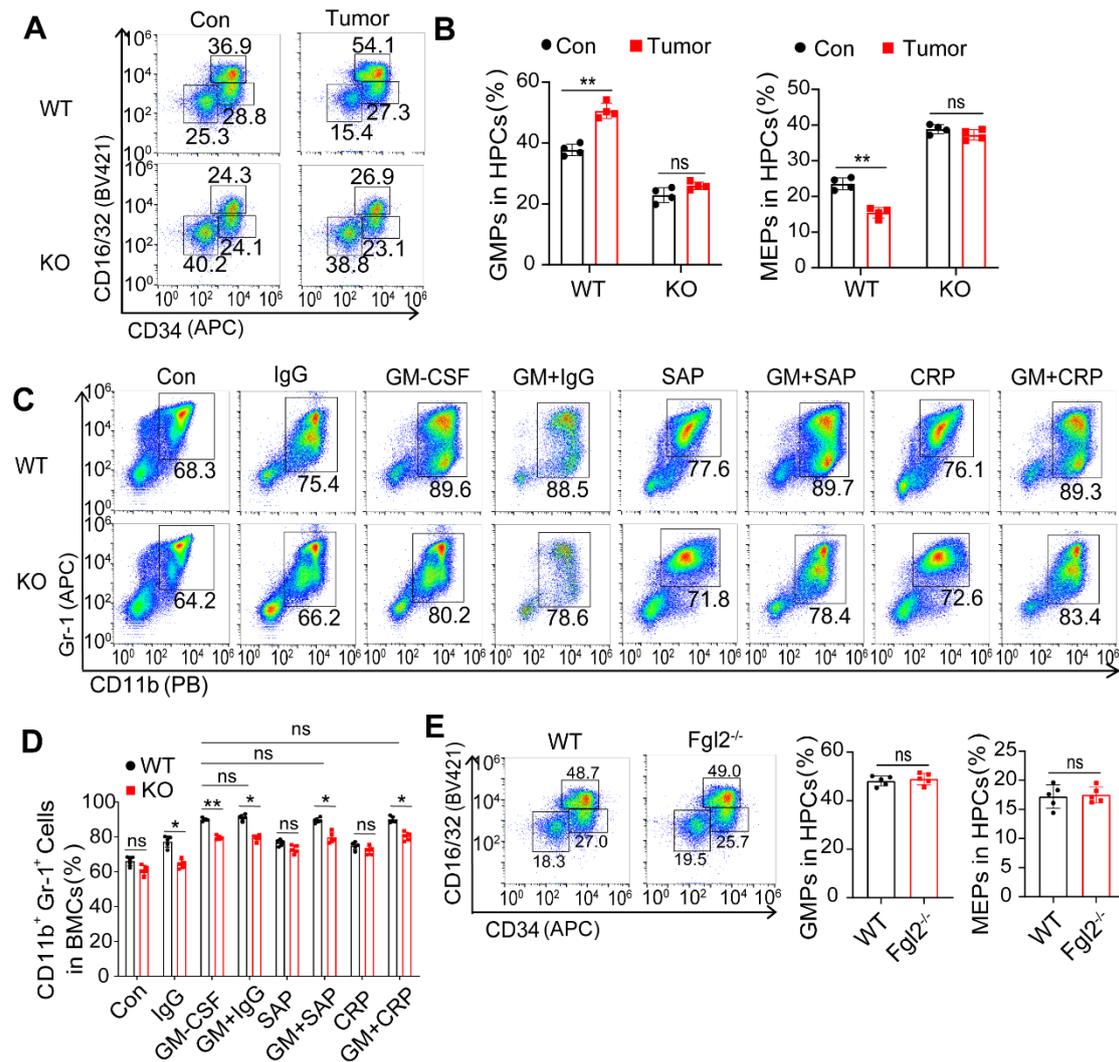


Figure S3. Fc γ RIIB promotes the differentiation of GMPs from HPCs independent of IgG, SAP, CRP and Fc γ 2. (A) The percentages of GMPs, CMPs and MEPs in HPCs from WT and KO tumor free or tumor bearing mice were analyzed. (B) Statistical results of A, $n = 5$. (C) WT and KO BMs were treated with GM-CSF/IL-6 (20 ng/mL) to induce MDSCs differentiation for 72h in the presence or absence of IgG (1 μ g/mL), SAP (20 ng/mL) or CRP (20 ng/mL). Cells were then harvested and MDSCs were assessed *via* FCM. (D) Statistical results of (C), $n = 4$. (E) WT or Fc γ 2^{-/-} mice were injected subcutaneously with 10⁶ MC38 cells, mice were sacrificed at day 14 post-grafting of MC38 cells and the percentages of GMP, CMP and MEP in HPCs were analyzed, $n = 5$. Data are expressed as means \pm SD. **P < 0.01, Mann-Whitney test. ns, no significant difference.

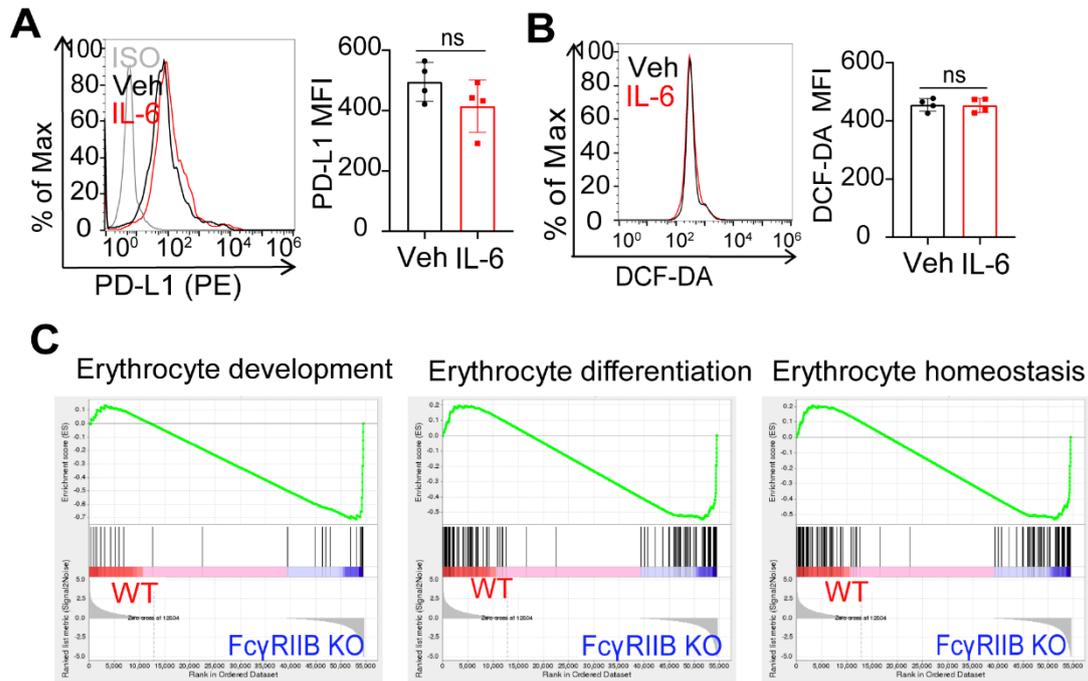


Figure S4. Fc γ RIIB deficiency promotes HPCs differentiation to MEPs through impairing Stat3 pathway. (A and B) Isolated Gr-1⁺ cells from KO mice were treated with IL-6 (20ng/mL) for 48 h, the expression of PD-L1, and DCF-DA were measured, $n = 4$. (C) Enrichment plot of the HALLMARK erythrocyte development, erythrocyte differentiation and erythrocyte homeostasis signaling pathway for the comparison between WT or Fc γ RIIB KO HPCs. Data are expressed as means \pm SD. Mann-Whitney test. *ns*, no significant difference.

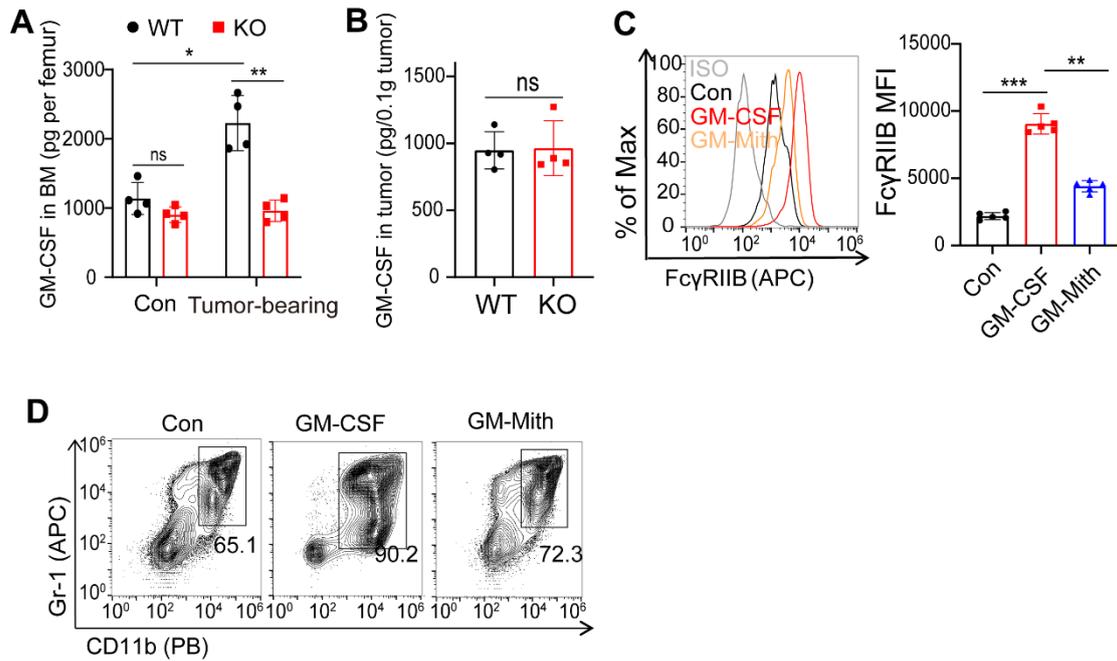


Figure S5. Tumor-bearing WT mice express high levels of GM-CSF in BM. (A and B) WT or KO mice were injected subcutaneously with 10^6 MC38 cells, mice were sacrificed at day 21 post-grafting, the GM-CSF expression levels in BM and tumor tissues ($n = 4$) were measured using a Mouse GM-CSF ELISA kit, $n = 4$. (C) THP1 cells were treated with GM-CSF (20ng/mL) for 48 h, in the presence or absence of Mith and FcγRIIB expression were assessed using FCM; $n = 5$. (D) BM cells were treated with GM-CSF in the presence or absence of 1 μ l PBS (Con) or Mithramycin A (Mith, 20 nM) for 48 h, the percentages of MDSCs were assessed. Data are expressed as means \pm SD. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, Mann-Whitney test. *ns*, no significant difference.

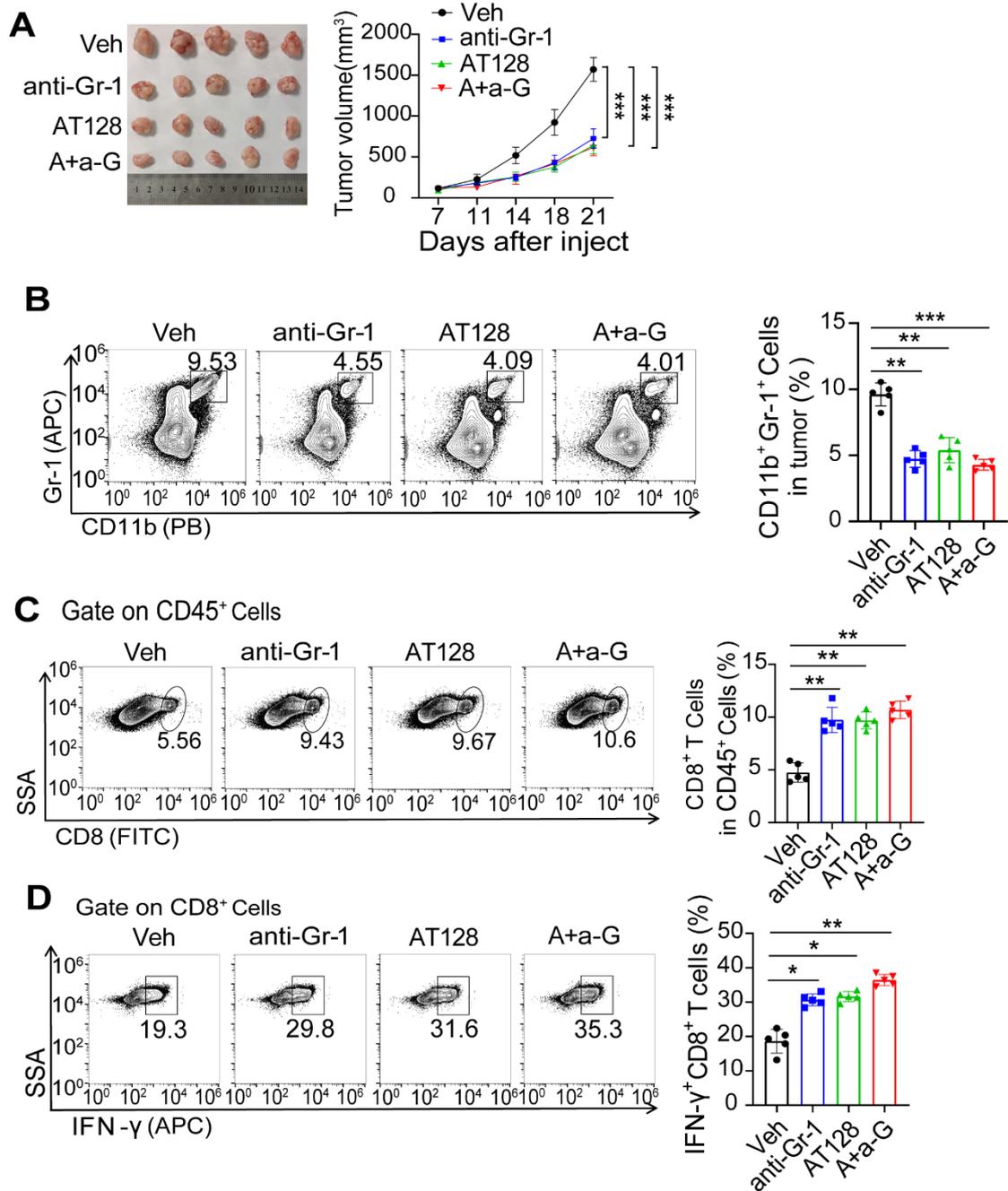


Figure S6. Blocking FcγRIIB signaling decreases MDSC infiltration and promotes CD8⁺ T cell activity. (A-D) WT mice were injected subcutaneously with MC38 tumor cell. After 7 days, tumor-bearing mice were injected with PBS (Veh), 250 μg of the anti-mouse Gr-1 antibody, 250 μg anti-mouse FcγRIIB antibody (AT128) or combined treatment (A+a-G). Tumor growth was monitored for 21 days (A). All mice were euthanized on day 21 after tumor injection, the percentages of MDSCs, the percentages of CD8⁺ T cells in tumor tissues (B), and the percentage of CD8⁺ T cells (C) producing IFN-γ (D, $n = 5$) in tumor tissues were analyzed by FCM. One-way ANOVA with Tukey multiple comparison posttest was used to evaluate statistical significance. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$. *ns*, no significant difference.

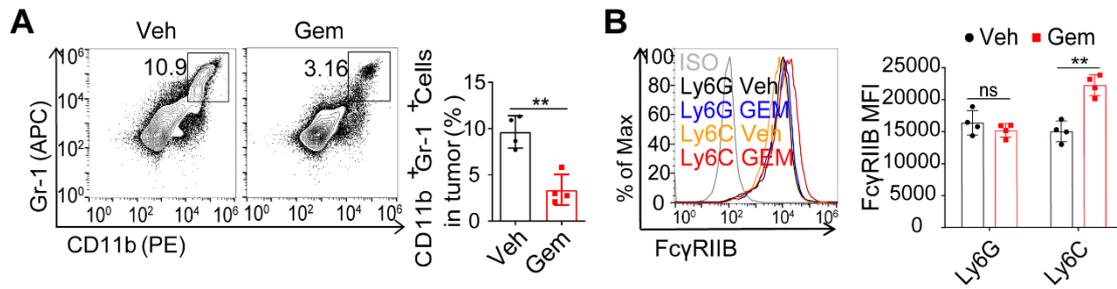


Figure S7. Fc γ RIIB is upregulated in tumor-infiltrating MDSCs from mice received gemcitabine treatment. (A) WT mice were injected subcutaneously with MC38 tumor cell. After 7 days, tumor-bearing mice were injected *i. p.* with 100 μ l PBS (Veh), gemcitabine (Gem, 50 mg/kg), the percentages of CD11b⁺Gr1⁺ in MC38 tumor tissues was analyzed 21 days after tumor inoculation, $n = 4$. (B) The expression of Fc γ RIIB on gMDSCs and mMDSCs from MC38 tumor were determined, $n = 4$. Data are expressed as means \pm SD. ** $P < 0.01$, by Mann-Whitney test. *ns*, no significant difference.