

Review



2021; 11(16): 8057-8075. doi: 10.7150/thno.59677

# Emerging role of circulating tumor cells in immunotherapy

Alexey Rzhevskiy<sup>1,2,3<sup>IC</sup></sup>, Alina Kapitannikova<sup>2</sup>, Polina Malinina<sup>4</sup>, Arthur Volovetsky<sup>2,5</sup>, Hamidreza Aboulkheyr Es<sup>6</sup>, Arutha Kulasinghe<sup>7,8</sup>, Jean Paul Thiery<sup>2,9</sup>, Anna Maslennikova<sup>5,10</sup>, Andrei V. Zvyagin<sup>1,2,11</sup> and Majid Ebrahimi Warkiani<sup>2,6</sup>

- ARC Centre of Excellence for Nanoscale BioPhotonics, Macquarie University, Sydney, NSW 2109, Australia 1
- Institute of Molecular Medicine, Sechenov First Moscow State Medical University, 119991 Moscow, Russia 2
- Institute for Urology and Reproductive Health, Sechenov University, Moscow 119991, Russia 3.
- Privolzhsky Research Medical University, 10/1, Minini Pozharsky Square, Nizhny Novgorod 603005, Russia 4
- Lobachevsky State University of Nizhny Novgorod, Gagarina Avenue 23, Nizhny Novgorod 603950, Russia 5.
- School of Biomedical Engineering, University of Technology Sydney, 2007 Sydney, Australia 6.
- Queensland University of Technology, Centre for Genomics and Personalised Health, School of Biomedical Sciences, Faculty of Health, Woolloongabba, 7
- OLD 4102, Australia 8 Translational Research Institute, Woolloongabba, QLD 4102 Australia
- 9.
- Guangzhou Institutes of Biomedicine and Health, Guangzhou, People's Republic of China 10. The Chair of Cancer, Radiotherapy and Radiologic Diagnostics, Privolzhsky Research Medical University, Nizhniy Novgorod. Russia 603005
- 11. IBCh Shemyakin Ovchinnikov Institute of BioOrganic Chemistry of the Russian Academy of Sciences, Miklukho Maklai Street, 16, Moscow, Russia

🖂 Corresponding authors: alexey.rzhevskiy@hdr.mq.edu.au; andrei.zvyagin@mq.edu.au; Majid.Warkiani@uts.edu.au. School of Biomedical Engineering, University Technology Sydney, Sydney, New South Wales 2007, Australia

© The author(s). This is an open access article distributed under the terms of the Creative Commons Attribution License (https://creativecommons.org/licenses/by/4.0/). See http://ivyspring.com/terms for full terms and conditions

Received: 2021.02.22; Accepted: 2021.06.17; Published: 2021.07.06

#### Abstract

Over the last few years, immunotherapy, in particular, immune checkpoint inhibitor therapy, has revolutionized the treatment of several types of cancer. At the same time, the uptake in clinical oncology has been slow owing to the high cost of treatment, associated toxicity profiles and variability of the response to treatment between patients. In response, personalized approaches based on predictive biomarkers have emerged as new tools for patient stratification to achieve effective immunotherapy. Recently, the enumeration and molecular analysis of circulating tumor cells (CTCs) have been highlighted as prognostic biomarkers for the management of cancer patients during chemotherapy and for targeted therapy in a personalized manner. The expression of immune checkpoints on CTCs has been reported in a number of solid tumor types and has provided new insight into cancer immunotherapy management. In this review, we discuss recent advances in the identification of immune checkpoints using CTCs and shed light on the potential applications of CTCs towards the identification of predictive biomarkers for immunotherapy.

Key words: Immunotherapy, circulating tumor cells, immune checkpoint inhibitors, cancer, personalized medicine

## Introduction

Circulating tumor cells (CTCs) were first discovered in 1869 by an Australian physician, Thomas Ashworth, who identified cells that were morphologically similar to those of the primary tumor in the blood of metastatic cancer patients [1]. CTCs were defined as single cells or cell clusters that are metastatic precursor cells responsible for the development of metastasis [2]. They have gained

significant attention as a promising biomarker for the diagnosis and prognosis of malignant tumors. However, at the early stages of cancer, CTCs are rare, typically several CTCs per milliliter of peripheral blood on an excessively high background of normal blood cells [3]. This necessitates enrichment of CTCs to apply analytical protocols. Numerous methods have been introduced for the enrichment of CTCs,

which are classified based on their operational principle into affinity-based and label-free isolation approaches (Figure 1).

The practical value of CTCs for clinical applications has been considered limited owing to inefficient CTC capture methodologies [4]. The CellSearch FDA-approved (Menarini Silicon Biosystems) technology has been applied for CTC enumeration across a number of tumor types [5]. In the CellSearch studies, EpCAM-positive CTCs counts were associated with progression-free survival (PFS) and overall survival (OS) in metastatic prostate, colorectal and breast cancer. However, later, it was identified that this approach had a relatively low efficiency of identifying CTCs at the early stages of various cancer types [6]. On the other hand, the rapid development of microelectromechanical systems over the last 20 years has considerably advanced the CTC enrichment technologies, which has resulted in an increase of CTC's yield available for analysis and processing. As a result, several viable approaches for epitope dependent/independent CTC enumeration

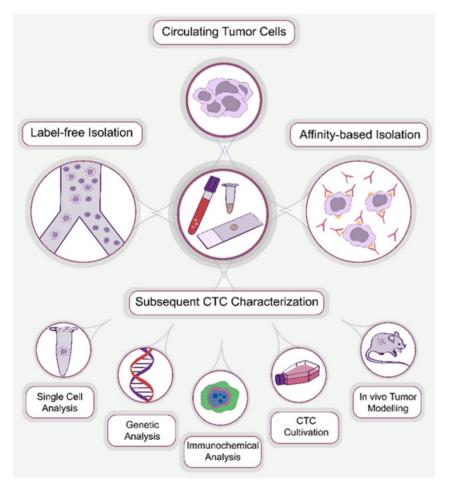


Figure 1. CTC processing methods. Generally, the approaches used for the enrichment of CTCs can be divided into two main groups: affinity-based and affinity-independent (label-free). After enrichment, CTCs are subjected to subsequent characterization, including single cell, genetic and immunocytochemical analysis, cultivation and in vivo tumor modeling.

and detection have emerged. The extensive development of CTC enrichment technologies has reached a point at which the main challenge has shifted towards molecular and genetic analysis of CTCs captured by different technologies, identification of CTC subpopulations and correlative analysis of the total CTC enumeration and subpopulations.

In turn, immunotherapy has revolutionized the field of oncology by providing a mechanism for the activation of immune cells that can recognize and destroy cancer cells [7]. This is realized by identifying and applying immune checkpoint inhibitors (ICIs) that target immune checkpoint proteins expressed by immune cells and cancer cells. This leads to downregulation of immune cell activation and surveillance [8], which leads to suppressed T-cell activity. The described mechanism is mediated through the binding of cytotoxic T-lymphocyte-associated antigen 4 (CTLA-4) carried by T-cells to inhibitory proteins CD80/CD86 located on the surface of antigen-presenting cells, which are predominantly

situated in lymph nodes. Alternatively, T-cell suppression can be achieved bv binding а programmed death 1 (PD-1) protein, also expressed on the surface of T-cells, to its ligand (PD-L1) on the surface of either T-cells or tumor cells predominantly located in the tumor microenvironment [9]. Numerous drugs that block either CTLA-4 or PD-L1/PD-1 interactions have been developed and tested in various clinical trials, including ipilimumab and tremelimumab for CTLA-4, pembrolizumab nivolumab, and cemiplimab for PD-1. and atezolizumab, durvalumab and avelumab for PD-L1. Thus, assessment of the status of immune checkpoints in patients is thought to be one of the crucial aspects in personalized immunotherapy [10, 11]. Recent studies have highlighted that CTCs frequently express PD-L1, which could provide a useful and non-invasive means to assess PD-L1 status in real-time [12, 13].

In this review, we discuss the emerging role of CTCs as a biomarker in evaluating the efficiency of immunotherapy in various types of solid tumors, particularly with ICIs either alone or in combination with other immunotherapy approaches. Additionally, the diagnostic and prognostic potential and the role of CTCs in personalized cancer treatment is discussed. The review is focused on urologic, gynecologic, breast, lung, head and neck, gastrointestinal cancers and melanoma.

# Search strategy

PubMed has been chosen as the data base of relevant publications. For the current review, full-text articles in English published in peer reviewed journals were selected. Also, ClinicalTrials.gov was searched for relevant clinical trials in which cancer immunotherapy was performed along with the qualitative or quantitative assessment of CTCs. The databases were searched up to December 2020. The keywords used in the literature search were as follows: circulating tumor cells, CTC(s), immunotherapy, immune checkpoint inhibitor(s), immune checkpoint blockade, immune checkpoint(s), programmed cell death protein 1, PD-1, cytotoxic T-lymphocyte-associated protein 4, CTLA-4, programmed death-ligand 1, PD-L1, bladder cancer, urothelial carcinoma, transitional cell carcinoma, prostate cancer, kidney cancer, renal cancer, renal cell carcinoma, ovarian cancer, endometrial cancer, cervical cancer, breast cancer, head and neck cancer, squamous head and neck cell carcinoma, oropharyngeal cancer, hypopharyngeal cancer, laryngeal cancer, lip cancer, oral cavity cancer, nasopharyngeal cancer, paranasal sinus cancer, nasal cavity cancer, salivary gland cancer, thyroid cancer, esophageal cancer, gastric or stomach cancer, colorectal cancer, pancreatic cancer, liver cancer, melanoma.

# **Urologic cancers**

Immunotherapy of the non-muscle-invasive form of bladder cancer through instillation of the bladder with Bacillus Calmette-Guerin (BCG) vaccine has been a standard of care for urologic cancers worldwide for decades [14]. Recently, along with FDA approval of ICIs for the treatment of urologic cancers, promising results have been reported in studies addressing the ICI treatment of renal cell carcinoma [15], urothelial [16] cancer and the use of therapeutic vaccines for the treatment of prostate cancer (PCa) [17]. Tumor mutation burden (TMB) and microsatellite instability (MSI) have been considered as promising biomarkers for the prediction of immunotherapy outcomes in urologic cancers [18]. CTCs have also been actively investigated as predictive biomarkers of an immunotherapy response by measuring the expression of immune checkpoints on CTCs including PD-L1.

In urologic cancers, CTCs have been mostly investigated as predictive biomarkers in the PCa immunotherapy regimens, in which the immune checkpoint blockade (ICB) has attracted significant attention. However, monotherapy with ICB has demonstrated modest efficiency [19]. At the same time, it was suggested that castration-resistant PCa expressing and rogen receptor splice variant 7 (AR-V7) might be associated with a higher level of MSI and subsequently higher TMB [20]. AR-V7 is an isoform of the androgen receptor [21] and can be a cause of an aggressive form of the disease and resistance to therapies, including hormonal those with enzalutamide and abiraterone [22]. This calls for the development of efficient therapies of AR-V7 positive PCa to cover the unmet need, in which case ICB immunotherapy is considered a promising treatment modality.

Further, combined ICI therapies, particularly with anti-cancer vaccines [23], are of significant interest. In a preclinical study, Fu et al. [24] identified an immunotherapeutic mechanism of the anti-cancer vaccines that might induce an antigen-specific IFNy-secretion by Т cells in the tumor microenvironment. This leads to an increment of PD-L1 expression, with a subsequent uprise in PD-1 antibody blockade, as demonstrated by the administration of the IFNy-inducing cancer vaccine and nivolumab in combined therapy. Therefore, such a combined approach may be beneficial in PCa immunotherapy.

In a recent study by Rekoske *et al.* [25], alterations in PD-L1 expression on CTCs of PCa patients at various stages of the disease mediated by vaccination with DNA vaccine encoding prostatic acid phosphatase were associated with elevation of the immune response and prolonged PFS. A significant increase of PD-L1 expression levels by CTCs was detected three months post-treatment, with comparable expression levels at the pre-treatment and one year post-treatment. A perspective approach of combination immunotherapy using anti-tumor vaccination and PD-1 blockade by pembrolizumab, coupled with CTCs as predictive biomarkers of PD-L1 expression, is currently being assessed in an ongoing clinical trial (NCT02499835).

A phase-2 clinical trial was conducted to evaluate the efficiency of combined ICB treatment of AR-V7 positive metastatic PCa with ipilimumab and nivolumab [26]. In the study, CTCs were assessed for markers such as ARV-7, DNA-repair gene mutations and phenotypic heterogeneity, and interrelation between the markers and an outcome of the combined ICB. The results showed that the presence of various mutations in DNA-repair-related genes in isolated CTCs and their phenotypic heterogeneity were associated with improved clinical outcomes in ARV-7 positive patients.

Furthermore, the prognostic significance of PD-L1 expression in CTCs has been recently reported. In a study by Satelly *et al.*[27], 30 metastatic PCa patients were recruited at different cycles of palliative chemotherapy and their blood was assessed for CTCs at random time points. The results showed that nuclear PD-L1 (nPD-L1) expression was associated with significantly worse PFS than OS. Furthermore, Yin *et al.* [28] reported no association between PCa stages and the expression of the antigen on CTCs, in which the expression level of PD-L1 was investigated in patients at different stages of cancer.

Urothelial bladder cancers were also identified to express PD-L1 [29]. In a recent study by Anantharaman et al. [30], CTCs were measured in 25 with metastatic patients (21 patients) and muscle-invasive (4 patients) forms of bladder cancer. Among them, CTCs were detected in blood samples of 20 patients, of which seven patients diagnosed with a metastatic form of bladder cancer showed PD-L1 expression on their CTCs. Further, four patients had more than one PD-L1 positive (PD-L1<sup>+</sup>) CTC per milliliter of blood, which was associated with a median OS of 194 days compared with 303 days in patients having 0-1 PD-L1+ CTCs per mL of blood. Noticeably, four patients had PD-L1 expression only on cytokeratin-negative cells, which may potentially provide a means for cancer cells to evade the immune system [31]. Moreover, seven patients received PD-1 checkpoint immunotherapy, and data regarding the therapy outcome was available for 5 patients.

# **Gynecologic cancers**

Ovarian cancer (OC) is one of the most common gynecologic malignancies in women, taking third place in occurrence worldwide after cervical cancer (CC) and endometrial cancer (EC), and is ranked first in mortality in developed countries [32]. In 70-80% of cases, the disease is diagnosed at its late stages, which leads to a 40-47% 5-year survival rate [33]. Among all histological types of OC, 85-90% of cases have been epithelial carcinomas [34]. EC is the second most common gynecologic cancer [32]. Morphologically, it is divided into two major subgroups: Type I includes the most common type-endometrioid cancer (60-70%) cases); Type II includes high-grade endometrioid cancer and rarer histological subtypes. Even though EC is diagnosed at an early stage, in most cases, up to 20% of tumors progress to the late stages, with a 15% 5-year survival rate [35]. CC is the fourth most common cancer in women, ranking after breast cancer (2.1 million cases), colorectal cancer (0.8

million) and lung cancer (0.7 million). The 5-year survival rate for women with CC is 66%.

The "gold standard" in gynecologic cancers therapy, which has had a significant impact on improving survival rates, is the combination of surgery, radiation and chemotherapy using drugs based on taxanes and platinum [36]. In the field of ovarian cancer therapy, chemotherapy with cisplatin is one of the most reliable therapies with a measurable clinical response, which is particularly effective in patients with early encapsulated epithelial or serous carcinoma [37]. In endometrial cancer, combination therapy with several drugs, and the best response rate at 57% with a median survival of 15.7 months, has been achieved by therapy with carboplatin and paclitaxel with the addition of doxorubicin [38]. At the initial stage of therapy, the tumor shows a high sensitivity to drugs. However, relapse usually occurs in approximately 80% of patients with ovarian cancer treated with paclitaxel or cisplatin, and in 45% of patients with endometrial cancer, also treated with cisplatin and paclitaxel [39, 40]. Also, some patients develop drug resistance. Several mechanisms to explain the resistance development have been proposed, including changes in the drug absorption mechanisms by tumor cells, apoptosis inhibition and increased DNA repair processes [41]. One of the reasons why drug resistance develops is associated with the role of some miRNAs' and has attracted the attention of many researchers, prompting them to look for alternative ways of tumor treatment [42]. As an alternative, ICB and adoptive cell therapy represent the most actively investigated types of immunotherapy for both ovarian and endometrium cancers and antitumor vaccines for ovarian cancer [43].

Recent data have revealed that various cancers with mismatched repair deficiency are susceptible to anti-PD-1/PD-L1 immunotherapy [44]. The role of PD-L1 expression on tumor cells, tissue or antigen-presenting cells in ovarian cancer is not yet well-understood [45-49]. Several studies have reported a favorable prognosis for patients with PD-L1 expression on tumor cells and outcomes [47, 49, 50], whereas other studies have reported a negative impact [45, 46]. Further investigation on the prognostic value of PD-L1 in ovarian cancer is required. The conclusions of clinical studies on PD-1/PD-L1 immunotherapy in ovarian cancers are very contradictory. Despite the immunotherapy efficiency was proven in many types of cancer, no immunotherapy has been approved for clinical applications in gynecologicl cancers to date. Platinum-resistant ovarian cancer is often referred to as a "cold tumor" due to decreased tumor infiltration

by immune cells. Although ovarian cancer has been proven to be immune-sensitive, monotherapy with ICIs showed low efficacy (KEYNOTE-100 trial [51], JAVELIN 200 trial [52]). When treatment with PD-1/PD-L1 inhibitors was combined with other therapeutic agents, an efficacy remained relatively low (phase II randomized trial NRG-003 [53]).

For endometrial cancer, the 11SP of immuno-oncological drugs is justified in first line as a mono-therapy for MSI tumors (KEYNOTE-158 [54]), as well as in 2 and subsequent lines after progression with platinum drugs in combination with tyrosine kinase inhibitors (KEYNOTE-775). Pembrolizumab monotherapy demonstrated durable antitumor activity and manageable safety in PD-L1+ patients with advanced CC (KEYNOTE-158 [54]), which can also be taken into account and used, among other aspects, to identify PD-L1 CTCs in this group of patients.

The study of CTCs as a diagnostic and prognostic biomarker of immunotherapy is the most prevalent in OC and EC. Depending on the type of OC, CTCs occur in 67-99% of patients and are sub-divided into several phenotypes classified by their invasiveness [55-58]. Over the last 10-12 years, the diagnostic and prognostic value of CTCs for assessing the dynamics of tumor development and monitoring the tumor response to the therapy in OC has been demonstrated in numerous studies [59-61]. For example, Buderath et al. [62] investigated CTCs as a potential biomarker for carboplatin therapy efficacy. In this study, CTCs were isolated from the peripheral blood and analyzed by RT-PCR to detect and evaluate the expression level of EpCAM, MUC-1, CA-125 and ERCC1, which was compared with the expression level of PD-L1/PD-L2 in serum to identify possible correlations. The following statistically significant relationship was found between PD-L2 (programmed cell death 1 ligand 2) and CTCs: the presence of CTCs before the therapy was associated with a lower level of PD-L2, whereas an absence of CTCs was associated with an increased level of PD-L2. There was a moderate correlation between the decrease in PD-L2 level and resistance to platinum chemotherapy. According to recent data, CTCs were detected in 33-75% of patients and occurred if the tumor invaded the cervix [63, 64]. Several EC biomarkers were identified through molecular profiling of isolated CTCs. Some of them, such as CTNNB1, GDF15, RUNX1, BRAF and PIK3CA, are believed to be worthwhile for further investigation as therapeutic targets. Moreover, the absence of CTCs was demonstrated after cycle of one standard chemotherapy in a study by Ni et al. [65]. No associations were found between the number of CTCs

and serum level of CA-125/HE4 or the expression of the other tumor biomarkers. The diagnostic and prognostic value of CTC in EC remains unclear, including monitoring the effects of various types of immunotherapies [63].

Several clinical trials of immunotherapeutic drugs have been performed to evaluate clinical applications of CTCs as a potential biomarker in treatment efficacy measurement. Berzovsky et al. have reported a clinical trial of a human epidermal growth factor receptor 2 (HER2)-targeted cancer vaccine [66]. It was based on autologous dendritic cells taken from patients and transfected with an adenovirus expressing non-signaling extracellular and transmembrane domains of HER2. Phase I of this study involved patients with the metastatic form of ovarian adenocarcinoma who failed to respond to at least one of the standard treatment regimens and had not previously received trastuzumab or other anti-HER2 therapy. In this study, a decrease in the number of CTCs by 40%, 83% and 100% at 12, 28 and 48 weeks was demonstrated.

In addition, clinical trials of GL-ONC1 oncolytic immunotherapy, i.e., genetically modified vaccinia oncolytic viruses (VOV), have been conducted (NCT02759588) [67] to retrospectively identify the clinical benefits of VOV monotherapy in patients with platinum-resistant OC. The comparative analysis included 11 patients and the following assaying: competence measurement of immune with neutralizing antibody titers, activity of virus-encoded glucuronidase, tumor response according to RECIST 1.1, prognostic nutrition index, and CTCs. As a result, CTCs were found as a prognostically significant marker: The absence of CTCs in peripheral blood directly correlated with the positive therapeutic effects of GL-ONC1. Generally, CTCs may be considered potentially significant biomarkers in the diagnosis and prognosis of OC. However, interrelations between CTCs different and immunotherapy types remain unclarified owing to the lack of data.

## **Breast cancer**

Although breast cancer (BC) is not traditionally considered immunogenic [68], increasing evidence suggests that its specific molecular subtypes, namely triple-negative and HER2-positive subtypes, often correlate with substantial infiltration of immune cells with a particular prognostic and even predictive value [73]. This has contributed to the ongoing assessment of ICI efficiency, especially with a triple-negative molecular subtype. In the KEYNOTE-086 study [69], an increase of objective response rate (ORR) was observed in untreated PD-L1<sup>+</sup> patients with metastatic triple-negative breast cancer (mTNBC) compared with patients receiving pembrolizumab in the second and subsequent lines of therapy. The use of pembrolizumab in combination with eribulin for mTNBC (KEYNOTE-150 / ENHANCE 1 trials [70]) in the first line of therapy has shown better results in PD-L1<sup>+</sup> patients, as well as the use of atezolizumab in combination with nab-paclitaxel (Impassion-130 [71]) as well as the use of pembrolizumab in combination with nab-paclitaxel (KEYNOTE - 355 [72]). Based on KEYNOTE-119 [73] trial, patients with overexpression of PD-L1 have also shown a good response to mono-immunotherapy with pembrolizumab in second and subsequent lines of therapy. The clinical efficacy of ICIs for the treatment of patients with HER2+ breast cancer remains to be determined. For instance, in the KATE-2 trial [74], the addition of atezolizumab to trastuzumab emtansine did not show clinically meaningful improvement in progression-free survival and was associated with more adverse events. Thus, untreated patients with locally advanced and metastatic TNBC expressing PD-L1 may benefit most from immunotherapy in combination with chemotherapy. However, it is worth mentioning that the clinical significance of PD-L1 as a biomarker for breast cancer has not been conclusively determined [75].

An implementation of PD-L1 as a reliable biomarker for selecting or excluding patients with BC for immunotherapy was complicated by several challenges, mainly related to molecular and cellular heterogeneity of the disease and the methodology used to measure PD-L1 expression. The assessment of PD-L1 expression in BC by immunohistochemical (IHC) methods has been widely studied. PD-L1 expression was significantly higher in invasive diseases than in normal breast tissue [82] and cancer in situ [76]. Additionally, a significant difference in PD-L1 expression was observed in different molecular subtype cohorts of BC favoring TNBC (up to 60% of PD-expression L1) [86]. These data appear to be consistent with the observation that the tumor expression of PD-L1 directly correlates with the number of stromal tumor-infiltrating lymphocytes (TILs) in this breast cancer subtype [87], which is known to be more often associated with TILs infiltration.

Evaluation of the PD-L1 expression by IHC methods is challenging because it is highly variable in tumor cells. For example, HER2-positivity was associated with a higher expression of PD-L1 (up to 50%) compared with HER2-negative BC in some studies [77, 78], whereas negligible differences have been reported in other studies [90]. The results of two meta-analyses studies confirmed a greater expression

of PD-L1 in TNBC [92] compared with non-TNBC subtypes but were not consistent in reporting a relationship between HER2 status and PD-L1 expression. The PD-L1 expression was associated unfavorable clinical with and prognostic characteristics, such as а lower degree of differentiation [94], high proliferative index [79], advanced stage [80], larger tumor size [81] and younger age [82]. However, conflicting data have been obtained regarding the association of PD-L1 expression with a better or worse clinical outcome. Some authors have reported lower disease-free survival and/or OS in cases of higher expression of PD-L1 by primary BC [83], especially in a TNBC subtype [84]. In contrast, other authors have reported the expression of PD-L1 was positively and independently associated with disease-free survival and/or OS in several primary BC cohorts [85-87].

The stromal compartment appears to make a significant contribution to the expression of PD-L1 in BC [88, 89]. However, no consistent data have been reported on the possible biological and clinical consequences of the differential expression of PD-L1 by CTCs or TILs. It has been suggested that evaluation of PD-L1 on stromal immune cells avoids false-negative results in the evaluation of PD-L1 status of breast cancer. This has the potential to increase the cohort of patients suitable for immunotherapy [89]. The results of a Javelin phase Ib study using avelumab [90] showed that the prognostic value of PD-L1 was probably more significant when PD-L1 was evaluated on TILs rather than on CTCs. A preliminary translational analysis of the randomized phase III study of Impassion130 showed that the assessment of TILs provided no additional predictive information above that provided by PD-L1 status [91].

Available data on spatial and temporal heterogeneity of PD-L1 [92] expression remain limited. These data have been largely obtained in analytical assaying of PD-L1 for metastases in the lymph nodes, in which the reliability of the determination of the immunity-related biomarkers was unclear and might be biased [93]. Jilaveanu et al. a weak correlation in have reported the immunocytochemical staining between the primary tumor and distant metastases. This was explained by the significantly higher expression of PD-L1 in distant metastatic foci than that in the primary tumor tissue and indicated that the primary tumor was not adequate for determining the expression of PD-L1 in the metastatic areas [94]. Real-time evaluation of the PD-L1 expression through liquid biopsy using immunocytochemical staining methods is a promising strategy that can potentially capture the dynamic nature of this biomarker compared with its evaluation

in primary tumor tissue or metastatic foci.

Mazel et al. [12] examined the frequency of PD-L1 expression in patients with triple-negative and HER2-positive BC. CTCs were detected using the FDA-approved CellSearch® system followed by immunocytochemical staining of the enriched material with a ready-made cocktail of antibodies from CellSearch® and analysis on the Celltracks II® platform. PD-L1 expressing CTCs were detected in 11/16 patients with BC (68.8%). According to the authors, this specific subset was identified as metastatic cells with a high potential to avoid T-cell mediated lysis - a potent target for immunotherapy. The proportion of PD-L1-positive CTCs varied from 0.2 to 100% in individual patients. In line with this finding, the results of other studies [95] have highlighted the percentage of the PD-L1-positive CTCs to be in the range from 0 to 100%. Thus, these studies demonstrated that PD-L1 was often expressed on circulating metastatic cells in triple-negative and HER2-positive BC patients, in which patients with a high percentage of PD-L1-positive CTCs had to be identified as potential candidates for anti-PD-L1 therapy.

In a study by Schott et al. [95], the authors used real-time liquid biopsy to determine PD-L1 and PD-L2 expression in the CTC of 72 patients with BC using the maintrac® method. The average amount of CTC was  $55/100 \mu$ L of blood (in the range from 5 to 805), CTC expressing PD-L1 were found in 94.5% of patients with BC; in patients expressing PD-L1 and PD-L2, the cell fraction of the PD-L1-positive CTCs was significantly higher than the fraction of the PD-L2-positive CTCs (54.6% versus 28.7%; p <0.001). In contrast with the study by Mazel *et al.*, Schott *et al.* were able to detect PD-L1-positive CTCs in patients without metastases, which might allow decisions to be already made when adjuvant therapy is prescribed. Moreover, patients with non-metastatic breast cancer had significantly more PD-L1-positive CTCs than patients without metastasis (median 75% versus 61.1%; p <0.05). The number of CTCs did not correlate with the other clinical and pathological parameters (age, size of the primary tumor, the presence of metastases in the lymph nodes, molecular subtype, predisposition. The authors performed a dynamic characterization of the PD-L1 expression on CTC and sequential monitoring of the treatment response in a patient with primary breast cancer. CTC numbers, as a signature of the successful immunotherapy, were decreased, and a percentage of the PD-L1-positive CTC was significantly reduced. After discontinuing ICI, the percentage of the PD-L1-positive CTC's continuously increased and a recurrence disease state was noted.

Thus, testing of PD-L1 in breast cancer tissue currently lacks standardization to encompass the diversity in the assays (IHC, gene expression), antibodies for testing, assessment systems and thresholds for classifying the PD-L1 expression (positive or negative), tumor microenvironment compartments included analysis (tumor cells, immune cells, or both) and nature of the tumor samples (primary, metastatic), along with a lack of appropriate widespread antibody platforms for testing PD-L1. Besides, there is currently no data on the effect of pre-analytic variables, including fixation time, type of fixative and storage on the reproducibility of testing PD-L1 in BC. The ratio between the expression of PD-L1 on CTC and the corresponding tumor tissue remains to be explored. An existing body of evidence suggests that the serum level of PD-L1 is directly related to the magnitude of the tumor burden [96] and the clinical outcome [97]. Thus, a liquid biopsy holds promise as a feasible strategy for dynamic assessment and sequential monitoring of the PD-L1 expression in patients with BC, avoiding the disadvantages of a solid tumor biopsy and potentially providing a real-time picture of PD-L1. Given the small number of studies, further experiments are necessary, particularly determining the relationship between the expression of PD-L1 by CTC and tumor tissue and in-depth dynamic characterization of the expression of PD-L1 CTC of BC during immunotherapy.

### Lung cancer

In the treatment of non-small cell lung cancer (NSCLC), immunotherapy takes an ever-stronger position from year to year. Starting with use nivolumab (PD-L1 all corner, second line therapy, CheckMate-017, CheckMate-057 [98]), pembrolizumab (PD-L1 tumor proportion score  $\geq 1\%$ , second and further lines, KEYNOTE-010 [99]) and atezolizumab (PD-L1 all corner, second and further lines, OAK [100]) after first line therapy with breakthrough results in increasing 5-year OS [101], there are currently а large number of immune-oncological first line treatment opportunities for both squamosus and not-squamosus mNSCLC. Pembrolizumab (KEYNOTE-024 [102]), atezolizumab (IMpower-110 [103]), or cemiplimab (EMPOWER-1 [104]) for PD-L1+ treatment-native patients can be used in monotherapy with significantly better tolerance and fewer adverse events. A recently published CITYSCAPE trial [105] has shown the efficacv of combination atezolizumab and tiragolumab for patients with hyper PD-L1 (≥50%) expression versus atezolizumab monotherapy with placebo. The use of immunotherapy in combination

standard platinum and/or pemetrexed with chemotherapy has shown an increase in overall survival with better results at a PD-L1 expression level ≥50% (KEYNOTE-189 [106], KEYNOTE-407 [107], IMpower-150 [108], IMpower-132 [109]) as well as the use of combinations of immuno-oncological drugs (CheckMate-227 [110]). Otherwise, at the same time, the use of a double combination of pembrolizumab + ipilimumab did not increase the median OS and PFS and was associated with a much higher incidence of grade 3-5 adverse events (KEYNOTE-598 [111]). Ongoing clinical trials on the use of immunotherapy in combination with chemoradiation therapy for patients with locally advanced NSCLC (IIIA-C stage, PACIFIC trial [112], KEYNOTE-799 [113]). Thus, the determination of PD-L1 expression in NSCLC directly affects the choice of therapy for the first and subsequent lines, the duration and quality of life of patients.

Lung cancer is one of the most common malignant neoplasms (12.9% of all new malignancies in the world) [114]. Late diagnosis occurs in 70-80% of cases and is associated with a low patient survival rate [115]. Quantifying and profiling CTCs are especially important for lung cancer patients. The relationship between the existence of CTCs in the peripheral blood of lung cancer patients and a poor prognosis has been demonstrated in most studies, regardless of the method of CTC isolation. The presence of CTC appears to correlate with the TNM stage [6, 116-118], lower disease-free and overall survival [119, 120]. These findings have been confirmed by meta-analyses involving a large number of patients [121, 122].

It is necessary to take into account that the detection of CTCs in the case of non-small cell lung cancer is difficult owing to their small number and the presence of non-epithelial characteristics [123]. A decrease in the number of CTCs was associated with a good response to cytostatic treatment and a longer and relapse-free survival [121, 122]. overall Additionally, the dynamics of a number of CTCs in the course of radiation therapy [124] and targeted therapy (gefitinib and erlotinib) [125, 126] also correlated with a tumor. With an increasing number of possible treatment regimens, including target therapy and immunotherapy, determination of the molecular profile of NSCLC is critical at every stage of the disease progression because it is necessary to identify biological changes that cause drug resistance and affect treatment decisions. To date, two anti-PD-1 (nivolumab and pembrolizumab) antibodies and one anti-PD-L1 antibody (atesolizumab) are registered by the FDA as drugs for the treatment of NSCLC [127, 128]. There are ongoing clinical trials on the use of these drugs in the first, second and third line of therapy in patients with NSCLC [129, 130]. Previous studies convincingly demonstrated that not all patients with advanced and disseminated NSCLC benefit from treatment with these drugs. Only 20% of the entire cohort of patients responded to the use of ICI, which emphasized a need for the correct selection of patients for this expensive therapy [131].

Clinically, the isolation and identification of PD-L1-positive CTCs have been introduced as a challenging issue in the case of NSCLC because neutrophils and other immature myeloid subsets have a low to absent expression of CD45 and also express PD-L1, which make it challenging to identify the CTCs and leads to potentially false-positive results [132]. Recently, PD-1 and PD-L1 expression across various subtypes of lung cancer cells, including squamous cell carcinoma, adenocarcinoma, and large cell carcinoma, were investigated [133]. In this study, a predictive value of PD-L1 and PD-1 expression by CTC was highlighted in patients with metastatic lung cancer before treatment and after three chemotherapy cycles. The presence of PD-1-positive CTCs correlated with shorter PFS; however, because of the small number of patients, final conclusions regarding the prognostic value of the expression of this marker cannot be made.

Changes in the CTC number in patients with non-resectable NSCLC treated with chemo-radiation therapy regimens have been reported in various studies. The presence of CTCs was detected in 100% of samples post-irradiation in comparison with that of 93% at the pre-treatment stage [134]. It was also found that an increased expression of PD-L1 by CTCs was associated with a poor prognosis. The results were consistent with those of Wang et al. [135], who reported on the effect of radiation therapy on the expression of PD-L1. In is study, whole blood from the non-metastatic NSCLC patients was collected before, during, and after the radiation or chemoradiation and processed using a microfluidic chip. PD-L1 expression in CTCs was assessed by immunofluorescence and qPCR and monitored through the course of treatment. PD-L1-positive CTCs were detected in 25 out of 38 samples (69.4%). After initiation of the radiation therapy, the proportion of significantly PD-L1-positive increased, CTCs indicating an up-regulation of PD-L1 in tumor cells in response to the radiation. Gene expression analysis revealed that the higher levels of PD-L1 were associated with poor prognosis. The authors concluded that CTCs can be used to monitor dynamic changes of PD-L1 during radiation therapy, which can potentially inform on the prognosis of a response to treatment.

Ilié et al. [136] reported on the enrichment of

CTCs and circulating white blood cells (WBCs) from peripheral blood samples from 106 NSCLC patients. The PD-L1 status of tumor tissue and CTCs, tumor-infiltrating WBC, and WBC of peripheral blood was studied, and their high interrelation was shown. A trend towards worse survival in patients receiving first-line cisplatin-based chemotherapy treatments, whose tumors expressed PD-L1 in CTCs or immune cells was found, similar to the effects of PD-L1 expression in matched-patient tumors. At the same time, in the study by Janning et al. [137], in contrast to the results reported by Ilié et al. [136], the percentage of the PD-L1-positive CTCs did not correlate with the percentage of PD-L1-positive in the biopsies determined by IHC. Also, the data demonstrated considerable heterogeneity in the PD-L1 status of CTCs in NSCLC patients.

Nicolazzo et al. [138] enrolled 24 patients at stage IV of NSCLC treated ICI Nivolumab. At a baseline and three months of treatment, the presence of CTCs and the expression of PD-L1 on their surface were found, and this was associated with poor treatment outcomes. Conversely, although CTCs were found in all patients six months after the treatment, at this time, patients could be dichotomized into two groups based on the PD-L1 expression of CTCs. All patients with PD-L1-negative CTCs had a clinical benefit, whereas all patients with PD-L1-positive CTCs had progressive disease. This suggested that the persistence of PD-L1-positive CTCs might mirror a mechanism of therapy resistance.

Guibert et al. [139] prospectively collected blood samples from patients with advanced NSCLC before nivolumab treatment and at the time of progression. CTCs were isolated using a cell size-based technology, PD-L1 expression was assessed and by immunofluorescence on CTCs and immunohistochemistry on tissue biopsies. CTCs were more often found to be PD-L1-positive than tissue (83% versus 41%), and no correlation was observed between tissue and CTC PD-L1 expression. A high CTC number before treatment was associated with an increased risk of death and progression. The presence of PD-L1-positive CTC pretreatment did not significantly correlate with the outcomes, but a higher baseline PD-L1-positive CTC number (≥1%) was observed in the "non-responders" group (PFS < 6months), and PD-L1-positive CTCs were seen in all patients at progression.

In a study by Kulasinghe *et al.*, CTCs were detected in 51.5% of blood samples of NSCLC patients, 64.7% of which were PD-L1-positive [140]. In the advanced stage NSCLC patient cohort, PFS was not found to be associated with CTCs before therapy, nor the presence of PD-L1 expression. The possible

predictive value of PD-L1-positive CTCs was also studied in patients receiving treatment with the ICIs pembrolizumab, nivolumab and avelumab [141]. In this study, a blood enrichment system based on different sizes of normal and tumor cells was used (Vortex HT chip). Because of the small number of patients, the study could not conclude the prognostic value of PD-L1 expression by the CTCs. Furthermore, Raimondi [142] demonstrated et al. that PD-L1-positive CTCs isolated from NSCLC patients were characterized by partial epithelial-mesenchymal phenotype and hypothesized transition that co-expression of PD-L1 and epithelial-mesenchymal transition cellular markers might represent a possible molecular background for the immune escape.

Analyzing a possible predictive and prognostic role of the expression of PD-L1/PD-1 by CTCs in patients with lung cancer, Raimondi et al. [143] noted the need to standardize the presentation of both clinical data and data regarding the blood processing (including the sampling, blood stabilization, storage time and temperature). Another problem pointed out by the authors of the review was the need to standardize methods for isolation of CTCs from samples of peripheral blood and their subsequent identification. Particularly, the most commonly used EpCAM-based CellSearch® System is designed to, first of all, identify cells of epithelial origin rather than CTCs, and therefore data collected with this system should be interpreted with caution. In addition, uniform approaches for the selection of antibodies utilized for the PD-L1 immunostaining are deemed necessary.

## Head and neck cancer

Head and neck squamous cell carcinoma (HNSCC) is traditionally associated with a high risk of recurrence and poor prognosis despite aggressive multimodal treatment strategies in clinical practice. Surgical methods, or radiation therapy, or chemotherapy, do not significantly increase patients' 5-year overall survival with a primary tumor, which remains at 40-50% [144]. Moreover, there is a lack of treatment options for patients with metastatic or recurrent disease; the median OS after the diagnosis is less than one year. Platinum-based chemotherapy and monoclonal antibody cetuximab targeting epidermal growth factor receptor are common choices for recurrent/metastatic HNSCC [145]. Nevertheless, the problem of treatment-related side effects has not been solved, and there is a significant amount of unmet need for improving treatment efficacy without further exacerbating toxicity.

The emergence of the human papillomavirus (HPV)-associated subset of HNSCC coupled with the

rapid development of immunotherapy has motivated novel, immune-based approaches to treatment. It was shown that HNSCC is an immunosuppressive disease, and patients have lower absolute lymphocyte counts than healthy subjects [146], impaired natural [147, killers cell activity 148], and poor antigen-presenting function [149]. Suppressive regulatory T cells secrete suppressive cytokines such as TGF-E and IL-10, express CTLA-4 and correlate with a poor prognosis, whereas the impairment of tumor-infiltrating lymphocytes (TILs) has a strong impact on clinical outcome, particularly in the case of HNSCC. Furthermore, HNSCC has a relatively high TMB [150], which is presumably owing to the production of altered and consequently antigenic proteins from mutated DNA, and can be predictive of the efficacy of ICIs [151].

The PD-1/PD-L1 pathway is a key mechanism of immune escape by tumor cells, so anti-PD1/PD-L1 agents boost the anti-tumor immune response and block the immunosuppressive signaling of tumors [152]. The biological rationale for targeting the anti-PD1/PD-L1 pathway in HNSCC has been reinforced recent large clinical by trials, demonstrating improved outcomes from **ICIs** with standard compared of care therapy. KEYNOTE-012 was a phase Ib trial that was the first to demonstrate durable responses to pembrolizumab in patients with platinum-refractory R/M HNSCC with  $\geq 1\%$  PD-L1 expression with an overall response rate of 16% [153, 154]. Soon after, data from the randomized phase III CheckMate 141 trial of 361 patients showed improved OS and quality of life relative to investigator's choice of the standard of care systemic therapy for the platinum-refractory disease regardless of HPV status, presence or no prior cetuximab therapy and PD-L1 expression [155]. The pembrolizumab activity in the platinum-refractory R/M HNSCC setting was subsequently confirmed in the Phase II KEYNOTE-055 [156] and Phase III KEYNOTE-040 trials [157]. In Phase III KEYNOTE-048 for the first-line recurrent/metastatic HNSCC, pembrolizumab significantly improved OS over EXTREME clinical trial with cetuximab. It is clear that such agents should be used to manage all patients who do not have contraindications. In this regard, the urgency of developing a correct method for assessing PD-L1 tumor status is increased.

In a study by Kulasinghe *et al.*, where blood samples from 23 patients with head and neck cancer (HNC) were examined for CTCs, CTC clusters were identified in 11/23 HNC samples. Notably, three of these patients had only CTC clusters and seven patients had both single and cluster CTCs. PD-L1 was found to be expressed (at least 1 CTC staining by immunofluorescence positive) in 6/11 (54.4%) samples. In two cases of HNC, all CTCs were PD-L1-positive. There was a significant difference in PFS between CTC-positive patients compared with the absence of CTCs in the investigated HNC cohort [140]. In another study by Kulasinghe *et al.* [158], CTCs clusters expressing PD-L1 were found in a patient suffering from supraglottic squamous cell carcinoma. A previously developed technique [5, 12] utilizing spiral microfluidic technology [159] was used in this study. The detected clusters exhibited a medium-high expression of PD-L1 compared with a panel of known HNC cell lines [158].

Many scientific groups have confirmed the importance of using molecular analysis for molecular characterization of CTCs [13, 160-162]. Real-time molecular analysis based on real-time PCR performed in a nucleic acid material (RNA or genomic DNA) isolated from the EpCAM-positive CTC fraction has been used to provide valuable information about the molecular characterization of CTC during gene expression [34], DNA methylation [35, 38] and the level of the DNA mutation [39]. In a study by Strati et al. [13], RT-qPCR analysis was applied for the analysis of PD-L1 mRNA transcripts to evaluate the increased expression of PD-L1 in the EpCAM-positive CTCs fraction. The authors showed that the detection of CTCs with the expression of PD-L1 at the completion of the final treatment in HNSCC patients correlated with a higher risk of progression and mortality compared with the negative analogs of PD-L1.

# Gastrointestinal cancers

According to global cancer statistic reports, gastrointestinal malignancies, an important component of solid tumors, have a heavier cancer-associated burden [163]. Currently, metastasis is the leading cause of malignancy-related deaths in gastrointestinal cancers [164]. Even at the early stage, a considerable cohort of patients that undergo resection treatment develop a metastatic disease within five years post-surgery [165]. This evidence points to a dormant metastatic process concurrent with the primary tumor genesis [166]. Additionally, tumor cells with metastatic potential are shed from the primary tumor site during surgery and are circulated in the system, resulting in distant metastasis [167, 168].

Speaking of immunotherapy, one should not forget that its role in the treatment of gastrointestinal tumors remains controversial despite a certain number of clinical studies. For PD-L1-positive esophageal and esophagogastric junction cancers, ICIs have shown their efficiency not only in second and subsequent lines of therapy (KEYNOTE-061 [169], KEYNOTE-059 [170], KEYNOTE-181 [171]), but recently in the first line in combination with chemotherapy. Thus, in KEYNOTE-590 study [172], pembrolizumab combined with chemotherapy provided superior OS, PFS, and ORR with a manageable safety profile in patients with untreated, advanced esophageal and esophagogastric junction cancer with hyper PD-L1 expression. On the contrary, in gastric cancer (GC), the use of pembrolizumab in previously untreated PD-L1<sup>+</sup> patients, both in monotherapy and in combination with chemotherapy, did not give any statistically significant benefit (KEYNOTE-062 [173]) and was effective only for patients with MSI-tumors. Adding nivolumab to standard first line therapy (XELOX\FOLFOX) in patients with HER2-negative GC with has led to significantly improved progression-free and overall survival over chemotherapy alone (CheckMate-649 [174]), whereas the use of similar therapy regimens without prior selection of patients according to the level of PD-L1 expression produced negative results in the ATTRACTION-4 [175] trial. For the case of hepatocellular carcinoma (HCC), there is a modest number of clinical studies, which, however, have shown the effectiveness of the use of immunotherapy in first line of therapy (atezolizumab + bevacizumab, IMBrave150 [176]) and in subsequent lines in combinations (ipilimumab + nivolumab/pembrolizumab, Abstract 330 [177]). For pancreatic and colorectal cancers, there is still no optimal cohort of patients for immunotherapy, as well as for PD-L1 expression testing.

The predictive value of PD-L1 IHC is still under debate because some patients with PD-L1 negative status tumors of various types of cancer, including GC, respond to anti-PD-1 therapy [178, 179]. In line with these observations, Cheng *et al.* [180] found that in GC patients with PD-L1-negative tumor tissue, the imaging flow cytometry signal analysis of isolated CTC was as varied as that of IHC staining, suggesting that the expression of CTC PD-L1 was useful in the immunophenotypic differential diagnosis of tumors and could be a potential candidate for anti-PD-1/ PD-L1 immune checkpoint therapy [143].

Because PD-L1-positive CTCs have been recognized as a biomarker for therapeutic efficacy of ICI in NSCLC, it is hypothesized that CTCs can also be a predictive marker in GC immune checkpoint therapy [181]. It was identified that after checkpoint blockade therapy, CTC enumeration can be a useful tool to predict poor prognosis in advanced GC patients [132]. In line with this, Yue *et al.* [182] demonstrated that the PD-L1 level in CTCs could be a potential predictor for PD-1/PD-L1 blockade therapies in patients with advanced gastrointestinal

tumors. The disease control rate in the patients with PD-L1-high CTC number was much higher than those with a lower level. Additionally, several PD-L1-high CTCs at the baseline had predictive values for PFS. Moreover, the dynamic changes in the number of PD-L1-positive CTCs can reflect the real-time response status. However, in this study, the changes in the total number of isolated CTCs, PD-L1-positive CTC, and PD-L1-high CTC significantly correlated with the clinical outcomes. This cohort study highlighted the advantage of using CTC-PD-L1 over the tumor PD-L1, which can provide baseline information for treatment response prediction in a time-efficient manner.

Similarly, Cheng and colleagues isolated CTCs from 32 GC patients and demonstrated that a total CTC pool and CTC-PD-L1 were highly correlated with the clinical outcome of checkpoint blockade therapy. Additionally, high number а PD-L1-positive CTCs in advanced GC patients was predictive of better five-year OS [180]. Taken together, the clinical evidence supports a potential application of PD-L1 expressing CTCs as a reliable prognostic biomarker for checkpoint blockade therapy in GC. Moreover, recent advances in single-cell molecular profiling of CTCs and tumor cells using next-generation sequencing may also provide a pool of tools to validate predictive biomarkers, including CTC-PD-L1.

In pancreatic ductal adenocarcinoma, an association between the presence of PD-L1 expression on isolated CTCs and patient outcome was described in few studies. The numeration and PD-L1 expression levels of CTC were analyzed in a clinical study involving 35 patients at various advanced stages of gastrointestinal tumors including pancreatic ductal adenocarcinoma: 74% (26/35) of patients had PD-L1-positive CTC and 60% (21/35) had at least one PD-L1-high CTCs [182]. Comparing the CTC number and expression of PD-L1 on isolated CTC before and after treatment with a PD-L1 inhibitor, patients with a stable-disease status showed a significant reduction in the number of CTC (25 versus 15) and PD-L1-high CTC (40% versus 6.67% of PD-L1 expression) in comparison with patients with progressive disease status. The results suggested that the abundance of PD-L1-high CTCs at a baseline might serve as a predictor to screen patients for PD-1/PD-L1 blockade therapies and measure the dynamic changes of CTC to indicate the therapeutic response of the initial cycle of treatment.

Recently, evaluation of the expression of PD-L1 on captured CTCs in patients with HCC opened a new way to manage patients undergoing immunotherapy. In a prospective cohort of 92 patients

(8 healthy control, 11 non-malignant liver disease, 73 HCC), the enumeration and characterization of the phenotype of captured CTCs were analyzed [183]. Considering the expression of PD-L1 on isolated CTCs, PD-L1<sup>+</sup> CTCs classified HCC patients at early-stage (3/39 with PD-L1<sup>+</sup> CTCs) and advanced/metastatic disease (23/34 with PD-L1+ CTCs) with sensitivity and specificity of 67.7% and 92.3%, respectively. Additionally, prior to the immunotherapy, 3 of 6 patients receiving anti-PD-1 therapy that successfully responded to therapy showed PD-L1-positive CTCs compared with only 1 of 3 non-responders.

Based on this evidence, in a larger cohort of 87 patients with various stages of HCC (49 early-stage, 22 locally advanced, and 16 metastatic), PD-L1+CTCs were identified in 4 of 49 early-stage patients, 12 of 22 (54.5%) locally advanced and 15 of 16 (93.8%) metastatic patients. Moreover, the analysis of the PD-L1 expression on CTCs could be used to accurately discriminate between early-stage and locally advanced/metastatic HCC (sensitivity at 71.1%, specificity at 91.8%) [184]. In 10 patients who received PD-1 inhibitor, the treatment-responsive patients (n = 5) demonstrated PD-L1<sup>+</sup> CTCs at a baseline compared with only 1 of 5 non-responders with a progressed disease within four months of starting treatment. This evidence indicates a robust positive association between PD-L1+ CTCs and positive treatment response in patients with HCC that received anti-PD-1 inhibitor.

# Melanoma

In melanoma, immunotherapy with ICIs has shown benefit in a subset of melanoma patients [185, 186]. Whilst targeted treatment such as BRAF and MEK inhibitors have shown transient responses, ICI therapies have led to long-lasting and durable responses. However, there remains a need for predictive biomarkers of response to ICI therapy. Several biomarkers have been introduced for guiding patient selection for immunotherapy, such as PD-L1 expression and TMB.

Thus, the use of pembrolizumab as a first line therapy significantly increased ORR and PFS in patients with expression PD-L1 rather than in its absence (KEYNOTE-001 [187]). The benefit from the use of nivolumab alone or in combination with ipilimumab (CheckMate 066 [188], CheckMate 067 [189]) was also higher in groups of patients with PD-L1 expression level ≥5%.

One of the challenges is the enrichment and isolation of CTCs and comprehensive characterization

of CTCs for biomarkers of response to therapy. This challenge has been met by the recent advances in microfluidic technology. Aya-Bonilla *et al.* [190] demonstrated that melanoma CTCs could be efficiently isolated and characterized using spiral chip technology. They recently compared two microfluidic technologies: Parsotix (Angle PLC) and ClearCell (Biolidics, Singapore) to capture and characterize melanoma CTCs. This study highlighted the heterogeneity of CTCs and multiple, non-overlapping populations [191]. Furthermore, facile detection of CTCs and the evaluation of PD-L1 expression has been reported in various studies [192].

CTCs have been used to quantify the response to therapy in melanoma. Hong et al. [193] analyzed a panel of 19-gene RNA signatures in enriched CTCs using microfluidic technologies, which were capable of monitoring the tumor burden in patients. In a study by Khattak et al. [194] using flow cytometry for the identification and characterization of CTCs based on melanoma markers (MCAM, MCSP) in metastatic melanoma, the authors found that PD-L1 expression buy CTCs was associated with a response to therapy and longer PFS. In a subsequent study by Khattak et al. [195], PD-L1-positive CTCs were identified in 64% of total CTCs and those patients had a significantly longer PFS. The survival at 12 months for patients with detectable PD-L1-positive CTCs was 76% compared with 22% for patients with PD-L1-negative CTCs. They also found that the expression of PD-L1 on CTCs was an independent predictive biomarker of PFS. In line with these findings, Yue et al. [182] applied a dynamic expression level of PD-L1 and demonstrated that the presence of PD-L1<sup>+</sup> CTCs at a baseline could serve as a predictor for screening patients for anti-PD-1/PD-L1 therapy and measuring CTC expression levels probably indicated real-time changes to therapies.

These data suggest that the PD-L1 expression on CTCs could be used to predict the response to anti-PD-1/PD-L1 therapy (e.g., pembrolizumab) in melanoma. However, larger validation studies are needed to identify whether liquid biopsy has the potential to be used as a tool for identifying patients likely to respond to therapy. These investigations need to include a direct comparison of liquid biopsy and solid tumor biopsy for PD-L1 expression scoring and measurement against the response to anti-PD-1/PD-L1 therapy to determine the clinical utility of this assay.

The studies, discussed in the current review, where immunotherapy was combined with CTC assessment are summarized in Table 1.

#### Table 1. Crucial findings on CTCs as biomarkers in cancer immunotherapy.

Type of cancer	Cancer form	Type of immunotherapy	Purpose of the study	CTC detection/isolation technique	CTC prognostic significance	Ref
Prostate cancer	Castration-resistant, none-metastatic form	Combination of anti-tumor vaccination and pembrolizumab	To analyze changes in checkpoint receptor expression on antigen-specific CD8+ T-cells and the effect of PD-1 blockade on immune response, and to analyze PD-L1 expression on CTCs.	Flow cytometry	PD-L1 expression on CTCs increased after vaccination, which was associated with the development of T-cell immunity and longer PFS.	[25]
	Metastatic castration-resistant form	Nivolumab, ipilimumab	To investigate if tumors of patients with AR-V7+ CTCs had more DNA-repair deficiency mutations, and therefore were potentially more sensitive ICB therapy.	AdnaTest	Presence of AR-V7+ CTCs was associated with CTCs heterogeneity, which correlated with likelihood of favorable response to immune-checkpoint inhibition.	[26]
Urothelial bladder cancer	Metastatic and muscle invasive forms	MPDL3280A anti-PD-L1 monoclonal antibody	To examine the efficiency of atezolizumab for the treatment of patients with urothelial bladder cancer. A biomarker analysis was performed, which included the analysis of CTCs and TILs for the expression of PD-L1.	N/A	The amount of PD-L1 <sup>+</sup> TILs was associated with worse median OS. The 43% response rate was achieved in patients with PD-L1 <sup>+</sup> IHC.	[29]
Ovarian cancer	Metastatic form	HER2-targeted cancer vaccine	A phase I clinical trial in patients, which included assessment of CTCs as one of the major biomarkers of therapy	N/D	A decrease in the number of CTCs by 40%, 83% and 100% at 12, 28 and 48 weeks post-treatment was identified.	
	Advanced platinum-resistant ovarian cancer	GL-ONC1 oncolytic immunotherapy	response. To identify clinical benefits of VOV monotherapy.	N/D	The absence of CTCs in peripheral blood directly correlated with the positive therapeutic effects of GL-ONC1 therapy.	[67]
Lung cancer (NSCLC)	TNM stages IIB-IV, including both metastatic and none-metastatic patients	Pembrolizumab, nivolumab, atezolizumab	To test an epitope-independent method (Parsortix© system) and utilize it to assess PD-L1 expression of CTCs from NSCLC patients undergoing PD-1/PD-L1 inhibitors therapy.	~	Upon disease progression, all patients demonstrated an increase in PD-L1+ CTCs, while no change or a decrease in PD-L1+ CTCs was observed in responding patients. An increase of PD-L1+ CTCs had a potential to predict resistance to PD-1/PD-L1 inhibitors.	[137]
	TNM stage IV, metastatic form	Nivolumab	To investigate if liquid biopsy might allow real-time sampling of patients for PD-L1 through the course of immunotherapy. PD-L1 expressing CTCs were assessed at baseline, at 3 and 6 months post-treatment.	Cellsearch system with anti-human B7-H1/PD-L1 conjugated antibody	A high CTC number was associated with poorer prognosis. At baseline and at 3 months of treatment, the presence of PD-L1* CTCs was associated with poor patient outcome.	[138]
	Advanced metastatic form	Nivolumab	To investigate if liquid biopsy might allow real-time sampling of patients for PD-L1 through the course of immunotherapy.	ISET technology	Patients with high CTC count experienced worse outcomes. The presence of PD-L1 <sup>+</sup> CTCs had no significant impact on PFS. Patients with PD-L1 <sup>+</sup> CTCs at baseline were more often non-responders compared to PD-1 1, patients	[139]
	Metastatic form	Pembrolizumab treatment	To determine if PD-L1 expression on CTCs can serve as a predictive biomarker of clinical benefit and response to treatment with the PD-1/PD-L1 inhibitor pembrolizumab.	Ficoll-Paque Density Gradient Media	compared to PD-L1- patients. PD-L1+ CTCs were identified in 64% of total CTCs. Patients with PD-L1+ CTCs had significantly longer PFS.	[195]

Type of cancer	Cancer form	Type of immunotherapy	Purpose of the study	CTC detection/isolation technique	CTC prognostic significance	Ref
Hepatocellular carcinoma	Early, locally advanced and metastatic stages	Anti-PD1 therapy (unspecified)	To detect PD-L1 expressing HCC CTCs, and investigate its role as a prognostic biomarker.	NanoVelcro Chip	The study indicated a strong positive association between the presence of PD-L1+ CTCs and positive treatment response. Analysis of PD-L1 expression on CTCs discriminated early stage from locally advanced/metastatic.	[183]
Different gastrointestinal tumors	Metastatic form	IBI308 PD-1 monoclonal antibody therapy	To determine if PD-L1 expression on CTCs could serve as an alternative biomarker of tumor response for predicting outcomes and monitoring PD-1 blockade therapies.	The Pep@MNPs isolation system	Before treatment: 74% of patinets had PD-L1+ CTCs, 60% had at least 1 high-positive PD-L1 CTC. After treatment: significant reduction of PD-L1+ CTCs and high-positive PD-L1 CTCs (40% vs. 6.67%). CTCs might be used as a baseline predictor to screen patients for PD-1/PD-L1 blockade therapies and monitor therapeutic response.	[182]

Abbreviations: AR-V7: androgen receptor splice variant 7; AR-V7\*: androgen receptor splice variant 7 positive; B7-H1: B7 homolog 1; CD8\*: cluster of differentiation 8 positive; CTC: circulating tumor cell; DNA: deoxyribonucleic acid; HCC: hepatocellular carcinoma; HER2: human epidermal growth factor receptor 2; ICB: immune checkpoint blockade; IHC: immunohistochemistry; NSCLC: non-small cell lung cancer; PD-1: programmed cell death protein 1; PD-L1: programmed death-ligand 1; PFS: progression-free survival; TIL: tumor-infiltrating lymphocyte; TNM: tumor, nodes and metastases; VOV: vaccinia oncolytic virus

## Summary and outlook

In this review, we discussed the role of immunotherapy in a number of solid malignancies and the need for the development of non-invasive liquid biopsy approaches for treatment management. Although for tumors of some localizations such as NSCLC, or TNBC, PD-L1 testing is really critical for using immuno-oncological drugs in first line treatment as mono-therapy or in combinations with the best efficacy and toxicity profile, for others such as melanoma, immunotherapy is effective in itself, is minimally associated with the level of PD-L1 expression and, apparently, does not require its dynamic assessment over time. The effectiveness of checkpoint inhibitors for other tumors such as colorectal and pancreatic cancer is still questionable and requires further research. Genomic subtypes of gastrointestinal tumors, in particular gastric cancer, differing in their genomic characteristics, create controversy in the response of such tumors to treatment with checkpoint inhibitors and cause mixed results in clinical trials. At the same time for head and neck cancers, it is important not to miss the optimal testing time to determine the better therapy for second and subsequent lines.

From a clinician's point of view, the main problem in testing CTCs is to determine the "ideal" cohort of patients for whom the time and financial costs of testing itself and subsequent immunotherapy will be commensurate with the increase in the effectiveness of the treatment. This is difficult until the indications for immunotherapy of solid tumors have not been finally determined and the localization of tumors for which such treatment is obviously ineffective is not completely ruled out. The clinical benefit of studying the expression of immune checkpoints on CTCs in real time is that it can probably predict the moment that resistance to therapy with checkpoint inhibitors is formed, but at the moment it is poorly understood and requires further discussion.

Immunotherapies have been hailed as a 'game-changer' in the field of cancer therapies. However, there remain some challenges in identifying which patients would best respond to ICB therapy. The challenge remains in assessing the dynamical genesis of a tumor using static, single timepoint tissue biopsy samples, because a repeat biopsy is often challenging to perform and carries associated risks for the patient. A solution for repeated measurements during the course of therapy could be the analysis of liquid biopsy samples of patients. The analysis of CTCs in the peripheral blood of patients represents such an approach, which has FDA approval for the enumeration of CTCs. Although the enumeration of CTCs has been associated with PFS and OS across a number of solid tumors, the characterization of these cells becomes increasingly more important. Recent reports have shown frequent expression of immune checkpoint proteins on CTCs such as PD-L1, which can be assessed prior to and during the course of therapy to understand dynamic tumor genesis in real time. While these data still emerge, clinical trials incorporating CTC enumeration, characterization and changes over the course of immunotherapy may be useful in determining whether CTCs can be used as a surrogate marker of response to therapy. Recent literature supports the notion that CTCs may provide an immune escape mechanism for tumors to evade the body's immune system and further investigation is required to determine the role of neutrophils and other immune cells which co-locate and cluster with CTCs. At the same time, standardisation of protocols for isolation/detection and analysis of CTCs is highly required.

#### Abbreviations

AR-V7: androgen receptor splice variant 7; AR-V7<sup>+</sup>: androgen receptor splice variant 7 positive; B7-H1: B7 homolog 1; CD8+: cluster of differentiation 8 positive; CTC: circulating tumor cell; DNA: deoxyribonucleic acid; HCC: hepatocellular carcinoma; HER2: human epidermal growth factor receptor 2; ICB: immune checkpoint blockade; IHC: immunohistochemistry; NSCLC: non-small cell lung cancer; PD-1: programmed cell death protein 1; programmed death-ligand PD-L1: 1; PFS: progression-free survival; TIL: tumor-infiltrating lymphocyte; TNM: tumor, nodes and metastases; VOV: vaccinia oncolytic virus.

#### Acknowledgments

In this review study, preparation of the parts on urologic cancers, gynecologic cancers, and breast cancer, was supported by the Ministry of Education and Science of the Russian Federation, grant No. 075-15-2019-192. Preparation of the parts on lung cancer, head and neck cancer, gastrointestinal cancers, and melanoma, was supported by Russian Foundation for Basic Research, project number 19-34-60034.

## **Competing Interests**

The authors have declared that no competing interest exists.

### References

- Ashworth T. A case of cancer in which cells similar to those in the tumours were seen in the blood after death. The Medical Journal of Australia. 1869; 14: 146.
- Massagué J, Obenauf ACJN. Metastatic colonization by circulating tumour cells. Nature. 2016; 529: 298-306.
- Alix-Panabières C, Pantel KJNRC. Challenges in circulating tumour cell research. Mol Oncol. 2014; 14: 623-31.
- Abalde-Cela S, Piairo P, Diéguez LJAc. The significance of circulating tumour cells in the clinic. Acta Cytol. 2019; 63: 466-78.
- Allard WJ, Matera J, Miller MC, Repollet M, Connelly MC, Rao C, *et al.* Tumor cells circulate in the peripheral blood of all major carcinomas but not in healthy subjects or patients with nonmalignant diseases. Clin Cancer Res. 2004; 10: 6897-904.
- Farace F, Massard C, Vimond N, Drusch F, Jacques N, Billiot F, et al. A direct comparison of CellSearch and ISET for circulating tumour-cell detection in patients with metastatic carcinomas. Br J Cancer. 2011; 105: 847-53.
- Rizvi NA, Hellmann MD, Snyder A, Kvistborg P, Makarov V, Havel JJ, et al. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. Science. 2015; 348: 124-8.
- Darvin P, Toor SM, Nair VS, Elkord E. Immune checkpoint inhibitors: recent progress and potential biomarkers. Exp Mol Med. 2018; 50: 1-11.
- Topalian SLJJ. Targeting immune checkpoints in cancer therapy. JAMA. 2017; 318: 1647-8.

- Sadeghi Rad H, Bazaz SR, Monkman J, Ebrahimi Warkiani M, Rezaei N, O'Byrne K, *et al.* The evolving landscape of predictive biomarkers in immunooncology with a focus on spatial technologies. Clin Transl Immunology. 2020; 9: e1215.
- Sadeghi Rad H, Monkman J, Warkiani ME, Ladwa R, O'Byrne K, Rezaei N, et al. Understanding the tumor microenvironment for effective immunotherapy. Med Res Rev. 2020; 41: 1474-98.
- 12. Mazel M, Jacot W, Pantel K, Bartkowiak K, Topart D, Cayrefourcq L, *et al.* Frequent expression of PD-L1 on circulating breast cancer cells. Mol Oncol. 2015; 9: 1773-82.
- Strati A, Koutsodontis G, Papaxoinis G, Angelidis I, Zavridou M, Economopoulou P, et al. Prognostic significance of PD-L1 expression on circulating tumor cells in patients with head and neck squamous cell carcinoma. Ann Oncol. 2017; 28: 1923-33.
- 14. Wartensee S, Ackermann D, Studer UE, Zingg EJ. Treatment of carcinoma in situ of the bladder with BCG. Helv Chir Acta. 1989; 9: 69-72.
- Ratta R, Zappasodi R, Raggi D, Grassi P, Verzoni E, Necchi A, et al. Immunotherapy advances in uro-genital malignancies. Crit Rev Oncol Hematol. 2016; 105: 52-64.
- Donin NM, Lenis AT, Holden S, Drakaki A, Pantuck A, Belldegrun A, et al. Immunotherapy for the treatment of urothelial carcinoma. J Urol. 2017; 197: 14-22.
- Redman JM, Gulley JL, Madan RA. Combining immunotherapies for the treatment of prostate cancer. Urologic Oncology: Seminars and Original Investigations: Elsevier; 2017. p. 694-700.
- Monkman J, Warkiani M, Ladwa R, O'Byrne K, Rezaei N, Kulasinghe A. Understanding the tumor microenvironment for effective immunotherapy. Med Res Rev. 2020; 41: 1474-98
- Robinson D, Van Allen EM, Wu Y-M, Schultz N, Lonigro RJ, Mosquera J-M, et al. Integrative clinical genomics of advanced prostate cancer. Cell. 2015; 161: 1215-28.
- Joshi H, Pinski JK. Association of ARV7 expression with molecular and clinical characteristics in prostate cancer. American Society of Clinical Oncology. J Clin Oncol. 2016; 34, suppl: 109-109.
- Antonarakis E, Armstrong A, Dehm S, Luo JJPc, diseases p. Androgen receptor variant-driven prostate cancer: clinical implications and therapeutic targeting. Prostate Cancer Prostatic Dis. 2016; 19: 231-41.
- Antonarakis ES, Lu C, Wang H, Luber B, Nakazawa M, Roeser JC, et al. AR-V7 and resistance to enzalutamide and abiraterone in prostate cancer. N Engl J Med. 2014; 371: 1028-38.
- Rekoske BT, Smith HA, Olson BM, Maricque BB, McNeel DGJCir. PD-1 or PD-L1 Blockade Restores Antitumor Efficacy Following SSX2 Epitope-Modified DNA Vaccine Immunization. Cancer Immunol Res. 2015; 3: 946-55.
- Fu J, Malm I-J, Kadayakkara DK, Levitsky H, Pardoll D, Kim YJJCr. Preclinical evidence that PD1 blockade cooperates with cancer vaccine TEGVAX to elicit regression of established tumors. Cancer Res. 2014; 74: 4042-52.
- Rekoske BT, Olson BM, McNeel DGJO. Antitumor vaccination of prostate cancer patients elicits PD-1/PD-L1 regulated antigen-specific immune responses. Oncoimmunology. 2016; 5: e1165377.
- Boudadi K, Suzman DL, Anagnostou V, Fu W, Luber B, Wang H, et al. Ipilimumab plus nivolumab and DNA-repair defects in AR-V7-expressing metastatic prostate cancer. Oncotarget. 2018; 9: 28561.
- Satelli A, Batth IS, Brownlee Z, Rojas C, Meng QH, Kopetz S, et al. Potential role of nuclear PD-L1 expression in cell-surface vimentin positive circulating tumor cells as a prognostic marker in cancer patients. Sci Rep. 2016; 6: 28910.
- Yin C, Wang Y, Ji J, Cai B, Chen H, Yang Z, et al. Molecular profiling of pooled circulating tumor cells from prostate cancer patients using a dual-antibody-functionalized microfluidic device. Anal Chem. 2018; 90: 3744-51.
- Powles T, Eder JP, Fine GD, Braiteh FS, Loriot Y, Cruz C, et al. MPDL3280A (anti-PD-L1) treatment leads to clinical activity in metastatic bladder cancer. Nature. 2014; 515: 558.
- Anantharaman A, Friedlander T, Lu D, Krupa R, Premasekharan G, Hough J, et al. Programmed death-ligand 1 (PD-L1) characterization of circulating tumor cells (CTCs) in muscle invasive and metastatic bladder cancer patients. BMC Cancer. 2016; 16: 744.
- Wu P, Wu D, Li L, Chai Y, Huang JJPo. PD-L1 and survival in solid tumors: a meta-analysis. PLoS One. 2015; 10.
- Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2018; 68: 394-424.
- Momenimovahed Z, Tiznobaik A, Taheri S, Salehiniya H. Ovarian cancer in the world: epidemiology and risk factors. Int J Womens Health. 2019; 11: 287-99.
- Castiglia M, Listì A, Incorvaia L, Chiantera V, Russo A. Liquid Biopsy in Gynecological Cancers. In: Russo A, Giordano A, Rolfo C, editors. Liquid Biopsy in Cancer Patients: The Hand Lens for Tumor Evolution. Cham: Springer International Publishing; 2017. p. 85-93.
- Bokhman JV. Two pathogenetic types of endometrial carcinoma. Gynecol Oncol. 1983; 15: 10-7.
- Poonawalla IB, Parikh RC, Du XL, VonVille HM, Lairson DR. Cost Effectiveness of Chemotherapeutic Agents and Targeted Biologics in Ovarian Cancer: A Systematic Review. Pharmacoeconomics. 2015; 33: 1155-85.

- Lawrie TA, Winter-Roach BA, Heus P, Kitchener HC. Adjuvant (post-surgery) chemotherapy for early stage epithelial ovarian cancer. Cochrane Database Syst Rev. 2015; CD004706.
- Carey MS, Gawlik C, Fung-Kee-Fung M, Chambers A, Oliver T, Cancer Care Ontario Practice Guidelines Initiative Gynecology Cancer Disease Site G. Systematic review of systemic therapy for advanced or recurrent endometrial cancer. Gynecol Oncol. 2006; 101: 158-67.
- Pokhriyal R, Hariprasad R, Kumar L, Hariprasad G. Chemotherapy Resistance in Advanced Ovarian Cancer Patients. Biomark Cancer. 2019; 11: 1179299X19860815.
- Gadducci A, Cosio S, Genazzani AR. Old and new perspectives in the pharmacological treatment of advanced or recurrent endometrial cancer: Hormonal therapy, chemotherapy and molecularly targeted therapies. Crit Rev Oncol Hematol. 2006; 58: 242-56.
- Stordal B, Davey M. Understanding cisplatin resistance using cellular models. IUBMB Life. 2007; 59: 696-9.
- Fang L, Wang H, Li P. Systematic analysis reveals a lncRNA-mRNA co-expression network associated with platinum resistance in high-grade serous ovarian cancer. Invest New Drugs. 2018; 36: 187-94.
- Lynam S, Lugade AA, Odunsi K. Immunotherapy for Gynecologic Cancer: Current Applications and Future Directions. Clin Obstet Gynecol. 2020; 63: 48-63.
- Le Saux O, Ray-Coquard I, Labidi-Galy SI. Challenges for immunotherapy for the treatment of platinum resistant ovarian cancer. Semin Cancer Biol. 2020.
- Hamanishi J, Mandai M, Iwasaki M, Okazaki T, Tanaka Y, Yamaguchi K, et al. Programmed cell death 1 ligand 1 and tumor-infiltrating CD8+ T lymphocytes are prognostic factors of human ovarian cancer. Proc Natl Acad Sci U S A. 2007; 104: 3360-5.
- Chatterjee J, Dai W, Aziz NHA, Teo PY, Wahba J, Phelps DL, et al. Clinical Use of Programmed Cell Death-1 and Its Ligand Expression as Discriminatory and Predictive Markers in Ovarian Cancer. Clin Cancer Res. 2017; 23: 3453-60.
- Wang Q, Lou W, Di W, Wu X. Prognostic value of tumor PD-L1 expression combined with CD8(+) tumor infiltrating lymphocytes in high grade serous ovarian cancer. Int Immunopharmacol. 2017; 52: 7-14.
- Aust S, Felix S, Auer K, Bachmayr-Heyda A, Kenner L, Dekan S, et al. Absence of PD-L1 on tumor cells is associated with reduced MHC I expression and PD-L1 expression increases in recurrent serous ovarian cancer. Sci Rep. 2017; 7: 42929.
- Darb-Esfahani S, Kunze CA, Kulbe H, Sehouli J, Wienert S, Lindner J, *et al.* Prognostic impact of programmed cell death-1 (PD-1) and PD-ligand 1 (PD-L1) expression in cancer cells and tumor-infiltrating lymphocytes in ovarian high grade serous carcinoma. Oncotarget. 2016; 7: 1486-99.
- Webb JR, Milne K, Kroeger DR, Nelson BH. PD-L1 expression is associated with tumor-infiltrating T cells and favorable prognosis in high-grade serous ovarian cancer. Gynecol Oncol. 2016; 141: 293-302.
- Matulonis UA, Shapira-Frommer R, Santin AD, Lisyanskaya AS, Pignata S, Vergote I, et al. Antitumor activity and safety of pembrolizumab in patients with advanced recurrent ovarian cancer: results from the phase II KEYNOTE-100 study. Ann Oncol. 2019; 30: 1080-7.
- Tumeh PC, Harview CL, Yearley JH, Shintaku IP, Taylor EJ, Robert L, et al. PD-1 blockade induces responses by inhibiting adaptive immune resistance. Nature. 2014; 515: 568-71.
- Zamarin D, Burger RA, Sill MW, Powell DJ, Jr., Lankes HA, Feldman MD, et al. Randomized Phase II Trial of Nivolumab Versus Nivolumab and Ipilimumab for Recurrent or Persistent Ovarian Cancer: An NRG Oncology Study. J Clin Oncol. 2020; 38: 1814-23.
- Borcoman E, Le Tourneau C. Keynote-158 study, FDA granted accelerated approval of pembrolizumab for the treatment of patients with advanced PD-L1-positive cervical cancer. Ann Transl Med. 2020; 8: 1611.
- Windmuller C, Zech D, Avril S, Boxberg M, Dawidek T, Schmalfeldt B, et al. CXCR3 mediates ascites-directed tumor cell migration and predicts poor outcome in ovarian cancer patients. Oncogenesis. 2017; 6: e331.
- Banys M, Solomayer EF, Becker S, Krawczyk N, Gardanis K, Staebler A, *et al.* Disseminated tumor cells in bone marrow may affect prognosis of patients with gynecologic malignancies. Int J Gynecol Cancer. 2009; 19: 948-52.
- He W, Kularatne SA, Kalli KR, Prendergast FG, Amato RJ, Klee GG, et al. Quantitation of circulating tumor cells in blood samples from ovarian and prostate cancer patients using tumor-specific fluorescent ligands. Int J Cancer. 2008; 123: 1968-73.
- Fan T, Zhao Q, Chen JJ, Chen WT, Pearl ML. Clinical significance of circulating tumor cells detected by an invasion assay in peripheral blood of patients with ovarian cancer. Gynecol Oncol. 2009; 112: 185-91.
- Krebs MG, Metcalf RL, Carter L, Brady G, Blackhall FH, Dive C. Molecular analysis of circulating tumour cells-biology and biomarkers. Nat Rev Clin Oncol. 2014; 11: 129-44.
- Poveda A, Kaye SB, McCormack R, Wang S, Parekh T, Ricci D, et al. Circulating tumor cells predict progression free survival and overall survival in patients with relapsed/recurrent advanced ovarian cancer. Gynecol Oncol. 2011; 122: 567-72.
- Pearl ML, Zhao Q, Yang J, Dong H, Tulley S, Zhang Q, et al. Prognostic analysis of invasive circulating tumor cells (iCTCs) in epithelial ovarian cancer. Gynecol Oncol. 2014; 134: 581-90.
- Buderath P, Schwich E, Jensen C, Horn PA, Kimmig R, Kasimir-Bauer S, et al. Soluble Programmed Death Receptor Ligands sPD-L1 and sPD-L2 as Liquid

Biopsy Markers for Prognosis and Platinum Response in Epithelial Ovarian Cancer. Front Oncol. 2019; 9: 1015.

- Kiss I, Kolostova K, Matkowski R, Jedryka M, Czekanski A, Pavlasek J, *et al.* Correlation Between Disease Stage and the Presence of Viable Circulating Tumor Cells in Endometrial Cancer. Anticancer Res. 2018; 38: 2983-7.
- Carvalho MJ, Laranjo M, Abrantes AM, Torgal I, Botelho MF, Oliveira CF. Clinical translation for endometrial cancer stem cells hypothesis. Cancer Metastasis Rev. 2015; 34: 401-16.
- Ni T, Sun X, Shan B, Wang J, Liu Y, Gu SL, et al. Detection of circulating tumour cells may add value in endometrial cancer management. Eur J Obstet Gynecol Reprod Biol. 2016; 207: 1-4.
- Berzofsky JA, Hoyoung M. Abstract A004: HER2 cancer vaccine phase I clinical trial shows clinical benefit in 54% of evaluable patients. Cancer Immunol Res. 2019; 7: A004.
- Manyam M. 120 Factors associated with extended survival for patients with platinum-resistant ovarian cancer (PROC) treated with modified vaccinia oncolytic virotherapy (VOV). International Journal of Gynecologic Cancer. 2019; 29.
- Luen SJ, Savas P, Fox SB, Salgado R, Loi S. Tumour-infiltrating lymphocytes and the emerging role of immunotherapy in breast cancer. Pathology. 2017; 49: 141-55.
- Adams S, Loi S, Toppmeyer D, Cescon DW, De Laurentiis M, Nanda R, et al. Pembrolizumab monotherapy for previously untreated, PD-L1-positive, metastatic triple-negative breast cancer: cohort B of the phase II KEYNOTE-086 study. Ann Oncol. 2019; 30: 405-11.
- Tolaney SM, Kalinsky K, Kaklamani VG, D'Adamo DR, Aktan G, Tsai ML, et al. Eribulin Plus Pembrolizumab in Patients with Metastatic Triple-Negative Breast Cancer (ENHANCE 1): A Phase Ib/II Study. Clin Cancer Res. 2021.
- Schmid P, Adams S, Rugo HS, Schneeweiss A, Barrios CH, Iwata H, et al. Atezolizumab and nab-paclitaxel in advanced triple-negative breast cancer. N Engl J Med. 2018; 379: 2108-21.
- Cortes J, Cescon DW, Rugo HS, Nowecki Z, Im SA, Yusof MM, et al. Pembrolizumab plus chemotherapy versus placebo plus chemotherapy for previously untreated locally recurrent inoperable or metastatic triple-negative breast cancer (KEYNOTE-355): a randomised, placebo-controlled, double-blind, phase 3 clinical trial. Lancet. 2020; 396: 1817-28.
- Winer EP, Lipatov O, Im SA, Goncalves A, Munoz-Couselo E, Lee KS, et al. Pembrolizumab versus investigator-choice chemotherapy for metastatic triple-negative breast cancer (KEYNOTE-119): a randomised, open-label, phase 3 trial. Lancet Oncol. 2021; 22: 499-511.
- Emens LA, Esteva FJ, Beresford M, Saura C, De Laurentiis M, Kim SB, et al. Trastuzumab emtansine plus atezolizumab versus trastuzumab emtansine plus placebo in previously treated, HER2-positive advanced breast cancer (KATE2): a phase 2, multicentre, randomised, double-blind trial. Lancet Oncol. 2020; 21: 1283-95.
- Miglietta F, Griguolo G, Guarneri V, Dieci MV. Programmed Cell Death Ligand 1 in Breast Cancer: Technical Aspects, Prognostic Implications, and Predictive Value. Oncologist. 2019; 24: e1055-e69.
- Thompson E, Taube JM, Elwood H, Sharma R, Meeker A, Warzecha HN, et al. The immune microenvironment of breast ductal carcinoma in situ. Mod Pathol. 2016; 29: 249-58.
- Sabatier R, Finetti P, Mamessier E, Adelaide J, Chaffanet M, Ali HR, et al. Prognostic and predictive value of PDL1 expression in breast cancer. Oncotarget. 2015; 6: 5449-64.
- Dill EA, Gru AA, Atkins KA, Friedman LA, Moore ME, Bullock TN, et al. PD-L1 Expression and Intratumoral Heterogeneity Across Breast Cancer Subtypes and Stages: An Assessment of 245 Primary and 40 Metastatic Tumors. Am J Surg Pathol. 2017; 41: 334-42.
- Ghebeh H, Tulbah A, Mohammed S, Elkum N, Bin Amer SM, Al-Tweigeri T, et al. Expression of B7-H1 in breast cancer patients is strongly associated with high proliferative Ki-67-expressing tumor cells. Int J Cancer. 2007; 121: 751-8.
- Li F, Ren Y, Wang Z. Programmed death 1 Ligand 1 expression in breast cancer and its association with patients' clinical parameters. J Cancer Res Ther. 2018; 14: 150-4.
- Qin T, Zeng YD, Qin G, Xu F, Lu JB, Fang WF, et al. High PD-L1 expression was associated with poor prognosis in 870 Chinese patients with breast cancer. Oncotarget. 2015; 6: 33972-81.
- Muenst S, Schaerli AR, Gao F, Daster S, Trella E, Droeser RA, et al. Expression of programmed death ligand 1 (PD-L1) is associated with poor prognosis in human breast cancer. Breast Cancer Res Treat. 2014; 146: 15-24.
- Tsang JY, Au WL, Lo KY, Ni YB, Hlaing T, Hu J, et al. PD-L1 expression and tumor infiltrating PD-1+ lymphocytes associated with outcome in HER2+ breast cancer patients. Breast Cancer Res Treat. 2017; 162: 19-30.
- Adams TA, Vail PJ, Ruiz A, Mollaee M, McCue PA, Knudsen ES, et al. Composite analysis of immunological and metabolic markers defines novel subtypes of triple negative breast cancer. Mod Pathol. 2018; 31: 288-98.
- Sun WY, Lee YK, Koo JS. Expression of PD-L1 in triple-negative breast cancer based on different immunohistochemical antibodies. J Transl Med. 2016; 14: 173.
- Mardones MA, Grosser D, Levin MK, Daoud Y, Palucka K, O'Shaughnessy J, et al. Abstract P2-04-20: PD-L1 expression in triple negative breast cancer (TNBC) is associated with improved outcomes. Cancer Research. 2017; 77: P2-04.

- Barrett MT, Lenkiewicz E, Malasi S, Basu A, Yearley JH, Annamalai L, et al. The association of genomic lesions and PD-1/PD-L1 expression in resected triple-negative breast cancers. Breast Cancer Res. 2018; 20: 71.
- Ali HR, Glont SE, Blows FM, Provenzano E, Dawson SJ, Liu B, et al. PD-L1 protein expression in breast cancer is rare, enriched in basal-like tumours and associated with infiltrating lymphocytes. Ann Oncol. 2015; 26: 1488-93.
- Tawfik O, Kimler BF, Karnik T, Shehata P. Clinicopathological correlation of PD-L1 expression in primary and metastatic breast cancer and infiltrating immune cells. Hum Pathol. 2018; 80: 170-8.
- Dirix LY, Takacs I, Jerusalem G, Nikolinakos P, Arkenau HT, Forero-Torres A, et al. Avelumab, an anti-PD-L1 antibody, in patients with locally advanced or metastatic breast cancer: a phase 1b JAVELIN Solid Tumor study. Breast Cancer Res Treat. 2018; 167: 671-86.
- Emens LA, Loi S, Rugo HS, Schneeweiss A, Diéras V, Iwata H, et al. Abstract GS1-04: IMpassion130: Efficacy in immune biomarker subgroups from the global, randomized, double-blind, placebo-controlled, phase III study of atezolizumab + <em>nab</em>-paclitaxel in patients with treatment-naïve, locally advanced or metastatic triple-negative breast cancer. Cancer Research. 2019; 79: GS1-04.
- 92. Rossi E, Fabbri F. CTCs 2020: Great Expectations or Unreasonable Dreams. Cells. 2019; 8.
- Li M, Li A, Zhou S, Xu Y, Xiao Y, Bi R, *et al*. Heterogeneity of PD-L1 expression in primary tumors and paired lymph node metastases of triple negative breast cancer. BMC Cancer. 2018; 18: 4.
- Jilaveanu LB, Shuch B, Zito CR, Parisi F, Barr M, Kluger Y, et al. PD-L1 Expression in Clear Cell Renal Cell Carcinoma: An Analysis of Nephrectomy and Sites of Metastases. J Cancer. 2014; 5: 166-72.
- 95. Schott DS, Pizon M, Pachmann U, Pachmann K. Sensitive detection of PD-L1 expression on circulating epithelial tumor cells (CETCs) could be a potential biomarker to select patients for treatment with PD-1/PD-L1 inhibitors in early and metastatic solid tumors. Oncotarget. 2017; 8: 72755-72.
- Kawaguchi K, Suzuki E, Yamaguchi A, Yamamoto M, Morita S, Toi M. Altered expression of major immune regulatory molecules in peripheral blood immune cells associated with breast cancer. Breast Cancer. 2017; 24: 111-20.
- Moku PR, Shepherd LE, Ali SM, Leitzel K, Parulekar WE, Zhu L, et al. Abstract PD3-10: Higher serum PD-L1 predicts for increased overall survival to lapatinib vs. trastuzumab in the phase 3 CCTG MA.31 trial. Cancer Research. 2019; 79: PD3-10.
- Horn L, Spigel DR, Vokes EE, Holgado E, Ready N, Steins M, et al. Nivolumab Versus Docetaxel in Previously Treated Patients With Advanced Non-Small-Cell Lung Cancer: Two-Year Outcomes From Two Randomized, Open-Label, Phase III Trials (CheckMate 017 and CheckMate 057). J Clin Oncol. 2017; 35: 3924-33.
- Herbst RS, Baas P, Kim DW, Felip E, Perez-Gracia JL, Han JY, et al. Pembrolizumab versus docetaxel for previously treated, PD-L1-positive, advanced non-small-cell lung cancer (KEYNOTE-010): a randomised controlled trial. Lancet. 2016; 387: 1540-50.
- 100. Rittmeyer A, Barlesi F, Waterkamp D, Park K, Ciardiello F, von Pawel J, et al. Atezolizumab versus docetaxel in patients with previously treated non-small-cell lung cancer (OAK): a phase 3, open-label, multicentre randomised controlled trial. Lancet. 2017; 389: 255-65.
- 101. Borghaei H, Gettinger S, Vokes EE, Chow LQM, Burgio MA, de Castro Carpeno J, et al. Five-Year Outcomes From the Randomized, Phase III Trials CheckMate 017 and 057: Nivolumab Versus Docetaxel in Previously Treated Non-Small-Cell Lung Cancer. J Clin Oncol. 2021; 39: 723-33.
- 102. Reck M, Rodriguez-Abreu D, Robinson AG, Hui R, Csoszi T, Fulop A, et al. Updated Analysis of KEYNOTE-024: Pembrolizumab Versus Platinum-Based Chemotherapy for Advanced Non-Small-Cell Lung Cancer With PD-L1 Tumor Proportion Score of 50% or Greater. J Clin Oncol. 2019; 37: 537-46.
- Herbst RS, Giaccone G, de Marinis F, Reinmuth N, Vergnenegre A, Barrios CH, et al. Atezolizumab for First-Line Treatment of PD-L1-Selected Patients with NSCLC. N Engl J Med. 2020; 383: 1328-39.
- 104. Sezer A, Kilickap S, Gumus M, Bondarenko I, Ozguroglu M, Gogishvili M, et al. Cemiplimab monotherapy for first-line treatment of advanced non-small-cell lung cancer with PD-L1 of at least 50%: a multicentre, open-label, global, phase 3, randomised, controlled trial. Lancet. 2021; 397: 592-604.
- 105. Rodriguez-Abreu D, Johnson ML, Hussein MA, Cobo M, Patel AJ, Secen NM, et al. Primary analysis of a randomized, double-blind, phase II study of the anti-TIGIT antibody tiragolumab (tira) plus atezolizumab (atezo) versus placebo plus atezo as first-line (1L) treatment in patients with PD-L1-selected NSCLC (CITYSCAPE). Journal of Clinical Oncology. 2020; 38: 9503-.
- 106. Gadgeel S, Rodriguez-Abreu D, Speranza G, Esteban E, Felip E, Domine M, et al. Updated Analysis From KEYNOTE-189: Pembrolizumab or Placebo Plus Pemetrexed and Platinum for Previously Untreated Metastatic Nonsquamous Non-Small-Cell Lung Cancer. J Clin Oncol. 2020; 38: 1505-17.
- 107. Paz-Ares L, Vicente D, Tafreshi A, Robinson A, Soto Parra H, Mazieres J, et al. A Randomized, Placebo-Controlled Trial of Pembrolizumab Plus Chemotherapy in Patients With Metastatic Squamous NSCLC: Protocol-Specified Final Analysis of KEYNOTE-407. J Thorac Oncol. 2020; 15: 1657-69.
- Socinski MA, Jotte RM, Cappuzzo F, Orlandi F, Stroyakovskiy D, Nogami N, et al. Atezolizumab for First-Line Treatment of Metastatic Nonsquamous NSCLC. N Engl J Med. 2018; 378: 2288-301.

- 109. Nishio M, Barlesi F, West H, Ball S, Bordoni R, Cobo M, et al. Atezolizumab Plus Chemotherapy for First-Line Treatment of Nonsquamous NSCLC: Results From the Randomized Phase 3 IMpower132 Trial. J Thorac Oncol. 2021; 16: 653-64.
- 110. Reck M, Schenker M, Lee KH, Provencio M, Nishio M, Lesniewski-Kmak K, et al. Nivolumab plus ipilimumab versus chemotherapy as first-line treatment in advanced non-small-cell lung cancer with high tumour mutational burden: patient-reported outcomes results from the randomised, open-label, phase III CheckMate 227 trial. Eur J Cancer. 2019; 116: 137-47.
- 111. Boyer M, Sendur MAN, Rodriguez-Abreu D, Park K, Lee DH, Cicin I, et al. Pembrolizumab Plus Ipilimumab or Placebo for Metastatic Non-Small-Cell Lung Cancer With PD-L1 Tumor Proportion Score >/= 50%: Randomized, Double-Blind Phase III KEYNOTE-598 Study. J Clin Oncol. 2021: JCO2003579.
- 112. Gray JE, Villegas A, Daniel D, Vicente D, Murakami S, Hui R, et al. Three-Year Overall Survival with Durvalumab after Chemoradiotherapy in Stage III NSCLC-Update from PACIFIC. J Thorac Oncol. 2020; 15: 288-93.
- 113. Reck M, Lee Kh, Frost N, Kowalski DM, Breder V, Pollock T, et al. OA02.03 Pembrolizumab Plus Platinum Chemotherapy and Radiotherapy in Unresectable, Locally Advanced, Stage III NSCLC: KEYNOTE-799. Journal of Thoracic Oncology. 2021; 16: S103-S4.
- Wong MCS, Lao XQ, Ho KF, Goggins WB, Tse SLA. Incidence and mortality of lung cancer: global trends and association with socioeconomic status. Sci Rep. 2017; 7: 14300.
- 115. Institute NNC. Cancer Stat Facts: Lung and Bronchus Cancer [Online]. . 2017.
- 116. Tanaka F, Yoneda K, Kondo N, Hashimoto M, Takuwa T, Matsumoto S, *et al.* Circulating tumor cell as a diagnostic marker in primary lung cancer. Clin Cancer Res. 2009; 15: 6980-6.
- 117. Krebs MG, Sloane R, Priest L, Lancashire L, Hou JM, Greystoke A, *et al.* Evaluation and prognostic significance of circulating tumor cells in patients with non-small-cell lung cancer. J Clin Oncol. 2011; 29: 1556-63.
- Earhart CM, Hughes CE, Gaster RS, Ooi CC, Wilson RJ, Zhou LY, et al. Isolation and mutational analysis of circulating tumor cells from lung cancer patients with magnetic sifters and biochips. Lab Chip. 2014; 14: 78-88.
- 119. Yin J, Wang Y, Yin H, Chen W, Jin G, Ma H, et al. Circulating Tumor Cells Enriched by the Depletion of Leukocytes with Bi-Antibodies in Non-Small Cell Lung Cancer: Potential Clinical Application. PLoS One. 2015; 10: e0137076.
- 120. Zhang Z, Xiao Y, Zhao J, Chen M, Xu Y, Zhong W, et al. Relationship between circulating tumour cell count and prognosis following chemotherapy in patients with advanced non-small-cell lung cancer. Respirology. 2016; 21: 519-25.
- Wu ZX, Liu Z, Jiang HL, Pan HM, Han WD. Circulating tumor cells predict survival benefit from chemotherapy in patients with lung cancer. Oncotarget. 2016; 7: 67586-96.
- 122. Muinelo-Romay L, Vieito M, Abalo A, Nocelo MA, Baron F, Anido U, et al. Evaluation of Circulating Tumor Cells and Related Events as Prognostic Factors and Surrogate Biomarkers in Advanced NSCLC Patients Receiving First-Line Systemic Treatment. Cancers (Basel). 2014; 6: 153-65.
- Lianidou ES, Markou A, Strati A. The Role of CTCs as Tumor Biomarkers. Adv Exp Med Biol. 2015; 867: 341-67.
- 124. Dorsey JF, Kao GD, MacArthur KM, Ju M, Steinmetz D, Wileyto EP, et al. Tracking viable circulating tumor cells (CTCs) in the peripheral blood of non-small cell lung cancer (NSCLC) patients undergoing definitive radiation therapy: pilot study results. Cancer. 2015; 121: 139-49.
- 125. Morabito A, Sandomenico C, Costanzo R, Montanino A, Caraco C, De Lutio E, *et al.* Positron emission tomography and circulating tumor cells to monitor a dramatic response to gefitinib. J Thorac Oncol. 2012; 7: e27-8.
- 126. Yanagita M, Redig AJ, Paweletz CP, Dahlberg SE, O'Connell A, Feeney N, et al. A Prospective Evaluation of Circulating Tumor Cells and Cell-Free DNA in EGFR-Mutant Non-Small Cell Lung Cancer Patients Treated with Erlotinib on a Phase II Trial. Clin Cancer Res. 2016; 22: 6010-20.
- 127. First Anti-PD-L1 Drug Approved for NSCLC. Cancer Discov. 2016; 6: OF1.
- 128. Sul J, Blumenthal GM, Jiang X, He K, Keegan P, Pazdur R. FDA Approval Summary: Pembrolizumab for the Treatment of Patients With Metastatic Non-Small Cell Lung Cancer Whose Tumors Express Programmed Death-Ligand 1. Oncologist. 2016; 21: 643-50.
- Bylicki O, Barazzutti H, Paleiron N, Margery J, Assie JB, Chouaid C. First-Line Treatment of Non-Small-Cell Lung Cancer (NSCLC) with Immune Checkpoint Inhibitors. BioDrugs. 2019; 33: 159-71.
- Malhotra J, Jabbour SK, Aisner J. Current state of immunotherapy for non-small cell lung cancer. Transl Lung Cancer Res. 2017; 6: 196-211.
- 131. Califano R, Lal R, Lewanski C, Nicolson MC, Ottensmeier CH, Popat S, et al. Patient selection for anti-PD-1/PD-L1 therapy in advanced non-small-cell lung cancer: implications for clinical practice. Future Oncol. 2018; 14: 2415-31.
- 132. Schehr JL, Schultz ZD, Warrick JW, Guckenberger DJ, Pezzi HM, Sperger JM, et al. High Specificity in Circulating Tumor Cell Identification Is Required for Accurate Evaluation of Programmed Death-Ligand 1. PLoS One. 2016; 11: e0159397.
- 133. Kallergi G, Vetsika EK, Aggouraki D, Lagoudaki E, Koutsopoulos A, Koinis F, et al. Evaluation of PD-L1/PD-1 on circulating tumor cells in patients with advanced non-small cell lung cancer. Ther Adv Med Oncol. 2018; 10: 1758834017750121.
- 134. Adams DL, Adams DK, He J, Kalhor N, Zhang M, Xu T, et al. Sequential Tracking of PD-L1 Expression and RAD50 Induction in Circulating Tumor and

Stromal Cells of Lung Cancer Patients Undergoing Radiotherapy. Clin Cancer Res. 2017; 23: 5948-58.

- Wang Y, Kim TH, Fouladdel S, Zhang Z, Soni P, Qin A, et al. PD-L1 Expression in Circulating Tumor Cells Increases during Radio(chemo)therapy and Indicates Poor Prognosis in Non-small Cell Lung Cancer. Sci Rep. 2019; 9: 566.
- 136. Ilie M, Szafer-Glusman E, Hofman V, Chamorey E, Lalvee S, Selva E, et al. Detection of PD-L1 in circulating tumor cells and white blood cells from patients with advanced non-small-cell lung cancer. Ann Oncol. 2018; 29: 193-9.
- 137. Janning M, Kobus F, Babayan A, Wikman H, Velthaus JL, Bergmann S, et al. Determination of PD-L1 Expression in Circulating Tumor Cells of NSCLC Patients and Correlation with Response to PD-1/PD-L1 Inhibitors. Cancers (Basel). 2019; 11.
- Nicolazzo C, Raimondi C, Mancini M, Caponnetto S, Gradilone A, Gandini O, et al. Monitoring PD-L1 positive circulating tumor cells in non-small cell lung cancer patients treated with the PD-1 inhibitor Nivolumab. Sci Rep. 2016; 6: 31726.
- 139. Guibert N, Delaunay M, Lusque A, Boubekeur N, Rouquette I, Clermont E, et al. PD-L1 expression in circulating tumor cells of advanced non-small cell lung cancer patients treated with nivolumab. Lung Cancer. 2018; 120: 108-12.
- 140. Kulasinghe A, Kapeleris J, Kimberley R, Mattarollo SR, Thompson EW, Thiery JP, et al. The prognostic significance of circulating tumor cells in head and neck and non-small-cell lung cancer. Cancer Med. 2018; 7: 5910-9.
- 141. Dhar M, Wong J, Che J, Matsumoto M, Grogan T, Elashoff D, et al. Evaluation of PD-L1 expression on vortex-isolated circulating tumor cells in metastatic lung cancer. Scientific reports. 2018; 8: 2592.
- 142. Raimondi C, Carpino G, Nicolazzo C, Gradilone A, Gianni W, Gelibter A, et al. PD-L1 and epithelial-mesenchymal transition in circulating tumor cells from non-small cell lung cancer patients: A molecular shield to evade immune system? Oncoimmunology. 2017; 6: e1315488.
- 143. Kloten V, Lampignano R, Krahn T, Schlange T. Circulating Tumor Cell PD-L1 Expression as Biomarker for Therapeutic Efficacy of Immune Checkpoint Inhibition in NSCLC. Cells. 2019; 8.
- 144. Leemans CR, Braakhuis BJ, Brakenhoff RH. The molecular biology of head and neck cancer. Nat Rev Cancer. 2011; 11: 9-22.
- 145. Vermorken JB, Mesia R, Rivera F, Remenar E, Kawecki A, Rottey S, et al. Platinum-based chemotherapy plus cetuximab in head and neck cancer. N Engl J Med. 2008; 359: 1116-27.
- 146. Kuss I, Hathaway B, Ferris RL, Gooding W, Whiteside TLJCcr. Decreased absolute counts of T lymphocyte subsets and their relation to disease in squamous cell carcinoma of the head and neck. Clin Cancer Res. 2004; 10: 3755-62.
- Bauernhofer T, Kuss I, Henderson B, Baum AS, Whiteside TLJEjoi. Preferential apoptosis of CD56dim natural killer cell subset in patients with cancer. Eur J Immunol. 2003; 33: 119-24.
- 148. Dasgupta S, Bhattacharya-Chatterjee M, O'Malley BW, Chatterjee SKJTJoI. Inhibition of NK cell activity through TGF-β1 by down-regulation of NKG2D in a murine model of head and neck cancer. J Immunol. 2005; 175: 5541-50.
- 149. López-Albaitero A, Nayak JV, Ogino T, Machandia A, Gooding W, DeLeo AB, et al. Role of antigen-processing machinery in the in vitro resistance of squamous cell carcinoma of the head and neck cells to recognition by CTL. J Immunol. 2006; 176: 3402-9.
- 150. Lawrence MS, Stojanov P, Polak P, Kryukov GV, Cibulskis K, Sivachenko A, et al. Mutational heterogeneity in cancer and the search for new cancer-associated genes. Nature. 2013; 499: 214-8.
- Kammertoens T, Schüler T, Blankenstein TJTimm. Immunotherapy: target the stroma to hit the tumor. Trends Mol Med. 2005; 11: 225-31.
- Ferris RL. Immunology and Immunotherapy of Head and Neck Cancer. J Clin Oncol. 2015; 33: 3293-304.
- 153. Mehra R, Seiwert TY, Gupta S, Weiss J, Gluck I, Eder JP, *et al.* Efficacy and safety of pembrolizumab in recurrent/metastatic head and neck squamous cell carcinoma: pooled analyses after long-term follow-up in KEYNOTE-012. Br J Cancer. 2018; 119: 153-9.
- 154. Seiwert TY, Burtness B, Mehra R, Weiss J, Berger R, Eder JP, et al. Safety and clinical activity of pembrolizumab for treatment of recurrent or metastatic squamous cell carcinoma of the head and neck (KEYNOTE-012): an open-label, multicentre, phase 1b trial. Lancet Oncol. 2016; 17: 956-65.
- 155. Ferris RL, Blumenschein Jr G, Fayette J, Guigay J, Colevas AD, Licitra L, et al. Nivolumab for recurrent squamous-cell carcinoma of the head and neck. N Engl J Med. 2016; 375: 1856-67.
- 156. Bauml J, Seiwert TY, Pfister DG, Worden F, Liu SV, Gilbert J, et al. Pembrolizumab for platinum-and cetuximab-refractory head and neck cancer: results from a single-arm, phase II study. J Clin Oncol. 2017; 35: 1542.
- 157. Cohen EE, Soulières D, Le Tourneau C, Dinis J, Licitra L, Ahn M-J, et al. Pembrolizumab versus methotrexate, docetaxel, or cetuximab for recurrent or metastatic head-and-neck squamous cell carcinoma (KEYNOTE-040): a randomised, open-label, phase 3 study. Lancet. 2019; 393: 156-67.
- Kulasinghe A, Perry C, Kenny L, Warkiani ME, Nelson C, Punyadeera C. PD-L1 expressing circulating tumour cells in head and neck cancers. BMC Cancer. 2017; 17: 333.
- 159. Warkiani ME, Guan G, Luan KB, Lee WC, Bhagat AA, Chaudhuri PK, *et al.* Slanted spiral microfluidics for the ultra-fast, label-free isolation of circulating tumor cells. Lab Chip. 2014; 14: 128-37.
- 160. Markou A, Strati A, Malamos N, Georgoulias V, Lianidou ES. Molecular characterization of circulating tumor cells in breast cancer by a liquid bead array hybridization assay. Clin Chem. 2011; 57: 421-30.

- 161. Chimonidou M, Strati A, Malamos N, Georgoulias V, Lianidou ES. SOX17 promoter methylation in circulating tumor cells and matched cell-free DNA isolated from plasma of patients with breast cancer. Clin Chem. 2013; 59: 270-9.
- Lianidou ES, Markou A. Circulating tumor cells in breast cancer: detection systems, molecular characterization, and future challenges. Clin Chem. 2011; 57: 1242-55.
- 163. Fitzmaurice C, Abate D, Abbasi N, Abbastabar H, Abd-Allah F, Abdel-Rahman O, et al. Global, regional, and national cancer incidence, mortality, years of life lost, years lived with disability, and disability-adjusted life-years for 29 cancer groups, 1990 to 2017: a systematic analysis for the global burden of disease study. JAMA Oncol. 2019; 5: 1749-68.
- Pantel K, Brakenhoff RH. Dissecting the metastatic cascade. Nat Rev Cancer. 2004; 4: 448-56.
- Miller KD, Siegel RL, Lin CC, Mariotto AB, Kramer JL, Rowland JH, et al. Cancer treatment and survivorship statistics, 2016. CA Cancer J Clin. 2016; 66: 271-89.
- Klein C. Parallel progression of primary tumours and metastases. Nat Rev Cancer. 2009; 9: 302-12.
- Micalizzi DS, Maheswaran S, Haber DA. A conduit to metastasis: circulating tumor cell biology. Genes Dev. 2017; 31: 1827-40.
- Yang C, Chen F, Wang S, Xiong B. Circulating Tumor Cells in Gastrointestinal Cancers: Current Status and Future Perspectives. Front Oncol. 2019; 9: 1427-43.
- 169. Shitara K, Ozguroglu M, Bang YJ, Di Bartolomeo M, Mandala M, Ryu MH, et al. Pembrolizumab versus paclitaxel for previously treated, advanced gastric or gastro-oesophageal junction cancer (KEYNOTE-061): a randomised, open-label, controlled, phase 3 trial. Lancet. 2018; 392: 123-33.
- 170. Bang YJ, Kang YK, Catenacci DV, Muro K, Fuchs CS, Geva R, et al. Pembrolizumab alone or in combination with chemotherapy as first-line therapy for patients with advanced gastric or gastroesophageal junction adenocarcinoma: results from the phase II nonrandomized KEYNOTE-059 study. Gastric Cancer. 2019; 22: 828-37.
- 171. Kojima T, Shah MA, Muro K, Francois E, Adenis A, Hsu CH, et al. Randomized Phase III KEYNOTE-181 Study of Pembrolizumab Versus Chemotherapy in Advanced Esophageal Cancer. J Clin Oncol. 2020; 38: 4138-48.
- 172. Kato K, Shah MA, Enzinger P, Bennouna J, Shen L, Adenis A, et al. KEYNOTE-590: Phase III study of first-line chemotherapy with or without pembrolizumab for advanced esophageal cancer. Future Oncol. 2019; 15: 1057-66.
- 173. Shitara K, Van Cutsem E, Bang YJ, Fuchs C, Wyrwicz L, Lee KW, et al. Efficacy and Safety of Pembrolizumab or Pembrolizumab Plus Chemotherapy vs. Chemotherapy Alone for Patients With First-line, Advanced Gastric Cancer: The KEYNOTE-062 Phase 3 Randomized Clinical Trial. JAMA Oncol. 2020; 6: 1571-80.
- 174. Moehler M, Shitara K, Garrido M, Salman P, Shen L, Wyrwicz L, et al. LBA6\_PR Nivolumab (nivo) plus chemotherapy (chemo) versus chemo as first-line (1L) treatment for advanced gastric cancer/gastroesophageal junction cancer (GC/GEJC)/esophageal adenocarcinoma (EAC): First results of the CheckMate 649 study. Annals of Oncology. 2020; 31: S1191.
- 175. Boku N, Ryu MH, Oh DY, Oh SC, Chung H, Lee KW, et al. LBA7\_PR Nivolumab plus chemotherapy versus chemotherapy alone in patients with previously untreated advanced or recurrent gastric/gastroesophageal junction (G/GEJ) cancer: ATTRACTION-4 (ONO-4538-37) study. Annals of Oncology. 2020; 31: S1192.
- 176. Finn RS, Qin S, Ikeda M, Galle PR, Ducreux M, Kim T-Y, et al. IMbrave150: Updated overall survival (OS) data from a global, randomized, open-label phase III study of atezolizumab (atezo) + bevacizumab (bev) versus sorafenib (sor) in patients (pts) with unresectable hepatocellular carcinoma (HCC). Journal of Clinical Oncology. 2021; 39: 267-89.
- 177. Wong JSL, Kwok GW, Tang V, Li B, Leung RC-Y, Chiu JWY, et al. Ipilimumab and nivolumab/pembrolizumab in advanced hepatocellular carcinoma refractory to prior immune checkpoint inhibitors. Journal of Clinical Oncology. 2021; 39: 330-51.
- 178. Fuchs CS, Doi T, Jang RW, Muro K, Satoh T, Machado M, et al. Safety and efficacy of pembrolizumab monotherapy in patients with previously treated advanced gastric and gastroesophageal junction cancer: phase 2 clinical KEYNOTE-059 trial. JAMA Oncol. 2018; 4: e180013.
- 179. Gandidi S, Massi D, Mandala M.PD-L1 expression in cancer patients receiving anti PD-1/PD-L1 antibodies: A systemic review and meta-analysis. Crit Rev Oncol Hematol. 2016; 10-29.
- 180. Cheng B, Tong G, Wu X, Cai W, Li Z, Tong Z, et al. Enumeration And Characterization Of Circulating Tumor Cells And Its Application In Advanced Gastric Cancer. Onco Targets Ther. 2019; 12: 7887-7901.
- 181. Wang Y, Kim TH, Fouladdel S, Zhang Z, Soni P, Qin A, et al. PD-L1 expression in circulating tumor cells increases during radio (chemo) therapy and indicates poor prognosis in non-small cell lung cancer. Sci Rep. 2019; 9: 1-9.
- 182. Yue C, Jiang Y, Li P, Wang Y, Xue J, Li N, et al. Dynamic change of PD-L1 expression on circulating tumor cells in advanced solid tumor patients undergoing PD-1 blockade therapy. Oncoimmunology. 2018; 7: e1438111.
- Winograd P, Hou S, Sadeghi S, Finn R, DiPardo B, Li Q, et al. Evaluation of hepatocellular carcinoma circulating tumor cells expressing programmed death-ligand 1. 2018; 20: S2-S3.
- 184. Winograd P, Hou S, Court CM, Lee YT, Chen PJ, Zhu Y, et al. Hepatocellular Carcinoma-Circulating Tumor Cells Expressing PD-L1 Are Prognostic and

Potentially Associated With Response to Checkpoint Inhibitors. Hepatol Commun. 2020; 4: 1527-40.

- Larkin J, Chiarion-Sileni V, Gonzalez R, Grob JJ, Cowey CL, Lao CD, et al. Combined nivolumab and ipilimumab or monotherapy in untreated melanoma. N Engl J Med. 2015; 373: 23-34.
- Postow MA, Chesney J, Pavlick AC, Robert C, Grossmann K, McDermott D, et al. Nivolumab and ipilimumab versus ipilimumab in untreated melanoma. N Engl J Med. 2015; 372: 2006-17.
- 187. Hamid O, Robert C, Daud A, Hodi FS, Hwu WJ, Kefford R, et al. Five-year survival outcomes for patients with advanced melanoma treated with pembrolizumab in KEYNOTE-001. Ann Oncol. 2019; 30: 582-8.
- Robert C, Long GV, Brady B, Dutriaux C, Maio M, Mortier L, et al. Nivolumab in previously untreated melanoma without BRAF mutation. N Engl J Med. 2015; 372: 320-30.
- Larkin J, Chiarion-Sileni V, Gonzalez R, Grob JJ, Rutkowski P, Lao CD, et al. Five-Year Survival with Combined Nivolumab and Ipilimumab in Advanced Melanoma. N Engl J Med. 2019; 381: 1535-46.
- 190. Aya-Bonilla CA, Marsavela G, Freeman JB, Lomma C, Frank MH, Khattak MA, et al. Isolation and detection of circulating tumour cells from metastatic melanoma patients using a slanted spiral microfluidic device. Oncotarget. 2017; 8: 67355-68.
- 191. Aya-Bonilla CA, Morici M, Hong X, McEvoy AC, Sullivan RJ, Freeman J, et al. Detection and prognostic role of heterogeneous populations of melanoma circulating tumour cells. Br J Cancer. 2020; 122: 1059-67.
- 192. Po JW, Ma Y, Balakrishna B, Brungs D, Azimi F, de Souza P, et al. Immunomagnetic isolation of circulating melanoma cells and detection of PD-L1 status. PLoS One. 2019; 14: e0211866.
- 193. Hong X, Sullivan RJ, Kalinich M, Kwan TT, Giobbie-Hurder A, Pan S, et al. Molecular signatures of circulating melanoma cells for monitoring early response to immune checkpoint therapy. Proc Natl Acad Sci U S A. 2018; 115: 2467-72.
- 194. Khattak M, Gray E, Reid A, Periera M, McEvoy A, Aya-Bonilla C, et al. 82P PD-L1 expression on pre-treatment circulating tumour cells, but not serum VEGF, is predictive of response to pembrolizumab in melanoma. Ann Oncol. 2018; 29: 80.
- 195. Khattak MA, Reid A, Freeman J, Pereira M, McEvoy A, Lo J, et al. PD-L1 Expression on Circulating Tumor Cells May Be Predictive of Response to Pembrolizumab in Advanced Melanoma: Results from a Pilot Study. Oncologist. 2020; 25: e520.