

## **An exosomal-carried short periostin isoform induces cardiomyocyte proliferation**

Carolina Balbi<sup>1,2\*</sup>, Giuseppina Milano<sup>1,3</sup>, Tudor E. Fertig<sup>4</sup>, Edoardo Lazzarini<sup>5</sup>, Sara Bolis<sup>1,5</sup>, Yoshiaki Taniyama<sup>6</sup>, Fumihiko Sanada<sup>6</sup>, Dario Di Silvestre<sup>7</sup>, Pierluigi Mauri<sup>7</sup>, Mihaela Gherghiceanu<sup>4</sup>, Thomas F. Lüscher<sup>2,8</sup>, Lucio Barile<sup>5,9,10</sup>, Giuseppe Vassalli<sup>1,2,10\*</sup>.

<sup>1</sup>Laboratory of Cellular and Molecular Cardiology, Istituto Cardiocentro Ticino, Lugano, Switzerland;

<sup>2</sup>Center for Molecular Cardiology, University of Zurich, Zurich, Switzerland;

<sup>3</sup>Laboratory of Cardiovascular Research, Lausanne University Hospital, Lausanne, Switzerland;

<sup>4</sup>Victor Babes National Institute of Pathology, Bucharest, Romania;

<sup>5</sup>Laboratory for Cardiovascular Theranostics, Istituto Cardiocentro Ticino, Lugano, Switzerland;

<sup>6</sup>Department of Clinical Gene Therapy, Osaka University Graduate School of Medicine, Suita, Osaka, Japan;

<sup>7</sup>Proteomics and Metabolomic Lab, ITB-CNR, Segrate, Italy;

<sup>8</sup>Royal Brompton & Harefield Hospital, Imperial College, London, UK;

<sup>9</sup>Institute of Life Science, Scuola Superiore Sant'Anna, Pisa, Italy;

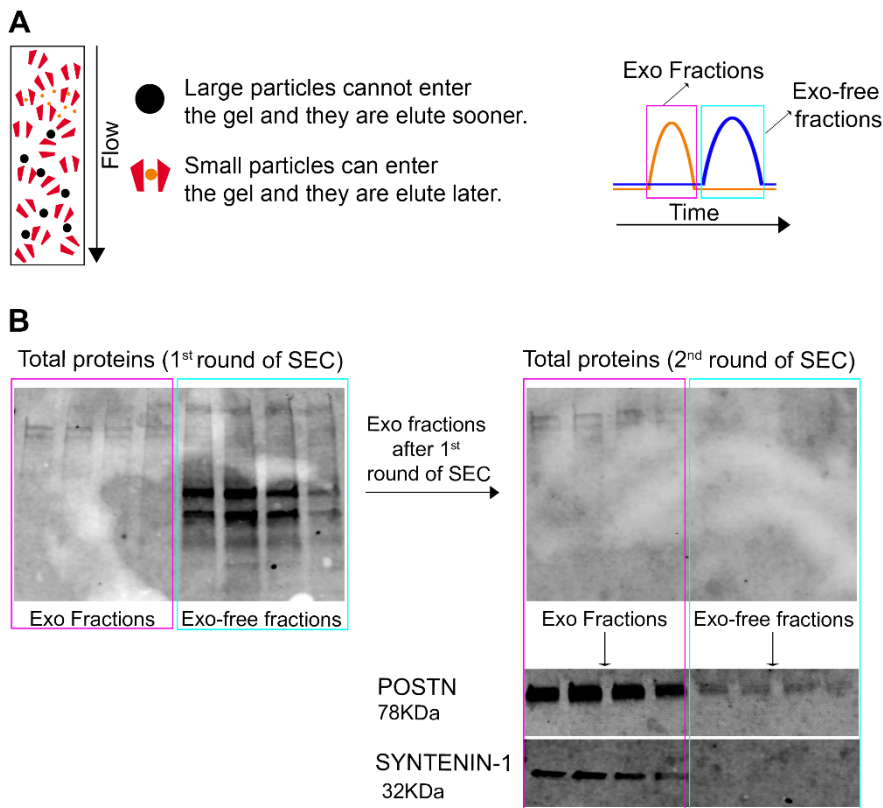
<sup>10</sup>Faculty of Biomedicine, Università della Svizzera Italiana (USI), Lugano, Switzerland.

### **\*Address correspondence to:**

Prof. Giuseppe Vassalli, Laboratory of Cellular and Molecular Cardiology, Istituto Cardiocentro Ticino, Via Tesserete 48, 6900 Lugano. Tel: +41 091 8053359. E-mail: [giuseppe.vassalli@cardiocentro.org](mailto:giuseppe.vassalli@cardiocentro.org)

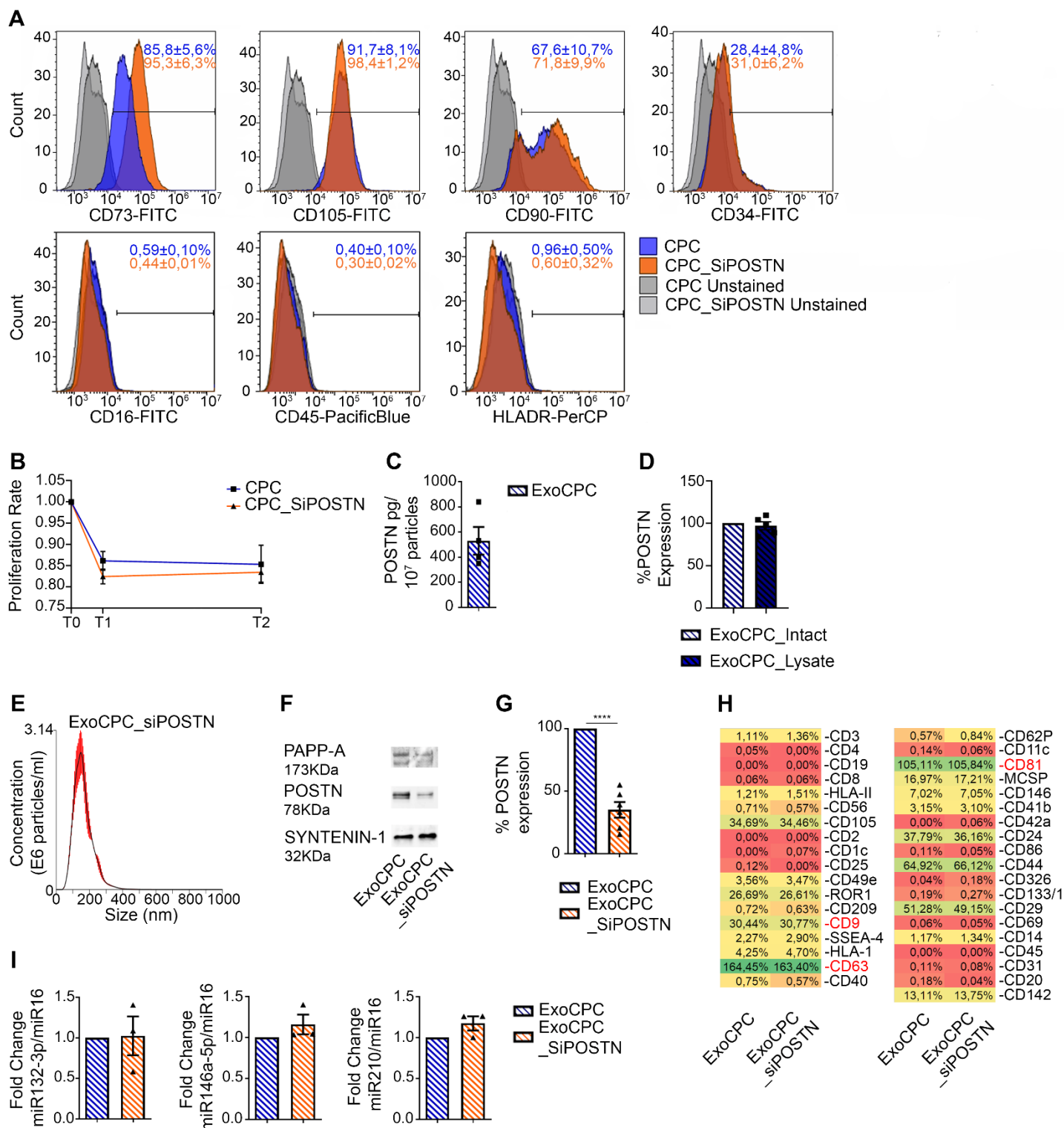
Dr. Carolina Balbi, Laboratory of Cellular and Molecular Cardiology, Istituto Cardiocentro Ticino, Via Tesserete 48, 6900 Lugano. Tel: +41 091 8053386. E-mail: [carolina.balbi@cardiocentro.org](mailto:carolina.balbi@cardiocentro.org)

## Supplementary Figures:



**Figure S1: Periostin enrichment in exosome-containing SEC fractions of CPC conditioned medium**

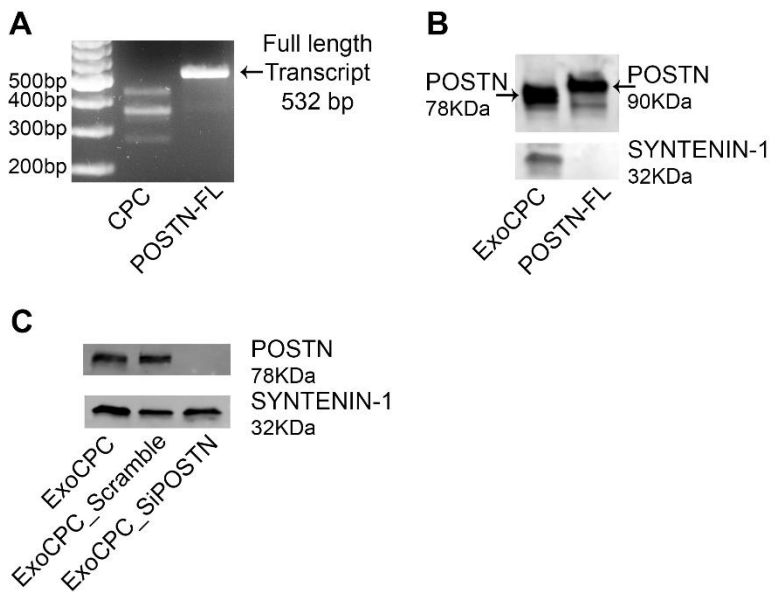
**A** Schematic of size exclusion chromatography (SEC) isolation principle. **B** Western blot analysis of total protein, periostin (POSTN) and exosome marker SYNTENIN-1. Left panel: Total protein staining after the first round of SEC purification. Right panel: Total protein, POSTN and SYNTENIN-1 after the second round of Exo SEC isolation. POSTN was enriched in Exo fractions, identified by high SYNTENIN-1 levels, vs. non-vesicular fractions.



**Figure S2: Characterization of periostin-silenced CPC and CPC-derived exosomes**

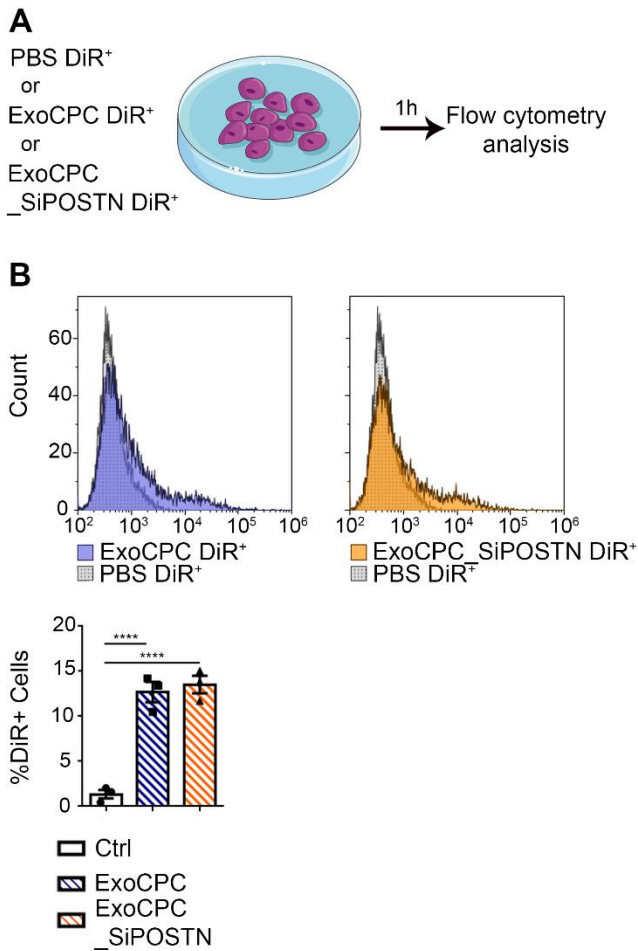
**A** Flow cytometry analysis of surface antigen expression in naïve CPC and CPC transfected with a siRNA against periostin (CPC\_SiPOSTN;  $n = 3$ ). **B** Proliferation rates of naïve CPC and CPC\_SiPOSTN at T0 (pre-transfection); T1 (24 hrs post-transfection) and T2 (5 days post-

transfection;  $n = 3$ ). **C** POSTN quantification (pg/ $10^7$  Exo particles) on intact ExoCPC by ELISA ( $n = 4$ ). **D** POSTN levels in ExoCPC lysate vs. intact ExoCPC, measured by ELISA (% of levels in intact ExoCPC;  $n = 5$ ). **E** Nanoparticle tracking analysis of Exo from CPC\_SiPOSTN (ExoCPC\_SiPOSTN). **F** Western analysis of pregnancy-associated plasma protein-A (PAPP-A)/pappalysin-1, POSTN, and SYNTENIN-1 in ExoCPC and ExoCPC\_SiPOSTN. **G** POSTN levels, normalized for SYNTENIN-1 levels, in ExoCPC\_SiPOSTN vs. naïve ExoCPC, as assessed by Western blotting (% of levels in naïve ExoCPC;  $n = 6$ ; \*\*\*\*  $p < 0.0001$ ). **H** MACSPlex analysis of surface marker expression in ExoCPC and ExoCPC\_SiPOSTN (green, yellow, and red colours indicate high, moderate, and low expression levels, respectively). **I** Real-time RT-PCR analysis of miR132-3p, miR146a-5p, and miR210 expression, in ExoCPC\_SiPOSTN vs. ExoCPC ( $n = 3$ ).



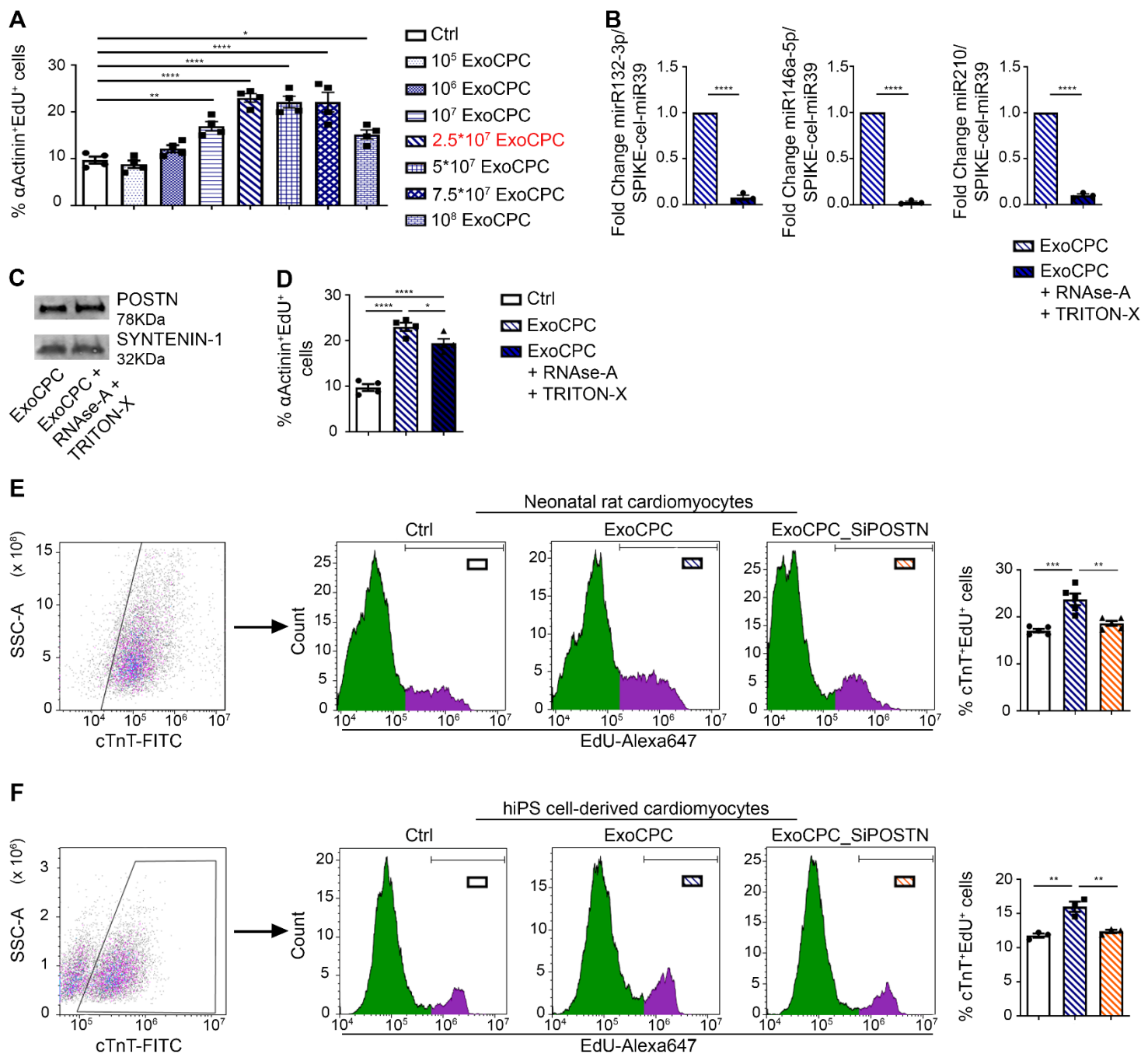
**Figure S3: Assessment of POSTN transcripts in CPC and POSTN protein levels in exosomes from naïve, scramble-siRNA transfected, and periostin-silenced transfected CPC**

**A** PCR analysis of human periostin (POSTN) on CPC cDNA. cDNA from a full-length human periostin (POSTN-FL) expression plasmid was used for comparison. **B** Western analysis of POSTN and SYNTENIN-1 (exosome marker) expression in ExoCPC. Recombinant human POSTN-FL was loaded for size comparison. **C** Western analysis of POSTN and SYNTENIN-1 expression in ExoCPC and in Exo from CPC transfected with scramble-siRNA (ExoCPC\_Scramble), or with siRNA against periostin (ExoCPC\_SiPOSTN).



**Figure S4: Periostin silencing does not affect exosome cellular uptake**

**A** Schematic of the experimental protocol. CPC-derived Exo (ExoCPC) were labelled with DiR fluorescent dye (ExoCPC DiR<sup>+</sup>) and re-purified by SEC to remove dye excess. DiR diluted in PBS at the same concentration used in ExoCPC, and processed with a SEC column, was used as a control (PBS DiR<sup>+</sup>). ExoCPC DiR<sup>+</sup> or PBS DiR<sup>+</sup> were added to cultured neonatal rat cardiomyocytes, which were analysed by flow cytometry 1 hour later. **B** Quantification of DiR<sup>+</sup> cells after treatment with naïve ExoCPC or Exo from CPC transfected with a siRNA against periostin (ExoCPC\_SiPOSTN;  $n = 3$ ; \*\*\*\*  $p < 0.0001$ ).



**Figure S5: Exosome dose-response study, effect of miRNAs on cell cycle activity, and periostin-dependent effect of exosomes on rat neonatal cardiomyocyte and hiPS cell-derived cardiomyocyte proliferation**

**A** Dose-response study of CPC-secreted Exo (ExoCPC) and neonatal rat cardiomyocyte cell cycle activity. Quantitative analysis of EdU-positive cardiomyocytes (% of cardiac  $\alpha$ -actinin-positive cells;  $n = 4$ ; \*  $p=0.0413$ ; \*\*  $p=0.0033$ ; \*\*\*\*  $p<0.0001$ ). A concentration of  $2.5 \times 10^7$  particles/well was found to be most effective. **B** Real-time RT-PCR analysis of miR132-3p, miR146a-5p, and miR210 levels

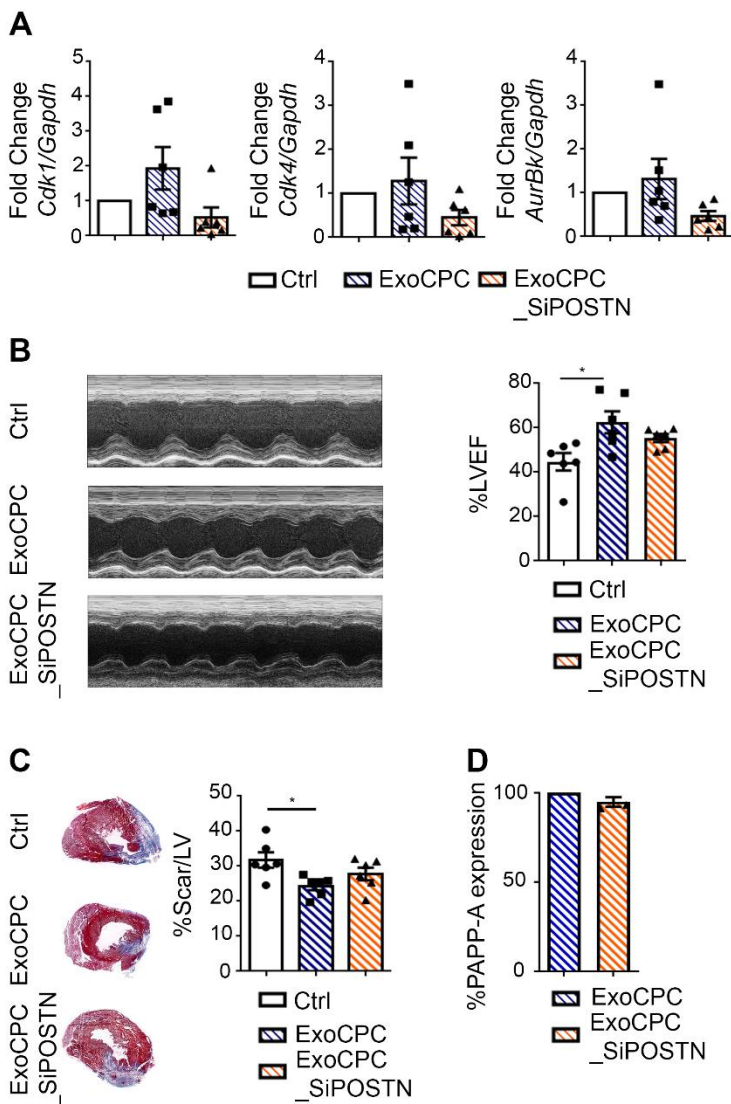
in both untreated and RNase-A-treated ExoCPC (fold-changes vs. ExoCPC;  $n = 3$ ; \*\*\*\*  $p < 0.0001$ ).

**C** Western analysis of POSTN and SYNTENIN-1 levels in both untreated and RNase-A-treated ExoCPC.

**D** Quantitative analysis of EdU-positive rat cardiomyocytes (% of  $\alpha$ Actinin-positive cells;  $n = 4$ ; \*  $p = 0.0417$ ; \*\*\*\*  $p < 0.0001$ ) treated with naïve CPC-derived Exo (ExoCPC), RNase-A treated ExoCPC or PBS (Ctrl).

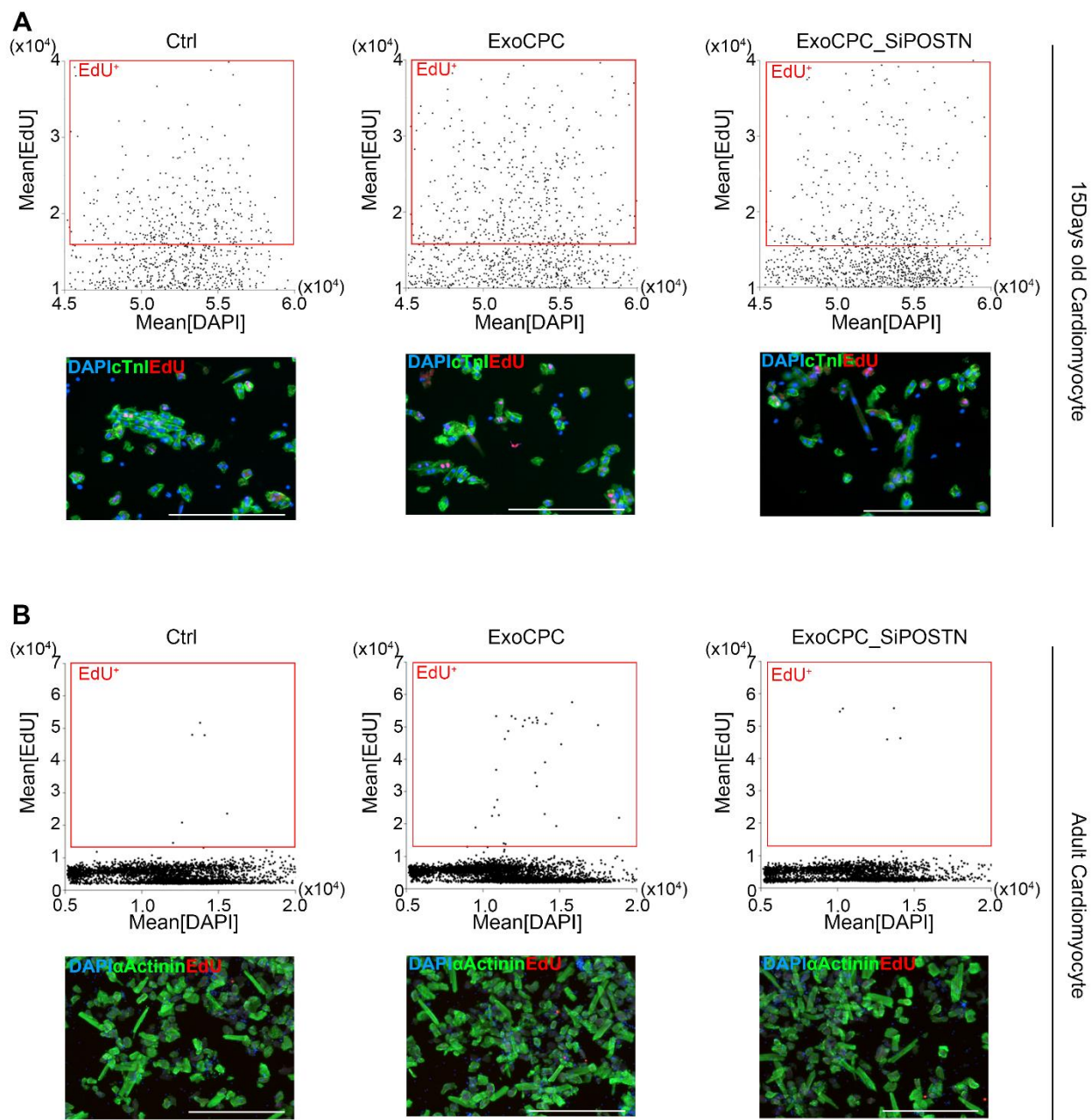
**E** Flow cytometric analysis of EdU<sup>+</sup> cardiomyocytes (% of cardiac-specific troponin T positive cells (TnT<sup>+</sup>)) treated with naïve ExoCPC, Exo from CPC transfected with a siRNA against POSTN (ExoCPC\_SiPOSTN), or PBS (Ctrl;  $n = 3$ ; \*\*  $p = 0.0019$ ; \*\*\*  $p = 0.0002$ ).

**F** Flow cytometry analysis of EdU<sup>+</sup> hiPS cell-derived cardiomyocytes (% of cTnT<sup>+</sup> cells) treated with naïve ExoCPC, ExoCPC\_SiPOSTN, or PBS (Ctrl;  $n = 3$ ; \*\*  $p = 0.0021$  and  $p = 0.0045$ ).



**Figure S6: Effects of CPC-derived exosomes in an *in vivo* model of adult rat myocardial infarction (MI).**

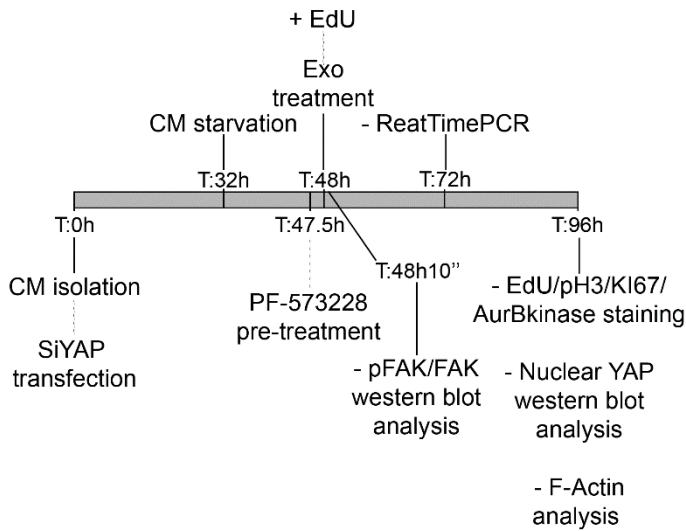
**A** Real-time RT-PCR analysis of *Cdk1*, *Cdk4*, and *AurBk* mRNA expression in dispersed isolated cardiomyocytes from rat hearts at day 14 post-MI. Infarcted rat hearts were injected intramyocardially with naïve ExoCPC, Exo from CPC transfected with a siRNA against periostin (ExoCPC\_SiPOSTN), or PBS (Ctrl; 6 animals/group). **B** Left panel: Representative M-mode echocardiographs at day 14 post-MI in the different groups. Right panel: Quantitative analysis of left ventricular ejection fraction (LVEF; %) at day 14 post-MI ( $n = 6$ ; \*  $p=0.0124$ ). **C** Representative heart sections at day 14 post-MI stained with Masson-trichrome. Quantitative analysis of infarct scar size (% of LV area;  $n = 6$ ; \*  $p=0.0251$ ). **D** Western blot analysis of pregnancy-associated plasma protein-A (PAPP-A) expression, normalized for SYNTENIN-1 expression, in naïve ExoCPC and ExoCPC\_SiPOSTN. (% ExoCPC;  $n = 3$ ).



**Figure S7: EdU nuclear incorporation analysis in dispersed isolated cardiomyocytes**

**A** Dispersed isolated neonatal rat cardiomyocyte (see Figure 5E for the schematic protocol of the treatment) immunostained for EdU (red) and cardiac-specific troponin I (cTnI; green). Scale bar: 200μm. Representative scatter plot of EdU+ nuclei on DAPI stained nuclei are shown. Quantitative analysis is shown in Figure 5E. **B** Dispersed isolated adult rat cardiomyocyte from hearts explanted at day 14 post-MI immunostained for EdU (red) and cardiac-specific sarcomeric α-actinin (green).

Scale bar: 200µm. Representative scatter plot of EdU+ nuclei on DAPI stained nuclei are shown. Quantitative analysis is shown in Figure 6C.



**Figure S8: *In vitro* experimental timeline**

Schematic of the experimental protocol of *in vitro* assays on neonatal rat cardiomyocytes. The relevant time points (T) are indicated.

#### Supplementary Tables:

**Supplementary Table 1**

| Accession        | Description                                                       | nPSM  |
|------------------|-------------------------------------------------------------------|-------|
| <b>30102</b>     | type I collagen [Homo sapiens]                                    | 185,6 |
| <b>119590947</b> | fibronectin 1, isoform CRA_I [Homo sapiens]                       | 184,2 |
| <b>762938</b>    | unnamed protein product [Homo sapiens]                            | 130,3 |
| <b>211904152</b> | glia-derived nexin isoform b precursor [Homo sapiens]             | 116,0 |
| <b>378404908</b> | glyceraldehyde-3-phosphate dehydrogenase isoform 2 [Homo sapiens] | 114,2 |
| <b>557786190</b> | <b>periostin isoform 7 precursor [Homo sapiens]</b>               | 98,4  |

|                  |                                                                                               |      |
|------------------|-----------------------------------------------------------------------------------------------|------|
| <b>119623487</b> | histone 1, H2bj, isoform CRA_b [Homo sapiens]                                                 | 57,8 |
| <b>194388798</b> | unnamed protein product [Homo sapiens]                                                        | 44,6 |
| <b>109150416</b> | peroxidasin homolog precursor [Homo sapiens]                                                  | 44,2 |
| <b>530366841</b> | PREDICTED: peroxidasin homolog isoform X1 [Homo sapiens]                                      | 42,5 |
| <b>86651742</b>  | tyrosine 3-monooxygenase/tryptophan 5-monooxygenase<br>activation protein zeta [Homo sapiens] | 42,4 |
| <b>119591511</b> | collagen, type VI, alpha 3, isoform CRA_c [Homo sapiens]                                      | 40,2 |
| <b>8101724</b>   | AF258350_1 canstatin, partial [Homo sapiens]                                                  | 40,0 |
| <b>133778299</b> | TUBB2B protein [Homo sapiens]                                                                 | 38,9 |
| <b>158256710</b> | unnamed protein product [Homo sapiens]                                                        | 37,9 |
| <b>89243632</b>  | ITGB1 protein [Homo sapiens]                                                                  | 35,7 |
| <b>530400269</b> | PREDICTED: myosin light polypeptide 6 isoform X6 [Homo<br>sapiens]                            | 34,4 |
| <b>87196339</b>  | collagen alpha-1(VI) chain precursor [Homo sapiens]                                           | 32,3 |
| <b>40226101</b>  | ACTG1 protein, partial [Homo sapiens]                                                         | 30,6 |
| <b>386997</b>    | prebeta-migrating plasminogen activator inhibitor, partial [Homo<br>sapiens]                  | 30,2 |
| <b>193787599</b> | unnamed protein product [Homo sapiens]                                                        | 28,7 |
| <b>16741721</b>  | Procollagen-lysine 1, 2-oxoglutarate 5-dioxygenase 1 [Homo<br>sapiens]                        | 27,5 |
| <b>119625664</b> | annexin A5, isoform CRA_c [Homo sapiens]                                                      | 24,3 |
| <b>340234</b>    | vimentin, partial [Homo sapiens]                                                              | 22,8 |
| <b>11935049</b>  | AF304164_1 keratin 1 [Homo sapiens]                                                           | 21,2 |
| <b>221042224</b> | unnamed protein product [Homo sapiens]                                                        | 21,2 |
| <b>34069</b>     | unnamed protein product [Homo sapiens]                                                        | 20,4 |
| <b>12667788</b>  | myosin-9 [Homo sapiens]                                                                       | 19,0 |

|                  |                                                                                  |      |
|------------------|----------------------------------------------------------------------------------|------|
| <b>34782802</b>  | PKM2 protein, partial [Homo sapiens]                                             | 18,8 |
| <b>194376948</b> | unnamed protein product [Homo sapiens]                                           | 18,3 |
| <b>148922238</b> | Thrombospondin 2 [Homo sapiens]                                                  | 17,7 |
| <b>119615401</b> | heparan sulfate proteoglycan 2 (perlecan), isoform CRA_b<br>[Homo sapiens]       | 16,0 |
| <b>282721077</b> | sushi repeat-containing protein SRPX isoform 3 precursor<br>[Homo sapiens]       | 15,5 |
| <b>119615036</b> | collagen, type I, alpha 1, isoform CRA_a [Homo sapiens]                          | 15,4 |
| <b>530425079</b> | PREDICTED: tubulin beta-4A chain isoform X2 [Homo sapiens]                       | 14,8 |
| <b>115527062</b> | collagen alpha-2(VI) chain isoform 2C2 precursor [Homo<br>sapiens]               | 12,9 |
| <b>84708830</b>  | TUBB6 protein [Homo sapiens]                                                     | 12,0 |
| <b>119616320</b> | hyaluronan and proteoglycan link protein 1, isoform CRA_b<br>[Homo sapiens]      | 11,8 |
| <b>197692249</b> | annexin I [Homo sapiens]                                                         | 10,3 |
| <b>49257389</b>  | CCDC80 protein [Homo sapiens]                                                    | 10,3 |
| <b>194384606</b> | unnamed protein product [Homo sapiens]                                           | 10,0 |
| <b>38197240</b>  | LAMB1 protein, partial [Homo sapiens]                                            | 8,4  |
| <b>157653329</b> | procollagen C-endopeptidase enhancer 1 precursor [Homo<br>sapiens]               | 8,3  |
| <b>3777617</b>   | serine protease [Homo sapiens]                                                   | 8,2  |
| <b>119616322</b> | EGF-like repeats and discoidin I-like domains 3, isoform CRA_a<br>[Homo sapiens] | 7,6  |
| <b>12653033</b>  | MYH10 protein [Homo sapiens]                                                     | 7,3  |
| <b>62089314</b>  | Pro-alpha-1 type V collagen variant [Homo sapiens]                               | 7,3  |
| <b>189054446</b> | unnamed protein product [Homo sapiens]                                           | 6,3  |

|                  |                                                                                   |     |
|------------------|-----------------------------------------------------------------------------------|-----|
| <b>194097352</b> | alpha-actinin-1 isoform c [Homo sapiens]                                          | 5,8 |
| <b>12653633</b>  | Lysyl oxidase-like 2 [Homo sapiens]                                               | 5,8 |
| <b>119582898</b> | MAM domain containing 2, isoform CRA_b [Homo sapiens]                             | 5,6 |
| <b>119590446</b> | nidogen 1, isoform CRA_b [Homo sapiens]                                           | 4,6 |
| <b>189054446</b> | unnamed protein product [Homo sapiens]                                            | 3,8 |
| <b>119607847</b> | pregnancy-associated plasma protein A, pappalysin 1, isoform CRA_c [Homo sapiens] | 3,6 |
| <b>119568019</b> | fibronectin type III domain containing 1 [Homo sapiens]                           | 3,5 |
| <b>30353925</b>  | CLTC protein [Homo sapiens]                                                       | 3,2 |
| <b>9309503</b>   | AC013451_1 LTBP-2 [Homo sapiens]                                                  | 2,9 |
| <b>553348</b>    | hexabrachion, partial [Homo sapiens]                                              | 2,8 |
| <b>119593150</b> | filamin A, alpha (actin binding protein 280), isoform CRA_a [Homo sapiens]        | 2,4 |
| <b>118572606</b> | hemicentin-1 precursor [Homo sapiens]                                             | 1,0 |

**Table S1: Proteomic Analysis.**

**Supplementary Table 2**

|            |               | Ctrl   |       | ExoCPC |       | ExoCPC_SiPOSTN |       |
|------------|---------------|--------|-------|--------|-------|----------------|-------|
|            |               | Mean   | SEM   | Mean   | SEM   | Mean           | SEM   |
| <b>CO</b>  | <b>mL/min</b> | 75,80  | 9,21  | 112,61 | 19,59 | 90,7           | 10,89 |
| <b>EF</b>  | <b>%</b>      | 44,69  | 3,92  | 62,50  | 4,95  | 55,41          | 1,842 |
| <b>FAC</b> | <b>%</b>      | 35,20  | 7,08  | 54,93  | 3,87  | 47,85          | 4,69  |
| <b>FS</b>  | <b>%</b>      | 10,91  | 1,97  | 19,13  | 2,06  | 16,39          | 2,44  |
| <b>SV</b>  | <b>µL</b>     | 242,41 | 22,35 | 310,81 | 49,52 | 258,48         | 27,50 |
| <b>Vd</b>  | <b>µL</b>     | 560,32 | 62,15 | 484,93 | 40,87 | 462,79         | 38,66 |
| <b>Vs</b>  | <b>µL</b>     | 317,91 | 58,30 | 174,12 | 16,00 | 207,31         | 12,84 |

**Table S2: Echocardiographic Data at Day 14 post-MI**

CO: Cardiac Output; EF: Ejection Fraction; FAC: Fractional Area Change; FS: Fractional Shortening;  
SV: Stroke Volume; Vd: Diastolic Volume; Vs: Systolic Volume.

**Supplementary Table 3**

| <b>Antibody</b>      | <b>Application</b> | <b>Dilution</b> | <b>Company</b>             | <b># Ref</b> |
|----------------------|--------------------|-----------------|----------------------------|--------------|
| <b>TSG101</b>        | WB                 | 1:1000          | Abcam                      | ab125011     |
| <b>SYNTENIN-1</b>    | WB                 | 1:1000          | Abcam                      | ab133267     |
| <b>CALNEXIN</b>      | WB                 | 1:300           | SantaCruz<br>Biotechnology | sc-70481     |
| <b>POSTN</b>         | WB                 | 1:300           | SantaCruz<br>Biotechnology | sc-398631    |
| <b>cTnl</b>          | IF                 | 1:200           | Abcam                      | ab47003      |
| <b>αSarcomeric</b>   | IF                 | 1:300           | Abcam                      | ab9465       |
| <b>pH3</b>           | IF                 | 1:300           | Abcam                      | ab14955      |
| <b>pAKT</b>          | WB                 | 1:1000          | Cell Signaling             | 4060         |
| <b>AKT</b>           | WB                 | 1:1000          | Cell Signaling             | 9272         |
| <b>GAPDH</b>         | WB                 | 1:5000          | Abcam                      | ab181602     |
| <b>CYCLIN D1</b>     | WB                 | 1:30000         | Abcam                      | ab134175     |
| <b>pFAK</b>          | WB                 | 1:1000          | Abcam                      | ab81298      |
| <b>FAK</b>           | WB                 | 1:1000          | Abcam                      | ab76496      |
| <b>YAP1</b>          | WB                 | 1:500           | Invitrogen                 | PA5-87568    |
| <b>H3</b>            | WB                 | 1:5000          | Abcam                      | ab176842     |
| <b>AuroraBKinase</b> | IF                 | 1:250           | BD                         | 611083       |
| <b>cTnT</b>          | IF                 | 1:100           | Miltenyi Biotec            | 130-119-674  |

|               |    |       |                            |           |
|---------------|----|-------|----------------------------|-----------|
| <b>CD63</b>   | IF | 1:100 | SantaCruz<br>Biotechnology | sc-365604 |
| <b>PAPP-A</b> | WB | 1:300 | HyTest Ltd                 | 4P41      |

**Table S3: Detailed information on antibodies used in the study.**