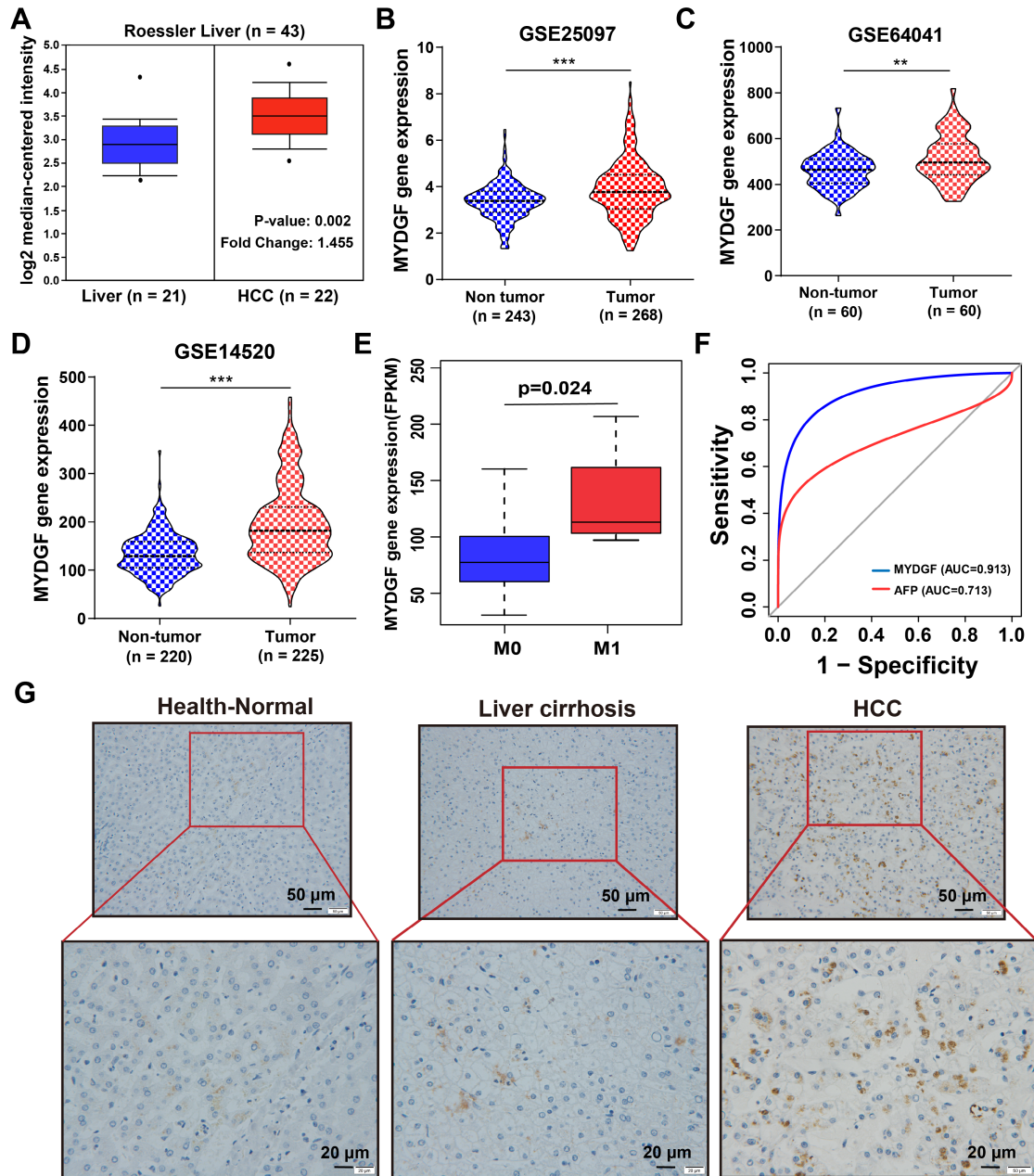


# 1 Supplementary Figures



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3 **Figure S1. MYDGF is upregulated in HCC.** (A) MYDGF gene expression in  
4 Roessler liver dataset from Oncomine database, containing 43 samples. (B)  
5 GSE25097 dataset from GEO database, which contains 243 non-tumor samples and  
6 268 HCC tumor samples. (C) GSE64041 dataset from GEO database which includes  
7 60 paired non-tumor and HCC tumor samples. (D) GSE14520 dataset from GEO  
8 database, it includes 220 non-tumor samples and 225 HCC tumor samples. (E) Gene

9 expression of MYDGF in no-distant metastasis (M0) and distant metastasis (M1)  
10 samples. (F) The ROC curve compared the ability of MYDGF to distinguish normal  
11 tissues from HCC samples at the genetic level. (G) Immunohistochemistry results  
12 showed that MYDGF expression is negative in normal liver tissue and cirrhosis  
13 samples, but specifically positive stain can be found in Stage I hepatocellular  
14 carcinoma samples.

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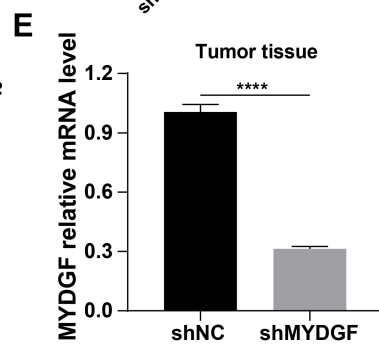
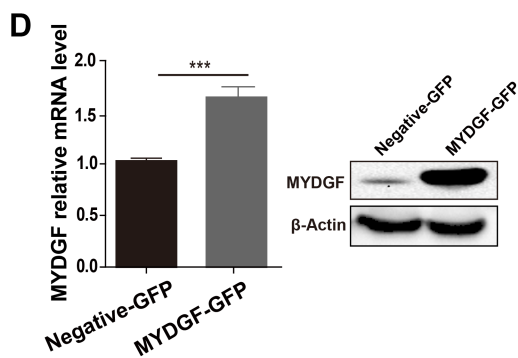
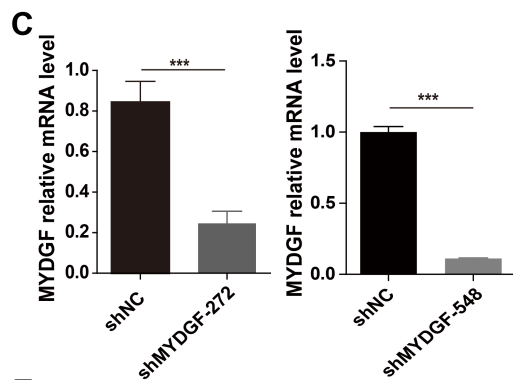
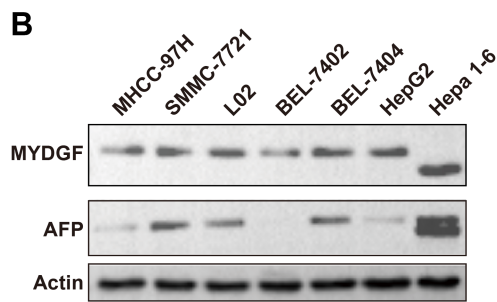
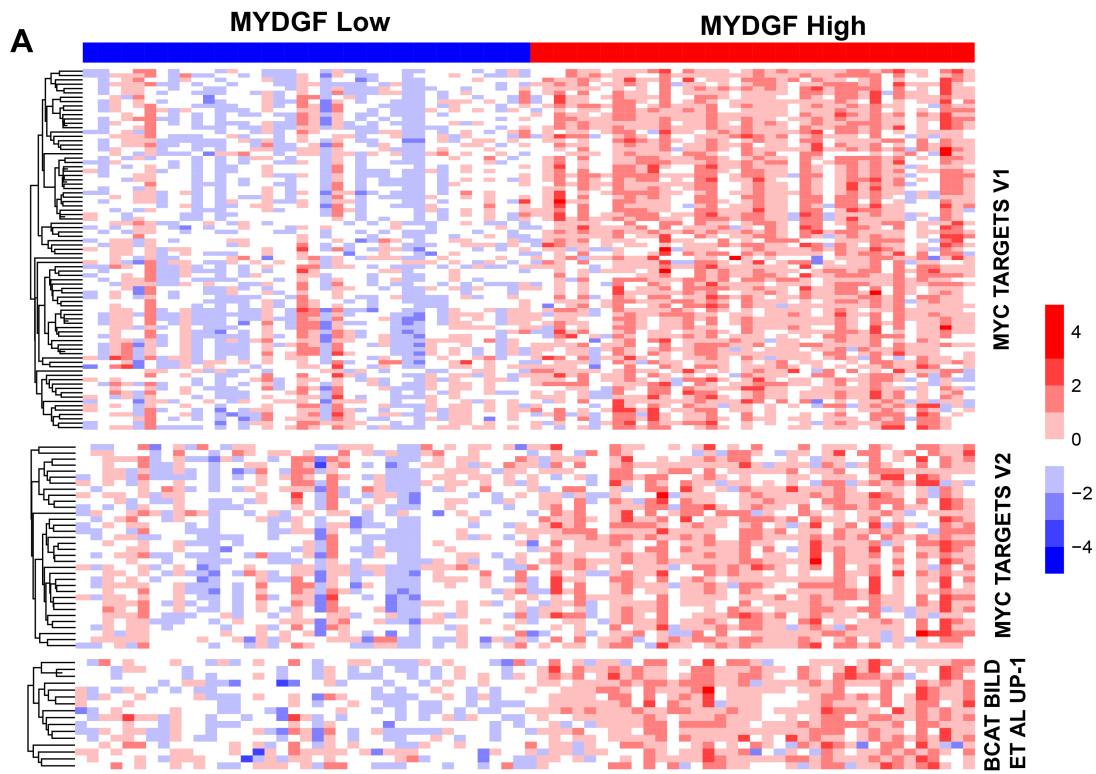
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32 **Figure S2. MYDGF promotes cell proliferation *in vitro*.** (A) MYDGF expression in  
 33 high and low groups of enriched signatures, include  
 34 “HALLMARK\_MYC\_TARGET\_V1”, “HALLMARK\_MYC\_TARGET\_V2” and  
 35 “BCAT\_BILD\_ET\_AL\_UP-1”. (B) MYDGF and AFP protein expression in 6 HCC

36 cancer cell lines and a normal liver cell line LO2. (C) Two sequences with the highest  
37 knockdown efficiency were selected for packaging lentiviruses for the construction of  
38 stable cell lines. The knockdown efficiency of MYDGF in Hepa1-6 cells is shown. (D)  
39 BEL-7404 was used to construct MYDGF overexpressing cell lines, and the  
40 efficiency of MYDGF overexpression was tested. (E) The expression level of  
41 MYDGF in the xenograft tumor tissue at the end of the experiment.

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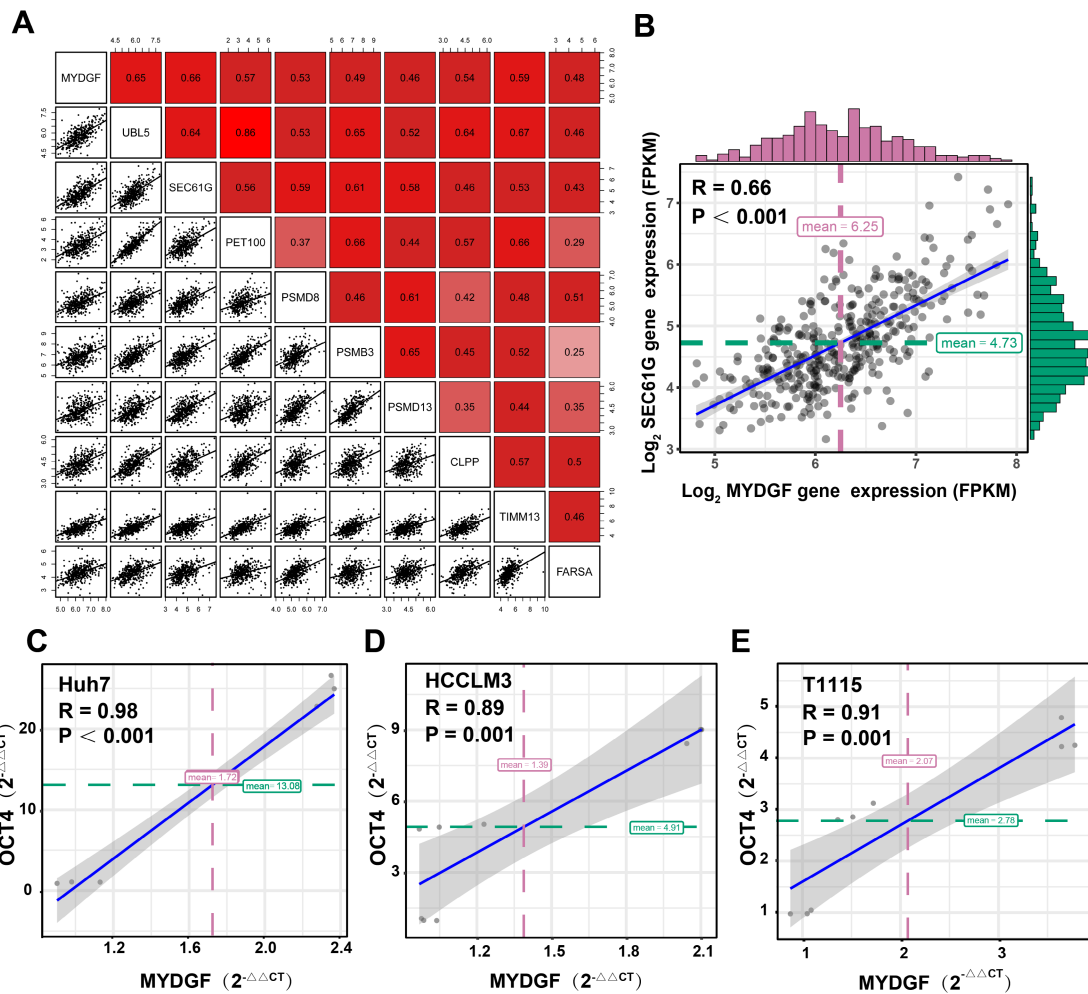
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59 **Figure S3. MYDGF expression is associated with stem cell related signatures and**

60 **genes.** (A) Correlation between MYDGF and top three leading-edge genes from each

61 stem cell related signatures in TCGA database. (B) The Most relevant gene was

62 shown in the scatter plot. Correlation value was calculated using person method. (C-E)

63 Correlation between MYDGF and stem cell marker OCT4 in three human HCC cell

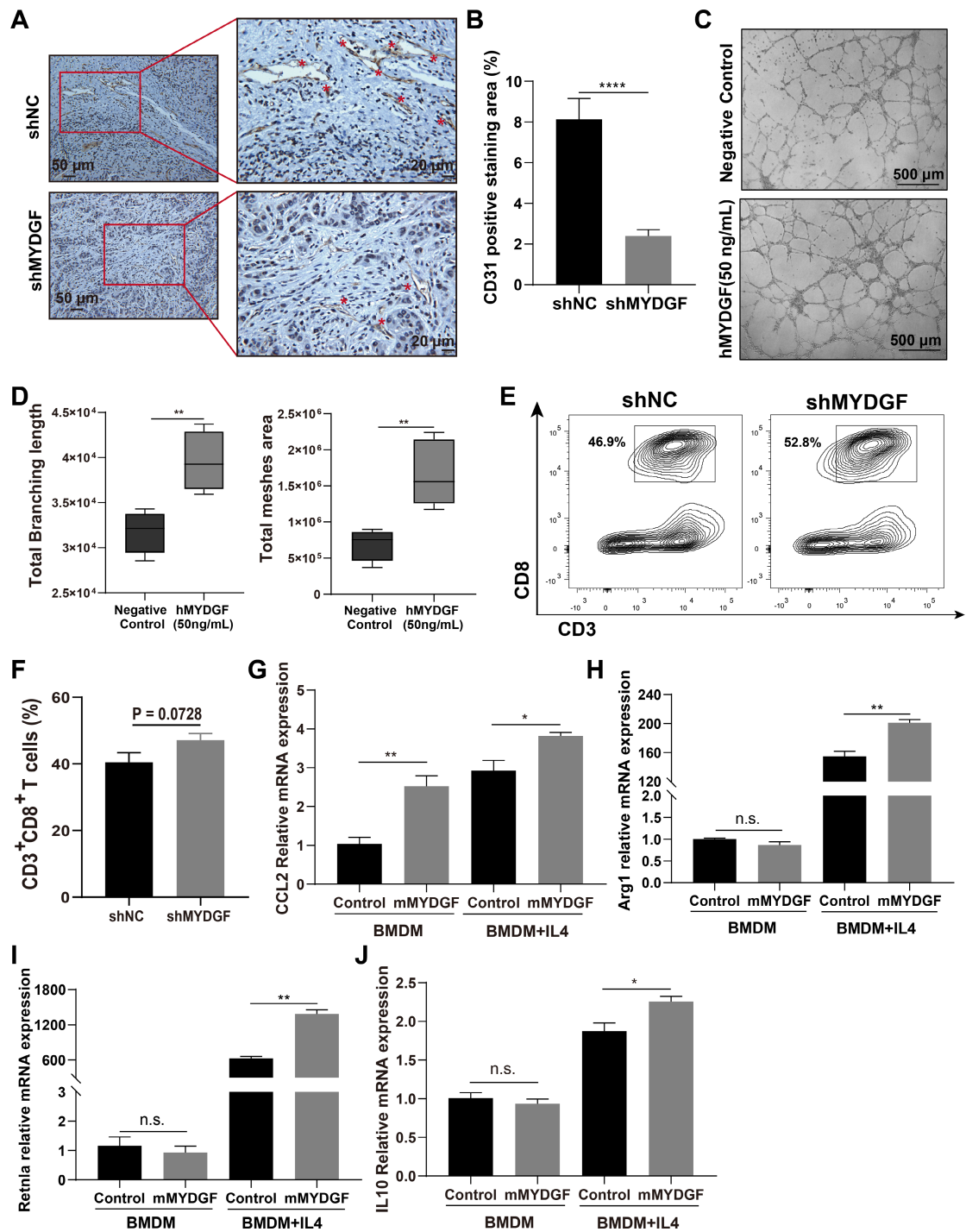
64 lines (Huh7, HCCLM3, T1115).

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70 **Figure S4. MYDGF indirectly promoted HCC development by remodeling TME.**

71 (A) Expression of CD31 in subcutaneous tumors of C57BL/6 mice was determined

72 using immunohistochemical staining, the scale bars are shown in the figures. (B)

73 CD31 positive staining area fraction was quantified by Image J software. (C) Tube

74 formation assay was performed using HUVEC cells cultured with 50 ng/mL human

75 MYDGF recombinant protein for 8 hours. Cells were examined using a Leica  
76 microscope. (D) Total tube length and total meshes area was calculated by Image J  
77 software. (E) The proportion of CD3<sup>+</sup>CD8<sup>+</sup> T cells infiltration in tumor tissue was  
78 detected by flow cytometry. (F) Quantitative statistics of CD3<sup>+</sup>CD8<sup>+</sup> T cells  
79 infiltration fraction. (G) CCL2 mRNA expression in BMDM and M2 type  
80 macrophages cultured with 100 ng/mL MYDGF recombinant protein. (H-J) Arg1,  
81 Retnla and IL-10 mRNA level in BMDM and M2 type macrophages cultured with  
82 100 ng/mL MYDGF recombinant protein.

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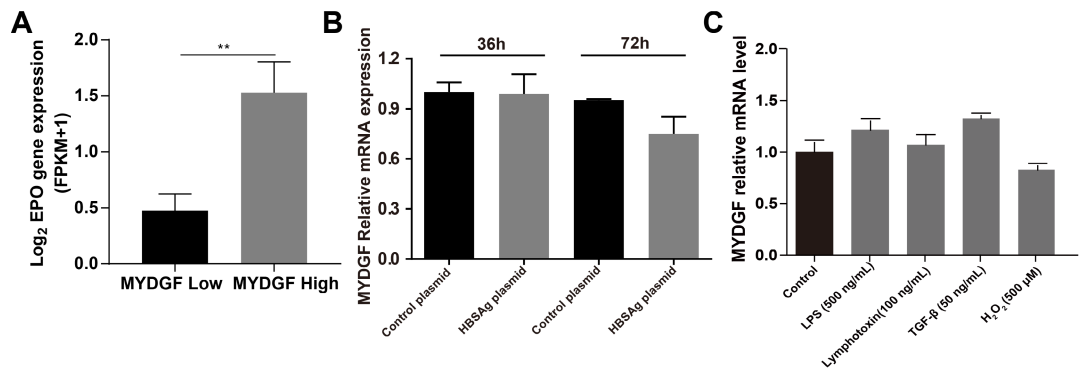
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**Figure S5. Hypoxia induces MYDGF expression in HCC.** (A) EPO gene expression in MYDGF<sup>high</sup> and MYDGF<sup>Low</sup> group from the "CELLULAR RESPONSE TO HYPOXIA" signature. (B) HepG2 cells were transfected with HBS-Ag plasmid and the mRNA expression of MYDGF at 36 hours and 72 hours was detected. (C) HepG2 cells were cultured with LPS, lymphotoxin, TGF-β and H<sub>2</sub>O<sub>2</sub> to simulate the factors that accelerate the progression of HCC in the microenvironment.



116 **Table 1. MYDGF siRNA sequences**

MYDGF(Mus)	Sequences	
	Sense (5'-3')	Antisense (5'-3')
siMYDGF-177	CCCGGGAACAAGUUUACAUTT	AUGUAAACUUGUUCCCGGGTT
siMYDGF-272	GCACUUUACCUGUACCAUCTT	GAUGGUACAGGUAAAGUGCTT
siMYDGF-351	GCUGAGAUCGAGUAUGCCATT	UGGCAUACUCGAUCUCAGCTT
siMYDGF-226	CCAACGAGCAAUGGCAGAUTT	AUCUGCCAUUGCUCGUUGGTT
siMYDGF-468	GCCUUCAAGCUGAGCUCUTT	AGAGCUCAGCUUUGAAGGCTT
siMYDGF-548	CCUUCAUGUCCACGUUCCUTT	AGGAACGUGGACAUGAAGGTT

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130 **Table 2. The primer sequences used for qPCR**

<b>Genes</b>	<b>Sequence (5' to 3')</b>
hMYDGF-F	CTGAAGTGAGTCCGGGAGC
hMYDGF-R	TGTTTGGGTTGGAGTTTGC
mMYDGF-F	ACATGTACATTCACCTACGCTT
mMYDGF-R	TGGTACAGGTAAAGTGCTGGC
mTNF-F	CCCTCACACTCAGATCATCTTCT
mTNF-R	GCTACGACGTGGGCTACAG
mIL-6-F	TAGTCCTTCCTACCCCAATTTCC
mIL-6-R	TTGGTCCTTAGCCACTCCTTC
mCCL2-F	TTAAAAACCTGGATCGGAACCAA
mCCL2-R	GCATTAGCTTCAGATTTACGGGT
mIL-10-F	GCTCTTACTGACTGGCATGAG
mIL-10-R	CGCAGCTCTAGGAGCATGTG
mArg1-F	CTCCAAGCCAAAGTCCTTAGAG
mArg1-R	AGGAGCTGTCATTAGGGACATC
mRetnla-F	CCAATCCAGCTAACTATCCCTCC
mRetnla-R	ACCCAGTAGCAGTCATCCCA
hGAPDH-F	CCTTCATTGACCTCAACTAC
hGAPDH-R	GAAGATGGTGATGGGATTTTC
mGAPDH-F	AGTGGCAAAGTGGAGATT
mGAPDH-R	GTGGAGTCATACTGGAACA