

**Research Paper** 

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# Differentiation of benign and malignant lymph nodes in pediatric patients on ferumoxytol-enhanced PET/MRI

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#### Abstract

The composition of lymph nodes in pediatric patients is different from that in adults. Most notably, normal lymph nodes in children contain less macrophages. Therefore, previously described biodistributions of iron oxide nanoparticles in benign and malignant lymph nodes of adult patients may not apply to children. The purpose of our study was to evaluate if the iron supplement ferumoxytol improves the differentiation of benign and malignant lymph nodes on <sup>18</sup>F-FDG PET/MRI.

**Methods:** We conducted a prospective clinical trial from May 2015 to December 2018 to investigate the value of ferumoxytol nanoparticles for staging of children with cancer with <sup>18</sup>F-FDG PET/MRI. Ferumoxytol is an FDA-approved iron supplement for the treatment of anemia and has been used "off-label" as an MRI contrast agent in this study. Forty-two children (7-18 years, 29 male, 13 female) received a <sup>18</sup>F-FDG PET/MRI at 2 (n=20) or 24 hours (h) (n=22) after intravenous injection of ferumoxytol (dose 5 mg Fe/kg). The morphology of benign and malignant lymph nodes on ferumoxytol-enhanced T2-FSE sequences at 2 and 24 h were compared using a linear regression analysis. In addition, ADCmean-values, SUV-ratio (SUV<sub>max</sub> lesion/SUV<sub>mean</sub> liver) and R2\*-relaxation rate of benign and malignant lymph nodes were compared with a Mann-Whitney-U test. The accuracy of different criteria was assessed with a receiver operating characteristics (ROC) curve. Follow-up imaging for at least 6 months served as the standard of reference.

**Results:** We examined a total of 613 lymph nodes, of which 464 (75.7%) were benign and 149 (24.3%) were malignant. On ferumoxytol-enhanced T2-FSE images, benign lymph nodes showed a hypointense hilum and hyperintense parenchyma, while malignant lymph nodes showed no discernible hilum. This pattern was not significantly different at 2 h and 24 h postcontrast (p=0.82). Benign and malignant lymph nodes showed significantly different ferumoxytol enhancement patterns, ADCmean values of 1578 and 852 ×10-6 mm<sup>2</sup>/s, mean SUV-ratios of 0.5 and 2.8, and mean R2\*-relaxation rate of 127.8 and 84.4 Hertz (Hz), respectively (all p<0.001). The accuracy of ADCmean, SUV-ratio and pattern (area under the curve (AUC): 0.99; 0.98; 0.97, respectively) was not significantly different (p=0.07). Compared to these three parameters, the accuracy of R2\* was significantly lower (AUC: 0.93; p=0.001).

**Conclusion:** Lymph nodes in children show different ferumoxytol-enhancement patterns on MRI than previously reported for adult patients. We found high accuracy (>90%) of ADCmean, SUV-ratio, pattern, and R2\* measurements for the characterization of benign and malignant lymph nodes in children. Ferumoxytol nanoparticle accumulation at the hilum can be used to diagnose a benign lymph node. In the future, the delivery of clinically applicable nanoparticles to the hilum of benign lymph nodes could be harnessed to deliver theranostic drugs for immune cell priming.

Key words: nanoparticles, PET/MRI, lymph nodes, cancer imaging, ferumoxytol

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### Introduction

Accurate detection of lymph node metastases is important for cancer staging of pediatric patients [1-3]. In the clinical realm, the morphology of lymph nodes is commonly used to estimate malignancy: Lymph nodes with a size of more than 1 cm and round shape are considered malignant on ultrasound [4], computed tomography (CT) [5, 6] or magnetic resonance imaging (MRI) [7]. Unsurprisingly, these morphological criteria demonstrate low specificity because malignant tumor cell infiltrations can be found in subcentimeter nodes and infections can cause enlarged benign nodes [6]. More recently, an increased glucose metabolism, as measured by an increased uptake of <sup>18</sup>F-fluorodeoxyglocose (<sup>18</sup>F-FDG) on a positron emission tomography (PET) scan, provided improved specificity for the detection of malignant lymph nodes in children with lymphoma [8], soft tissue sarcoma [9] and bone sarcoma [10], among others [11]. However, <sup>18</sup>F-FDG PET studies can still be difficult to interpret in children, because they often have inflammatory nodes [12]: Previous investigators reported an incidence of 27% false positive benign lymph nodes on <sup>18</sup>F-FDG PET studies in pediatric oncology patients [5, 13, 14] compared to 15% in adult patients [15]. Therefore, additional approaches to diagnose malignant lymph nodes more reliably would be highly valuable [16].

Iron oxide nanoparticles can improve the specificity of MR imaging for the differentiation of benign and malignant lymph nodes [17, 18]. In adult patients, benign lymph nodes showed significant T2- and T2\*-enhancement of both hilum and parenchyma at 24 hours (h) after intravenous infusion of iron oxide nanoparticles, while malignant lymph nodes showed no significant enhancement [17-21]. This difference was due to presumed nanoparticle phagocytosis by macrophages in benign lymph nodes and absent phagocytosis in metastatic lymph nodes, in which normal stroma cells were replaced by tumor cells [17].

However, this concept might not apply to pediatric patients due to well-described differences in the cellular composition of benign lymph nodes in children and adults [22, 23]. In children, lymph nodes are better vascularized than in adults [24], the hilum contains more vessels and minimal fat [25] and the parenchyma is mostly composed of T- and B-cells [26, 27]. Importantly, lymph nodes in children contain less macrophages [28]. Therefore, the iron oxide nanoparticle enhancement patterns of benign lymph nodes in children could be different from those described for adult patients in the literature [17, 18]. We hypothesized that the iron oxide nanoparticle enhancement of lymph nodes in children would be a function of vascularity rather than macrophage content. We investigated the iron oxide nanoparticle compound ferumoxytol, which is an FDA-approved iron supplement for the treatment of anemia, which can be used "off-label" as an MRI contrast agent. To knowledge, ferumoxytol our the value of nanoparticles for the differentiation of benign and malignant lymph nodes in pediatric patients has not been investigated. The purpose of our study was to evaluate if ferumoxytol administration improves the characterization of malignant lymph nodes in pediatric patients on <sup>18</sup>F-FDG PET/MRI.

### Methods

### **Patient population**

This prospective, non-randomized, health insurance portability and accountability act-compliant clinical trial (NCT01542879) was approved by our institutional review board and performed under an investigator-initiated investigative new drug application (111,154). We included pediatric cancer patients from May 2015 until December 2018 if they met the following inclusion criteria: 1) age 6 to 18 vears, 2) solid extracranial tumor, diagnosed based on standard imaging tests or biopsy, 3) willingness to give written informed consent. Patients were excluded if they had: 1) a primary lymphatic disease (i.e. lymphoma), 2) contraindications to MRI, 3) hemosiderosis or hemochromatosis, or 4) if they were pregnant. All patients or their legal representative gave written informed consent. We recruited 42 patients with biopsy proven sarcoma (osteosarcoma (n=11), Ewing sarcoma (n=4), soft tissue sarcoma including rhabdomyosarcoma (n=3), synovial sarcoma (n=3), undifferentiated sarcoma (n=2), gastrointestinal stromal tumor (n=1), desmoplastic small round cell tumor (n=3), malignant peripheral nerve sheath tumor (n=1)) as well as Langerhans cell histiocytosis (n=6), Wilms tumor (n=2), nasopharynxgeal carcinoma, ovarian steroid cell tumor, hepatocellular carcinoma, ovarian mixed germ cell tumor, neuroblastoma and lipoma (n=1, respectively). The patients included 13 girls (mean age 13.4 +/- 3.4 years; range: 7-18 years) and 29 boys (mean age 13.6 +/- 3.1 years; range: 8-18 years, Table S1).

### **PET/MR-imaging**

All patients underwent an integrated <sup>18</sup>F-FDG PET/MRI scan (Signa PET/MR, GE Healthcare, Milwaukee, WI) at either 2 h (n=20) or 24 h (n=22) after an intravenous infusion of the iron supplement ferumoxytol (Feraheme® Injection, AMAG Pharmaceuticals, Waltham, MA). We randomly assigned patients to scans at two time points, 2 h and 24 h postcontrast, because previous studies had used

delayed 24 hour scans for lymph node imaging [17, 19, 29]. As a sub-aim of our investigations, we wanted to evaluate if the same information could be obtained from an earlier postcontrast scan, which would be more practical in a clinical-translational setting.

Ferumoxytol was diluted 1:3 in saline and administered at a dose of 5 mg Fe/kg bodyweight over 15 minutes (min) as recommended by the food and drug administration (FDA) [3]. We carefully monitored the heart rate and blood pressure of the patients before, during and up to 30 min after the ferumoxytol infusion and after the patient finished the PET/MRI scan. We also asked the patients for any subjective side effects. At approximately 60 min before the planned PET/MRI scan (mean: 57.5 +/-10.1 min, range: 42-86 min), all patients received an intravenous injection of <sup>18</sup>F-FDG at a dose of 3 MBq/kg bodyweight (151.7 +/- 51.8 MBq, range: 62.9-270.1 MBq). After fasting for at least 4 h before the scan, the mean blood glucose level of all patients was 90.9 +/- 9.7 mg/dL (range: 71-109 mg/dL) at the time of the <sup>18</sup>F-FDG injection. The effective dose from the <sup>18</sup>F-FDG injection was calculated using the age-specific conversion factors published by the international commission on radiological protection [30].

Our imaging protocol included axial PET acquisition slabs (25 cm axial FOV, acquisition time of 3:30 min per slab) with simultaneous acquisition of the following axial MR sequences: T1-weighted Liver Acquisition with Volume Acquisition (LAVA), T2-weighted Fast Spin Echo (FSE) with fat saturation and diffusion-weighted imaging (DWI; b-value: 50/600). In addition, axial free-breathing periodically rotated overlapping parallel lines with enhanced reconstruction (PROPELLER) images of the chest and axial iterative decomposition of water and fat with echo asymmetry and least-squares estimation (IDEAL-IQ) images were obtained to create R2\* maps. Detailed sequence parameters for PET/MRI scans are given in Table S2.

We reconstructed PET data using a 3D time of flight iterative ordered subsets expectation maximization algorithm (24 subsets, 3 iterations, temporal resolution 400 ps, matrix 192 × 192; voxel size 2.8×2.8×2.8 mm). A two-point Dixon MR sequence was used for attenuation correction.

# Morphology of benign and malignant lymph nodes

To evaluate morphological characteristics of benign and malignant lymph nodes, two radiologists (AJT, FS) measured the size (short and long axis) of all visually detectable lymph nodes on T2-weighted FSE images in five anatomical regions (neck, axilla, mediastinum, abdomen, and groin) of each patient. Response evaluation criteria for solid tumors (RECIST) guidelines were used to categorize benign and malignant lymph nodes based on their length of the short axis of <10 mm or  $\geq$ 10 mm, respectively [31, 32]. In order to determine if ferumoxytol enhancement can add diagnostic information, two radiologists (HD, AJT; >20 years and 7 years of experience) jointly reviewed and determined qualitative enhancement patterns for all lymph nodes. Twenty-seven of our patients had received a clinical scan with unenhanced axial T2-weighted FSE sequences (same sequence parameter as PET/ MRI-protocol) within three weeks (range: 1-21 days) before the PET/MRI scan. These unenhanced scans served as controls to determine the enhancement signal of the lymph nodes after ferumoxytol administration in children.

# Quantitative evaluation of benign and malignant lymph nodes

Two operators (FS and AJT) outlined each lymph node with operator defined regions of interest (ROIs) and measured the following parameters: ADCmean, R2\* and SUV-ratio:

We reconstructed apparent diffusion coefficient (ADC) maps from DWI images (b=50/600) using the integrated OsiriX (OsiriX 8.2, Pixmeo, Geneva, Switzerland) software tool and measured the mean ADC value of each lymph node through an operator-defined region of interest (ROI), which covered the entire lymph node (i.e. the hilum and the parenchyma):

$$ADC_{mean} = -\frac{1}{b} \ln \left( \frac{Sdwi}{S50} \right)$$

We generated R2\* maps from multi-echo IDEAL-IQ sequences with the PET/MRI scanner software DV26. We measured the R2\* relaxation rate of each lymph node using OsiriX software through operator-defined ROIs. To quantify the uptake of ferumoxytol nanoparticles in lymph nodes, we calculated R2\* relaxation rates as R2\*=1/T2\*. R2\* relaxation rates are directly proportional to the tissue's iron content [33].

We measured the <sup>18</sup>F-FDG radiotracer uptake in lymph nodes and liver on the attenuation corrected PET data and calculated the maximum standardized uptake value (SUV<sub>max</sub>) of each lymph node with MIM software (MIM 6.5, Cleveland, OH).

$$SUV_{max} = \frac{tissue \ tracer \ activity \ (mCi/g)}{(injected \ dose(mCi)/patient \ body \ weight \ (kg))}$$

We normalized the  $SUV_{max}$  of each lymph node by the  $SUV_{mean}$  of the patient's liver as an internal standard of reference by calculating the SUV tumor-to-liver ratio. This allowed us to compare the <sup>18</sup>F-FDG uptake in lymph nodes of different patients, who might have received their PET scans at slightly different time points after the <sup>18</sup>F-FDG injection. A SUV-ratio above one indicates an increased tissue <sup>18</sup>F-FDG uptake in lymph nodes compared to liver as an internal reference standard.

#### Standard of reference

Follow up imaging for at least six months served as the standard of reference. A lymph node with benign imaging features that remained stable at 6 months was defined as benign. A lymph node with malignant features that either progressed or responded after chemotherapy was defined as malignant.

We obtained exemplary proof-of-concept histopathologies of a benign and a malignant lymph node in two of our patients, who had undergone a tumor resection along with a resection of lymph nodes one day after the ferumoxytol PET/MRI. The lymph node specimens were embedded in paraffin, sliced in 4-mm sections that were placed on glass slides. These slides were baked for 1 h at 60°C and afterwards deparaffinized in xylene and hydrated in a graded series of alcohol. Hydrogen peroxide was used to block endogenous peroxidase. Samples were stained on the Benchmark XT stainer (Roche Ventana, Oro Valley, AZ) with standard heat-induced epitope retrieval except as indicated, and diaminobenzidine (Enzo Biochem Inc., Farmingdale, NY) as the chromogen. CD68 (mouse monoclonal KP1 1:1600, Dako, Santa Clara, CA) was the antibody used for macrophage staining. Hematoxylin (VWR, Radnor, PA) was used for counterstaining. To detect intracytoplasmatic iron in macrophages, paraffin-embedded lymph node slides were stained with Prussian blue (Dako, Santa Clara, CA). Light microscopic images were captured on an Olympus BX45 microscope (Olympus, Center Valley, PA) with UPlanFL 4 /0.13, 20/0.50, and 100/1.25 oil immersion lenses and a SpotFLEX camera.

### **Statistical Analysis**

Differences in morphology between benign and malignant lymph nodes at 2 h and 24 h post ferumoxytol injection were compared with a linear regression analysis using a general linear model. ADCmean, SUV-ratio and R2\* of benign and malignant lymph nodes were compared with a Mann-Whitney-U test. ADCmean, SUV-ratio and R2\* as predictors were evaluated with size and p-value, 95% confidence interval (CI), odds ratio (OR), area under the curve (AUC), and 95%CI of AUC were reported and compared using a general linear model with a random effect for each patient. Due to multiple comparisons (n=12), the corrected criterion for significance testing used was p=0.004 (0.05 divided by 12).

### Results

The ferumoxytol infusion was well tolerated by all patients and we did not encounter any adverse events. The mean radiation exposure of our patients as part of the <sup>18</sup>F-FDG PET/MRI study was 2.8 mSv (range 1.2 - 5.2 mSv).

### Size of benign and malignant lymph nodes on ferumoxytol-enhanced MR scans

We detected a total of 613 lymph nodes in 42 patients. Of those, 464 (75.7%) were benign and 149 (24.3%) were malignant (Table S3). The mean short axis diameter was 5.5 +/- 3.5 mm for benign lymph nodes and 10.6 +/- 3.9 mm for malignant lymph nodes. Size is usually a major parameter for radiologists to assess if a lymph node is malignant, with larger than 10 mm considered malignant [4-6] and smaller than 5 mm being benign [6, 34]. Yet, our data revealed size to be an insufficient predictor of malignant lymph nodes smaller than 10 mm with a sensitivity of only 47.7%. However, size was an excellent predictor of benign etiology for lymph nodes smaller than 5 mm with a sensitivity of 94.0%. Thus, the state of malignancy of lymph nodes cannot be predicted accurately within the range of 5-10 mm. Therefore, additional imaging criteria like iron oxide nanoparticle enhancement pattern would be most useful for lymph nodes in this range of sizes.

# Morphology of benign and malignant lymph nodes on ferumoxytol-enhanced MR scans

Unenhanced T2-FSE images showed а hyperintense signal of both benign and malignant lymph nodes: 289 (87%) of 333 benign lymph nodes did not show a fatty hilum and only 44 (13%) benign lymph nodes did show a fatty hilum. By contrast, T2-FSE images after ferumoxytol infusion delineated a hypointense hilum in 362 out of 464 benign lymph nodes (78.0%), whereas 144 out of 149 malignant lymph nodes were homogenously hyperintense (96.6 %) (Figure 1, Table 1). This difference in ferumoxytolenhancement of benign and malignant lymph nodes was significant (p<0.001). The ferumoxytol enhancement pattern did not change significantly between 2 h and 24 h scans, neither for benign lymph nodes (p=0.01, NOTE: due to multiple testing significance level p=0.004) nor malignant lymph nodes (p=0.95, Figure S1). When considering nodal size, we found absent hilar enhancement in 79/79 (100%) malignant lymph nodes > 10 mm and positive hilar enhancement in 10/11 (91%) benign lymph nodes > 10 mm

(Figure S2). For sizes  $\geq 5 - \langle 10 \text{ mm}, \text{ pattern added}$  significant value compared to size alone (p $\langle 0.001 \rangle$ ) with a ten-fold higher chance of malignancy for lymph nodes with absent hilar enhancement (OR size: 1.4, OR pattern: 10.9; Figure 2, Table 2). We did not recognize the hypointense hilum in 74 (39.8%) out of 186 benign lymph nodes  $\langle 5 \text{ mm} \rangle$  (Figure S2). Furthermore, presence or absence of hilar enhancement yielded no additional information for nodes  $\langle 5 \text{ mm} \rangle$  in size, because the likelihood of them being malignant was so low with a sensitivity of 94.0%.

### Quantitative evaluation of benign and malignant lymph nodes in pediatric patients

There was no significant difference in quantitative data at 2 h and 24 h post ferumoxytol for ADCmean, SUV-ratio or R2\* data (p=0.21, p=0.63, p=0.8, respectively; Table S4). Therefore, we included imaging studies from all time points in further analyses:

Based on the previous work from Ustabasioglu et al. an ADCmean value less than  $1030 \times 10^{-6} \text{ mm}^2/\text{s}$  is considered restricted diffusion [35]. Diffusion weighted images revealed restricted diffusion in 118 of 149 (79.2%) malignant lymph nodes and unrestricted diffusion in 395 of 464 (85.1%) benign lymph nodes. The ADCmean value was 1578 +/- 420  $\times 10^{-6} \text{ mm}^2/\text{s}$  for benign lymph nodes and 852 +/- 206  $\times 10^{-6} \text{ mm}^2/\text{s}$  for malignant lymph nodes. This difference was statistically significant (p<0.001, Figure 3D-F, Table 3).

For <sup>18</sup>F-FDG PET data, the mean SUV-ratio was 0.53 +/- 0.29 for benign and 2.77 +/- 1.44 for malignant lymph nodes, which was also significantly different (p<0.001, Figure 3G-I, Table 3).

 Table 1. Distribution of pattern in benign and malignant lymph nodes

	Hilum ↓ Parenchyma ↑		Hilum ↑ Parenchyma ↑	
Sizes	benign	malignant	benign	malignant
< 5 mm	112	0	74	9
$\geq 5$ - $< 10~\mathrm{mm}$	240	5	27	64
≥10 mm	10	0	1	71
All	362	5	102	144

Table 2. Predictive value of quantitative parameters

Parameter	Odds	95% Conf	95% Confidence interval		AUC
	ratio	lower	upper	-	
ADCmean	0.99	0.99	0.99	< 0.001	0.989 (0.984-0.995)
SUV-ratio	10.82	4.18	28.03	< 0.001	0.979 (0.967-0.991)
Pattern	10.70	3.94	29.11	< 0.001	0.974 (0.961-0.987)
R2*	1.31	1.22	1.40	<0.001	0.933 (0.906-0.961)

ADCmean: Mean apparent diffusion coefficient; SUV: standardized uptake value; SUV-ratio: SUVmax lesion/SUVmean liver; R2\*: R2\*-relaxation rate; AUC: area under the curve

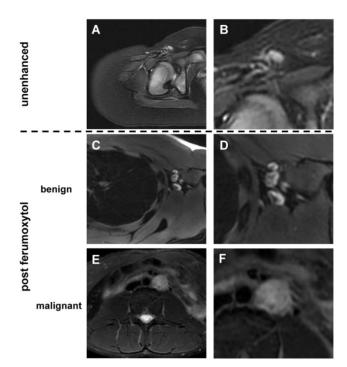


Figure 1. Morphology of benign and malignant lymph nodes on T2-weighted FSE images: (A) Normal lymph node in the right inguinal region demonstrates homogenous hyperintense signal before ferumoxytol administration. In young children, we found little or no fat at the lymph node hilum. (B) Enlarged view shows homogenous hyperintense lymph node with vessel entering the lymph node at the hilum. No significant fat is noted at the hilum. (C) After ferumoxytol administration, normal lymph node demonstrates dark (hypointense) enhancement of the hilum, but not the parenchyma. (D) Enlarged view demonstrates hypointense (dark) hilum and hyperintense (bright) parenchyma. (E) After ferumoxytol administration, malignant abdominal lymph node in the left para-aortic region demonstrates slightly inhomogeneous hyperintense T2-signal. (F) Enlarged view shows no evidence for hilar nanoparticle enhancement. FSE: fast spin echo.

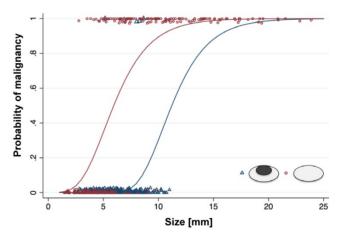


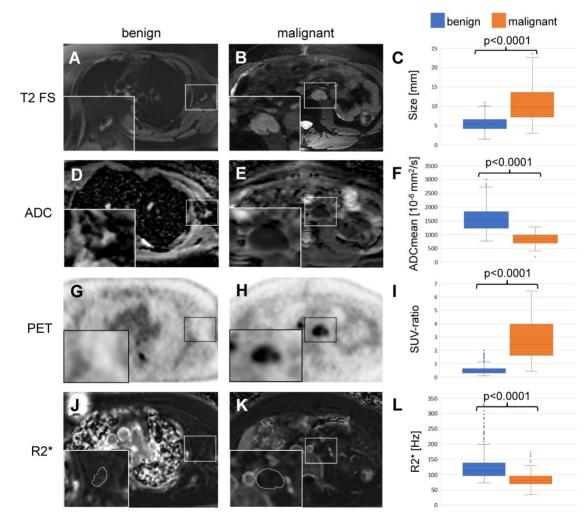
Figure 2. Probability of malignancy of lymph nodes with different size and morphology on ferumoxytol-enhanced T2-FSE images: For lymph nodes with a size of more than 10 mm, the probability of malignancy is high, and morphology on MRI adds little additional information. For lymph nodes with a size of less than 5 mm, the probability of malignancy is low, and morphology on MRI adds little additional information. For lymph nodes with a short axis diameter between 5-10 mm, morphology on ferumoxytol-enhanced MRI can help to differentiate benign and malignant nodes.

We measured the R2\*-relaxation rate of entire lymph nodes, including the hilum and the parenchyma. The mean R2\*-relaxation rate for benign lymph nodes was significantly higher (127.8 +/- 46.3 Hz) compared to malignant lymph nodes (84.4 +/- 23.4 Hz; p<0.001, Figure 3J-L, Table 3).

When we compared the different parameters and adjusted for size, ADCmean values were the best predictors of malignancy with an AUC of 0.99, followed closely by SUV-ratio, hilum enhancement pattern and R2\* values (AUC= 0.98, 0.97, 0.93, respectively). Between ADCmean, SUV-ratio and pattern there was no significant difference (p=0.07), however all three parameters were significantly different to R2\* (p=0.001, respectively; Table 2).  
 Table 3. Differentiation of benign and malignant lymph nodes in pediatric patients on ferumoxytol-enhanced <sup>18</sup>F- FDG-PET/MRI scans

	ADCmean [10-6 mm <sup>2</sup> /s]		SUV-ratio		R2* [Hz]	
Sizes	benign	malignant	benign	malignant	benign	malignant
< 5 mm	$1521.0 \pm$	851.7 ±	0.5 ±	$1.2 \pm 0.5$	$123.5 \pm$	$92.3\pm23.7$
	416.6	121.0	0.2		45.9	
$\geq 5 - \leq 10 \text{ mm}$	$1614.1 \pm$	$818.2 \pm$	$0.57 \pm$	$2.3 \pm 1.2$	$130.9 \pm$	$91.7 \pm 27.9$
	418.9	188.1	0.3		47.9	
≥10 mm	1539.3 ±	884.2 ±	0.7 ±	$3.4 \pm 1.4$	$128.3 \pm$	$76.7 \pm 17.9$
	432.7	227.4	0.3		32.5	
All	1578.7 ±	851.7 ±	0.5 ±	$2.8\pm1.5$	127.8 ±	$84.4\pm23.4$
	419.8	206.2	0.3		46.3	

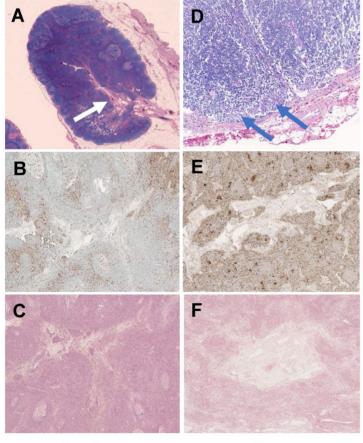
Mean +/- standard deviation; ADCmean: Mean apparent diffusion coefficient; SUV: standardized uptake value; SUV-ratio: SUVmax lesion/SUVmean liver; R2\*: R2\*-relaxation rate



**Figure 3. Differentiation of benign and malignant lymph nodes in pediatric patients:** Example of a benign lymph node in the left axillary of a 16 year old boy with Ewing's sarcoma (left row) and a malignant lymph node in the left para-aortic retroperitoneal region of a 14 year old boy with a desmoplastic small round cell tumor (middle row). (A, B) Ferumoxytol-enhanced T2-weighted FSE images show a hypointense hilum of the benign lymph node (A) and lack of hypointense enhancement of the malignant node (B). (C) The mean size of 464 benign and 149 malignant lymph nodes on T2-FSE images was significantly different (p<0.0001). (D, E) Apparent diffusion coefficient (ADC) maps demonstrate unrestricted diffusion (bright signal) of the benign lymph node (D) and restricted diffusion (dark signal) of the malignant lymph node (E). (F) The mean ADC value of 464 benign and 149 malignant lymph nodes on diffusion-weighted images was significantly different (p<0.0001). (G, H) <sup>18</sup>F-FDG PET images demonstrate background signal of the benign lymph node (G) and markedly increased FDG uptake of the malignant lymph node. (H) compared to normal background tissue. (I) The mean standardized uptake value (SUV) ratio of 464 benign and 149 malignant lymph nodes was significantly different (p<0.0001). (J, K) R2\* relaxation rate maps demonstrate increased R2\* (=shortened T2\*) of the benign lymph node uc to increased iron content and short R2\* (= long T2\*) of the malignant lymph nodes. The mane R2\* relaxation rate of 464 benign and 149 malignant lymph nodes was significantly different (p<0.0001). (<sup>18</sup>F-FDG: 2-deoxy-2-[F-18]fluoro-D-glucose; ADC mean: mean apparent diffusion coefficient; FSE: fast spin echo; PET: positron emission tomography; R2\*: R2\*-relaxation rate; SUV: standardized uptake value; SUV<sub>mean</sub> liver.

#### Histology

To obtain proof-of-concept of the normal architecture of lymph nodes in a child, we obtained H&E, CD68 and Prussian blue (iron) stains of two exemplary normal lymph nodes of two of our patients: In accordance with our imaging findings, we noted a vascularized hilum and the typical morphology of medulla, paracortex and cortex (Figure 4). The cortex contained multiple lymphoid follicles. We detected tumor cell infiltrates in the malignant lymph node. We found only few, scattered macrophages in the benign lymph node. Interestingly, we found a slightly increased number of macrophages in the malignant lymph node (Figure 4). Prussian blue stains performed on sections of both nodes show no cytoplasmic iron stored within these macrophages (Figure 4), confirming our hypothesis that the iron enhancement pattern of benign lymph nodes is a function of vascularity rather than macrophage uptake.



### benign

### malignant

**Figure 4. Histology of a representative benign and malignant lymph node after ferumoxytol infusion.** The benign lymph node demonstrates a prominent central hilum with vessels (A, white arrow) on H&E staining, few scattered macrophages (brown) on CD68-stains (B, 40x magnification) and no iron in macrophages on Prussian blue staining (C, 40x magnification). The malignant lymph node demonstrates tumor cell infiltration (D, blue arrows), an increased number of macrophages on CD68-staining (E, 40x magnification) and no iron necrophages on Prussian blue staining (E, 40x magnification) and no iron necrophages on Prussian blue staining (F, 40x magnification) and no iron necrophages on Prussian blue staining (F, 40x magnification). H&E: Hematoxylin and eosin.

### Discussion

Our data showed that ferumoxytol administration improves the characterization of intermediate size lymph nodes (5-10 mm) on <sup>18</sup>F-FDG PET/MR imaging studies. The absence of an enhancing hilum in these subcentimeter nodes was highly suggestive of malignancy.

Our finding of a hyperintense signal of malignant lymph nodes on ferumoxytol-enhanced T2-weighted images in pediatric patients is in accordance with previous studies in adult patients [17, 29]. The absent ferumoxytol-enhancement of malignant lymph nodes can be explained by lymphoid tissue replacement with malignant cells, which do not phagocytose iron oxide nanoparticles [17, 21, 36]. In pediatric patients with lymphoma and markedly enlarged malignant lymph nodes, Aghighi et al. reported a significant ferumoxytol enhancement

on T2-weighted MR images, which corresponded to ferumoxytol uptake by tumor associated macrophages on histology [37]. It has been well described that the number of tumor associated macrophages increases with increasing tumor size [38]. Absent ferumoxytol enhancement in subcentimeter lymph node metastases in our study could be either due to a lack of extravasation of nanoparticles into the lymph node interstitium or low quantity of tumor associated macrophages.

We found different ferumoxytol enhancement patterns of benign lymph nodes in children compared to those previously reported in adult patients [17]. In adult patients, benign lymph nodes demonstrated hypointense signal enhancement of the entire lymph node at 24 h injection [17, 29]. These nodes post demonstrated minimal or no T2\*-enhancement on relatively early postcontrast scans (5 h), and progressively increasing hypointense (dark) T2\* signal enhancement at 18 h and 24 h after intravenous injection of ferumoxtran-10 [17] and ferumoxytol [29]. Previous investigators hypothesized that the iron oxide nanoparticles were slowly phagocytosed by macrophages in normal lymph nodes [17]. However, we found a different ferumoxytol enhancement pattern of benign lymph nodes in pediatric patients. We noted a marked hypointense signal enhancement of the hilum of benign lymph nodes on T2- weighted scans. This hilar enhancement did not change between 2 h and 24 h after intravenous injection of ferumoxytol. Interestingly, Wells et al. showed in benign inguinal lymph nodes in adults that after an initial increase in R2\*, the pharmacokinetics in lymph nodes are

similar to the distribution of ferumoxytol in the blood pool [39]. It is well known from ultrasound studies [25] and histopathological investigations [22] that normal lymph nodes in children contain a highly vascularized hilum. As patients get older, the vascularity of the hilum decreases and the hilum is replaced by fat [22]. Interestingly, we did not find any ferumoxytol significant enhancement of the parenchyma of benign lymph nodes in our pediatric patients, neither at 2 h nor at 24 h, suggesting that macrophage uptake does not play a role in the pediatric lymph node enhancement on MRI. This can be explained by well-described differences in cellular composition of benign lymph nodes in children and adults [22, 23, 28]. In children, benign and reactive lymph nodes are better vascularized and mostly composed of T- and B-cells, with little macrophages.

Previous studies in adults show that the detection of small lymph node metastases with short axis diameter of <10 mm is a main limitation of both <sup>18</sup>F-FDG PET [34, 40] and MR [34, 41] due to a limited sensitivity of PET for detection of subcentimeter lesions and absence of functional information on unenhanced MR images. Studies in animal models have shown that nanoparticles can differentiate tumor containing sentinel lymph nodes and normal lymph nodes in cervical cancer [42] and oral squamous cell cancer [43] based on differential near infrared enhancement patterns. However, both nanoparticle compounds used in these studies were not clinically applicable in humans. We found that the FDA-approved iron supplement ferumoxytol added value for the diagnosis of lymph nodes with a diameter of 5-10 mm in humans. This is important as subcentimeter malignant lymph nodes might show only mild <sup>18</sup>F-FDG uptake. Positron emission tomography studies in humans revealed that tumor-targeted PET tracers increased the detection rates of small lymph node metastasis with diameters of less than 5 mm in humans with breast cancer [44] and prostate cancer [45]. However, tumor-targeted tracers have not been developed for bone or soft tissue sarcomas in children. Our approach to visualize the hilum with ferumoxytol was not useful for normal lymph nodes with a size of < 5 mm, likely due to limitations in anatomical resolution. However, our data indicate that the likelihood of malignancy in such small lymph nodes is extremely low and micrometastasis would not change the treatment or outcome of patients. Currently the standard of evaluating and detecting micrometastatic disease burden is through polymerase chain reaction and flow cytometry as it can detect as few as 1 tumor cell in 500 000 cells [46] which is more sensitive than any other clinical imaging technique. Yet, the presence of micrometastases in children with neuroblastoma [47] or Ewing sarcoma [48] did not affect the initial disease burden, event-free or overall survival in patients. Therefore, detection of micrometastases does not alter patient management. In patients with bone sarcomas, pulmonary metastases smaller than 3 mm are not referred to surgery because they are too small to be localized, cannot be reliably resected and because the minimal disease burden will likely not change overall outcome [49, 50].

Due to recent concerns about gadolinium deposition in the brain, there is a search for alternative MR contrast agents that are biodegradable. Iron oxide nanoparticles could fill this gap. The off-label use of iron supplements such as ferumoxytol as MR contrast agents is an example of a therapeutic drug that can be used for diagnostic purposes. Future studies could investigate, if the hilar accumulation of nanoparticles can facilitate priming of antigen presenting dendritic immune cells and enhance the anti-tumor effect of standard chemotherapy. Other studies showed that the tumor environment releases immunosuppressive cytokines [51]. Presentation of whole-tumor cell lysate vaccines caused limited activation of the immune system against the cancer [52]. Kohlhapp et al. showed that coupling tumor antigens to nanoparticles activated T-cells and reduced tumor growth [53]. Other groups showed that nanoparticles coupled with cancer antigen delayed tumor growth and prolonged survival [54]. Nanoparticles with a relatively small size of 25 nm induced antigen presentation and dendritic cell activation in lymph nodes [55].

Our results showed that ADCmean values, SUV-ratio, and hilar enhancement pattern had similar high prediction rates for lymph node malignancy. Several studies described the high diagnostic value of ADC maps in pediatric patients [56, 57]. Ferumoxytol-enhancement pattern and ADCmean yielded the same predictive information than SUV-values on PET, yet without any radiation exposure. This is very important because ionizing radiation exposure can be avoided and MR scanners are more widely available than PET scanners.

In our study, a few limitations should be noted: 1. Participants in our study had a variety of different tumor types. However, our sample is representative of pediatric patients typically investigated in a major Children's Hospital, especially since lymph node metastases are very rare in pediatric cancers. 2. Our study evaluated patients before start of chemotherapy or irradiation. Further studies have to evaluate ferumoxytol-enhancement patterns of benign and malignant lymph nodes after therapy. 3. As ferumoxytol has to be injected slowly over 15 min we did not acquire pre-contrast scans in all patients. To acquire two separate PET/MRI scans without and with ferumoxytol would not be feasible for routine clinical applications in children especially given the radiation dose associated with a PET scan.

### Conclusions

In summary, we found that MR enhancement pattern of lymph nodes with ferumoxytol are different in children compared to adults. For lymph nodes with a size of 5-10 mm, the presence of an enhancing hilum can reassure the diagnosis of a benign lymph node.

### Abbreviations

<sup>18</sup>F-FDG: 2-deoxy-2-[F-18]fluoro-D-glucose; ADC: apparent diffusion coefficient; AUC: area under the curve; CI: confidence interval; CT: computed tomography; DWI: diffusion weighted imaging; FOV: field of view; IDEAL-IQ: iterative decomposition of water and fat with echo asymmetry and least-squares estimation; IND: investigational new drug; LAVA: liver acquisition with volume acquisition; MRI: magnetic resonance imaging; OR: odds ratio; PET: positron emission tomography; PROPELLER: periodically rotated overlapping parallel lines with enhanced reconstruction; R2\*: R2\*-relaxation rate; RECIST: response evaluation criteria for solid tumors; ROC: receiver operating curve; SUV: standardized uptake value; SUV-ratio: SUVmax lesion/SUVmean liver; T2-FSE: T2-weighted fast spin echo.

### **Supplementary Material**

Supplementary figures and tables. http://www.thno.org/v10p3612s1.pdf

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### **Competing Interests**

The authors have declared that no competing interest exists.

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