

**Figure S1.** Chemical synthesis of MQs. **(A)** Reverse-phase liquid chromatography of the crude synthesis of MQ1 on Waters X-bridge C18, 19 × 250 mm, 10  $\mu$ m, flow of 15 mL/min. The gradient was 10 to 40% of solvent B in 30 min. **(B)** Reverse-phase liquid chromatography of crude oxidized MQ1 on a Waters Sunfire 10  $\mu$ m, 10 × 250 mm, 5 mL/min, same gradient. **(C)** Analytical chromatography of the pure oxidized MQ1 performed on Waters X-bridge C18, 4.6 × 150 mm, 3.5  $\mu$ m. The gradients were 10 to 50% of solvent B in 40 min. Insert peak zoom. **(D)** Reverse-phase liquid chromatography of the crude synthesis of 6-azidohexanoic-MQ1 on Waters X-bridge C18, 19 × 250 mm, 10  $\mu$ m, flow of 15 mL/min. The gradient was 10 to 40% of solvent B in 30 min. **(E)** Reverse-phase liquid chromatography of crude oxidized 6-azidohexanoic-MQ1 on a Waters Sunfire 10  $\mu$ m, 10 × 250 mm, 5 mL/min, 5 mL/min, same gradient. **(F)** Analytical chromatography of the pure oxidized MQ1 performed on Q1 performed on Waters X-bridge C18, 4.6 × 150 mm, 3.5  $\mu$ m. The gradient was 10 to 50% of solvent B in 30 min. **(E)** Reverse-phase liquid chromatography of crude oxidized 6-azidohexanoic-MQ1 on a Waters Sunfire 10  $\mu$ m, 10 × 250 mm, 5 mL/min, same gradient. **(F)** Analytical chromatography of the pure oxidized MQ1 performed on Waters X-bridge C18, 4.6 × 150 mm, 3.5  $\mu$ m. The gradient was 10 to 50% of solvent MQ1 on a Waters Sunfire 10  $\mu$ m, 10 × 250 mm, 5 mL/min, same gradient. **(F)** Analytical chromatography of the pure oxidized MQ1 performed on Waters X-bridge C18, 4.6 × 150 mm, 3.5  $\mu$ m. The gradient was 10 to 50% of solvent MQ1 on Waters X-bridge C18, 4.6 × 150 mm, 5.5  $\mu$ m. The gradient was 10 to 50% of the pure oxidized MQ1 performed on Waters X-bridge C18, 4.6 × 150 mm, 3.5  $\mu$ m. The gradient was 10 to 50% of solvent MQ1 performed on Waters X-bridge C18, 4.6 × 150 mm, 3.5  $\mu$ m. The gradient was 10 to 50% of solvent MQ1 performed on Waters X-bridge C18, 4.6 × 150 mm, 3.5  $\mu$ m.

solvent B in 40 min. Insert peak zoom. (G) Reverse-phase liquid chromatography of deferoxamine-DBCO-MQ1 on Waters X-bridge C18, 4.6 × 150 mm, 3.5  $\mu$ m. The gradient was 20 to 60% of solvent B in 40 min. (H) Zoom of the analytical chromatography of the pure deferoxamine-DBCO-MQ1 performed on Waters X-bridge C18, 4.6 × 150 mm, 3.5  $\mu$ m. Reverse-phase liquid chromatography of AFDye-488-DBCO-MQ1 (I) or Cy5.5-DBCO-MQ1 (J) on Waters X-bridge C18, 4.6 × 150 mm, 3.5  $\mu$ m. The gradient was 30 to 60% of solvent B in 30 min. solvents are A (H<sub>2</sub>O, TFA 0.1%) and B (acetonitrile, TFA 0.1%). Color bar indicates collected fractions. (K) Reverse-phase liquid chromatography of the crude synthesis of MQ1 variant on Waters X-bridge C18, 19 × 250 mm, 10  $\mu$ m, flow of 15 mL/min. The gradient was 10 to 40% of solvent B in 30 min. (L) Reverse-phase liquid chromatography of crude oxidized MQ1 variant on a Waters Sunfire 10  $\mu$ m, 10 × 250 mm, 5 mL/min, same gradient.



**Figure S2**. **(A)** Representative iTLC chromatograms of fresh <sup>89</sup>Zr-DFO-MQ1 and free <sup>89</sup>Zr. iTLC chromatograms of blood sampling 1.5 h after injection (**B**) and 7 days after injection (**C**). **(D)** *In vitro* stability of <sup>89</sup>Zr-DFO-MQ1 in bovine serum and PBS determined by iTLC. **(E)** RadioHPLC characterization of fresh <sup>89</sup>Zr-DFO-MQ1. **(F)** RadioHPLC characterization of blood sampling at 7 days post injection.





**Figure S3.** (A) Binding curve on hV2R of MQ1, Cy5.5-MQ1 and AFDye-488-MQ1. B) Labeling of hV2R expressed in stable CHO cell line with increasing doses of AFDye-488-MQ1. Control without AFDye-488-MQ1 looks like the 10 nM cell labeling. Specificity is determined by the last panel in presence of 3.4  $\mu$ M MQ1. (C) Selectivity of 30 nM AFDye-488-MQ1 to hV2R labeling compared to hV1aR, hV1bR, hOTR and rOTR expressed stably in CHO cell lines. (D) and (E) are the same than (B) and (C) but using the Cy5.5-MQ1 probe.

Table \$	S1: Blood	sample	preparation	for PK	determination
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Sample										
100 μL of rat plasma										
Dilution 400 µL of TFA 0.1%										
Solid Phase Extraction Oasis HLB 1 cc 30 mg cartridge (Waters) / SpeedDisk (Biotage)										
500 μL										
2 x 500 μL H₂O/MeOH (9/1) + TFA 0.1%										
lution 2 x 150 µL H <sub>2</sub> O/MeOH (2/8) + TFA 0.1%										
Evaporation to dryness under N <sub>2</sub> at 40°C										
Reduction/Alkylation										
Reduction 25 µL DTT 100 mM (in AB 50 mM)-incubation 10 min at 90°C.										
Alkylation 25 µL IAA 225 mM (in AB 50 mM)-incubation 30 min at RT to darkness										
ddition 10 µl H <sub>2</sub> O/ACN (9/1) 0.5% HCOOH										
Centrifugation 20 000 g /15 min										
40 µL										
Injection 10 µL into the UPLC-MS/MS system										
trifluoroacetic acid	AB	ammonium bicarbonate								
methanol	IAA	iodoacetamide								
dithiothreitol	RT	room temperature								
water	ACN	acetonitrile								
	100 $\mu$ L of rat plasma 400 $\mu$ L of TFA 0.1% traction Oasis HLB 1 cc 3 500 $\mu$ L 2 x 500 $\mu$ L H <sub>2</sub> O/MeOH (9/ 2 x 150 $\mu$ L H <sub>2</sub> O/MeOH (2/3 to dryness under N <sub>2</sub> at 40° vlation 25 $\mu$ L DTT 100 mM (in AB 25 $\mu$ L IAA 225 mM (in AB 10 $\mu$ I H <sub>2</sub> O/ACN (9/1) 0.5% 20 000 g /15 min 40 $\mu$ L 10 $\mu$ L into the UPLC-MS/M trifluoroacetic acid methanol dithiothreitol water	100 µL of rat plasma400 µL of TFA 0.1%traction Oasis HLB 1 cc 30 mg cartrid500 µL2 x 500 µL H <sub>2</sub> O/MeOH (9/1) + TFA 0.1°2 x 150 µL H <sub>2</sub> O/MeOH (2/8) + TFA 0.1°to dryness under N <sub>2</sub> at 40°Crlation25 µL DTT 100 mM (in AB 50 mM)-incu25 µL IAA 225 mM (in AB 50 mM)-incu10 µl H <sub>2</sub> O/ACN (9/1) 0.5% HCOOH20 000 g /15 min40 µL10 µL into the UPLC-MS/MS systemtrifluoroacetic acidABmethanolIAAdithiothreitolRTwaterACN								

	Transitions monitored	Role	
	673.03 > 731.62	quantification	
	673.03 > 775.09		
	747.58 > 836.10	specificity	
	841.02 > 975.05		
MQ1 variant	666.97 > 735.39	Used for internal standard (IS) normalization	
	740.87 > 840.60	IS specificity	

 Table S2: parent ion > product ion transition monitored

		Plasma	
	MQ1 plasma	concentration	Diuresis values
Times	concentrations	with modelling	with modelling
(h)	µg/ml	(µg/mL)	(mL/h/kg)
1	1.709	1.305	27.69
2	0.486	0.783	80.74
3	0.259	0.606	55.34
4	1.239	0.469	37.93
5	0.257	0.363	26.00
6	0.378	0.281	17.82
8	0.150	0.169	14.79
10	0.299	0.178	14.12
12	0.219	0.173	13.48
16	0.154	0.164	12.28
24	0.096	0.148	10.20
48	0.056	0.108	5.83
72	0.092	0.079	3.34
96	0.074	0.058	1.91

Table S3. PK values and PK PD concentrations under modeling.