Supporting Data

Rod-shape theranostic nanoparticles facilitate antiretroviral drug biodistribution and activity in human immunodeficiency virus susceptible cells and tissues

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Results

SR1: XPS Analysis

BSNRs and LuBSNRs particles were analysis by XPS and data were shown in the survey plot (supplementary Figure S5A, E) exhibiting the present of key elements, such as O, C, Bi, and S. This shows that the particle surface is of the highest quality in terms of chemistry and stability, free from unreacted end precursors. The presence of the O peak is due to the adsorbed oxides on the particles surface.[1] C1s with standard binding energy value of ~ 285 eV is used as a reference.[2] In the Bi region, the high resolution XPS spectra (supplementary Figure S5C) validations two characteristic peaks at 158.3 and 163.6 eV, which can be exchangeable, respectively, to the binding energies of Bi 4f7/2 and Bi 4f5/2 in BSNRs particles.[2, 3] The broad peak at around 225.43 eV (supplementary Figure S5B) corresponds to the binding energy of S2s. The experimental values are analyzed to be in nearby with the data reported previously. [4] LuBSNRs particles' most intense peak occurs at 157.8 eV (Bi 4f7/2). This peak and corresponding doublet at 163.1 eV (Bi 4f5/2) indicates that the Bi/S ratio in the compound is greater at the surface or is indicative of BiS coordination. Lu signals appeared but did not rise above the level of the background signal which indicates Lu was not available on the surface of particles (supplementary Figure S5H).

SR2: Morphology and particle size characterization

The surface morphology of BSNR particles was studied using AFM. The size and size distribution of these BSNRs demonstrated uniform size and distribution (**Figure S6A-B**). Surface morphology of the NRPV nanocrystals was determined using AFM, SEM and

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TEM. The particles were uniform and monodisperse with a characteristic rectangular shape. SEM images of pure NRPV nanocrystals are shown in **Figure S6C-E**.

SR3: In vitro bismuth and RPV correlation study in macrophage

Figure S9: Panels A-D show various two parameter comparisons for data obtained for bismuth and RPV concentrations in macrophages treated with BSNRs-RPV particles at 12.5 µg/mL or 25.0 µg/mL concentrations. **Panel A** shows the correlation, between the uptake of bismuth, after macrophages were treated with BSNRs-RPV particles at a low concentration (12.5 µg/mL) or at a higher concentration (25 µg/mL). As expected, this correlation was very good with r = 0.9982. We also examined the relationship between the uptake of RPV in macrophages treated with a low (12.5 μ g/mL) or high (25 μ g/mL) concentration of BSNRs-RPV, and found a correlation with r = 0.8350, as shown in panel B. Panel C shows the correlation between bismuth and RPV uptake over time in macrophages treated with either 12.5 µg/mL or 12.5 µg/mL BSNRs-RPV particles, while **panel D** displays the correlation in cells treated with 25 µg/mL BSNRs-RPV particles. The bismuth and RPV data from the lower concentration of BSNRs-RPV particles showed a positive correlation with r = 0.7570 while the higher concentration showed a positive correlation with r = 0.9690. **Panels E-H** show the various two parameter comparisons for data obtained during retention experiments. Macrophages were treated with either a high (25 µg/mL) or low (12.5 µg/mL) concentration of BSNRs-RPV particles for 8 hours before treatment was removed and replaced by treatment free media. The retention of bismuth in macrophages treated with 12.5 µg/mL BSNRs-RPV positively correlated with the amount of RPV retained in cells over time in macrophages treated with 25 μ g/mL BSNRs-RPV particles, with r = 0.9721. The comparison between bismuth

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concentrations retained in macrophages treated with the high concentration of BSNRs-RPV particles and bismuth concentration retained in cells treated with the lower BSNRs-RPV particles dose yielded an "r" value of 0.9353. **Panel G** shows the relationship between RPV and bismuth retention in cells treated with a low dose of BSNRs-RPV particles. This also demonstrated a significant, positive correlation with r = 0.8093. The final **panel**, **H**, shows the positive correlation between the two parameters in cells treated with a high concentration of BSNRs-RPV particles (r = 0.8200).

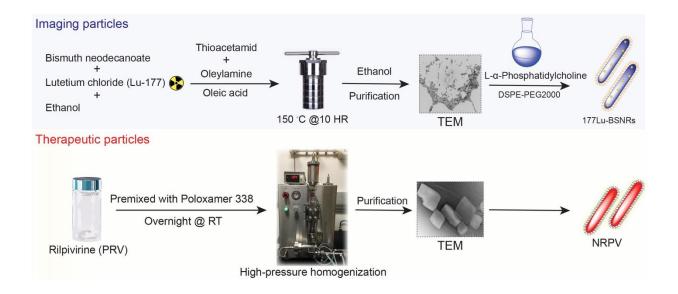


Figure S1. Schematic of production of imaging and therapeutic particles. Schematic illustration of the two-step production, synthesis, and lipid functionalization of BSNRs imaging particles via solvothermal technique. Therapeutic NRPV particles were prepared by a top-down high-pressure homogenization approach using poloxamer 338 as a surfactant and stabilizing agent. Representative transmission electron microscopy (TEM) images of both types of particles are depicted in this schematic.

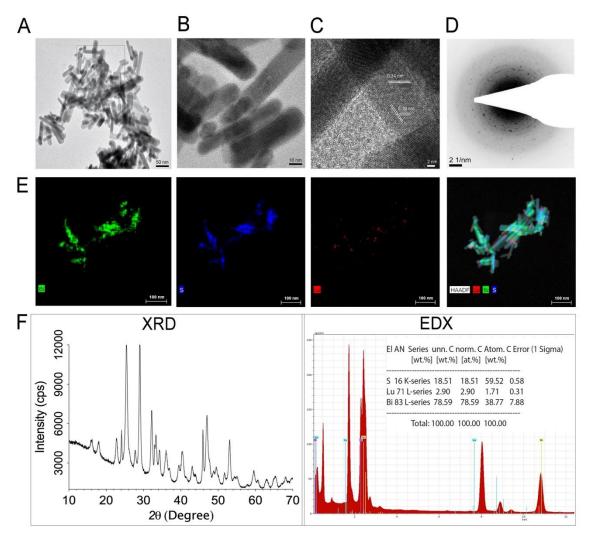


Figure S2. Synthesis and characterization of LuBSNRs. Low resolution TEM images of LuBSNRs are illustrated in (A). High resolution TEM images of twin LuBSNRs particles with lattice planes are shown in (B-C) with corresponding electron diffraction patterns. The selected area electron diffraction (SAED) pattern of LuBSNRs are shown in panel (D). High-angle annular dark-field electron microscopy showed element localization within the particles. Bismuth (green), sulfur (blue) and lutetium (red) (E). XRD patterns (F left panel) and quantitative measurements of particle element distribution were made by energy dispersive X-ray spectroscopy (F, right panel). Chemical composition of LuBSNRs was assessed by X-ray fluorescence. Typical particles consisted of 87.72% bismuth, 11.29% sulfur and 0.975 % lutetium by mass (**XRF data: Supplementary figure S4**). Molar ratio of synthesis was 1:1:0.1 (Bismuth neodecanoate: thioacetamide: lutetium (III) chloride, anhydrous, LuCl₃) with a typical synthesis involving, 8 mM (5.8g) bismuth neodecanoate, 8 mM (600 mg) thioacetamide, and 0.8 mM (224 mg) LuCl₃.

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			SQX Ca	alculation Resu	ılt		
Sample : BiS Application : F-U_Solid_N_021 Matching library :			Sample type : Metal & Alloy Sample film corr. : File : BiS		Date analyzed : 2019- 6-28 11:36 Balance : Impurity corr. : CupFiCot		
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Figure S3. XRF (X-ray fluorescence) analysis of BSNRs.

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			SQX Ca	alculation Resu	ılt		
Sample : LuBiS Application : F-U_Solid_N_020 Matching library :			Sample type : Metal & Alloy Sample film corr. : File : LuBiS		Date analyzed : 2019- 6-28 11:15 Balance : Impurity corr. : CupFiCot		
1	S	11.2982	mass%	0.07030	S-KA	33.9524	12.0977
2	Lu	0.9756	mass%	0.07762	Lu-LB1	0.7009	0.8335
3	Bi	87.7262	mass%	0.08037	Bi-LA	156.2487	74.9502
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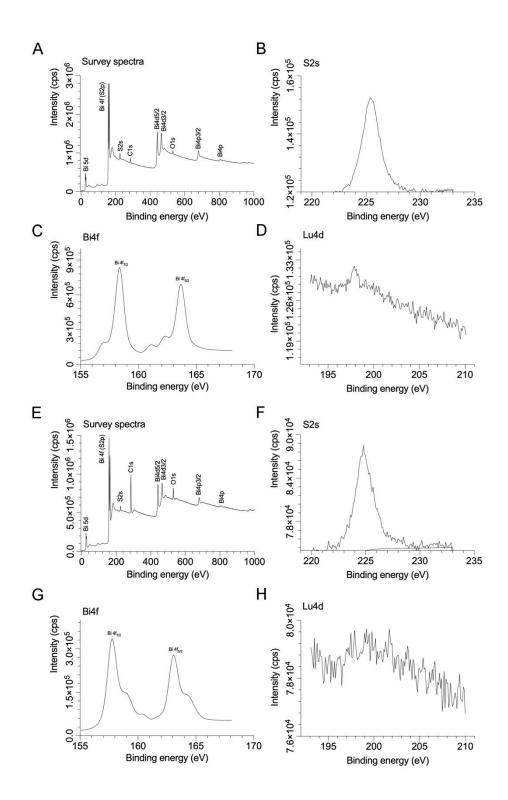


Figure S5. X-Ray Photoelectron Spectroscopy (XPS) analysis of Lu/BSNRs. Survey spectra (A, E), S region, (B, F), Bi (B, F) and Lu regions (D, H) of BSNRs and LuBSNRs particles, respectively are illustrated.

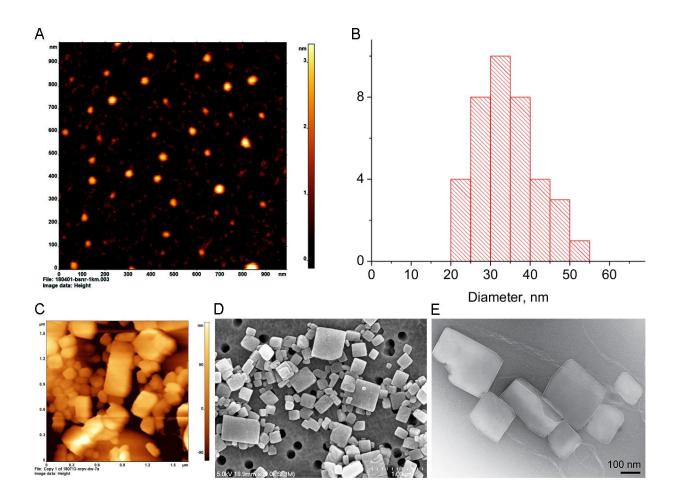


Figure S6. Morphological analysis of particles. AFM image of prepared BSNRs dispersed in cyclohexane (A). Corresponding size distribution histogram of BSNRs particles (B) and NRPV particles (C). SEM (D), and TEM (E) images of the NRPV particles are shown.

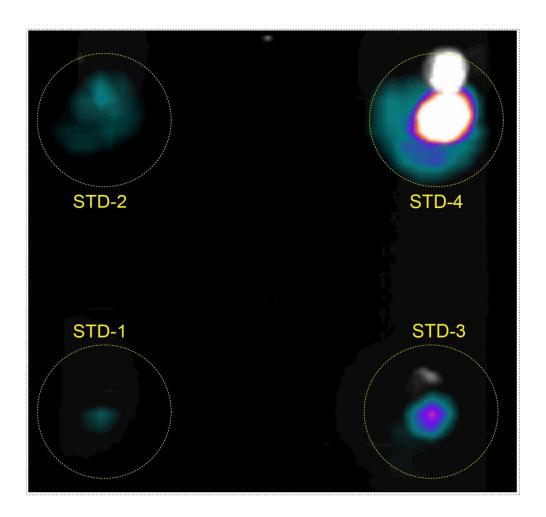


Figure S7. Phantom preparation. SPECT/CT phantom experiment using four known concentrations of ¹⁷⁷LuBSNRs particles are illustrated (STD 1 (~12.5 μ Ci); STD -2 (~ 25 μ Ci); STD-3 (~30 μ Ci); STD-4 (~50 μ Ci).

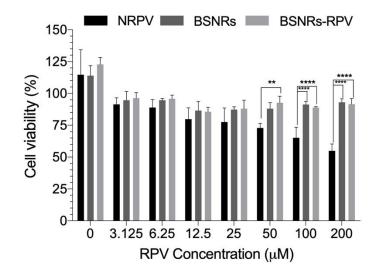


Figure S8. Cell viability. The analyses were performed with 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT); a cell viability assay performed in macrophages after treatment with NRPV, BSNRs-RPV or BSNRs particles at various concentrations (RPV). The total time of analysis was 8 hours. Significant differences were determined by a 2-way ANOVA followed by the Tukey's post hoc analysis. Stars denote significant differences compared to NRPV. **p < 0.01; ***p < 0.001; ****p < 0.001.

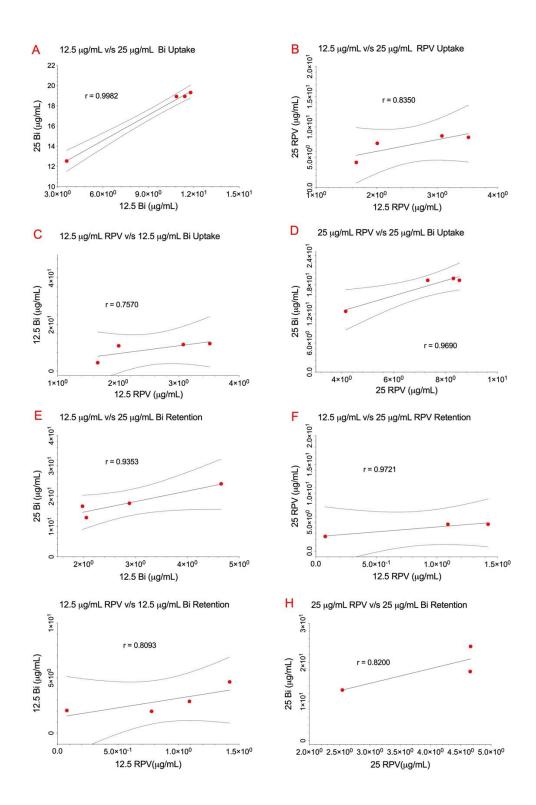


Figure S9. *In vitro* **uptake and retention.** Cross validation of metal and drug contents in macrophages by Pearson correlation analysis (Note: Panel "A" and "G" were also presented in Figure 2 of the manuscript).

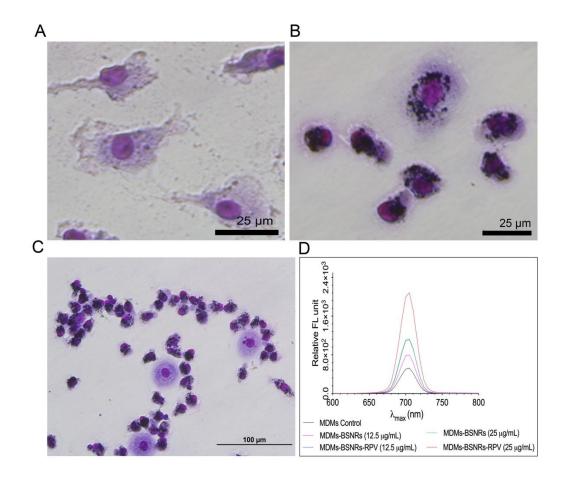


Figure S10. Light microscopy images of BSNR particles in macrophages (Giemsa stain). The black dots observed in the purple (Giemsa) stained cytoplasm are BSNRs particles. Untreated macrophages (A) and particle treated macrophages (25 μg/mL based on bismuth content) (B). Low magnification image of particle treated cells (C). Staining was carried out according to the manufacturer's instructions (Sigma Aldrich, May-Grünwald, MG500-500 mL and Giemsa stain Modified Solution, 48900-100ML-F). Uptake in macrophages was reconfirmed by fluorescence spectra analysis of particle-loaded cells (D).

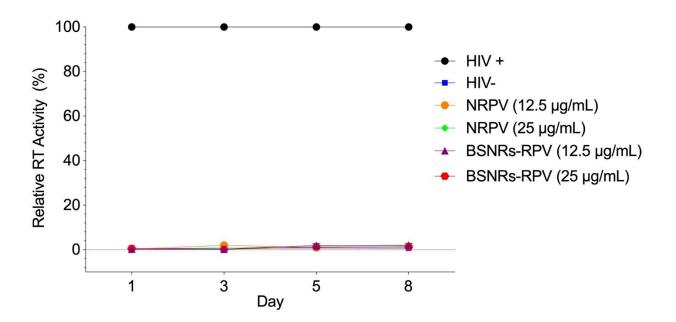


Figure S11. Antiretroviral activity as determined by HIV-1 reverse transcriptase. Antiretroviral activity was determined in macrophages treated for 8 hours with NRPV or BSNRs-RPV particles (12.5 and $25 \,\mu$ g/mL based on Bismuth contents) and then infected with HIV-1_{ADA} 1, 3, 5 or 8 days after particles treatment at a multiplicity of infection (MOI) of 0.1. At 10 days after infection, the media was collected from wells and progeny HIV virion production was determined by HIV reverse transcriptase (RT) activity using beta scintillation spectroscopy as previously described.[5]

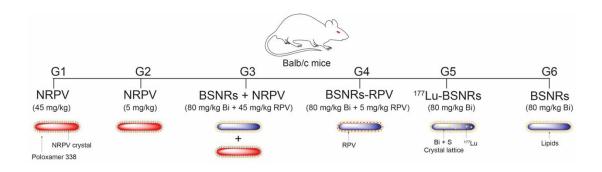


Figure S12. Design of animal experiments with different treatment groups.

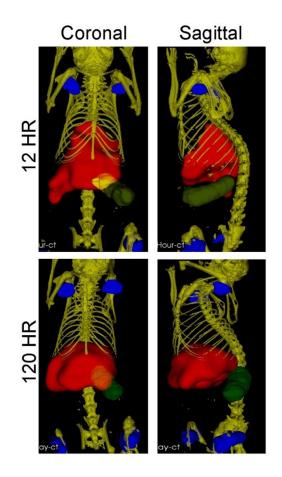


Figure S13. SPECT regions of interest (ROIs). SPECT 3D images of ¹⁷⁷LuBSNRs particles injected into Balb/c mice were quantified by drawing 3D regions-of-interest (ROI) to obtain radioactivity counts per volume of tissue. Shown here are representative ROIs for various organs. Regions of interest were drawn using VivoQuant v3.5 (Invicro Imaging Software) based on anatomy. Color code; liver = red, spleen = green, lymph nodes = blue and skeleton = yellow.

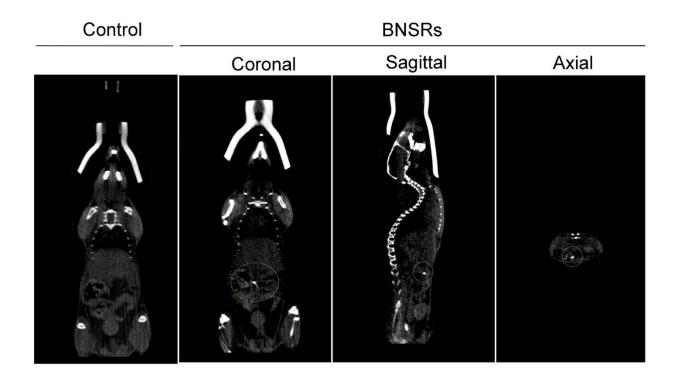


Figure S14. CT imaging using Xstrahl small animal radiation research platform (sarrp). BSNRs provide CT contrast in the system as represented by the images. CT coronal, sagittal and axial views of a mouse following intravenous injection of BSNRs solution (80 mg/kg) are shown. BSNRs treated mouse were shown the small signal for gut area highlighted in yellow ring. Two animals (One injected BSNRs solution at a dose of 80 mg/kg and other untreated) were analyzed 2 days after treatment using Xstrahl Inc. (Xstrahl, Suwanee, GA, USA) small animal radiation research platform (sarrp). Micro CT image is acquired in a pancake geometry. [6] The X-ray tube operates at a focal spot of 1 mm with tube voltage of 60 kV which gives an output of 1cGy of radiation over 1 minute. The images are acquired at a frame rate of 30 fps. After that the images are analyzed using (3D) Slicer with the DICOM module (https://www.slicer.org).

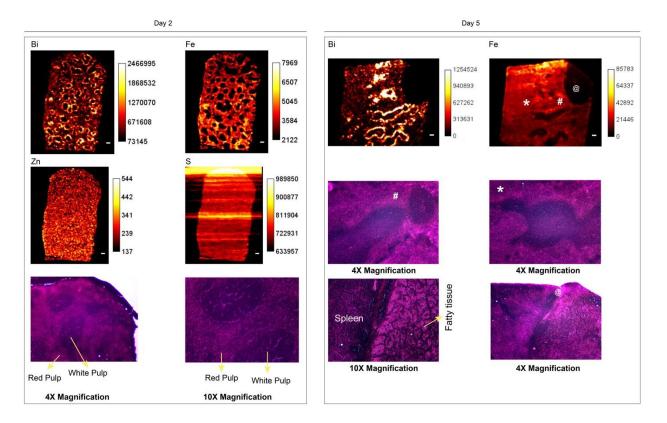


Figure S15. LA-ICP-MS images of BSNRs (G6) in spleen tissue. Spleen tissue from animals treated with BSNRs (G6) was collected on days 2 and 5 post-injection. Tissue sections were stained with H&E and analyzed by LA-ICP-MS. Bi, Fe, Zn and S levels were compared with background levels. Signal to background ratio for Bi was found to be highest in the macrophage rich marginal zone of the spleen tissue and the red pulp region with higher levels seen at day 5 as compared to day 2. Fe allows for distinction between the red and white pulp. Zn is used as an internal standard for signal quality. The white bar corresponds to 10 pixels (500 μ m).

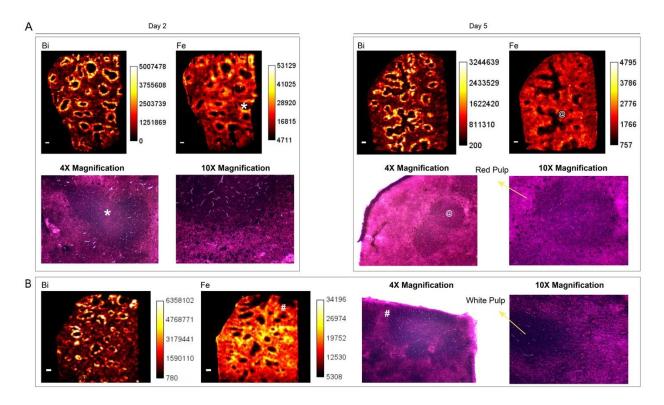
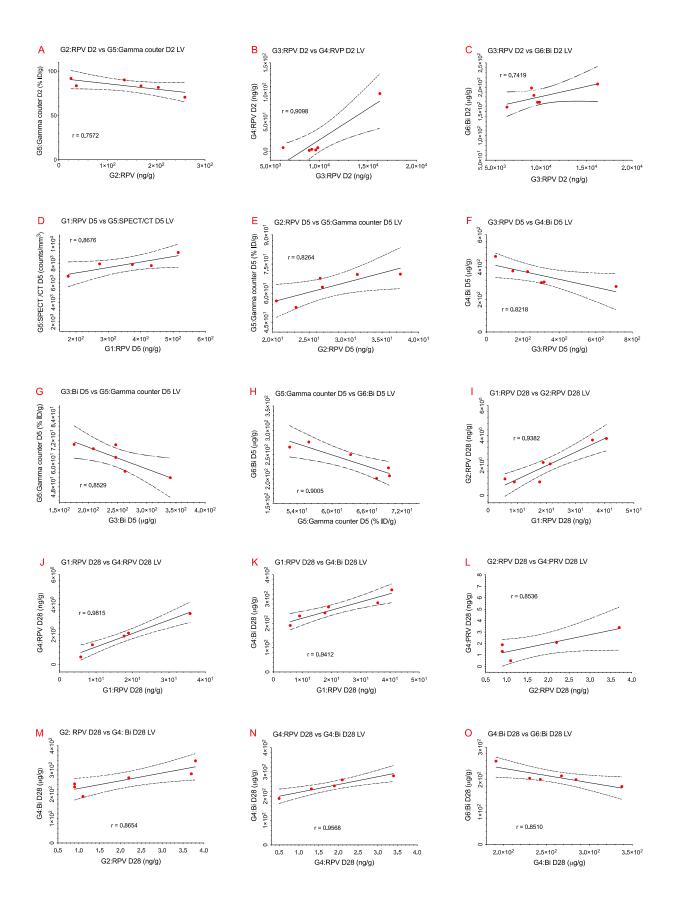
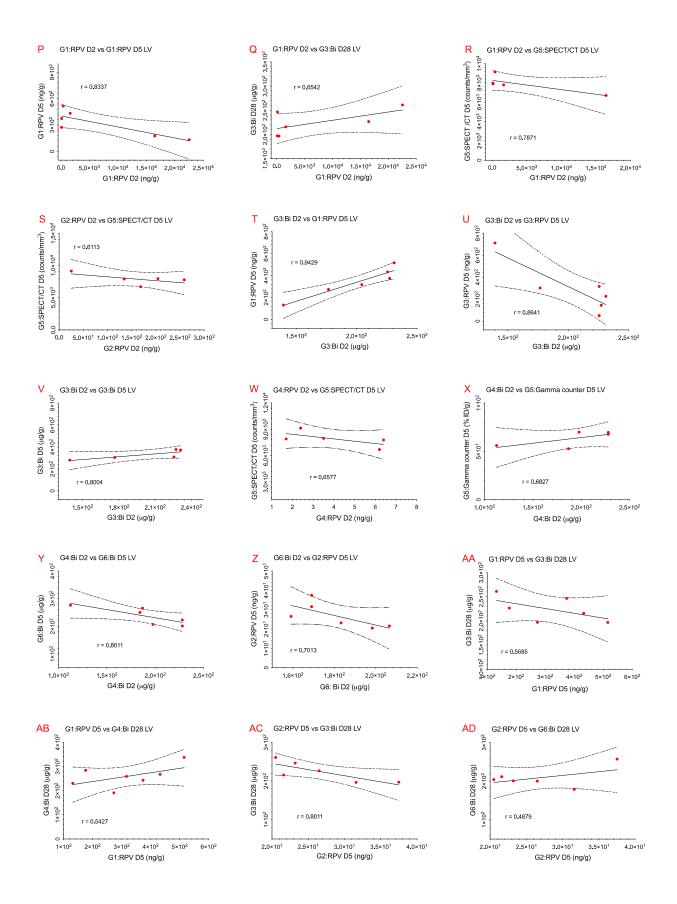
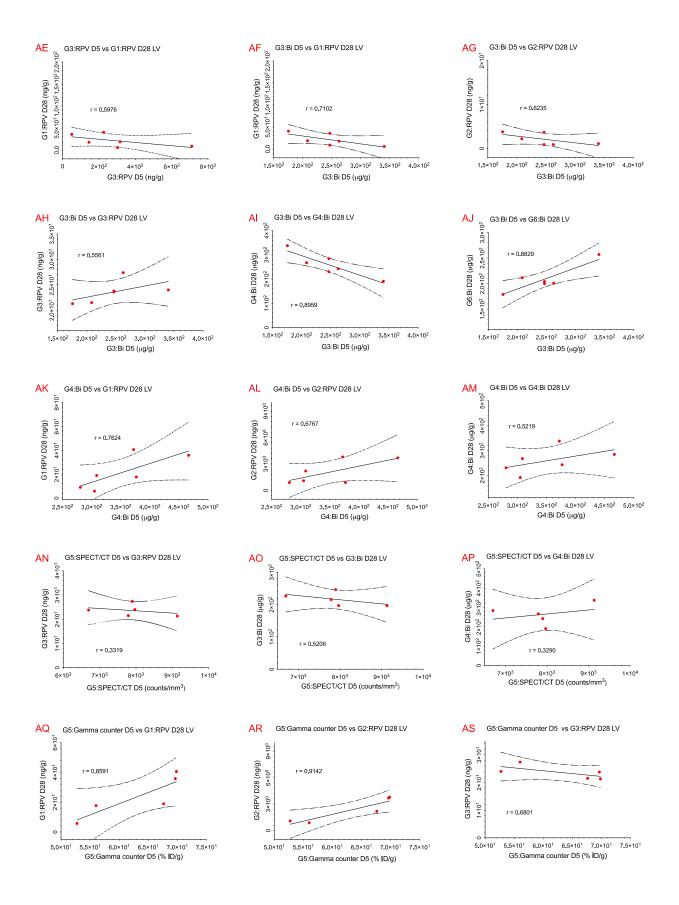


Figure S16. LA-ICP-MS images of BSNRs-RPV (G4) and BSNRs and NRPV(G3) treated spleen tissue. Spleens from animals treated with BSNRs-RPV(G4) were collected on days 2 and 5 after injection (A). Spleen tissue from animals treated with BSNRs plus NRPV(G3) was collected on Day 2 post-injection (B). Tissue sections were stained with H&E and analyzed by LA-ICP-MS. Bi and Fe levels were compared with background levels. Signal to background ratio for Bi was found to be highest in the macrophage rich marginal zone of the spleen tissue and the red pulp region with higher levels seen at day 5 as compared to day 2. Fe allows for distinction between the red and white pulp. The white bar corresponds to 10 pixels (500 µm).







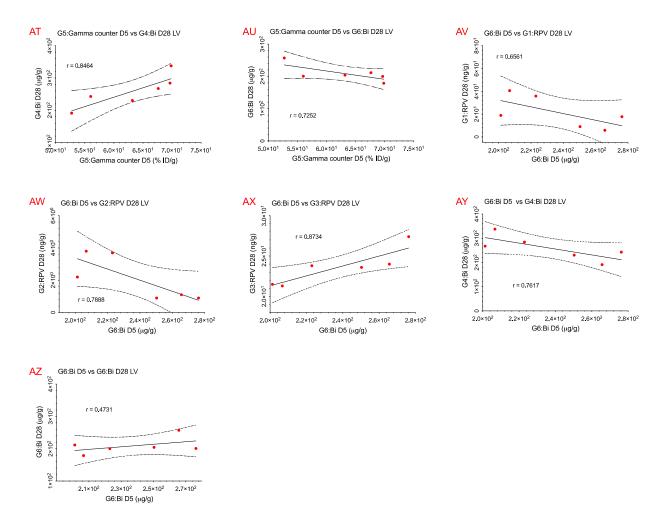
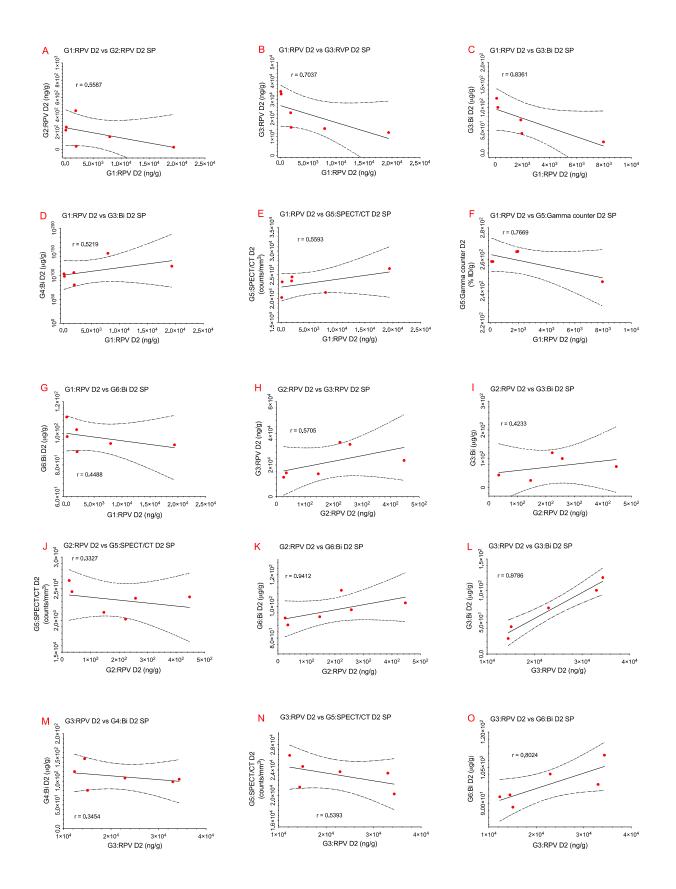
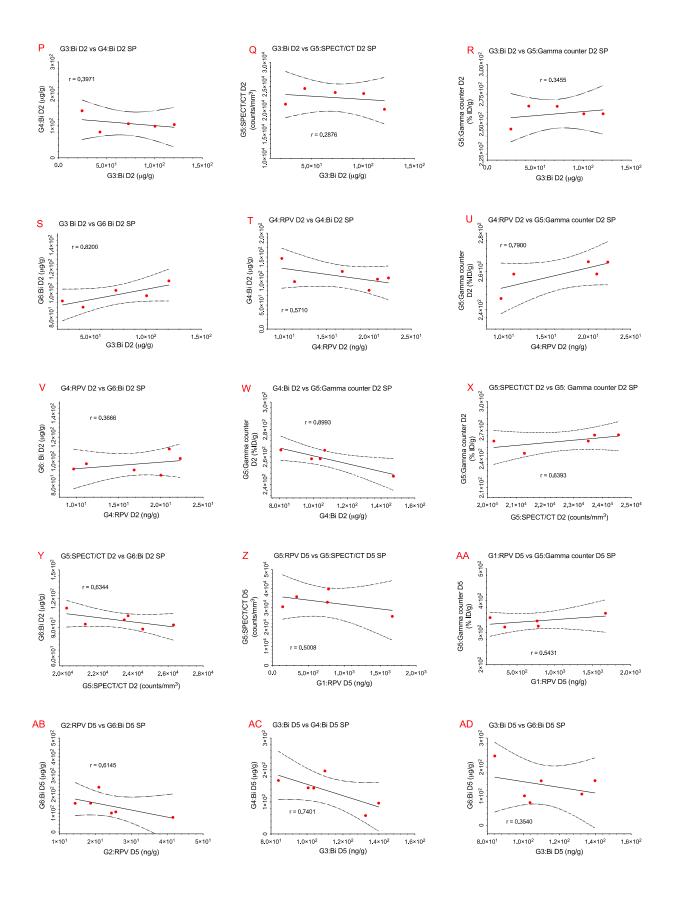
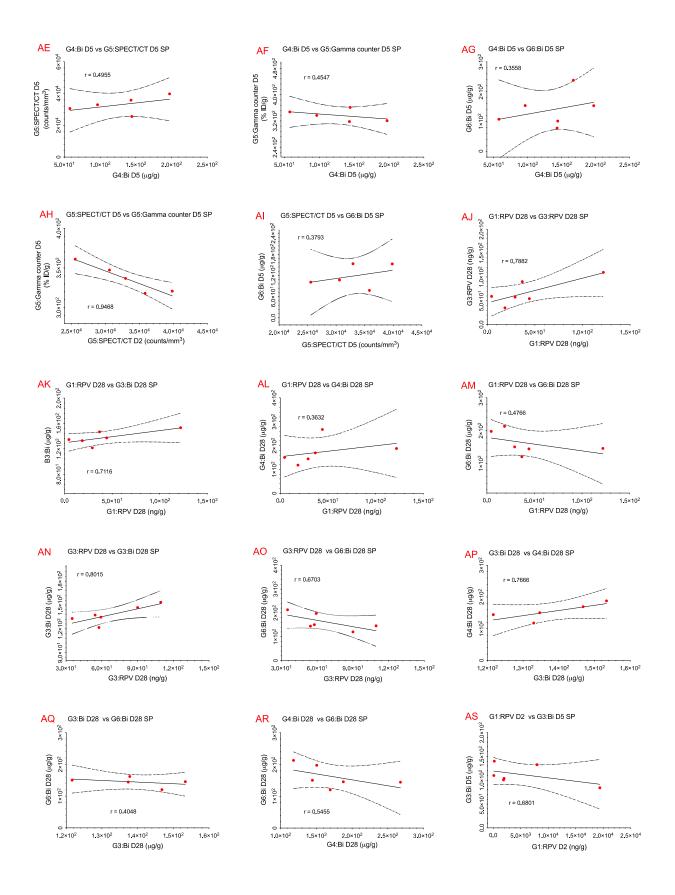
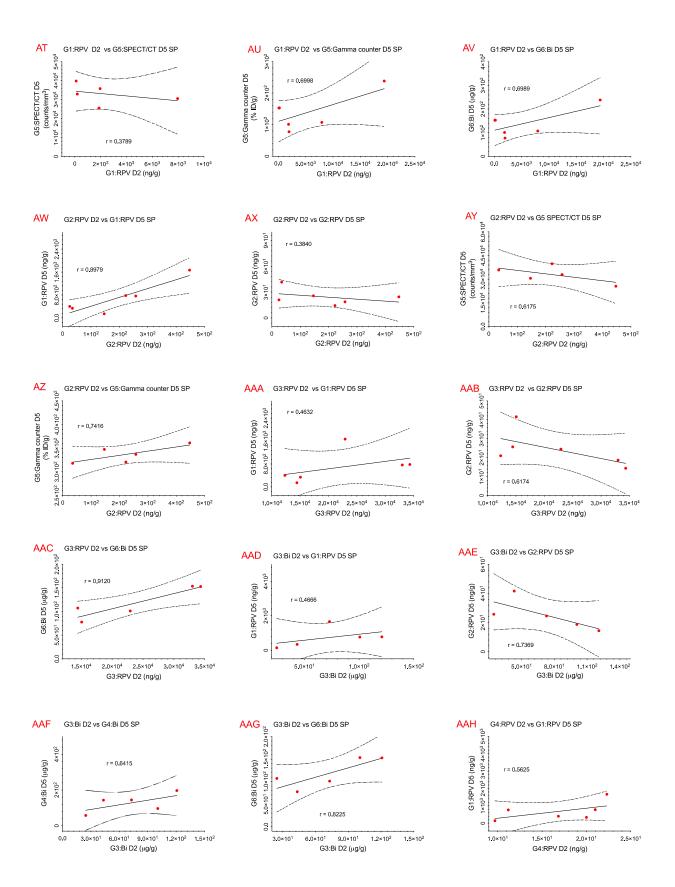


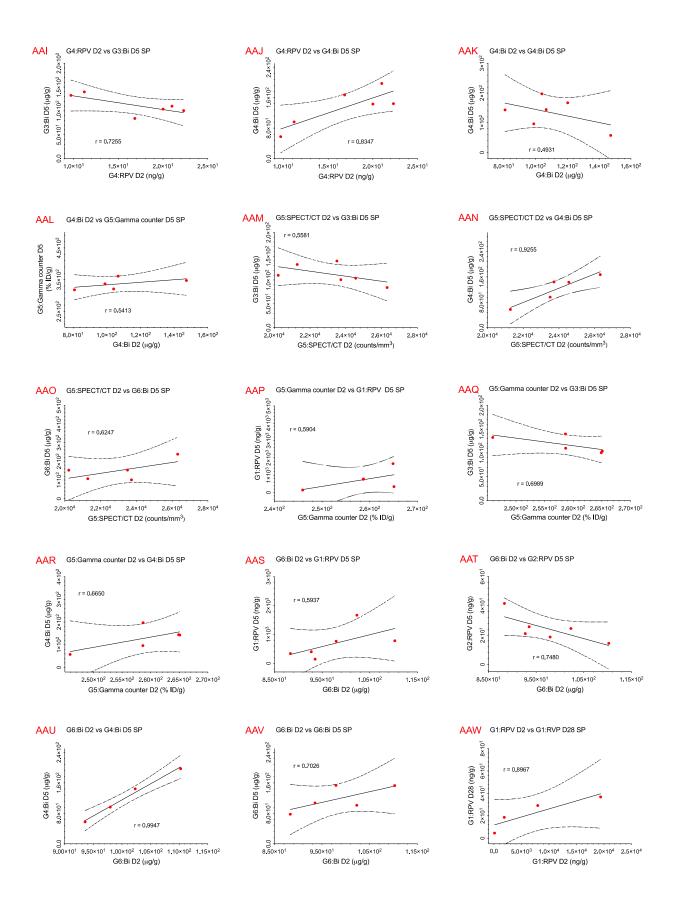
Figure S17. Pearson's correlation analysis of liver tissue. Correlation data were considered from bismuth content (ICP-MS), *ex-vivo* radioactivity (Gamma counter), radioactivity counts per volume of tissue (SPECT/CT ROI), and RPV drug content (UPLC-MS/MS) from days 2, 5 and 28. Only comparisons with a Pearson's coefficient greater than 0.5 (r > 0.5) are shown.

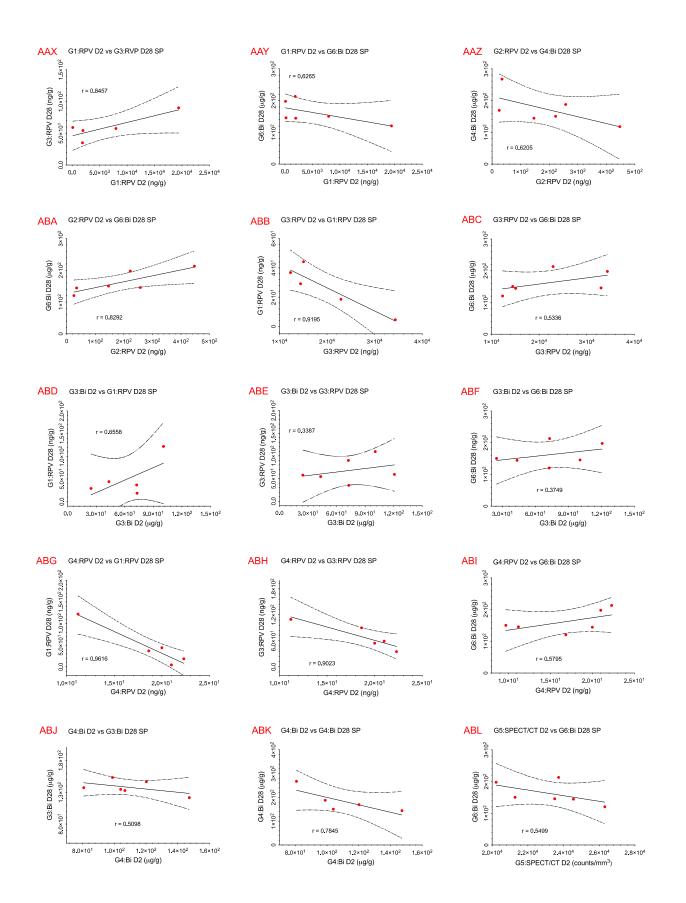


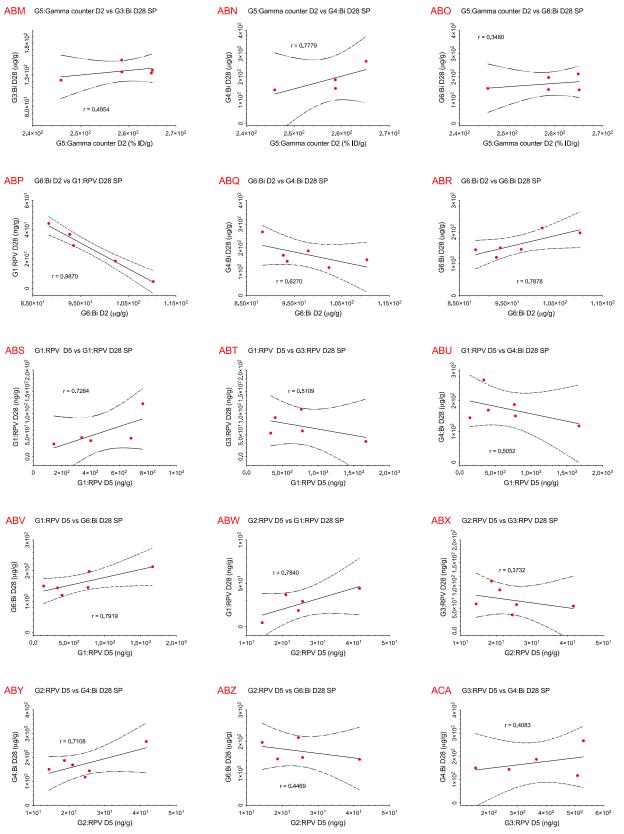












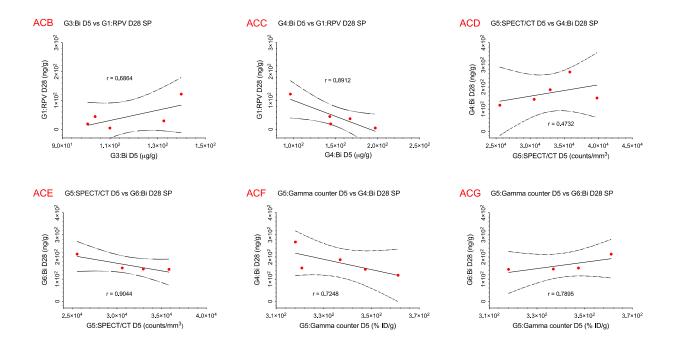
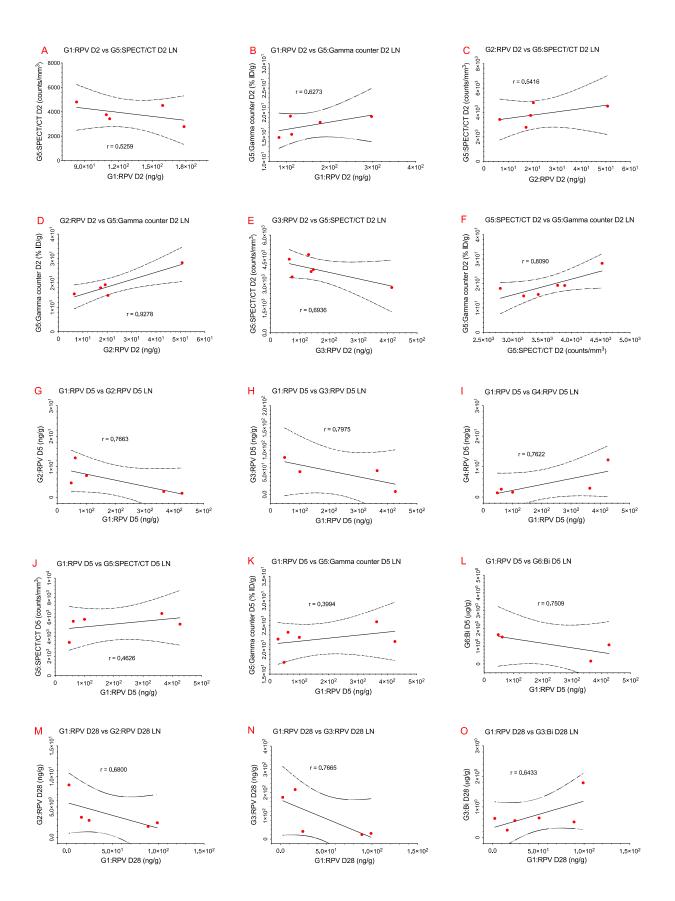
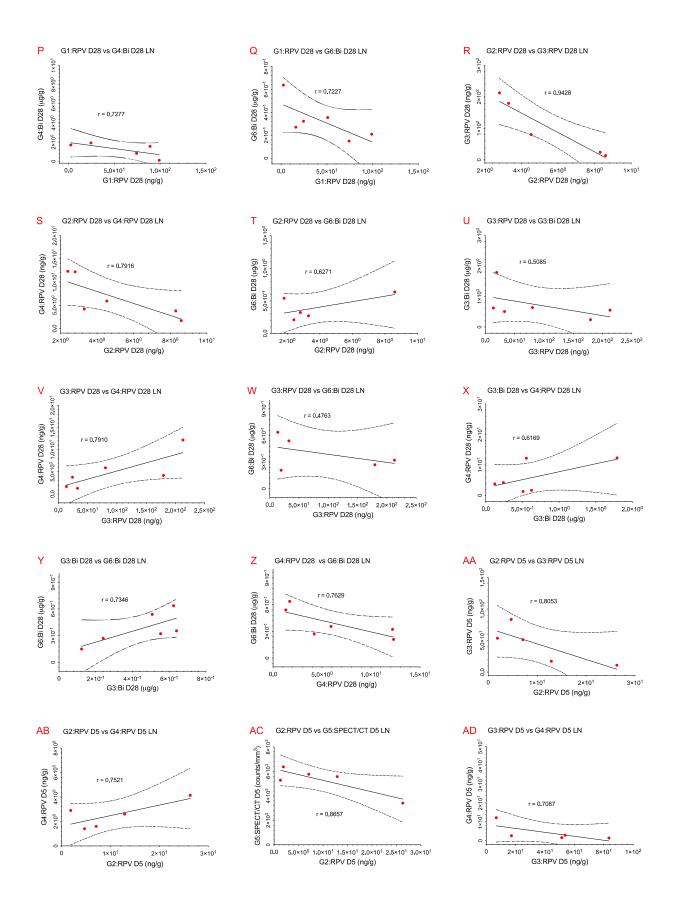
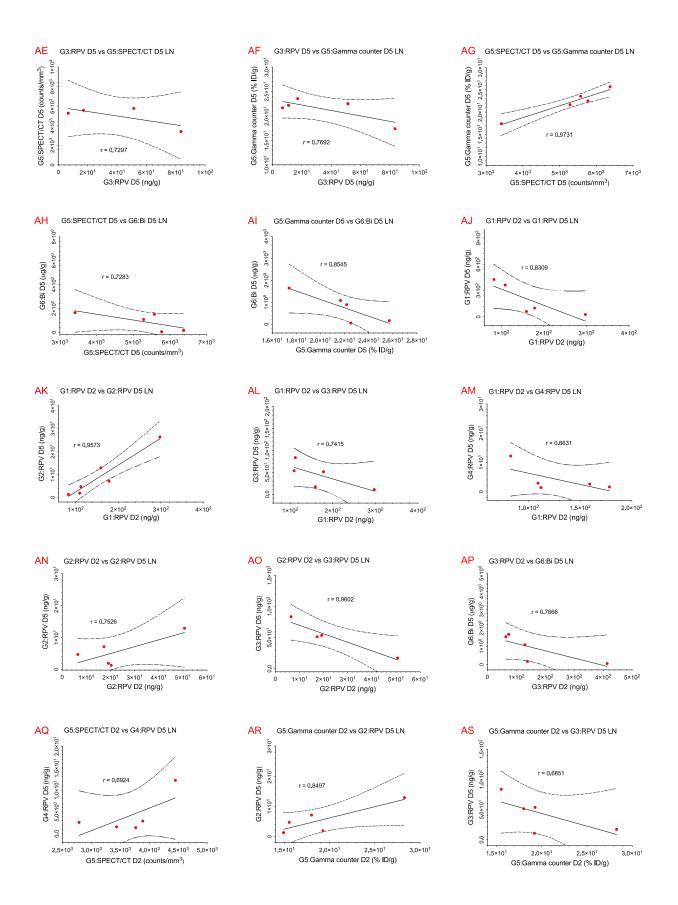
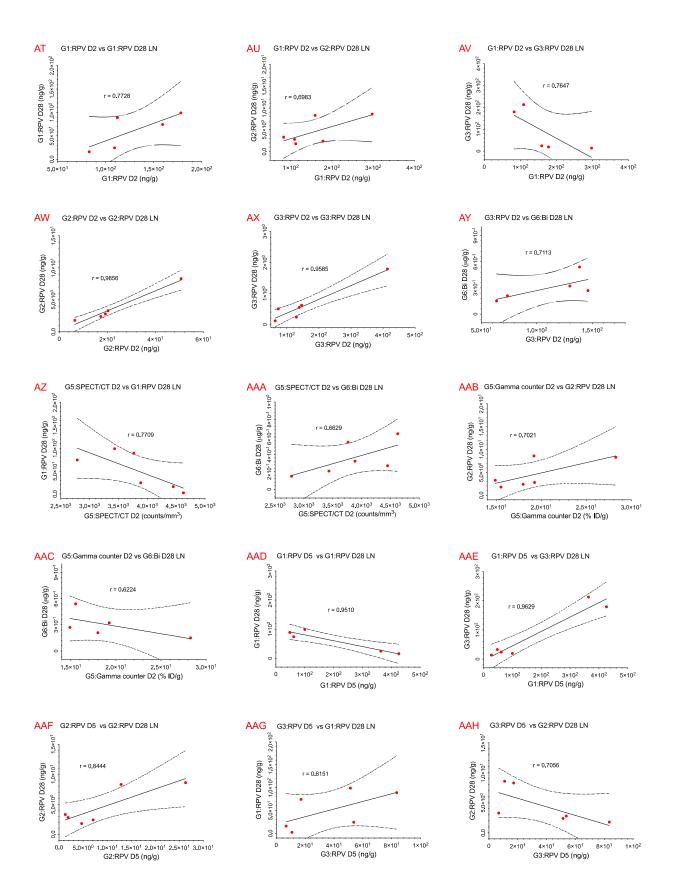


Figure S18. Pearson's correlation analysis of spleen tissue. Correlation data were considered from Bi content (ICP-MS), *ex-vivo* radioactivity (Gamma counter), radioactivity counts per volume of tissue (SPECT/CT, ROI), and RPV drug content (UPLC-MS/MS) from days 2, 5 and 28. Only comparisons with a Pearson's coefficient greater than 0.5 (r > 0.5) are shown.









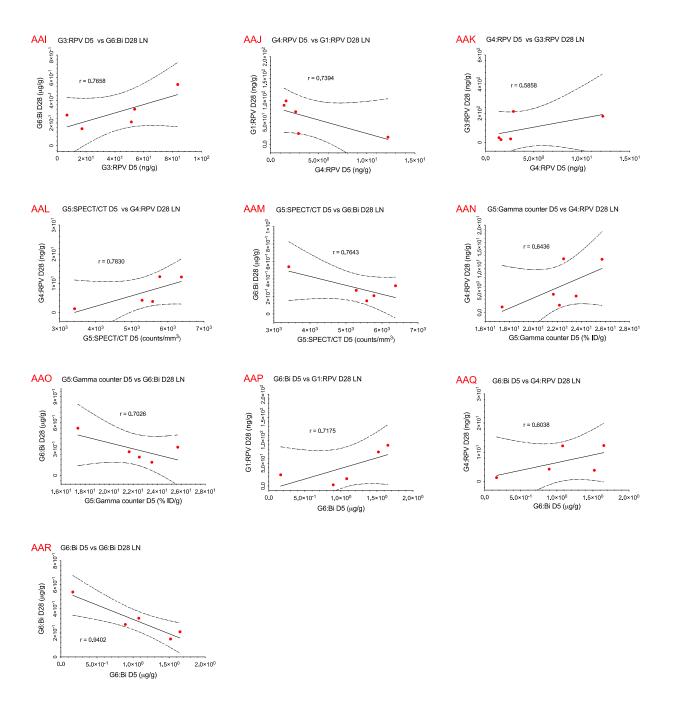


Figure S19. Pearson's correlation analysis of lymph node tissue. Comparison data sets were considered from Bi content (ICP-MS), ex-vivo radioactivity (Gamma counter), radioactivity counts per volume of tissue (SPECT/CT, ROI), and RPV drug content (UPLC-MS/MS) from days 2, 5 and 28. Only comparisons with a Pearson's coefficient greater than 0.5 (r > 0.5) are shown.

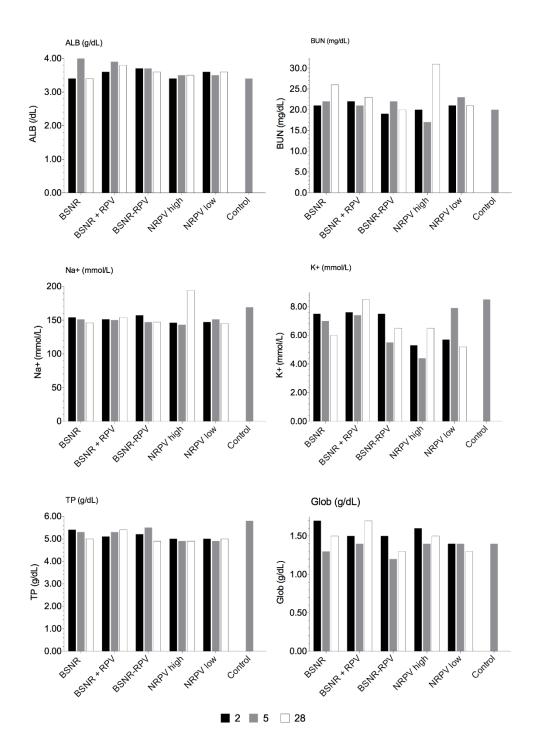


Figure S20. Serum chemistry analysis from treated mice. Comprehensive serum chemistry analysis of mice 2, 5, and 28 days post administration (IV) of NRPV, BSNRs-RPV and BSNRs particles. (ALB: albumin, BUN: blood urea nitrogen, Na+: sodium, K+: potassium, Glob: globulin TP: total protein)

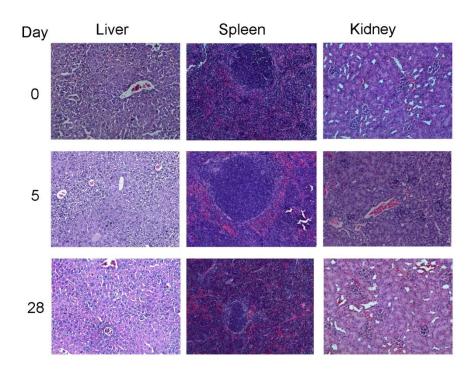


Figure S21. Tissue histopathology of hematoxylin and eosin stained liver, spleen and kidney sections.

References

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