

Supplemental Data

Manuscript title:

**Small heterodimer partner regulates circadian cytochromes p450
and drug-induced hepatotoxicity**

Authors:

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Materials and Methods

Materials

Acetaminophen (APAP), phenacetin, collagenase IV, mitoxantrone (MTX) and NADPH were purchased from Sigma-Aldrich (St. Louis, MO). NAC-APAP and Cys-APAP were purchased from Toronto Research Chemicals (North York, Ontario, Canada). Aminophylline, 1,3-dimethyluric acid (1,3-DMU), resorufin, coumarin, testosterone, tolbutamide, retinoic acid, alamethicin, 7-hydroxycoumain, reduced glutathione (GSH), arachidonic acid, and 4-nitrophenol were purchase from Aladdin (Shanghai, China). 15 α -hydroxytestosterone (15 α -OH testosterone) and 6 β -hydroxyltestosterone (6 β -OH testosterone) were obtained from Steraloids (Wilton, NH, USA). 20-hydroxy arachidonic acid (20-HETE) was purchased from MedChem Express (Monmouth Junction, NJ, USA). Aflatoxin B1 (AFB1) was purchased from Fermentek Ltd. (Jerusalem, Israel). Pentoxyresorufin was purchased from ApexBio Technology (Houston, TX, USA).

Plasmids

pRL-TK vector was purchased from Promega (Madison, WI). mCyp1a2 (2 kb)-Luc, mCyp2a5 (2 kb)-Luc, mCyp2a5 (1500)-Luc, mCyp2a5 (600)-Luc, mCyp2a5 (100)-Luc, mCyp2b10 (2.5 kb)-Luc, mCyp2b10 (2.3 kb)-Luc, mCyp2b10 (2 kb)-Luc, mCyp2b10 (600)-Luc, mCyp2c38 (2 kb)-Luc, mCyp2c38 (1200)-Luc, mCyp2c38 (700)-Luc, mCyp2c38 (80)-Luc, mCyp2e1(2 kb)-Luc, mCyp3a11 (2 kb)-Luc, mCyp3a11 (1650)-Luc, mCyp3a11 (1400)-Luc, mCyp3a11 (1000)-Luc, mCyp3a11 (400)-Luc, mCyp4a10(2 kb)-Luc, mCyp4a10(1500)-Luc, mCyp4a10(1350)-Luc, mCyp4a10(180)-Luc, mCyp4a14(2 kb)-Luc, mCyp4a14(1700)-Luc, mCyp4a14(700)-Luc, mCyp4a14(400)-Luc, mCyp4a14(200)-Luc, pcDNA-His-mNr1d1, pcDNA-mDec2, pcDNA-Flag-mE4bp4, pcDNA-mLrh-1, pcDNA-HA-mShp, pcDNA-mHnf1 α , pcDNA-mHnf4 α and siRNA were synthesized by Transsheep (Shanghai, China).

Table S1. siRNA sequences used in this study.

Gene	Forward (5'-3' sequence)	Reverse (3'-5' sequence)
siE4BP4	GCACAAGCUUCGGAUAAAATT	UUUAAUCCGAAGCUUGUGCTT
siRev-erb α	CUUCGUUGUUAACGUGAATT	UUCACGUUGAACACGAAGTT
siLrh-1	GCAGAAGAAAGCCUCAUUTT	AAUGAGGGCUUUCUUCUGCTT
siHnf4 α	CCUGCAGGUUUAGCCGACAAUGUGU	ACACAUUGUCGGCUAAACCGCAGG
siHnf1 α	CACCCAUUCUGAAAGAGCUGGAGAA	UUCUCCAGCUCUUCAGAAUGGGUG
siDec2	CAGCCUUAACUGAGCAGCAGCAUCA	UGAUGCUGCUGCUCAGUUAAGGCUG
Negative	UUCUCCGAACGUGUCACGUTT	ACGUGACACGUUCGGAGAATT

Table S2. Mouse primer sequences for quantitative real-time PCR (qPCR).

Gene	Gene bank no.	Forward (5'-3' sequence)	Reverse (3'-5' sequence)
Cyp1a1	NM_009992.4	GGTTAACCATGACCGGGAAC	TGCCCAAACCAAGAGAGTGA
Cyp1a2	NM_009993.3	AGTACATCTCCTTAGCCCCAG	GGTCCGGGTGGATTCTTCAG
Cyp1b1	NM_009994.1	TTCATCTCCATCCTCGCTCA	CAGCCTTCTCAGTTTGACG
Cyp2a4	NM_009997.2	AACGGTGCTTTCATTGACCC	CCTCATAGTCAAAGCGGTCCC
Cyp2a5	NM_007812.4	GGACAAAGAGTTCCTGTCACTGCTTC	GTGTTCCACTTTCTTGGTTATGAAGTCC
Cyp2a12	NM_133657.1	TTAGCTCCATTGTCTTCGGC	GGTACTTCATAACTGAATGG
Cyp2a22	NM_001101467.1	ATCAAGAGGGAGGGAGAGCC	GACCCTTTCTGGATGAGTGACA
Cyp2b9	NM_010000.2	GACTTTGGGATGGGAAAGAG	CCAAACACAATGGAGCAGAT
Cyp2b10	NM_009999.4	AGACATCCTTATTCTATGCC	CTGCATACTGGAGCACTTCATT
Cyp2b13	NM_007813.2	TTTTCTTCCAGTGTGTTACAGCC	AACGCAGGAAGTTCATCTG
Cyp2c29	NM_007815.3	TGGTCCACCCAAAAGAAATTGA	GCAGAGAGGCCAAATCCATTCA
Cyp2c37	NM_010001.2	CACGAGGCGTTTCTCACTCA	AGGGCTGCTCAGAATCTTTGT
Cyp2c38	NM_010002.3	CACTATGGAGACAGAGGTCTA	CCAAATACAGAGTGAAAACG
Cyp2c39	NM_010003.2	GGAGACAGAGCTGTGGC	TAAAAACAATGCCAAGGCCG
Cyp2c40	NM_010004.2	TCCGGTTTTTGACAAGGTTTCTAC	TGCCCAAGTTCCTCAAGGTATTC
Cyp2c44	NM_001001446.3	GGCTATGATGTGGTGAAGGAAGC	TCGCAGGAGTTTCCACCTCTCT
Cyp2c50	NM_001167877.1	ATGGAAAAACACAAGGCGCTTC	GCAACCAAGGGGTGCTCATT
Cyp2c54	NM_206537.2	TTGACCATGGGGATGTCTTT	ATCCAATGCCATTCCATTA
Cyp2c55	NM_028089.3	AATGATCTGGGGGTGATTTTCAG	GCGATCCTCGATGCTCCTC
Cyp2c65	NM_028191.2	ACAACCCAAAGACGGAGTTTAC	CTGTCCCAGCAGCAAACAGAT
Cyp2c66	NM_001011707.1	GGAGTTTGCTGGAAGAGGAACC	GCGTCTCATCTCTTCCAGGTC
Cyp2c69	NM_001104525.1	TGTAGTCTTGGTGCTTTGTCTG	ACAATAGGCTGTGAGCCAAAATA
Cyp2c70	NM_145499.2	AGTATGGCCCTGTGTTTACTGT	GCCTTGGCTGGTTCTACTGAG
Cyp2d9	NM_010006.2	AGTCTCTGGCTTAATTCCTGAGGTT	CGCAAGAGTATCGGGAATGC
Cyp2d10	NM_010005.3	TTCCGGTTGCCCCCTACCCC	TCAGCCTGCCCTGCCTACCA
Cyp2d11	NM_001104531.1	TCTCAGTGCCTGATGGACAG	CACAGAGCTGGTAGGGGAAG
Cyp2d22	NM_001163472.1	CAGTGGTTGTAATAATGGGCT	GCTAGGACTATACCTTGAGAGCG
Cyp2d26	NM_029562.2	TTCAAAAGCCTGGAAGCAGT	TCCACCAGAAGCAGGAAGAT
Cyp2d40	NM_023623.2	GTCGAGCATGTCTGGGTGAG	CCATCCGGTACTGAGAAGCTA
Cyp2e1	NM_021282.2	CGTTGCCCTTGCTGTCTGGA	AAGAAAGGAATTGGGAAAGGTCC
Cyp2f2	NM_007817.2	TGCGTCACGCCTTCCTTATT	ACATCTGTGCCCTTGGGTATC
Cyp2g1	NM_013809.1	AGGAGGAGGCTGGCTACC	TCAGGTATAAGGTGGGATCTATGG
Cyp2j5	NM_010007.4	CAGACATGGAAGGAGCAAAGG	GAATGCGCTCCTCCAAGCT
Cyp2j6	NM_010008.4	ATGCTCGCTGCTACCGGCTC	GTGCTTCTTTGATTAAGGGC
Cyp2r1	NM_177382.4	TGGTGAGGTAAATGAGGCTTTC	TGCCAGTGCTCCAGTCTTC
Cyp2s1	NM_028775.3	GGTCACGGAGTTTTCTTTGCC	ATAGGGCAAACGGATGCCAA
Cyp2u1	NM_027816.3	GGCTGAAGTGTTCAAGTACC	CGTATGCAAACCTCCTCGATG

Gene	Gene bank no.	Forward (5'-3' sequence)	Reverse (3'-5' sequence)
Cyp3a11	NM_007818.3	CGCCTCTCCTTGCTGTCA	CTTTGCCTTCTGCCTCAAGT
Cyp3a13	NM_007819.4	CCTCTGCCTTTCTTGGGGACGAT	CCGCCGGTTTGTGAAGGTAGAGT
Cyp3a16	NM_007820.2	CTGAAATTAAGCAGAGA	ATCACGCTCCAGTTATGA
Cyp3a25	NM_019792.2	CTTCACTGTCCAGCCTTGTGAA	AATTGGTTCCTGCTGATCTTC
Cyp3a41	NM_017396.3	GTGGAGAAAGCCAAAGGGATT	GAAGACCAAAGGATCAAAAAAGTCA
Cyp3a59	NM_001105160.1	ACTACTTTGATGGCATGTGGA	GCCCAAGAGTCGGACGG
Cyp4a10	NM_010011.3	TTCCCTGATGGACGCTCTTTA	GCAAACCTGGAAGGGTCAAAC
Cyp4a12a/b	NM_172306.2	GGGGAGATCAGACCCAAAAGC	ATTCGTCGGTGTGAAACCAT
Cyp4a14	NM_007822.2	CCCAAAGGTATCACAGCCACAA	CAGCAATTCAAAGCGGAGCAG
Cyp4a31	NM_201640.3	AGCAGTCCCATCACCGCCC	TGCTGGAACCATGACTGTCCGTTT
Cyp4a32	NM_001100181.1	AAGGCGAACAAAGAACTAA	GCAAACCATACCCAATCC
Cyp4b1	NM_007823.2	CTGCATGGCCCTTTATCCTA	GAAGCACTCCTTCATGCACA
Cyp4f13	NM_130882.1	CGACAGCAACTGCCAAGAGT	CAGTGAGATAGTACAGGAGGTCC
Cyp4f14	NM_001357788.1	ACTGGCTTATGGGTCACGTG	ACCCACCAAACGAGTCAATTC
Cyp4f15	NM_134127.1	CCCCCAAATGGAATTGGTTCT	ACCAGTTAGTCACCTCCTTC
Cyp4f16	NM_024442.1	CTGCGGCTAAGTGTGTCTGGGC	TGGAAAGTCTGGCCATTTGCGTA
Cyp4f17	NM_001101445.1	TGACACCTGCATTCCACTTTG	GCAGTTCCTTGGTGGTACG
Cyp4f18	NM_024444.2	TGTTTTGAGAACTCTCTCGCC	TGGAATATACGGATGACTGGG
Cyp4v3	NM_133969.2	TCCGAGTTTTCCCATCTGTC	CGGTGTAGTGCCTAGGGAAT
Cyp4x1	NM_001003947.1	GCCGTGAAGCTCTACCTGAG	GGTCTTTGGGTCTGTTCTGC
Cyp7a1	NM_007824.2	AGCAACTAAACAACCTGCCAGTACTA	GTCCGGATATTCAAGGATGCA
Cyp7b1	NM_007825.4	AATTGGACAGCTTGGTCTGC	TTCTCGGATGATGCTGGAGT
Cyp8a1	NM_008968.3	ATGCCATCAACAGCATCAAA	AAACTCAGGAACCTCTGTGTCC
Cyp8b1	NM_010012.3	GCCTTCAAGTATGATCGGTTCCCT	GATCTTCTTGCCCGACTTGTAGA
Cyp17a1	NM_007809.3	CATCCCACACAAGGCTAACA	CAGTGCCAGAGATTGATGA
Cyp20a1	NM_030013.3	CTGGACTTCGCCATCTTCG	CCGGGTAGAGGTACAGCAC
Cyp21a1	NM_009995.2	AGGAATTCTCCTTCTCACTTGT	TCTGTACCAACGTGCTGTCC
Cyp26a1	NM_007811.2	CGAGAAGAGATAAAGAGCAAG	TCAGAGCAACCCGAAACC
Cyp26b1	NM_175475.3	GCAAGATCCTACTGGGCGAAC	TTGGGCAGGTAGCTCTCAAGT
Cyp27a1	NM_024264.5	GTGGACAACCTCCTTTGGGA	TTGCTCTCCTTGTGCGATGAA
Cyp39a1	NM_018887.4	ATCCAGCCACACTCAATACCC	GGAGCCATACTCAAAGCCCTC
Cyp46a1	NM_010010.1	ATGCCAGGGAAGAGAAAGC	CTTTGAGGATCTGCGTGAG
Cyp51a1	NM_020010.2	GTCTACCTGTTCCGTCTCGC	TCATATGCAACTCCCTTCCC
Ugt1a1	NM_201645.2	GCTTCTCCGTACCTTCTGTTG	GCTGCTGAATAACTCCAAGCAT
Ugt1a2	NM_013701.3	ATGGACACGGGACTATGTGTG	CATGGGTAACACCAGCACTTTT
Ugt1a5	NM_201643.2	TGAGAAGGTGCTAGTGTTCCT	GGGAACGGCATAGACTTTGAA
Ugt1a6a	NM_145079.3	GTTTCTCTTCTAGTGCTTTGGG	CCTCGTTCCTGAGATGTTCTAC
Ugt1a9	NM_201644.2	TTTCGATGTGTGCGGCTTAAC	GGTTCGAGTCTTTCCCTTGAA
Ugt2a1	NM_053184.2	TTGAGGAGTTTCATTTGGTGTCC	GGGAAAACCGCAAGGAATAGATA
Ugt2a2	NM_001024148.1	TGTGGTAGTTTGGCCTACAGA	TTGTGTACGAAACCGGAATCTC

Gene	Gene bank no.	Forward (5'-3' sequence)	Reverse (3'-5' sequence)
Ugt2a3	NM_028094.3	GATGTTGAGTTTCCTCGTCCAT	GCCTGAGTATCTCCACAAAACCT
Ugt2b1	NM_152811.1	TTGATGTCGTTCTAGCAGATGC	TCAAAAAGCAACACCTGCAAC
Ugt2b34	NM_153598.2	TGAAGTGATGGTTCTGAGACCT	ACTGCTTTGGCAGCTCATAAAT
Ugt2b35	NM_172881.3	CTTGTACAGAGGGGCCATGAA	TGGGACCTCAAAGTCCATTC
Ugt2b36	NM_001029867.1	CTACAAGTGGCTTCCCCAAAA	ATGGATCGCCTCGTAGAGTCC
Ugt2b37	NM_053215.3	ACAGCTTTTGAATGTGTGGACT	GCGATTAGCTCCCCACCAA
Ugt2b38	NM_133894.2	TGCGCCACAAAAGGGCTAA	ACACAAGAGAGTAGGAAGCCG
Ugt2b5	NM_009467.3	TGTCTATGAGGCCATCTATC	ATATTCAGTGTAAACAGCTGC
Ugt3a1	NM_207216.2	ATGGCTGCACATCGGAGTTG	AGAATTTGCGACACACGGTTC
Ugt3a2	NM_144845.3	TGATAAGCCGTGTGTCGCA	AGTCGATGCCTAAGATCAGCA
Ugt8a	NM_011674.5	ACTCCATATTTTCATGCTCCTGTG	AGGCCGATGCTAGTGTCTTGA
Sult1a1	NM_133670.1	CACAAGGGTCTCTCCTTAGC	TGACAGCGGAACGTGAAGTC
Sult1b1	NM_001356943.1	GAGGTGCTCCATCTCAGGTCA	GCAGGCTTCAATCAGAACGA
Sult1c2	NM_026935.4	GGCGAACCATCATTCAACACC	AGGAAACCATGCAGTCTTTAGC
Sult1d1	NM_016771.3	GGGATGCAATCTACAAACGAGT	CCACATCTTTGGCATTCCGTG
Sult1e1	NM_023135.2	ATGGAGACTTCTATGCCTGAGT	ACACAACCTCACTAATCCAGGTG
Sult2a1	NM_001111296.2	CCAGGGTCACTCGGAACTTA	GCTCAAACCATGATCCGAAT
Sult2a7	NM_001184981.1	TTGTTATGAGAGAAGACACAGTCGT	GCAGACAATCTCAACAAGCCAG
Sult2a8	NM_175250.5	GACCTTACTCCATGTAAAAGCCTAT	AGTTGCTGAAGCAGGGTAATTGAG
Sult5a1	NM_020564.3	ATGACTGAGCGCATGAACACC	CCACAAGTGACCCTCACAGA
Sult6b1	NM_001163625.1	AACTTTCCGAGCTTTGGATGC	TTGAACCACACTTCGGGTAGG
Lxra	NM_001177730.1	AGGAGTGTGACTTCGCAAA	CTCTTCTTGCCGCTTCAGTTT
Ppara	NM_013839.4	AGGCCGTTGCCACTGTTGAG	AGCCCTCTTCATCCCCAAGC
Vdr	NM_009504.4	GCATCCAAAAGGTCATCGGC	AGCGCAACATGATCACCTCA
Ahr	NM_013464.4	AGCCGGTGCAGAAAACAGTAA	AGGCGGTCTAACTCTGTGTTC
Car	NM_009803.5	CCCTGACAGACCCGGAGTTA	GCCGAGACTGTTGTTCCATAAT
Pxr	NM_010936.3	GAGATCTCCATGTGCAGCCA	CTCCAGCTCTCCTCAGGTCT
Nrf2	NM_010902.4	TAGATGACCATGAGTCGCTTGC	GCCAAACTTGCTCCATGTCC
Dec2	NM_024469.2	CCCTCATTTGCAAGAGAGACAG	AGGTATCCTTGGTATCGTCTCG
E4bp4	NM_017373.3	CTTTCAGGACTACCAGACATCCAA	GATGCAACTTCCGGCTACCA
Shp	NM_011850.3	CGATCCTCTTCAACCCAGATG	AGGGCTCCAAGACTTCACACA
Rev-erba	NM_145434.4	TTTTTCGCCGGAGCATCCAA	ATCTCGGCAAGCATCCGTTG
Lrh-1	NM_030676.3	GAACTGTCCAAAACCAAAAAGG	CTTCCAGCTTCATCCCAAC
Hnf4a	NM_008261.3	GACCATGGGCAATGACACGTCC	TTGAGCCGTCACAGCTCGA
Cyclophilin b	NM_011149.2	TCCACACCCTTTTCCGGTCC	CAAAGGAAGACGACGGAGC

Table S3. The probe substrates, monitored metabolites, and incubation conditions for the measurements of enzymatic activities.

P450 isoform	Substrate	Concentration (μM)	Incubation time (h)	Metabolite
Cyp1a2	Phenacetin	50	2	Acetaminophen
Cyp2a4	Testosterone	100	2	15 α -OH Testosterone
Cyp2a5	Coumarin	5	2	7-OH Coumarin
Cyp2b10	Pentoxeresorufin	10	1	resorufin
Cyp2c39	Retinoic Acid	100	2	4-OH Retinoic acid
Cyp2e1	4-Nitrophenol	50	2	4-OH Nitrophenol
Cyp3a11	Testosterone	100	2	6 β -OH Testosterone
Cyp4a	Arachidonic acid	100	2	20-HETE

Table S4. Chromatographic and mass spectrometric parameters for quantification of chemicals in this study.

Analyte	Retention time (min)	[M+H] ⁺	[M-H] ⁻	Analytical column	Analytical gradient
Phenacetin	2.45	180.101		Luna Omega polar C18 1.6 μm, 100 × 2.1 mm	10% B at 0-1.0min, 10-90% B at 1.0-3.5 min, 90% B 3.5min, and 90-10 % B at 4.5-5.0 min.
APAP	2.61	152.070			
APAP-Cys	2.70	271.075			
APAP-NAC	2.72	313.081			
APAP-G	2.10	328.100			
APAP-S	2.30	232.145			
Aflatoxin B1	3.54	313.216			
Coumarin	3.19	147.056			
7-OH Coumarin	2.75	163.053			
Theophylline	2.41	181.245			
1,3-DMU	1.93	197.234			
Mitoxantrone	2.96	445.424			
Testosterone	4.75	289.212			
6β-	2.37	305.212			
15α-	2.73	305.212			
Retinoic acid	3.74	301.139		ACE Ultracore 2 .5 Supper C18, 50 × 2.1 mm	10% B at 0-1.0min, 10-90% B at 1.0-3.5 min, 90% B 3.5min, and 90-10 % B at 4.5-5.0 min.
4-OH retinoic acid	2.89	317.140			
Arachidonic acid	4.22	303.234			
19/20-HETE	2.89	319.140			
4-Nitrophenol	2.74		138.020	ACE Excel 2 C18-PFP, 50 × 2.1 mm	
4-nitrocatechol	2.65		154.100		
Resorufin					λ _{ex} = 530 nm, λ _{em} = 580 nm

Table S5. Oligonucleotide sequences for EMSA assays.

Oligonucleotide	Forward (5'-3' sequence)
Cyp2a5-E4bp4RE (-927/-901)	CAGTAGGGCTTTTGTAATTAGAACTT
Cyp2b10-RevRE (-1963/-1937)	ATTATGAGTTCAAAGTCAGCCTGGGAT
Cyp2c38-Lrh-1RE (-1066/-1040)	GTGCATGGTTCAAGGTCAAGAAATTGT
Cyp2c38-Hnf4 α RE (-143/-117)	CTATCAAGGAACAAAGTTTATTTGAAT
Cyp4a10-RevRE (-1105/-1078)	AGAGTTCCTTGGTCATGGTGTGTTGCT
Cyp4a14-RevRE (-1715/-1688)	GCTTACCATAATATGGGTCAAACAAA

Table S6. Primer sequences for CHIP assays.

Gene	Forward (5'-3' sequence)	Reverse (3'-5' sequence)
Cyp2a5_E4BP4	AATAACAGGAGCAATGAG	GACAACATCTGAGGGTCT
Cyp2a5_Distal	AAAGGCAGATTGAAGTTTAG	TTCTCCTGATAGTAATGGT
Cyp3a11_Hnf1 α	ATCTTGGTGGGTATGTGG	TGTGGCCTTGAAGTCTAA
Cyp3a11_Distal	CCAGGTGGAACAGAAGGG	GCAATGGTGTGACGCTTT
Cyp2b10_Rev-erba	CACATCTGTGGTCCCAGTA	TCCCAGGTGTGAGGATTTTCAG
Cyp2b10_Distal	GAAGTTCTGCTGTGGGTC	GTAACATTACTATCTAGCCTCT
Cyp2c38_Lrh-1	ATTGACATTCTAAGCATTAC	TTCTTGACCTTGAACCAT
Cyp2c38_Hnf4 α	GTAGCAATACTTGGACAT	TGTTGAGTAACCCATTAG
Cyp2c38_Distal	GAATACCAAATGGCAGAG	CACCAACAATGGAGGAGT
Cyp7a1_Lrh-1/Hnf4 α	GCTTATCGACTATTGCAGCTCTCT	CTGGCCTTGAACCTAAGTCCATCT
Cyp4a10_Rev-erba	TGTAAGCCAGGCCCAATT	GGTGCCCATGTATTTCTC
Cyp4a10_Distal	TGTAAGGGTGGCATAAAG	TAATGGGACATGGTCTGG

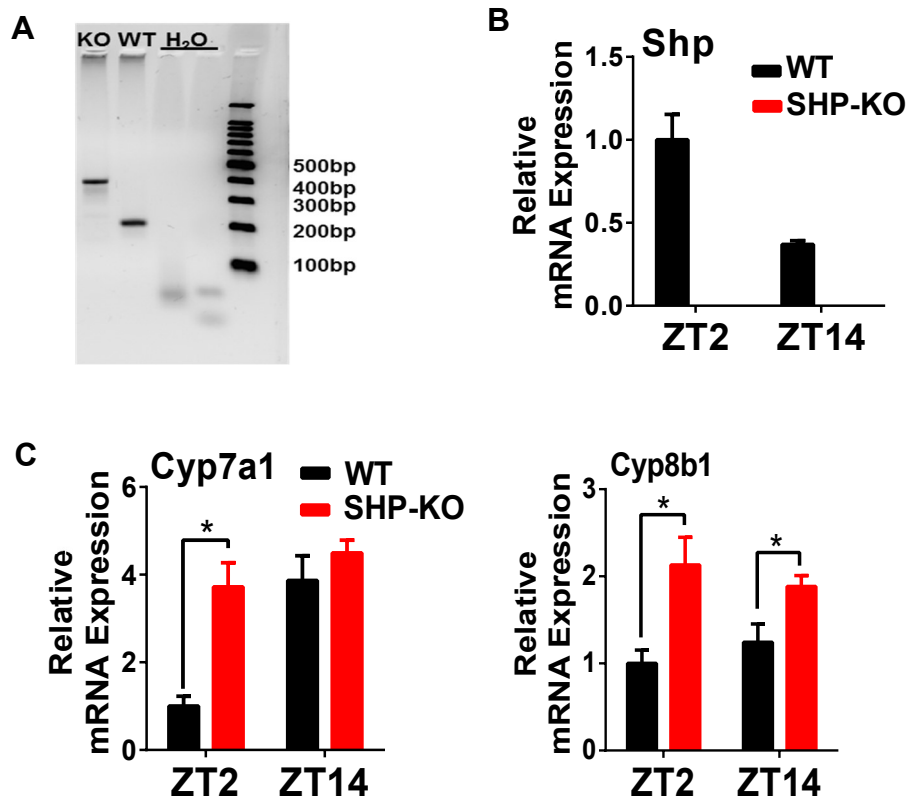


Figure S1. (A) Genotypes of SHP-knockout (SHP-KO) and wild-type (WT) mice were identified using genomic DNAs prepared from mouse tail. Expression of hepatic SHP (B), Cyp7a1 and Cyp8b1 (C) at ZT2 and ZT14 in WT and SHP-KO mice. mRNA levels were measured by qPCR and normalized to cyclophilin b. Data are mean \pm SD from $n = 5$ mice for each time point, * $p < 0.05$ versus WT group.

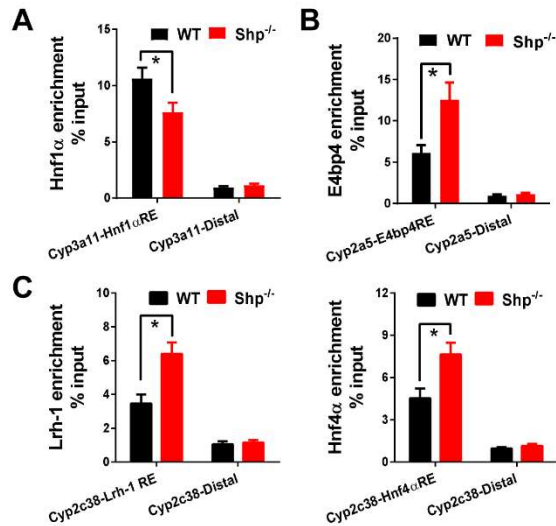


Figure S2. (A) ChIP assays showing the relative enrichment of Hnf1 α to the promoter region of Cyp3a11 in the livers of WT and SHP^{-/-} mice. (B) ChIP assays showing the relative enrichment of E4bp4 to the promoter region of Cyp2a5 in the livers of WT and SHP^{-/-} mice. (C) ChIP assays showing the relative enrichment of Lrh-1 and Hnf4 α to the promoter region of Cyp2c38 in the livers of WT and SHP^{-/-} mice. Data are presented as mean \pm SD (n = 5). *p < 0.05 versus IgG.

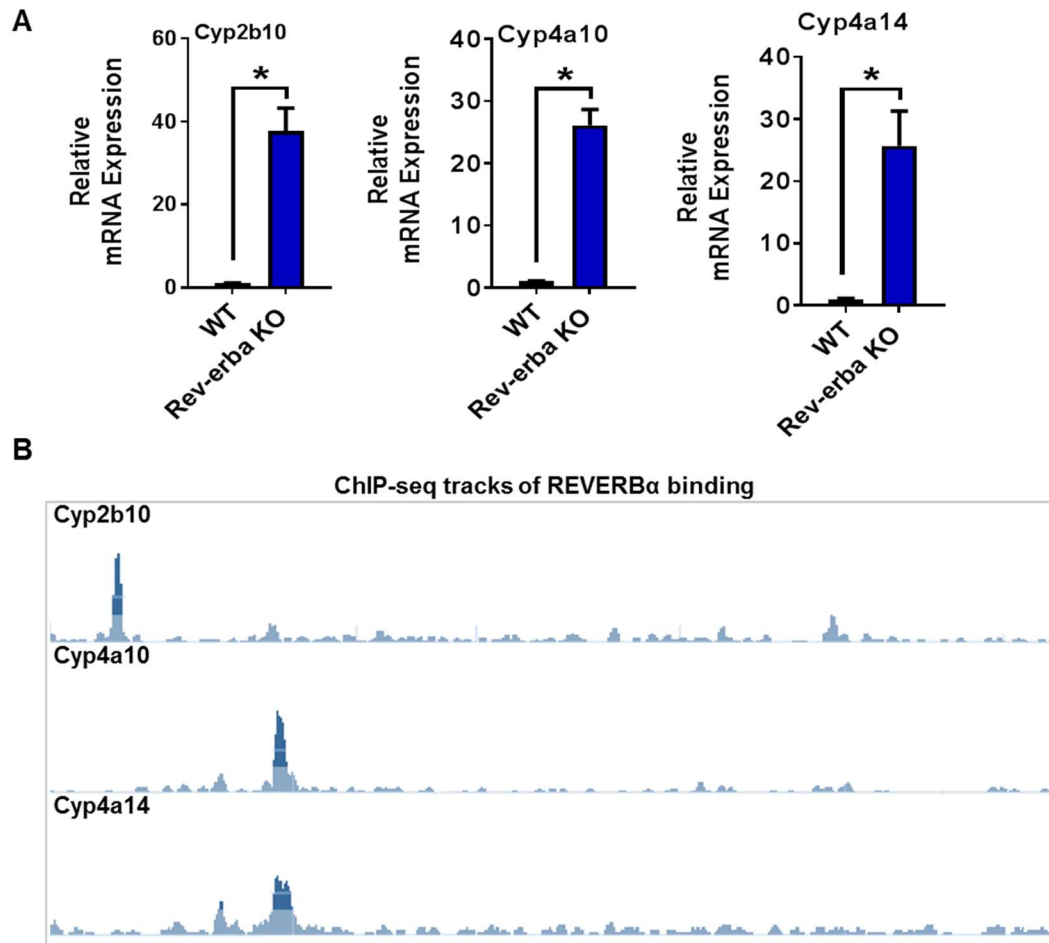


Figure S3. Rev-erba α regulates Cyp2b10, Cyp4a10 and Cyp4a14. (A) Cyp2b10, Cyp4a10 and Cyp4a14 mRNA expression in the livers from WT and Rev-erba KO mice. (B) Representative ChIP-seq tracks of REVERB α binding on Cyp2b10, Cyp4a10 and Cyp4a14 genes. The data were retrieved from the gene expression omnibus database (GSE59486 and GSE67973)

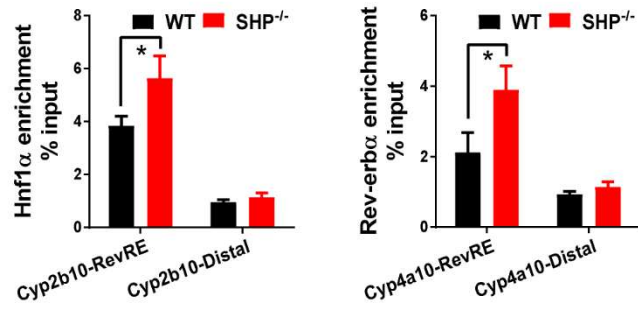


Figure S4. ChIP assays showing the relative enrichment of Rev-erba to the promoter region of Cyp2b10 and Cyp4a10 in liver of WT and SHP^{-/-} mice. Data are presented as mean \pm SD ($n = 5$). * $p < 0.05$ versus IgG.

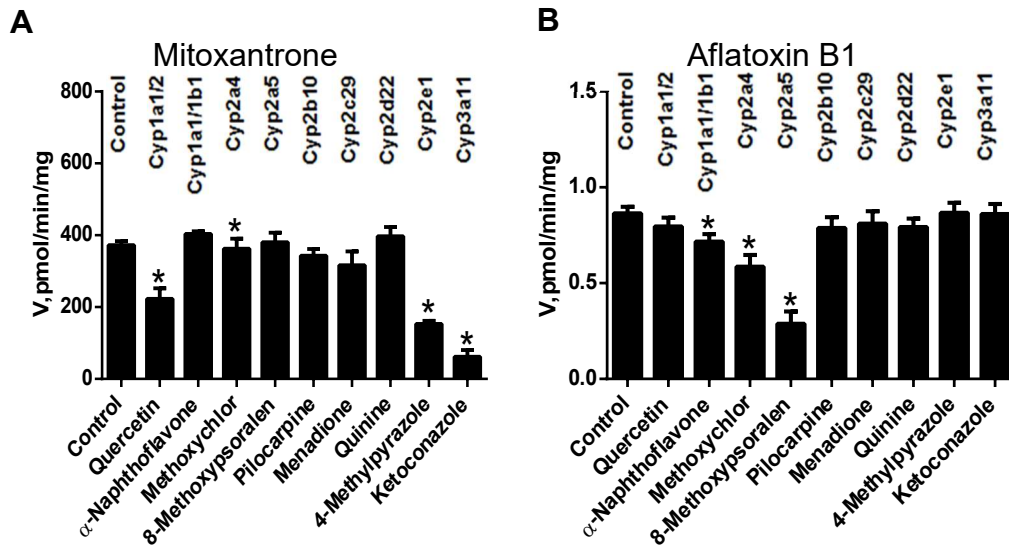


Figure S5. The effects of CYP enzyme inhibitors on mitoxantrone (**A**) and aflatoxin B1 (**B**) metabolism. The metabolic rates of mitoxantrone and aflatoxin B1 were measured after incubation for 2 h with mouse liver microsomes (1 mg/ml) at 37 °C with DMSO (control), quercetin (2 μM), α-naphthoflavone (10 μM), methoxychlor (2 μM), 8-methoxypsoralen (5 μM), pilocarpine (10 μM), menadione (10 μM), quinine (10 μM), 4-methylpyrazole (20 μM), or ketoconazole (2 μM). Data are presented as mean ± SD ($n = 5$). * $p < 0.05$ versus control.

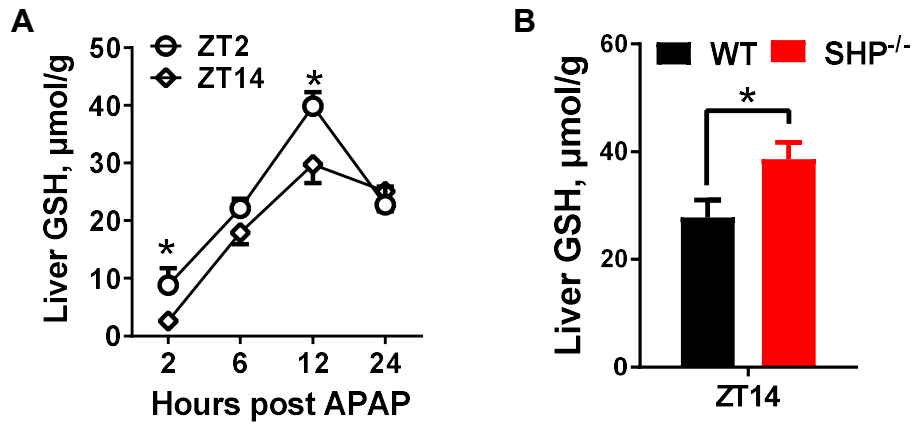


Figure S6. (A) Liver levels of GSH determined at various time points after APAP administration (500 mg/kg, i.p.) at ZT2 and ZT14. Data are presented as mean \pm SD ($n = 5$). * $p < 0.05$ versus ZT14. **(B)** Liver GSH levels in WT and SHP^{-/-} mice 12 h after APAP administration (500 mg/kg, i.p.) at ZT14. Data are presented as mean \pm SD ($n = 5$). * $p < 0.05$.

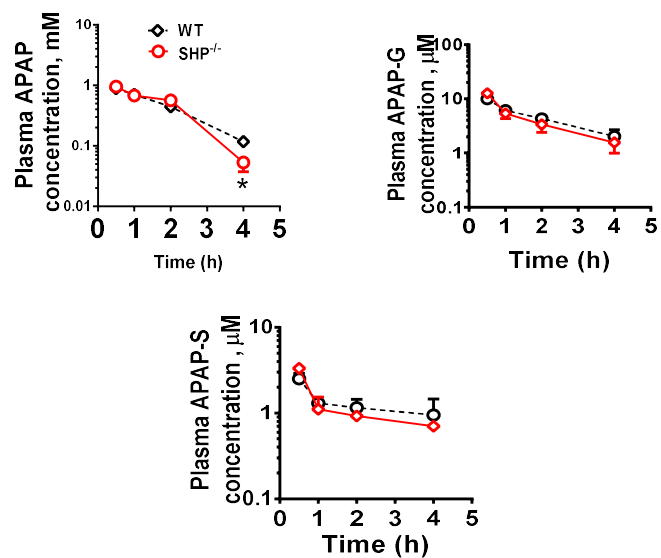


Figure S7. Plasma concentrations of acetaminophen (APAP), acetaminophen-O-glucuronide (APAP-G) and acetaminophen-O-sulfate (APAP-S) in WT and SHP^{-/-} mice at 0.5, 1, 2, and 4 h after APAP treatment (500 mg/kg, i.p.). Data are presented as mean \pm SD ($n = 3$).

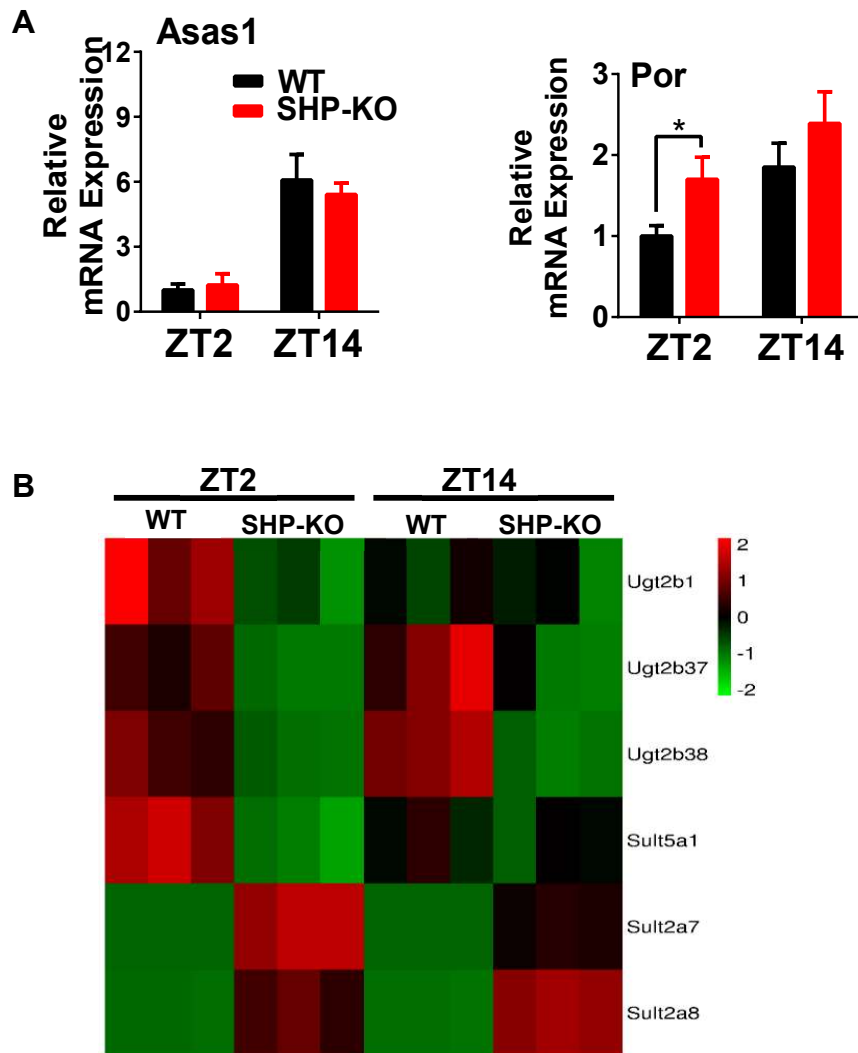


Figure S8. (A) Expression of hepatic *Alas1* and *Por* at ZT2 and ZT14 in WT and SHP-KO mice. **(B)** Heatmap of relative mRNA levels of UGT and SULT at ZT2 and ZT14 in WT and SHP-KO mice (Fold change >2). mRNA levels were measured by qPCR and normalized to cyclophilin b. Data are mean \pm SD from $n = 5$ mice for each time point, * $p < 0.05$ versus WT group.

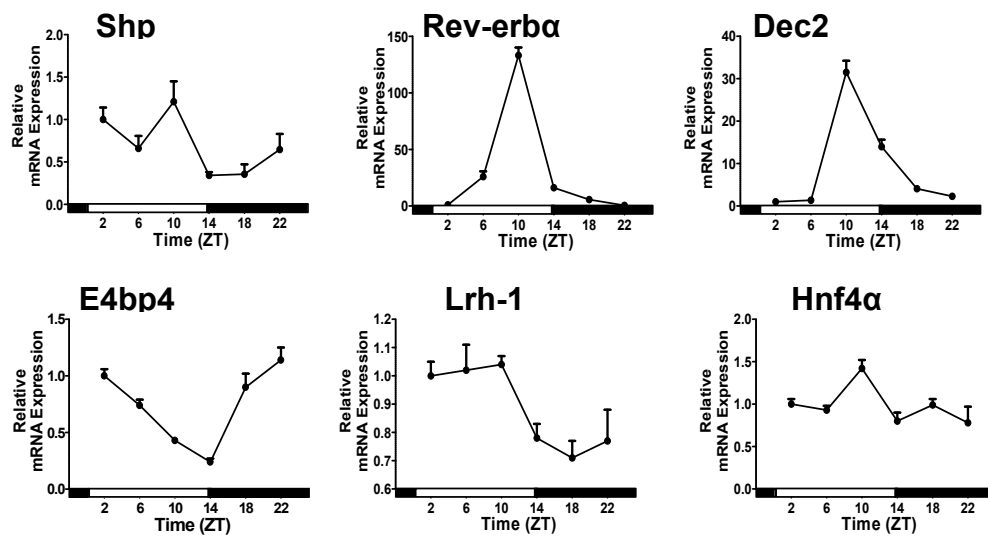


FIGURE S9. Circadian expression of Shp, Rev-Erb α , Dec2, E4bp4, Lrh-1 and Hnf4 α in WT mice. mRNA levels were measured by qPCR and normalized to *cyclophilin b*. Data are mean \pm SD ($n = 5$).

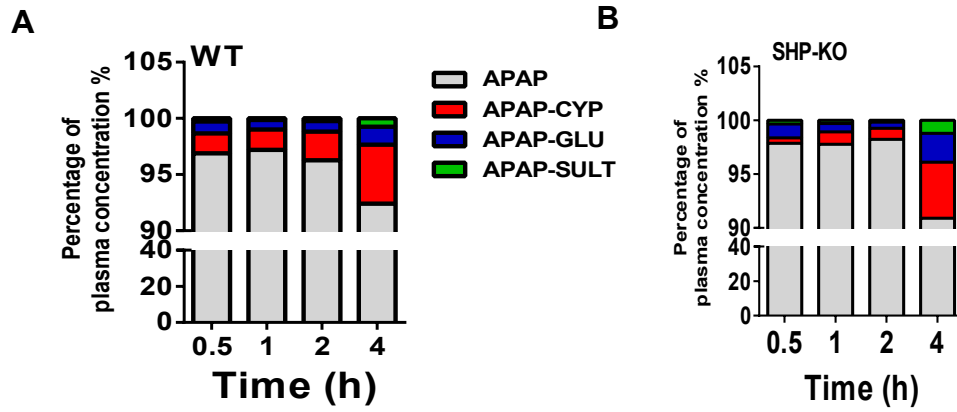


Figure S10. Percentages of APAP, APAP-CYP (APAP-Cys and APAP-NAC), APAP-G and APAP-S in WT **(A)** and SHP-KO mice **(B)** plasma at 0.5, 1, 2, and 4 h after APAP treatment (500 mg/kg, i.p.). APAP, acetaminophen; APAP-S, acetaminophen-O-sulfate; APAP-Cys, 3-cysteinyacetaminophen; APAP-NAC-, 3-N-acetylcysteinyacetaminophen; APAP-G, acetaminophen-O-glucuronide. Data are presented as mean \pm SD ($n = 3$).

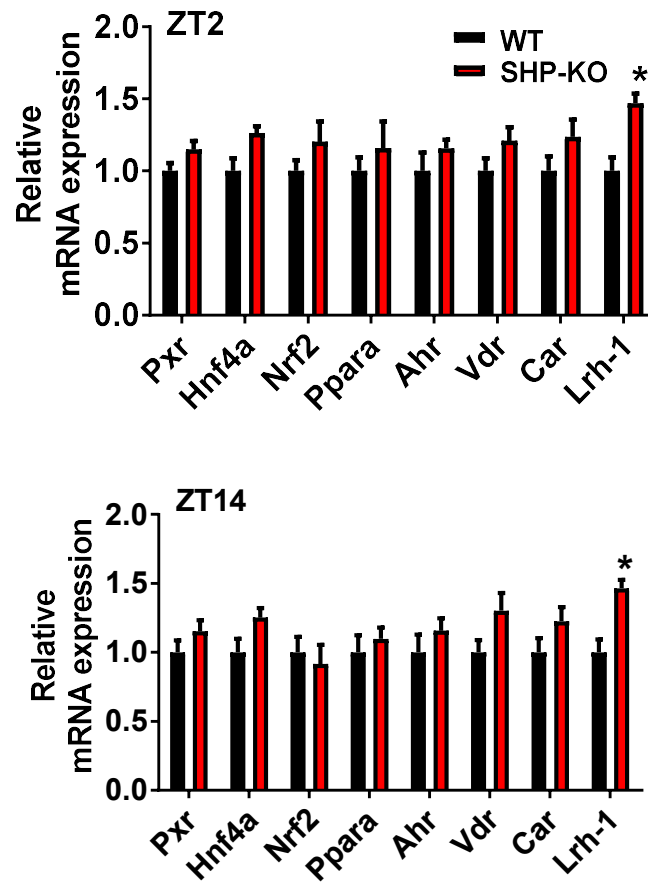


Figure S11. Expression of hepatic nuclear receptors/transcriptional factors involved in xenobiotic metabolism at ZT2 and ZT14 in WT and SHP-KO mice. mRNA levels were measured by qPCR and normalized to *cyclophilin b*. Data are mean \pm SD ($n = 5$). * $p < 0.05$ versus WT group.

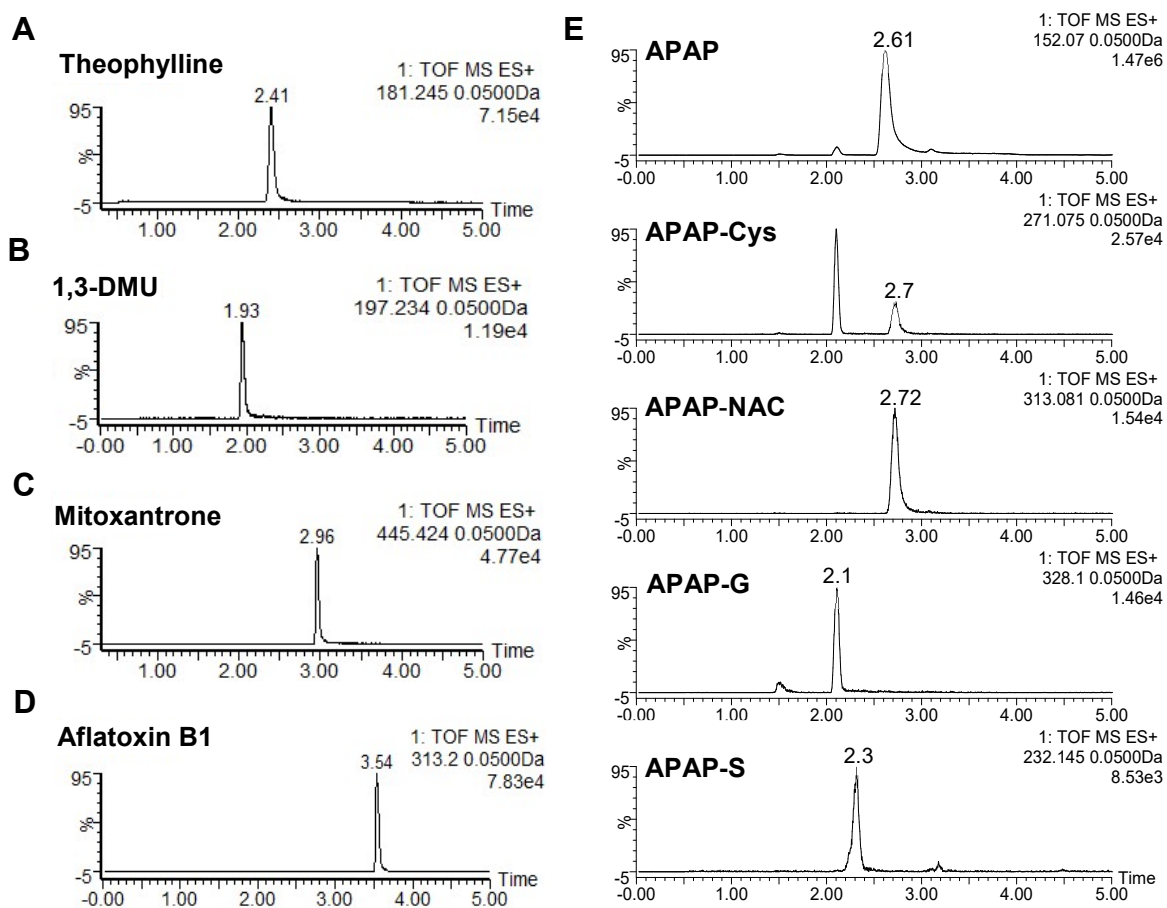


Figure S12. Representative chromatograms for quantification of drugs/xenobiotics and their metabolites. 1,3-DMU, 1,3-dimethyluric acid; APAP, acetaminophen; APAP-S, acetaminophen-O-sulfate; APAP-Cys, 3-cysteinyacetaminophen; APAP-NAC, 3-N-acetylcysteinyacetaminophen; APAP-G, acetaminophen-O-glucuronide.