

Research Paper

Magnetically Responsive Smart Nanoparticles for Cancer Treatment with a Combination of Magnetic Hyperthermia and Remote-Control Drug Release

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Abstract

We report the synthesis of smart nanoparticles (NPs) that generate heat in response to an alternating current magnetic field (ACMF) and that sequentially release an anticancer drug (doxorubicin, DOX). We further study the *in vivo* therapeutic efficacy of the combination of magnetic hyperthermia (MHT) and chemotherapy using the smart NPs for the treatment of multiple myeloma. The smart NPs are composed of a polymer with a glass-transition temperature (T_g) of 44°C, which contains clustered Fe₃O₄ NPs and DOX. The clustered Fe₃O₄ NPs produce heat when the ACMF is applied and rise above 44°C, which softens the polymer phase and leads to the release of DOX. The combination of MHT and chemotherapy using the smart NPs destroys cancer cells in the entire tumor and achieves a complete cure in one treatment without the recurrence of malignancy. Furthermore, the smart NPs have no significant toxicity.

Key words: smart nanoparticles, alternating current magnetic field, cancer cells

Introduction

Magnetic hyperthermia treatment (MHT) is a cancer therapy that relies on the heat produced by magnetic nanoparticles (NPs) under an alternating current magnetic field (ACMF), and it has the potential to realize a local, scarless, and economical treatment with fewer side effects.¹⁻⁷ The magnetic NPs must heat the tumor to a temperature sufficient to destroy the tumor cells (above 42.5°C) with exposure to harmless ACMF. The ACMF is considered harmless if the product of the amplitude (H) and frequency (f) of the field is less than 5×10^9 A m⁻¹ s⁻¹.⁸ For clinical

applications of MHT, magnetic NPs must be developed with high ACMF-to-heat conversion efficiency, which is governed by their specific absorption rate (SAR).

Controlling the composition of the magnetic NPs is an effective method for increasing the SAR. However, considering the potential toxicity to the body, the number of usable elements is severely limited.⁹ From this perspective, magnetite (Fe₃O₄) is the most promising material because of its low toxicity and relatively high saturation magnetization (M_s).^{4,9} Fur-

thermore, Fe_3O_4 is already used as a contrast agent for magnetic resonance imaging (MRI) in clinical practice.¹ However, individual Fe_3O_4 NPs must be concentrated at an unrealistically high level (~ 5 mg/mL) to heat the tumor cells to the extinction temperature by the application of ACMF.⁸

Recently, it has been shown that the clustering of magnetic NPs induces a significant increase in the magnetic moment, and consequently, the clustered magnetic NPs have a much higher M_s and SAR than individual magnetic NPs.^{10–12} Therefore, clusters of magnetic NPs provide an effective method for increasing SAR without introducing harmful elements.

Few studies have reported the in vivo therapeutic efficacy of MHT.^{12–16} According to these studies, MHT can inhibit the growth of tumor, but it does not provide a complete cure.^{12–15} This finding indicates that MHT cannot completely kill the cancer cells and that the residual cancer cells regrow.

Chemotherapy is a typical cancer therapeutic. However, this treatment cannot completely kill cancer cells, and serious side effects may develop even if the anticancer drugs are injected directly into the tumor because the anticancer drugs may escape into the healthy tissue surrounding tumor.¹⁷

Combining MHT and chemotherapy may overcome the problems in the individual treatments while producing the following synergistic effects: (1) MHT shrinks cancer cells and increases the space between the cells, which would allow anticancer drugs to spread throughout the tumor and destroy the cancer cells in the entire tumor; (2) the utilization of heat generated by the magnetic NPs under ACMF as a trigger for the drug release enables the gradual release of anticancer drugs from the NPs, which prevents the anticancer drugs from leaving the tumor and reduces side effects; and (3) MHT can potentiate the effect of the anticancer drug, which enhances the destruction of cancer cells.

For drug release using heat as a trigger, the drug carriers should ideally retain their load at body temperature ($\sim 37^\circ\text{C}$) and release the drug within a locally heated tumor at the therapeutic temperature for hyperthermia (42.5 – 45°C).¹⁸ Current thermoresponsive drug carriers include liposome^{19,20} or polymer NPs that exhibit a lower critical solution temperature.^{21–24} To release the drugs from these drug carriers in vivo, heat is generally applied using temperature-controlled water sacks or miniature annular-phased array microwave applicators.¹⁸ Unfortunately, these methods heat normal hypodermal tissues before heating deep tumors. Therefore, considering the damage to normal hypodermal tissues, it is extremely difficult to increase the tumor temperature to the therapeutic temperature for hyperthermia.

Furthermore, these heating methods have poor control over the tumor temperature.

On the other hand, ACMF heating has the following advantages over conventional heating methods: ^{25–42} (1) ACMF penetrates deep into the tissue, and it can heat deep tumors without damaging normal hypodermal tissues; and (2) adjusting the H and f of ACMF allows for easy regulation of the tumor temperature.^{42–45} Therefore, ACMF is a suitable trigger for drug release. ACMF-responsive drug carriers enable the remote, on-demand release of anticancer drugs, which reducing the side effects possible in conventional treatment methods.

Herein, we present the synthesis of smart NPs that effectively produce heat in response to harmless ACMF ($H = 8$ kA m^{-1} , $f = 230$ kHz, $Hf = 1.8 \times 10^9$ A m^{-1} s^{-1}) and sequentially release doxorubicin (DOX), an anticancer drug. Furthermore, we report the in vivo therapeutic efficacy of combining MHT and chemotherapy using the above smart NPs for treating multiple myeloma, which remains an incurable malignancy.

Results and Discussion

Synthesis and characterization of smart NPs

We designed the ACMF-responsive smart NPs by combining the DOX and clustered Fe_3O_4 NPs core within a containing polymer with a glass-transition temperature (T_g) of 44°C (Fig. 1A). During exposure to the ACMF, the inner Fe_3O_4 NPs cluster produces heat until at least the T_g , leading to the softening of the polymer phase, which allows the release of DOX.

The ACMF-responsive smart NPs were synthesized as follows (Fig. 2A). First, we polymerized pyrrole-3-carboxylic acid (PyCOOH) using iron (III) ion as a catalyst in an aqueous solution of DOX and polyvinyl alcohol (PVA) to synthesize DOX-containing carboxylic polypyrrole (DOX/PPyCOOH) NPs. In this reaction, DOX is contained within the NPs by the π - π interaction between PPyCOOH and DOX. In addition, the carboxylic acid groups of the DOX/PPyCOOH NPs are used for later surface modification. Second, we synthesized Fe_3O_4 NPs within the DOX/PPyCOOH NPs by reducing part of the used iron (III) ions to iron (II) ions with hydrazine. Hydrazine also serves as a catalyst to promote the formation of Fe_3O_4 by making the pH of the solution alkaline. The products of this reaction are called Fe_3O_4 /DOX/PPyCOOH NPs. Finally, to increase the NP retention in the tumor, we modified the NPs with polyethylene glycol (PEG) and folic acid (FA) by the amidation reaction of Fe_3O_4 /DOX/PPyCOOH NPs with amine- and FA-heterobifunctionalized PEG (FA-PEG-NH₂). In particular, PEG prevents the

phagocytosis by macrophages, and FA specifically binds to the folate receptors overexpressed in the cancer cells.^{46–49} The products obtained by this modification are called Fe₃O₄/DOX/PPy-PEG-FA NPs.

The transmission electron microscope (TEM) images of the Fe₃O₄/DOX/PPy-PEG-FA NPs confirm that the Fe₃O₄ NPs are clustered. The Fe₃O₄ NPs have a mean particle size of 10.5 nm, and the mean cluster size is 47.4 nm (Supplementary Material: Fig. S1†). Furthermore, the clustered Fe₃O₄ NPs are coated with a layer of PPy-PEG-FA with a thickness of ~10 nm. Thus, the mean size of the Fe₃O₄/DOX/PPy-PEG-FA NPs is ~70 nm.

The inorganic phase of the Fe₃O₄/DOX/PPy-PEG-FA NPs is composed of single-phase Fe₃O₄ as shown by the X-ray diffraction (XRD) pattern (Supplementary Material: Fig. S2). The crystallite sizes are 15.5, 13.8, 17.5, 16.6 and 16.1 nm, respectively, as determined from the Scherrer equation using the 220, 311, 400, 511 and 440 reflections. Thus, the average crystallite size is 15.9 nm.

Supplementary Material: Fig. S3 shows the Fourier-transform infrared (FTIR) spectra of PPyCOOH, DOX, Fe₃O₄/DOX/PPyCOOH NPs, and Fe₃O₄/DOX/PPy-PEG-FA NPs. In the Fe₃O₄/DOX/PPyCOOH NP spectrum, the bands corresponding to the amine of

DOX and PPyCOOH are observed at 3640–3310 cm⁻¹.⁵⁰ In addition, the bands attributed to the carboxylic acid of DOX and PPyCOOH are observed at 1790 and 1640 cm⁻¹, respectively. In the Fe₃O₄/DOX/PPy-PEG-FA NP spectrum, the bands attributed to the carboxylic acid of PPyCOOH disappear, indicating that the amidation between Fe₃O₄/DOX/PPyCOOH NPs and FA-PEG-NH₂ has occurred. Furthermore, the band attributed to the aromatic hydrocarbon of FA appears at 845 cm⁻¹. These findings confirm that the NPs are successfully modified with FA and PEG.

The hydrodynamic diameters of the Fe₃O₄/DOX/PPyCOOH NPs and Fe₃O₄/DOX/PPy-PEG-FA NPs in water were 51 ± 8 and 67 ± 13 nm, respectively, as measured by dynamic light scattering (DLS) (Supplementary Material: Fig. S4). The hydrodynamic diameter of the Fe₃O₄/DOX/PPy-PEG-FA NPs was larger than that of the Fe₃O₄/DOX/PPyCOOH NPs by ~16 nm because of the FA and PEG modification. In addition, the hydrodynamic diameters of Fe₃O₄/DOX/PPy-PEG-FA NPs are consistent with the size estimated from the TEM images. The zeta potentials of the Fe₃O₄/DOX/PPyCOOH NPs and Fe₃O₄/DOX/PPy-PEG-FA NPs were -0.97 and -1.87 mV, respectively.

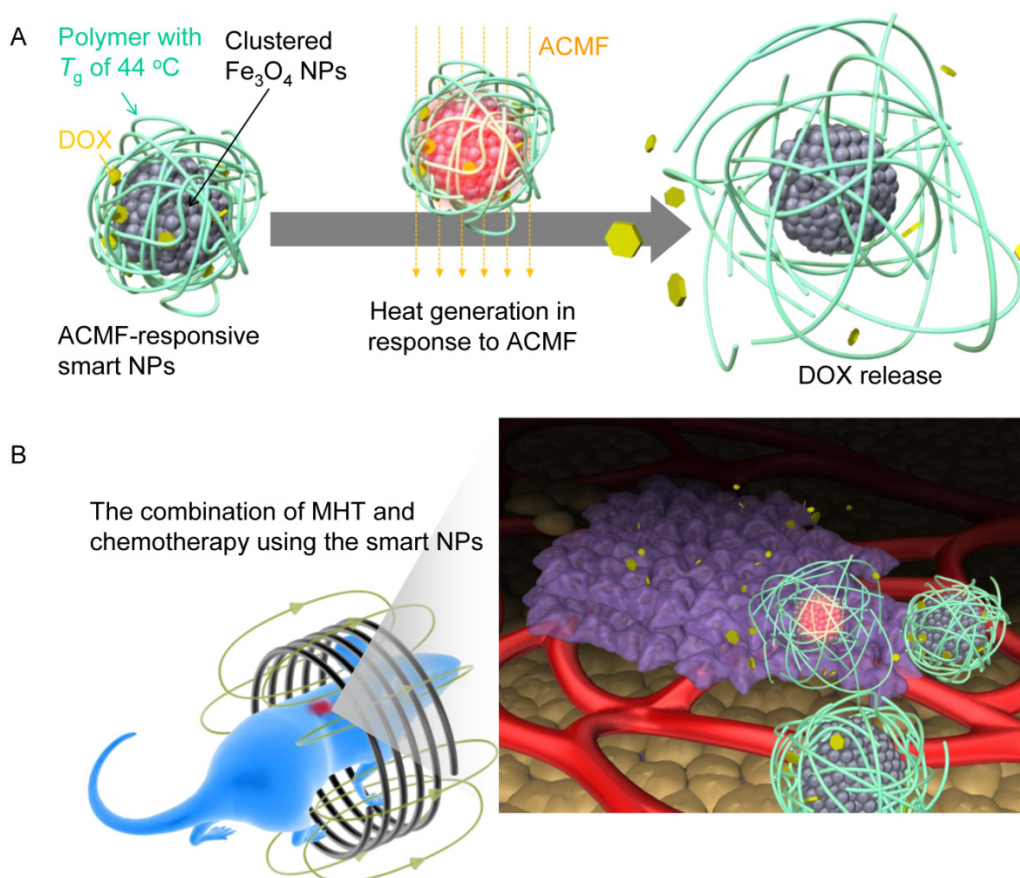


Figure 1. (A) Illustration of smart NPs that produce heat in response to ACMF and sequentially release DOX. (B) Illustration of cancer treatment with the combination of MHT and chemotherapy using the smart NPs.

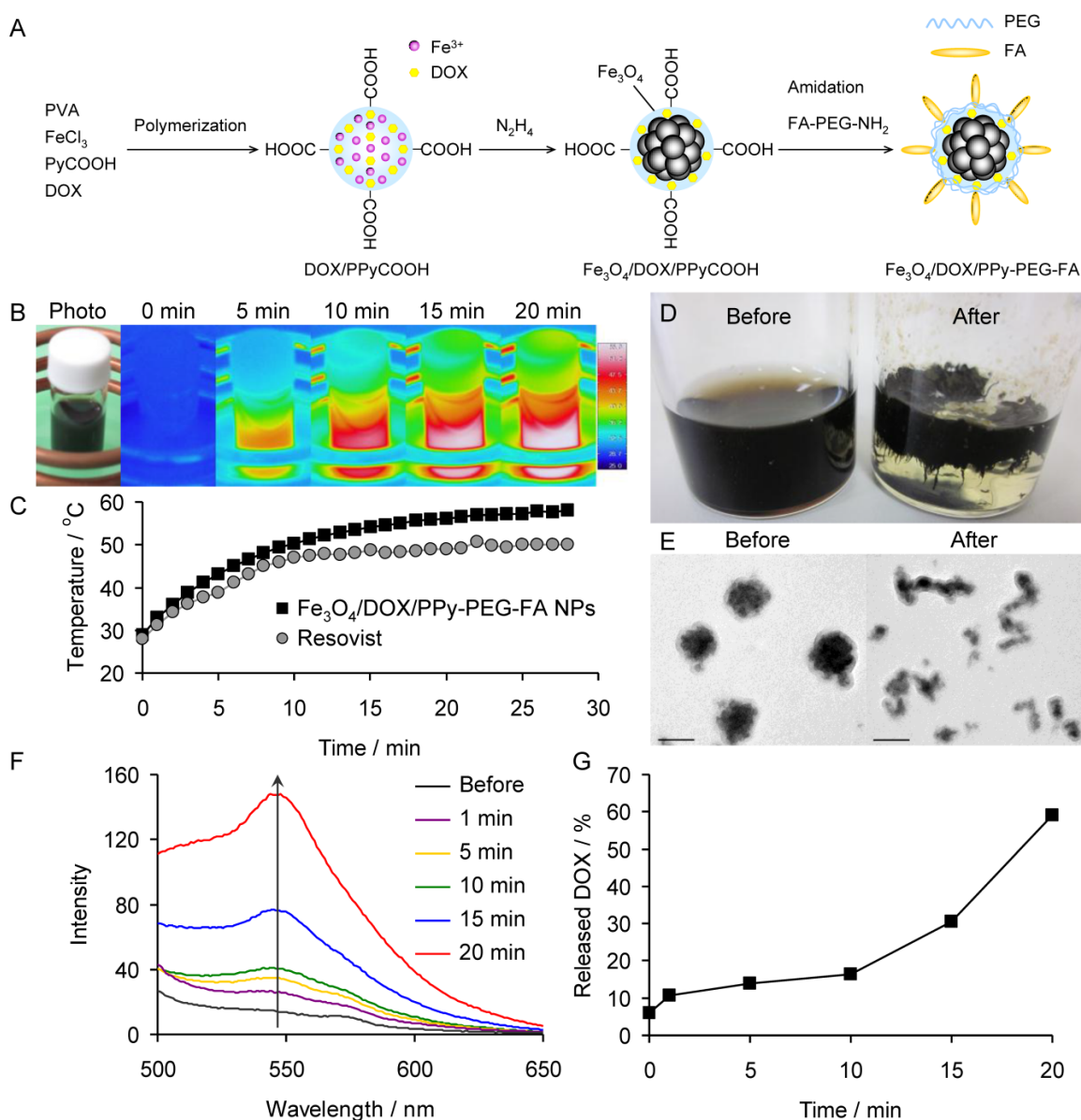


Figure 2. (A) Schematic showing the synthesis of $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs. (B) Photograph and thermal images of the aqueous dispersion of $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs under ACMF. (C) Temperature change of the aqueous dispersion of $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs with respect to ACMF exposure time. (D) Photographs and (E) TEM images of $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs before and after the application of ACMF. (F) Fluorescence spectra of the supernatant of the aqueous dispersion of $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs before and after the application of ACMF. (G) Proportion of DOX released from $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs with respect to ACMF exposure time.

The organic phase proportions in the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPyCOOH}$ NPs and $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs were 57 and 80%, respectively, as measured by thermogravimetry (TG) (Supplementary Material: Fig. S5). Thus, the fraction of FA-PEG in the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs is estimated to be 23%. Based on this estimate, the abundance of FA-PEG is 2.3×10^{-5} mol/g.

The differential scanning calorimetry (DSC) curve of the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs demonstrates that the T_g was 44°C (Supplementary Material: Fig. S6).

We estimated the DOX content in the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs from the absorbance at the wavelength of 480 nm, which is attributed to the

light absorption by DOX, using the calibration curve method (Supplementary Material: Fig. S7). This evaluation demonstrated that the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs had a DOX content of 42.6 mg/g.

The magnetic properties of the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs were measured using a vibrating sample magnetometer (VSM) at room temperature. The M_s of the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs was 16.7 emu/g (Supplementary Material: Fig. S8A). The corrected M_s is 83.5 emu/g because the proportion of the Fe_3O_4 phase in the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs was 20% as shown from the TG result (Supplementary Material: Fig. S6). Furthermore, the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs have a coercivity (H_c) of 15.2 Oe and a remanent magnetization

(M_r) of 1.25 emu/g (Supplementary Material: Fig. S8B), indicating that they are ferromagnetic. Fe_3O_4 NPs with a particle size of ~ 15 nm generally exhibit superparamagnetism, which shows neither H_c nor M_r . Actually, both H_c and M_r of Fe_3O_4 NPs synthesized by mixing FeCl_3 with hydrazine, which is the similar method as $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs, are zero (Figs. S8C and S8D). However, the clustering of Fe_3O_4 NPs produces strong magnetic dipole-dipole attractions between the Fe_3O_4 NPs, which induces ferromagnetic behavior (Figs. S8A and S8B).⁵¹ Furthermore, ferromagnetic NPs exhibit higher SARs than superparamagnetic NPs.⁴² In particular, ferromagnetic NPs with a H_c of ~ 15 Oe represent the maximum SAR, which is five times higher than that of superparamagnetic NPs.⁴³ Therefore, $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs are more useful for MHT than typical superparamagnetic NPs.

We evaluated the SAR of the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs in water and compared it to the SAR of Resovist[®], a clinical MRI contrast agent, by exposing the aqueous solutions (3 mg/mL) of $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs and Resovist[®] to ACMF. Both the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs and Resovist[®] generated heat and increased the water temperature by ~ 29 and $\sim 22^\circ\text{C}$ for 25 min, respectively (Fig. 2B and 2C). The SAR normalized to the Fe_3O_4 amount is calculated by

$$\text{SAR} = (C/m) (dT/dt),$$

where C is the specific heat capacity of water, m is the concentration (g/L of Fe_3O_4) of magnetic NPs in solution, and dT/dt is the slope of the curve for the first minute of ACMF exposure.¹ The SAR average and minimum values of the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs and Resovist[®] were 487, 336, 360 and 236 W/g, respectively. These findings demonstrated that the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs have a significantly higher ACMF-to-heat conversion efficiency than Resovist[®].

The photographs and TEM images of the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs before and after the ACMF exposure show that the polymer phase of the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs is softened by the ACMF exposure. This confirms that the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs were heated to the T_g ($\sim 44^\circ\text{C}$) by the heat generation of the Fe_3O_4 phase (Fig. 2D and 2E).

After exposure to the ACMF, we determined the amount of DOX released from the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs in aqueous solution (0.25 mg/mL) by measuring the fluorescence intensity of the supernatant DOX, which was collected by centrifugation. The fluorescence intensity of the supernatant DOX

increased with ACMF exposure time (Fig. 2F). Based on the calibration curve (Supplementary Material: Fig. S9A), the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs released 6.3 $\mu\text{g}/\text{mL}$ of DOX after 20 min of exposure (Fig. S9B), indicating that $\sim 60\%$ of the DOX content in the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs was released by the ACMF application (Fig. 2G). Furthermore we investigated the relationship between the proportion of DOX released from $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs and temperature (Fig. S9C). $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs began to release DOX at 39°C , and then the proportion of released DOX was drastically increased with temperature. More than 60% of the DOX content in the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs was released at T_g ($\sim 44^\circ\text{C}$), and the proportion of released DOX reached about 90% at 46°C . Generally, DOX leaks from tumor in the case of rapid intratumoral injection. The $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs is useful for preventing the leakage of DOX from tumor because they gradually release DOX.

In vivo therapeutic efficacy

To assess the in vivo therapeutic efficacy of the combination of MHT and chemotherapy, we injected the mice bearing subcutaneous xenograft tumors derived from the RPMI8226 multiple myeloma cell line intratumorally with the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs (5 mg/kg) and then exposed the mice to the ACMF (Fig. 1B). Fig. 3A shows the thermal image of the mice exposed to the ACMF for 20 min after the injection of $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs. The tumor was locally heated to the therapeutic temperature for hyperthermia (above 42.5°C), although the normal tissues were regulated at the survival temperature for cells. Fig. 3B shows the change of the tumor temperature of the mice injected with $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs, DOX-free NPs (designated as $\text{Fe}_3\text{O}_4/\text{PPy-PEG-FA}$ NPs), and no NPs with respect to the ACMF exposure time ($n = 5$). The tumor temperature of the mice injected with $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs or $\text{Fe}_3\text{O}_4/\text{PPy-PEG-FA}$ NPs drastically increased immediately after the application of the ACMF, and then it gradually increased with time. After the 7 min of ACMF exposure, both of these tumor temperatures reached 44°C . The unadministered mice exposed to the ACMF for 20 min showed little difference in the tumor temperature, indicating that the ACMF has only a limited effect on the elevation of body temperature. The follow-up photographs show that the mouse treated with $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs has a scab on their tumors 4 days after treatment, and their tumors completely disappear within 8 days (Fig. 3C).

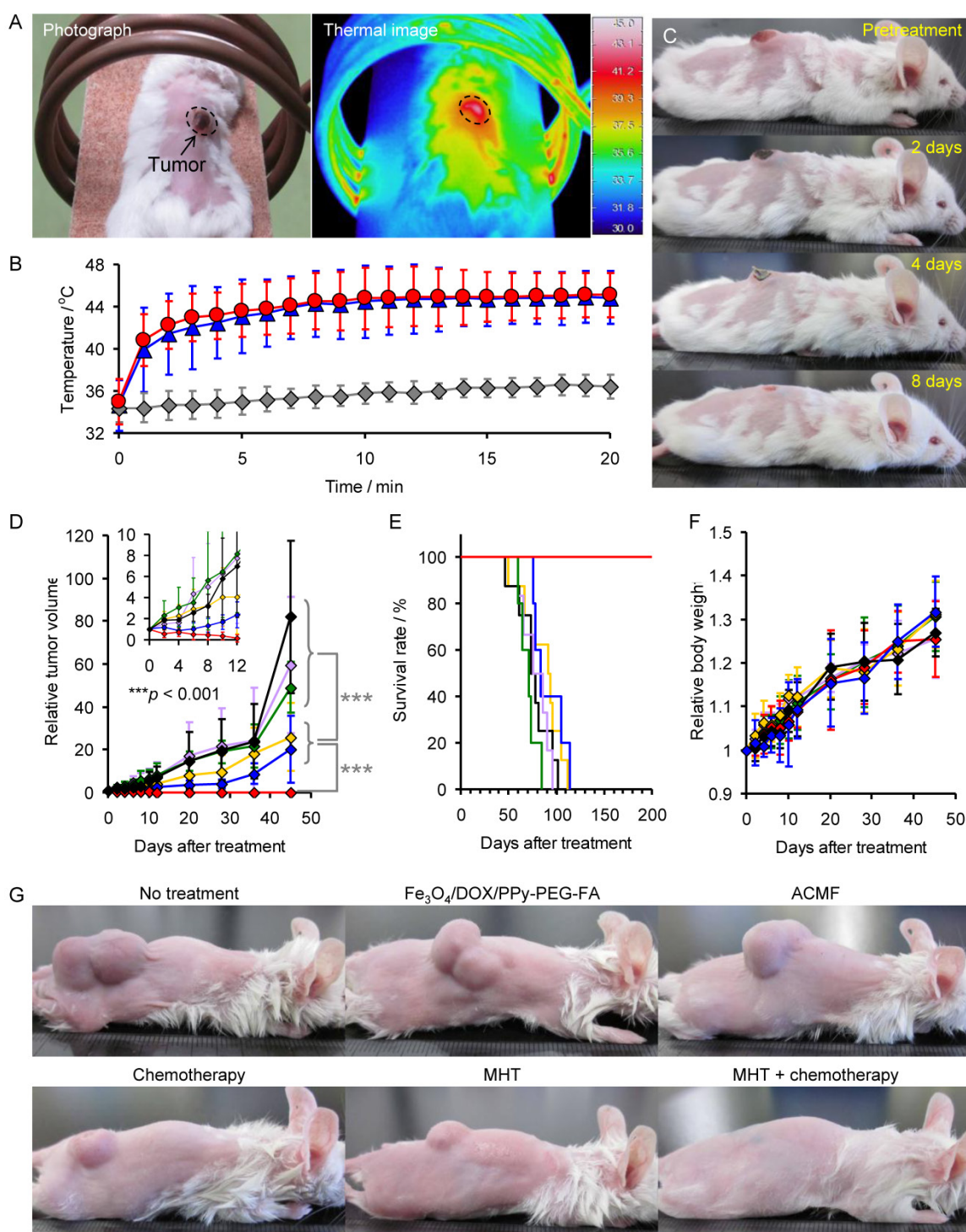


Figure 3. (A) Photograph and thermal image of a mouse exposed to ACMF for 20 min after injection with Fe₃O₄/DOX/PPy-PEG-FA NPs. (B) Average change of the tumor temperature of the mice injected with Fe₃O₄/DOX/PPy-PEG-FA NPs, Fe₃O₄/PPy-PEG-FA NPs, and no NPs with respect to ACMF exposure time (n = 5). (C) Follow-up photographs of mouse exposed to ACMF for 20 min after injection with Fe₃O₄/DOX/PPy-PEG-FA NPs. Change of (D) tumor volume, (E) survival rate, and (F) body weight: non-treated mice (black), mice treated with chemotherapy (yellow), mice exposed to ACMF (green), mice injected with Fe₃O₄/DOX/PPy-PEG-FA NPs intratumorally (purple), mice treated with MHT (blue), and mice treated with the combination of MHT and chemotherapy (red). The inset in Fig. 3D shows the magnified view for the first 12 days after treatment. (G) Photographs of non-treated mice, mice treated with chemotherapy, mice exposed to ACMF, mice injected with Fe₃O₄/DOX/PPy-PEG-FA NPs intratumorally, mice treated with MHT, and mice treated with the combination of MHT and chemotherapy 45 days after treatment.

Furthermore, we compared the tumor volume, survival rate, and the body weight of the mice exposed to the ACMF after intratumoral injection of Fe₃O₄/DOX/PPy-PEG-FA NPs (designated as mice treated with the combination of MHT and chemo-

therapy) with those of the control groups: non-treated mice; mice injected with DOX intratumorally (designated as mice treated with chemotherapy); mice exposed to ACMF; mice injected with Fe₃O₄/DOX/PPy-PEG-FA NPs intratumorally; mice

exposed to ACMF after the intratumoral injection of $\text{Fe}_3\text{O}_4/\text{PPy-PEG-FA}$ NPs (designated as mice treated with MHT) (Fig. 3D, 3E, and 3F). Forty-five days after treatment, the tumor volume of the mice treated with chemotherapy or MHT was significantly smaller than those of the non-treated mice, the mice exposed to ACMF, and the mice injected with $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs (Fig. 3D). Thus, chemotherapy and MHT inhibit the tumor growth, but they cannot cure the tumor completely. For the mice treated with the combination of MHT and chemotherapy, the tumor volume gradually decreased as time passes and it reached zero within 12 days after treatment as shown in the inset of Fig. 3D. The tumor volume remained in zero 45 days after treatment (Fig. 3D), indicating that the combination of MHT and chemotherapy prevented recurrence and completely cured the tumor. The photographs 45 days after treatment also show that both MHT and chemotherapy inhibited the tu-

mor growth, but the combination of MHT and chemotherapy eliminates the tumor completely (Fig. 3G). Furthermore, the survival rate for mice treated with the combination of MHT and chemotherapy was 100% 200 days after the treatment, while the non-treated mice, the mice exposed to ACMF, and the mice injected with $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs all died within about 100 days (Fig. 3E). MHT and chemotherapy somewhat prolonged the survival period. There was no significant difference in the body weight between the groups (Fig. 3F).

We investigated how the therapeutic efficacy was enhanced from the combination of MHT and chemotherapy using histological analyses of the tumors 24 h after treatment. In the non-treated mice, the mice exposed to ACMF, and the mice injected with $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs, the viable cancer cells were densely packed in the tumors as shown in region V (Fig. 4).

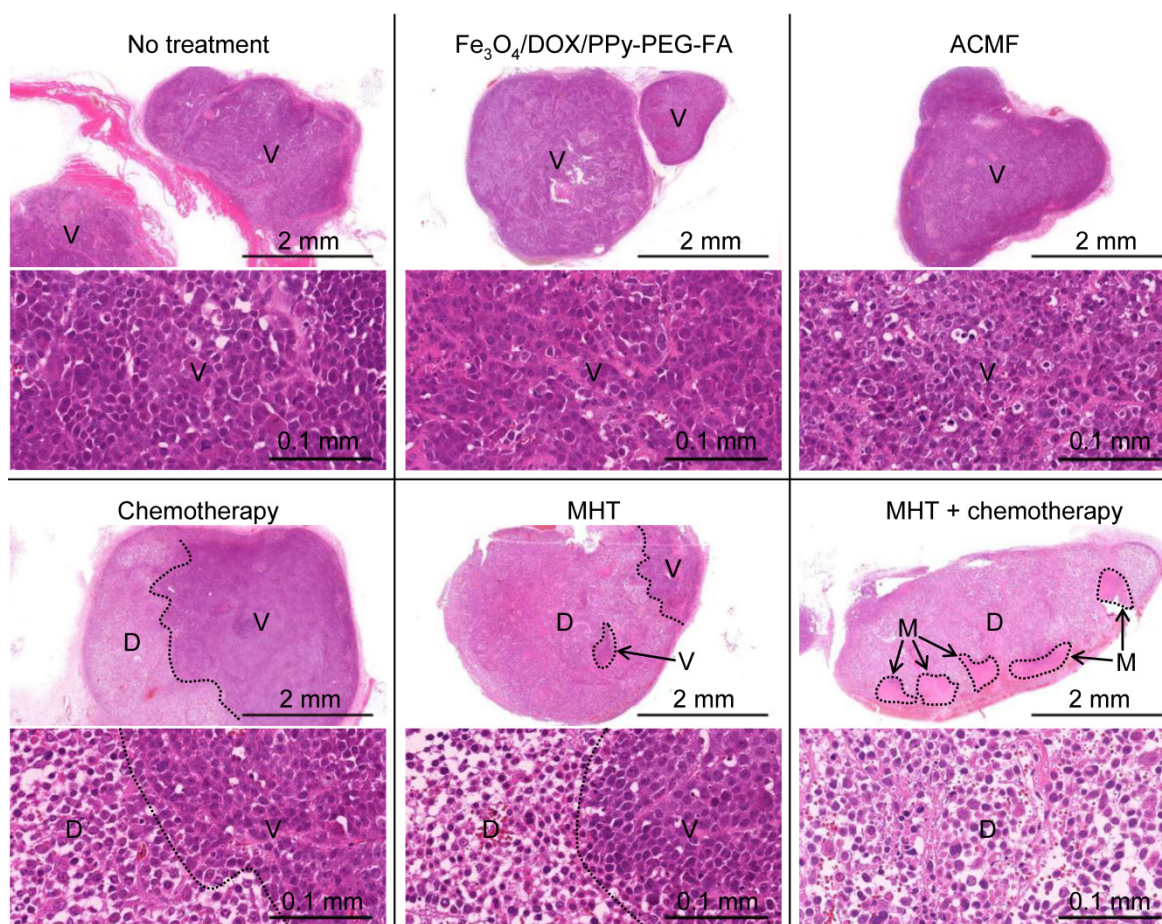


Figure 4. Histological findings of tumors of non-treated mice, mice treated with chemotherapy, mice exposed to ACMF, mice injected with $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs intratumorally, mice treated with MHT, and mice treated with the combination of MHT and chemotherapy 24 h after treatment: viable tumor tissue (V); destroyed tumor tissue (D); normal muscle tissue (M).

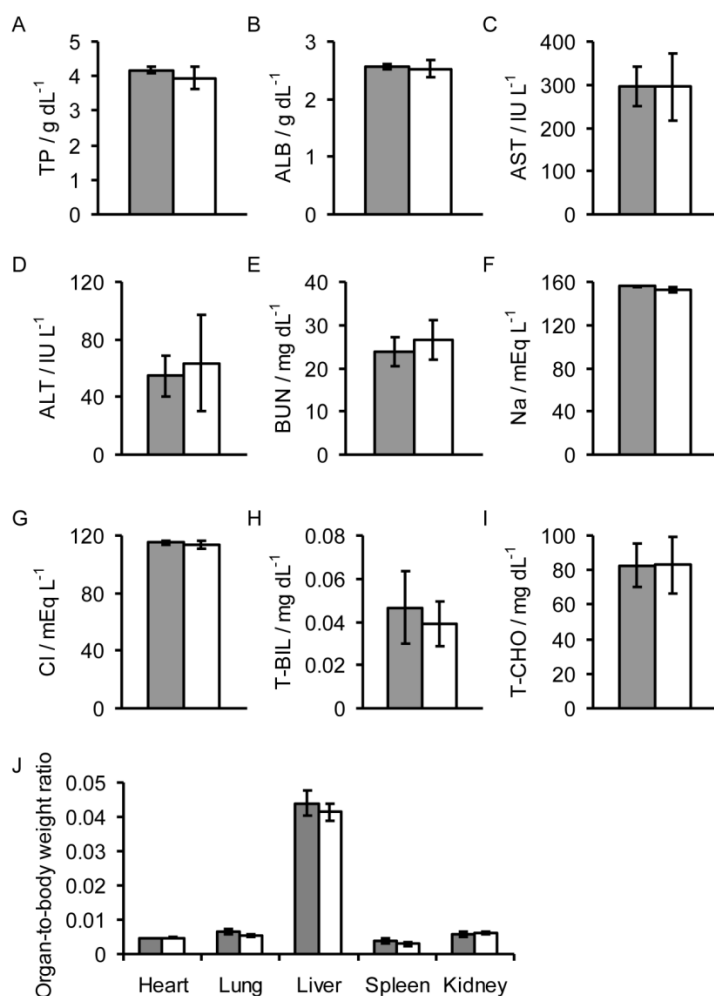


Figure 5. Biochemical assays in the sera and organ-to-body weight ratios of mice injected with $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs (gray) and non-treated mice (white). The following measurements were taken 24 h after injection: (A) total protein (TP), (B) albumen (ALB), (C) aspartate aminotransferase (AST), (D) alanine aminotransferase (ALT), (E) blood urea nitrogen (BUN), (F) sodium (Na), (G) chlorine (Cl), (H) total bilirubin (T-BIL), and (I) total cholesterol (T-CHO) values, and the (J) organ-to-body weight ratios.

For the mice treated with chemotherapy, the cancer cells were only partly destroyed as shown in the region D, indicating that DOX did not spread throughout the tumor. MHT destroyed cancer cells in a broader tumor area than that of chemotherapy; however, all cancer cells were not destroyed. Furthermore, MHT shrank the cytoplasm of the cancer cells and widened the space between cancer cells (Supplementary Material: Fig. S10). The similar phenomenon has been observed in light-mediated treatment.⁵² For the mice treated with the combination of MHT and chemotherapy, the cancer cells were fully destroyed throughout the tumor. This probably occurred because MHT widened the space between the cancer cells, allowing DOX to spread throughout the tumor. Furthermore, the combination of MHT and chemotherapy significantly widened the space between the cancer cells compared to only MHT (Supplementary Material: Fig. S10), which indicates that the combination treatment caused more serious

damage to the tumor. Thus, the combination of MHT and chemotherapy can more effectively destroy cancer cells than MHT. The data from the tumor volume, survival rate, and the histological findings demonstrates that the combination of MHT and chemotherapy creates a synergistic effect, which enhances the therapeutic efficacy. Moreover, the muscle tissues extended to the tumor suffered no damage from the combination of MHT and chemotherapy (Supplementary Material: Fig. S11). Thus, the combination of MHT and chemotherapy destroys the tumor without damaging normal tissues. Furthermore, Prussian blue nuclear fast red staining of the tumor section of the mouse treated with a combination of MHT and chemotherapy shows that $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs are distributed throughout the entire tumor and their distribution is relatively uniform. (Supplementary Material: Fig. S12).

Toxicity assay

To assess the toxicity of the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs, we measured the organ-to-body weight ratio, and the amount of total protein (TP), albumen (ALB), aspartate aminotransferase (AST), alanine aminotransferase (ALT), blood urea nitrogen (BUN), Na, Cl, total bilirubin (T-BIL), and total cholesterol (T-CHO) in the sera of the mice 24 h after intravenous injection of the NPs. There was no significant difference in the values between the mice injected with NPs and the non-treated mice (Fig. 5). Thus, $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs have no significant toxicity.

Conclusions

The $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs effectively produced heat when exposed to a harmless ACMF and sequentially released DOX. MHT or chemotherapy could not cure the tumor, but the treatment inhibited the tumor growth. On the other hand, the combination of MHT and chemotherapy using the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs completely cured the tumor without recurrence of malignancy. The combination of MHT and chemotherapy destroyed the cancer cells in the entire tumor, causing more serious damage to tumor than MHT. Thus, the combination of MHT and chemotherapy creates a synergistic effect, which destroys the cancer cells. Furthermore, the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs have no significant toxicity. In this study, we administered the $\text{Fe}_3\text{O}_4/\text{DOX}/\text{PPy-PEG-FA}$ NPs to the mice by intratumoral injection as the first step for investigating the

therapeutic efficacy of the combination of MHT and chemotherapy using the Fe₃O₄/DOX/PPy-PEG-FA NPs. In the future, we will assess the therapeutic efficacy by intravenous injection of the Fe₃O₄/DOX/PPy-PEG-FA NPs.

Experimental

Materials

PyCOOH, hydrazine monohydrate, 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (EDAC), and *N*-hydroxysuccinimide (NHS) were purchased from Tokyo Chemical Industry (Tokyo). PVA ($M_w = 40,000$) and iron (III) chloride hexahydrate were purchased from Sigma-Aldrich (MO, USA). DOX was purchased from Toronto Research Chemicals Inc. (NY, USA). FA-PEG-NH₂ ($M_w = 10,000$) was purchased from Nanocs (NY, USA).

Synthesis of Fe₃O₄/DOX/PPyCOOH NPs

DOX/PPyCOOH NPs were synthesized by reference to the synthesis method of DOX-free PPy NPs without carboxylic acid.⁵³ PVA (0.94 mmol) was dissolved in distilled water (10 mL) at 85°C for 60 min. After cooling the aqueous solution of PVA to room temperature, iron (III) chloride hexahydrate (0.32 mmol) was added to the solution, which was then stirred at room temperature for 1 h. DOX (3.5 mmol) and PyCOOH (0.28 mmol) were added to the above solution, which was then stirred at room temperature for 72 h in the dark. Hydrazine monohydrate (10.3 mmol) was added to the solution of DOX/PPyCOOH NPs, which was then stirred for 24 h in the dark.^{12,42–45,54} The product was collected from the dispersion by centrifugation (20,000 g, 5 h) and re-dispersed in distilled water. This sequence was repeated three times.

Synthesis of Fe₃O₄/DOX/PPy-PEG-FA NPs

FA-PEG-NH₂ (20 nmol), EDAC (20 nmol), and NHS (20 nmol) were added to the aqueous dispersion of Fe₃O₄/DOX/PPyCOOH NPs, which was then stirred at room temperature for 24 h in the dark. The product was collected from the dispersion by centrifugation (20,000 g, 5 h) and redispersed in distilled water. This sequence was repeated three times.

Characterization

The size and shape of Fe₃O₄/DOX/PPy-PEG-FA NPs were observed by TEM (H-760, Hitachi, Tokyo). The hydrodynamic diameter and zeta potential were measured by DLS (NICOMP 380 ZLS, Showa Denko, Tokyo). The FTIR spectra were recorded with a FTIR spectrometer (Nexus 470, Nicolet, Madison, WI). The amount of organic phase was measured using TG (Thermo plus EVO TG8120, Rigaku, Tokyo, Japan) at

a heating rate of 10°C/min from room temperature to 800°C under an oxygen atmosphere. The crystalline phases were analyzed by XRD using CuK α radiation with a monochromator (Rigaku SmartLab, Rigaku, Tokyo). The T_g was measured by DSC (TA Instruments, Japan, Tokyo). The magnetic properties were measured with a vibrating sample magnetometer (Type 5, Toei Kogyo, Tokyo) at room temperature. For comparison, we synthesized uncoated Fe₃O₄ NPs by mixing iron (III) chloride hexahydrate (0.32 mmol) with hydrazine monohydrate (10.3 mmol), which is the similar method as Fe₃O₄/DOX/PPy-PEG-FA NPs.

Estimation of DOX content in Fe₃O₄/DOX/PPy-PEG-FA NPs

The absorbance of the aqueous dispersion of Fe₃O₄/DOX/PPy-PEG-FA NPs (0.5 mg/mL) was measured using an ultraviolet-visible (UV-Vis) spectrophotometer (U-3000, Hitachi, Tokyo). To prepare the calibration curve, the absorbance of DOX was measured at various concentrations (0.001, 0.025, 0.05, 0.1, and 0.2 mg/mL). The DOX content in Fe₃O₄/DOX/PPy-PEG-FA NPs was determined by substituting the absorbance of the aqueous dispersion of Fe₃O₄/DOX/PPy-PEG-FA NPs in the calibration curve.

The heat generation of Fe₃O₄/DOX/PPy-PEG-FA NPs in water

The aqueous dispersions (3 mg/mL) of Fe₃O₄/DOX/PPy-PEG-FA NPs and Resovist were placed in an ACMF with $H = 8 \text{ kA m}^{-1}$ and $f = 230 \text{ kHz}$ ($Hf = 1.8 \times 10^9 \text{ A m}^{-1} \text{ s}^{-1}$) for 20 min. An ACMF was generated using a transistor inverter with field coils (2.4 kW, EasyHeat, Ambrell, NY). Thermal images were taken using a Thermograph (Thermo Gear G100EX, NEC Avio Infrared Technologies, Tokyo). The SAR values of Fe₃O₄/DOX/PPy-PEG-FA NPs and Resovist normalized to the Fe₃O₄ amounts were calculated according to the following equation:

$$\text{SAR} = (C/m) (dT/dt),$$

where C is the specific heat capacity of water, m is the concentration (g/L of Fe₃O₄) of the magnetic NPs in solution, and dT/dt is the slope of the curve during the first minute of ACMF exposure.

The estimation of the amount of DOX released from Fe₃O₄/DOX/PPy-PEG-FA NPs by the application of ACMF

The aqueous dispersion of Fe₃O₄/DOX/PPy-PEG-FA NPs (0.25 mg/mL) was placed in an ACMF with $H = 8 \text{ kA m}^{-1}$ and $f = 230 \text{ kHz}$ ($Hf = 1.8 \times 10^9 \text{ A m}^{-1} \text{ s}^{-1}$) for 1, 5, 10, 15, or 20 min. The supernatant was collected by centrifugation (20,000 g, 5 h). The amount of DOX released from

Fe₃O₄/DOX/PPy-PEG-FA NPs was estimated by substituting the fluorescence intensity of the supernatant DOX in the calibration curve, which was prepared by measuring the fluorescence intensity of DOX aqueous solutions (5, 10, 50, and 100 µg/mL).

Ethics statement

The study protocol was approved by the Animal Care and Use Committee of the University of Tokushima (Tokushima).

Animals

Female CB17/Icr-Prkdcscid mice aged 4 weeks were purchased from Charles River Laboratories (Yokohama) and maintained in a specific pathogen-free facility in our Animal Resources Center. To eradicate residual natural killer (NK) cells, mice were injected intraperitoneally with 10 µL of rabbit anti-asialo GM1 antiserum (Wako Pure Chemicals, Osaka) 1 day before tumor inoculation.⁵⁵ Inoculation with human multiple myeloma cells (5×10⁶) was accomplished by subcutaneous injection into the back of the mice.

In vivo therapeutic efficacy of the combination of MHT and chemotherapy

The therapeutic efficacy of the combination of MHT and chemotherapy was investigated using mice with tumor volumes of 130 mm³. Tumor volumes were calculated by

$$V = AB^2\pi/6,$$

where A is the long and B is the short lateral diameter of the tumor. The mice were divided into 6 groups: no treatment ($n = 8$); intratumoral administration of 5 mg/kg of Fe₃O₄/DOX/PPy-PEG-FA NPs (1 mg/mL) ($n = 6$); application of ACMF ($n = 5$); intratumoral administration of 0.13 mg/kg of DOX, which is equivalent to the amount of DOX released from 5 mg/kg of Fe₃O₄/DOX/PPy-PEG-FA NPs (1 mg/mL) ($n = 8$); application of ACMF 24 h after intravenous administration of 5 mg/kg of Fe₃O₄/PPy-PEG-FA NPs (1 mg/mL) ($n = 5$); and application of ACMF 24 h after intravenous administration of 5 mg/kg of Fe₃O₄/PPy-PEG-FA NPs ($n = 5$). The H , f , and application time of ACMF were 8 kA m⁻¹, 230 kHz, and 20 min, respectively. Fe₃O₄/DOX/PPy-PEG-FA NPs, Fe₃O₄/PPy-PEG-FA NPs, and DOX were injected in one shot. The mice were killed when the tumor volume reached 15,000 mm³.

Histological analyses

The tissue samples obtained for histology were immediately immersed in a 4% paraformaldehyde solution for 24 h. Fixed tissues were encased in paraffin blocks. Tissue sections (2 µm) were cut from the

paraffin blocks and affixed to Superfrost Microscopy Slides. Hematoxylin and eosin staining of tissue sections was carried out in a conventional manner.

Toxicity evaluation

Fe₃O₄/DOX/PPy-PEG-FA NPs (5 mg/kg) were injected intravenously into mice ($n = 5$ per cohort). Blood samples were collected 24 h after injection, and serum was obtained by centrifugation of whole blood at 3,000 rpm for 15 min. The biochemical parameters (TP, ALB, AST, ALT, BUN, Na, Cl, T-BIL, and T-CHO) in the serum were determined using an automated biochemical analyzer (Hitachi 7180, Tokyo). Furthermore, 24 h after injection, the heart, liver, lung, kidney, and spleen were enucleated, and their weights were measured using an electric balance.

Supplementary Material

Fig.S1 – Fig.S12.

<http://www.thno.org/v04p0834s1.pdf>

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Competing Interests

The authors have declared that no competing interest exists.

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