


Research Paper

## DiR-labeled Embryonic Stem Cells for Targeted Imaging of *in vivo* Gastric Cancer Cells

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### Abstract

Embryonic stem (ES) cells have great potential in applications such as disease modeling, pharmacological screening and stem cell therapies. Up to date, there is no related report on the use of ES cells as tracking and contrast reagents of cancer cells *in vivo*. Herein we report that DiR-labeled murine ES cells can recognize and target gastric cancer cells *in vivo*. DiR-labeled murine ES (mES) cells ( $5 \times 10^6$ ) were intravenously injected into gastric tumor-bearing mice. The biodistribution of DiR-labeled mES cells was monitored by IVIS imaging within 24 h. Major organs were harvested and analyzed by immunofluorescence staining and Western blotting. Chemotaxis assay was employed to investigate the chemotaxis of ES cells tracking cancer cells. Fluorescent imaging results showed that DiR-labeled mES cells targeted gastric cancer tissue *in vivo* as early as 10 min post-injection, reaching a peak at 2h post-injection. Immunofluorescence staining and Western blotting results showed gastric cancer tissues specifically expressed SSEA-1. *In vitro* migration tests confirmed that mES cells actively moved to test sites with different concentration of CXCL12 in a dose-dependent manner. In conclusion, DiR-labeled mES cells may be used for gastric cancer targeted imaging *in vivo*, and have great potential in applications such as identifying and imaging of early gastric cancer in near future.

Key words: murine embryonic stem cells; gastric cancer cells; target imaging; migration; chemotaxis.

### Introduction

Since the firstly isolation of the murine embryonic stem (mES) cells in the 1980s, ES cells show their pluripotent nature, differentiating into cell types of all primary germ lineages [1-3]. In recent years, researchers have paid more and more attention on the *in vivo* monitoring and therapy potential of stem cells [4-6]. Generally, stem cell therapies consist of two aspects, one is the *in vivo* targeted migration of stem cells to the target tissues, and another is the prolifera-

tion and differentiation of stem cells in the target tissues to regenerate new organs [7-10]. Those applications have promising prospects in regenerative medicine.

Stomach malignancy is currently the fourteenth most common cancer in the United States and the second most common cancer in China [11, 12]. Gastric cancer is still the second most common cause of cancer-related death in the world. It remains difficult to

cure effectively even in Western countries, primarily because most patients are identified at the advanced stages of the malignancy [13]. Thus, early recognition and tracking of gastric cancer cells *in vivo* would be of particular significance. Our group has tried to establish an early gastric cancer pre-warning system since 2005 [14]. We hoped to find early gastric cancer cells *in vivo* by multi-mode targeted imaging techniques [15–18]. However, our efforts were stalled by a lack of specific gastric cancer biomarkers. This is one reason why the search for an alternative way to recognize and track early gastric cancer cells *in vivo* has become a central subject in this field.

Among all the imaging techniques, near-infrared (NIR) imaging and bioluminescence imaging (BLI) have become the most popular modalities [19, 20]. NIR imaging has many advantages over other imaging means, because it can penetrate biological tissues such as skin and blood more efficiently than visible light [21, 22]. BLI combined with NIR light has been found to enable real-time observation of stem cell trafficking and *in vivo* gene transfer.

DiR dye is a lipophilic, NIR fluorescent cyanine dye ideal for staining cytoplasmic membrane. The two long 18-carbon chains of DiR dye can insert into the cell membrane, resulting in specific and stable cell staining with negligible dye transfer between cells. The NIR property of DiR dye makes it ideal for *in vivo* imaging because of significantly reduced autofluorescence from the animal at higher wavelength [23]. In this study, mES cells were labeled with NIR DiR dyes, *in vivo* distribution of DiR-labeled mES cells (DiR-mES cells) was monitored by an IVIS imaging system, and the chemotaxis mechanism of ES cells tracking cancer cells was investigated. The significant finding is that tumor tissues or metastatic cancer cells *in vivo* can be tracked by using mES cells as tracking and contrast reagents.

## Materials and Methods

All animal experiments (NO.SYXK2007-0025) were approved by the Institutional Animal Care and Use Committee of Shanghai Jiao Tong University.

### Feeder-free cultured murine embryonic stem (mES) cells

Murine embryonic stem (GFP-SV129, mES) cells were provided by the Shanghai Institute of Digestive Disease, Renji Hospital. The mES cells were cultured with completed medium, which is composed of Knockout-DMEM (Gibco) supplemented with fetal bovine serum (FBS, Gibco), non-essential amino acids (NEAA, Gibco), L-glutamine (Gibco),  $\beta$ -mercaptoethanol (Gibco) and recombinant human

leukemia inhibitory factor (LIF, Chemicon). The mES cells were routinely passaged every 2 days, and the medium was changed on alternate days. The feeder-free mES cells were prepared by using the direct transition method: mES cells were split onto newly gelatinized plates without feeders and incubated for 30 min, and then the supernatant culture medium were collected and transferred into newly gelatinized plates without feeders and cultured until mitosis. The mES cells were continuously cultured for three to four or more splits by this method to eliminate all feeders.

### Labeling mES cells with DiR and *in vitro* cell imaging

The mES cells were washed three times with PBS, trypsinized with 0.05% trypsin-ethylene diaminetetra acetic acid (EDTA; Gibco-Invitrogen). The mES cells were incubated with 3.5  $\mu$ g/mL DiR buffer for 30 min at 37°C according to the protocol of Xeno-Light DiR (Caliper Lifesciences). DiR-labeled mES cells were used as the DiR(+) test group, mES cells which were not incubated with DiR were washed with PBS (pH 7.0) and marked with DiR(-) control group. Then the DiR(+) and DiR(-) cells were centrifuged for 3 min at 1000 rpm and 4°C, and washed twice with PBS buffer and examined for viability using a Typan Blue Staining Cell Viability Assay Kit (Beyotime). Finally, DiR(+) and DiR(-) cells ( $5 \times 10^6$ ) were resuspended in 0.2 mL PBS buffer, and performed fluorescence imaging in a 96-well black culture dish by IVIS system under 710 nm of excitation and 760 nm of emission. The DiR(+) and DiR(-) cells were continuously cultured and imaged at 4, 8, 12, and 24 h after first imaging under the same imaging conditions. The intensity of the region of interest (ROI) was plotted in units of the maximum number of photons per second per centimeter square per steradian (p/s/cm<sup>2</sup>/sr). In order to evaluate the changes of the fluorescent signals varied cell numbers,  $1 \times 10^6$ ,  $1 \times 10^5$ ,  $1 \times 10^4$ ,  $1 \times 10^3$  and  $1 \times 10^2$  DiR(+) cells were also counted and performed fluorescent imaging in a 96-well black culture dish by IVIS system under 710 nm of excitation and 760 nm of emission, the correlation between the number of DiR(+) cells and the intensity of fluorescent signals was plotted.

### Preparation of bioluminescence gastric cancer mouse models

Male athymic nude mice (4 weeks old,  $17 \pm 2$  g) were obtained from the Shanghai LAC Laboratory Animal Co. Ltd., Chinese Academy of Sciences (Shanghai, China, SCXK2007-0005). Bioluminescence mouse gastric cancer cell lines (MFC) ( $1 \times 10^6$ ) were injected subcutaneously into the right anterior flank

areas of the mice. Four weeks later, tumors grew to approximately 5 mm in diameter, and bioluminescence images of gastric cancer mouse models were taken using an IVIS system with the exposure time of 30 s.

### DiR-mES cells for targeted imaging of gastric cancer cells *in vivo*

DiR-mES cells ( $5 \times 10^6$ ) were intravenously injected into the gastric cancer mouse models ( $n=3$ ) as test group. In control group, gastric cancer mouse models ( $n=3$ ) were injected with DiR. These injected gastric cancer mouse models were monitored using the IVIS system at 0.5, 1, 2, 4, 6, 12, and 24h post-injection to obtain serial fluorescence images. The ideal filter conditions for DiR imaging were set at 710 nm of excitation and 760 nm of emission. Identical illumination settings (e.g., lamp voltage, filter, exposure time) were used in all animal imaging experiments. The mice were imaged at all-time points at lateral, supine and prone positions. Spleen, liver, lung, kidney, heart, and tumor tissues were collected for *ex vivo* imaging at 24 h post-injection. Gray-scale photographic images and fluorescent images of each sample were analyzed and overlaid using Living Image software (Xenogen, Alameda, CA). Regions of interest (ROI) were drawn over the signals, and average radiant efficiency was quantified in  $p/s/cm^2/sr$ .

### Immunofluorescence analysis and Western Blot analysis

For histological evaluation, the mice in the test and control groups were euthanized after imaging. Tumors and major organs (spleen, liver, lung, kidney, and heart) were excised, frozen, embedded, and sectioned into 8  $\mu m$  slices. The expression of SSEA-1 in these tissues were examined under a fluorescence microscope (Olympus IX71) after performing immunohistochemical staining of slices. GFP imaging of ultra-thin slices were also imaged simultaneously. Meanwhile, the expression of SSEA-1 in tumor tissues were evaluated in protein level. Tumor tissues and important organs (liver, lung, kidney, spleen, and heart) were excised and lysed in protein lysis buffer. Equal amounts of sample lysates were separated by sodium dodecylsulfate polyacrylamide gel electrophoresis (SDS-PAGE) and electrophoretically transferred to polyvinylidene difluoride (PVDF) membranes (Millipore). The membranes were blocked with 0.1% BSA in Tris-buffered saline Tween-20 (TBST) buffer and incubated overnight at 4°C with anti-SSEA-1 antibody. The membranes were washed with TBST buffer and incubated with horseradish

peroxidase-conjugated secondary antibodies. Enhanced chemiluminescence kits were used (Amersham, ECL kits).

### Chemotaxis analysis

The number of CXCR4-positive cells among mES cells was analyzed using a flow cytometer. PE-labeled anti-CXCR4 antibody (BioLegend) was used to sort for CXCR4-positive cells. Amount of CXCL12 in the culture supernatants of MFC cells was examined by enzyme-linked immunosorbent assay (ELISA) with commercial kits (R & D System). The migration ability of mES cells was evaluated using a 48-well modified Boyden chamber. The polycarbonate filter (12  $\mu m$  pore size, CN110416; Neuroprobe, Bethesda, MD) was precoated with fibronectin (5  $\mu g/mL$ ; Sigma Chemical, St Louis, MO). The mES cells were resuspended at  $5 \times 10^5/mL$  in the appropriate medium supplemented with 10% FBS and seeded in the upper chamber. Recombinant CXCL12 (R&D System) was used as a chemoattractant in the lower compartment. The chambers were incubated overnight at 37°C. Results were expressed as the percentage of migrating cells. Each experiment was performed in triplicate.

### Statistical Analysis

All data were presented in this paper as means result  $\pm$  S.D. Statistical differences were evaluated using the t-test and considered significance at  $P < 0.05$  level.

## Results and Discussion

### Morphological observation of cultured mES cells

As shown in Fig.1A and 1C, GFP-SV 129 mES cells cultured on the feeder cells formed typical large colonies with long-ellipse or long-shuttle shape, mES cells exhibited small enchyrama and big nucleoli with obvious stereoscopic traits and clear boundaries when cultured with feeder cells. It also had strong green fluorescence signals since this mES cell line carries the GFP marker, as shown in Fig.1B. Although mES cells in feeder-free layer also maintained a normal karyotype, remarkable changes in mES cell shape grown in a monolayer can be observed as shown in Fig.1D and 1F, the feeder-free mES cells also have strong green fluorescence signal (Fig.1E).

### *In vitro* fluorescence imaging of DiR-mES cells

The NIR property of DiR dye makes it ideal for *in vivo* imaging. After the mES cells were labeled with DiR, DiR-mES cells ( $5 \times 10^6$ ) were resuspended and

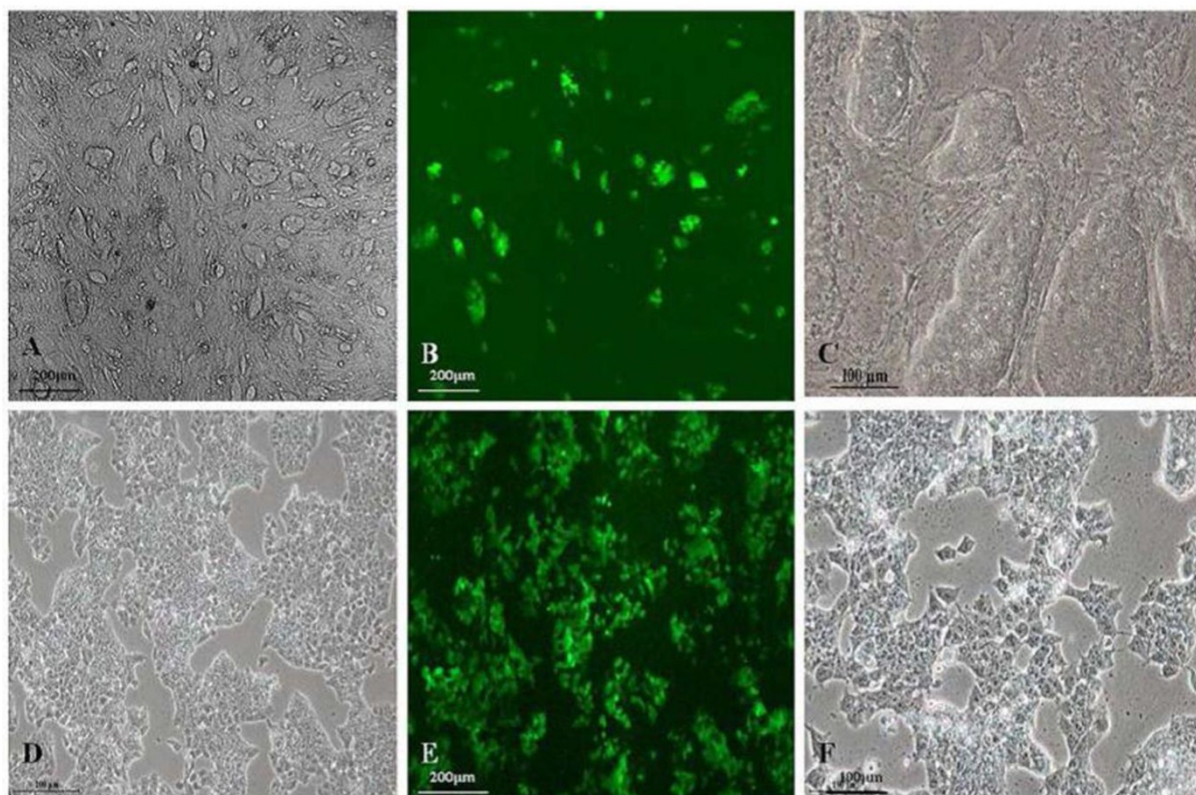


added into 96-well plate and performed fluorescence imaging at 0, 4, 8, 12, and 24 h, respectively. The results showed that the fluorescence signals of DiR(+) cells were stronger than that of DiR(-) cells. Although the fluorescent signals decreased over the time as shown in Fig.2A, it has statistical significance compared to the DiR(-) cells. Meanwhile, fluorescence imaging of different concentrations of DiR(+) cells were performed as shown in Fig.2B. The linear relationship ( $r^2=0.9939$ ) between Fluc activity and cell number confirms that the mES cells were successfully labeled with DiR dye. Fig.2C shows the result of trypan blue staining, which suggests that the DiR(+) mES cells had good viability similar with the DiR(-) mES cells. Fig.2D shows that DiR-mES cells grew very well after being labeled with DiR. These data highly suggest that DiR-mES cells had good cellular bioactivity and were suitable for *in vivo* imaging and tracking.

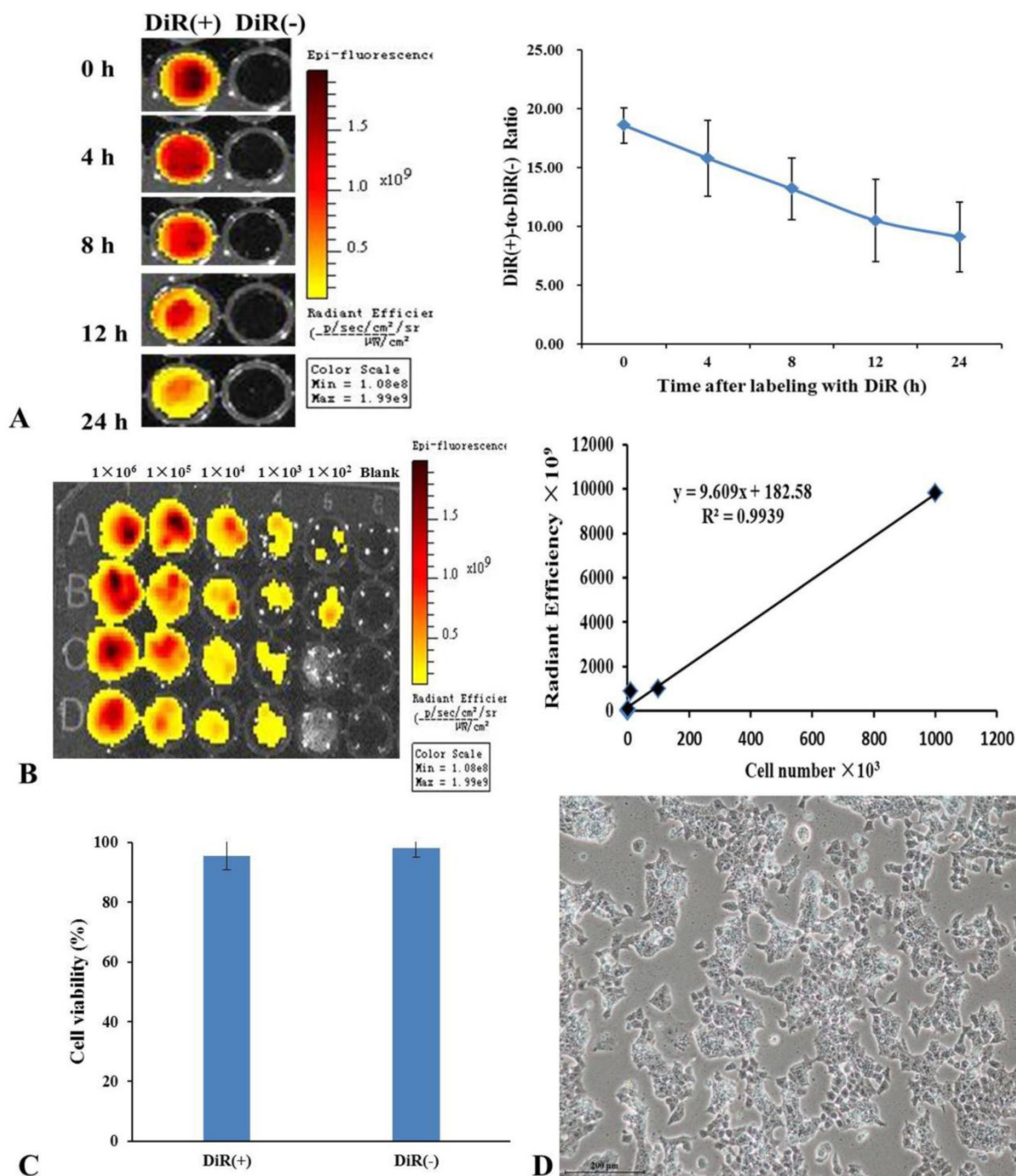
### ***In vivo* fluorescence imaging of DiR-mES cells in gastric cancer mouse models**

In order to assess the biodistribution of mES cells in bioluminescence gastric cancer nude mouse models. DiR-GFP-mES cells were intravenously injected

into gastric cancer mouse models as test group. In control group, gastric cancer mouse models were injected with DiR dye. The distribution of DiR-mES cells in gastric cancer nude mouse models was monitored within 24 h by IVIS imaging system at lateral, supine and prone positions. As shown in Fig.3, fluorescent signals in subcutaneous tumor tissues in test group were clearly differentiated from the surrounding tissues at 10 min post-injection at lateral position. Fluorescent signals in tumor tissues gradually increased, reaching a maximum value at 2 h post-injection. The higher tissue to background ratio (TBR) value suggests that the DiR-mES cells preferentially accumulated in tumor tissues and achieved their maximum value at 2 h post-injection. Conversely, no fluorescent signals could be observed in the tumor tissues in control group. In addition, the *in vivo* imaging of the mouse models was performed at supine position and prone position as shown in Supplementary Material: Fig.S1. These results show that DiR-mES cells mainly located in the tumor tissues, and very few DiR-mES cells located in the liver, lung and kidney organs, similar to several reports [24-26].

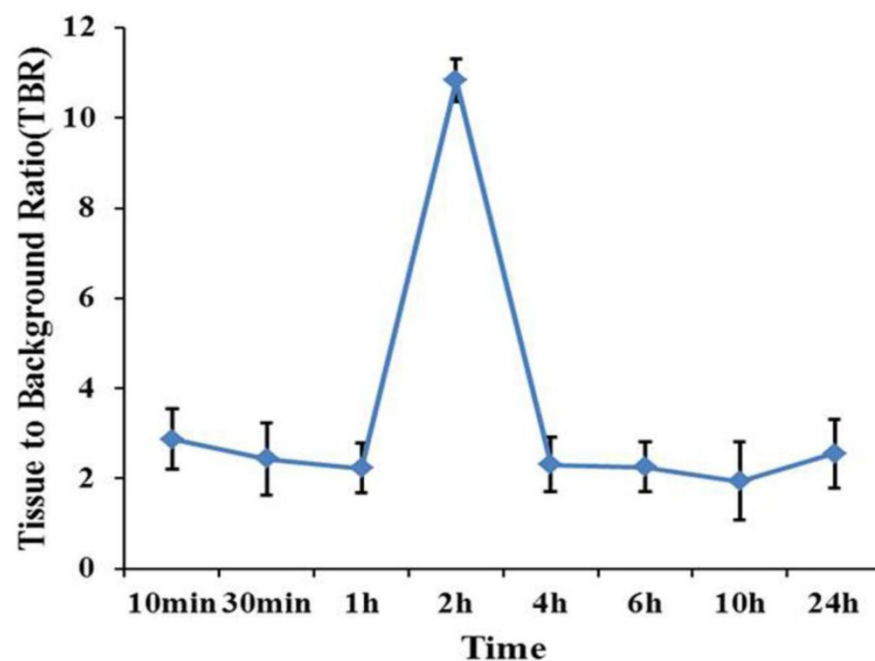
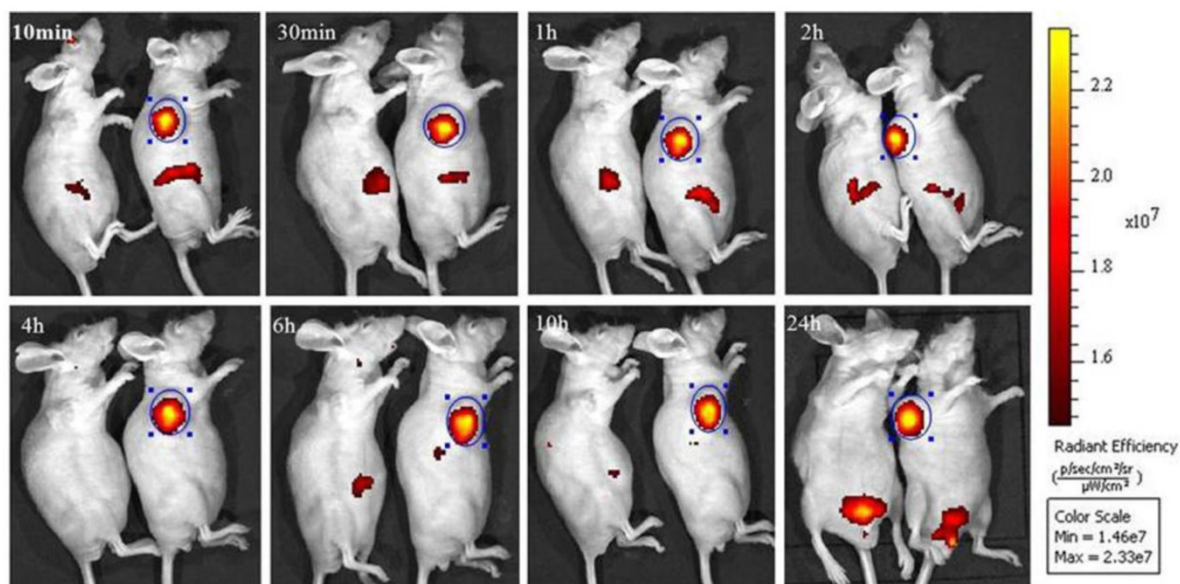


**Fig.1** Fluorescence microscope images of the morphology of GFP-SV129 mES cells. (A) GFP-SV129 mES cells exhibited typical large colonies and clearly boundaries with feeder cells. (B) mES cells expressed strong green fluorescence. (C) Magnification of mES cells cultured on feeder cells. (D) Feeder-free culture of mES cells grew in monolayer. (E) Feeder-free culture of mES cells exhibited green fluorescence. (F) Magnification of feeder-free culture of mES cells.



**Fig.2** (A) DiR-labeled mES cells were subjected to fluorescent imaging at different exposure time, the ratio of DiR-labeled mES cells to unlabeled mES cells shows that the DiR-mES cells had strong fluorescent signals within 24 h. (B) The quantitative analysis of fluorescence of DiR(+) cells showed a linear relationship ( $r^2=0.9939$ ) between Fluc activity and cell numbers. (C) Trypan blue staining shows that mES cells had good cell viability after being labeled with DiR. (D) mES cells grew very well after being labeled with DiR 24 h.





**Fig.3** *In vivo* fluorescence images of gastric cancer mouse models at 10 min, 30 min, 1 h, 2 h, 4 h, 6 h, 10 h, 24 h post-injection DiR-mES cells at lateral position. It shows that tumor tissues had strong fluorescence signals after injection DiR-mES cells as shown in blue circles. The tissue to background ratio (TBR) values show that DiR-mES cells accumulated in the tumor tissues and reached peak value at 2 h post-injection.

In order to track the distribution of DiR-mES cells in gastric cancer mouse models, the gastric cancer mouse models were performed bioluminescence and fluorescence imaging at 6 h post-injection. Fig.4A shows the NIR fluorescence imaging of the gastric cancer mouse models. Fig.4B shows the bioluminescence imaging of the gastric cancer mouse models. Fig.4C is a combination image of bioluminescence and

fluorescence imaging. These results fully confirm that DiR-mES cells could migrate to the tumor tissues precisely. As shown in Fig.4D, the *ex vivo* fluorescence imaging of major tissues in test and control groups shows that DiR-mES cells mainly existed in the tumor tissue of the test group. No obvious fluorescent signal were detected in the spleen and heart tissues of the test and control group, and weak fluorescent signals

were found in the lung, kidney and liver tissues of the test group. The reason for fluorescent signals found in liver tissue may be as following: DiR-mES cells were cleared from circulation by the liver and trapped within capillary beds in the liver [27]. As shown in Fig.4E, the radiant efficiency of tumor tissue in test group was the highest among all the tissues in the test and control groups, which further confirm that DiR-mES cells could preferentially accumulate in the tumor tissue and efficiently target and identify gastric cancer cells in gastric cancer mouse models.

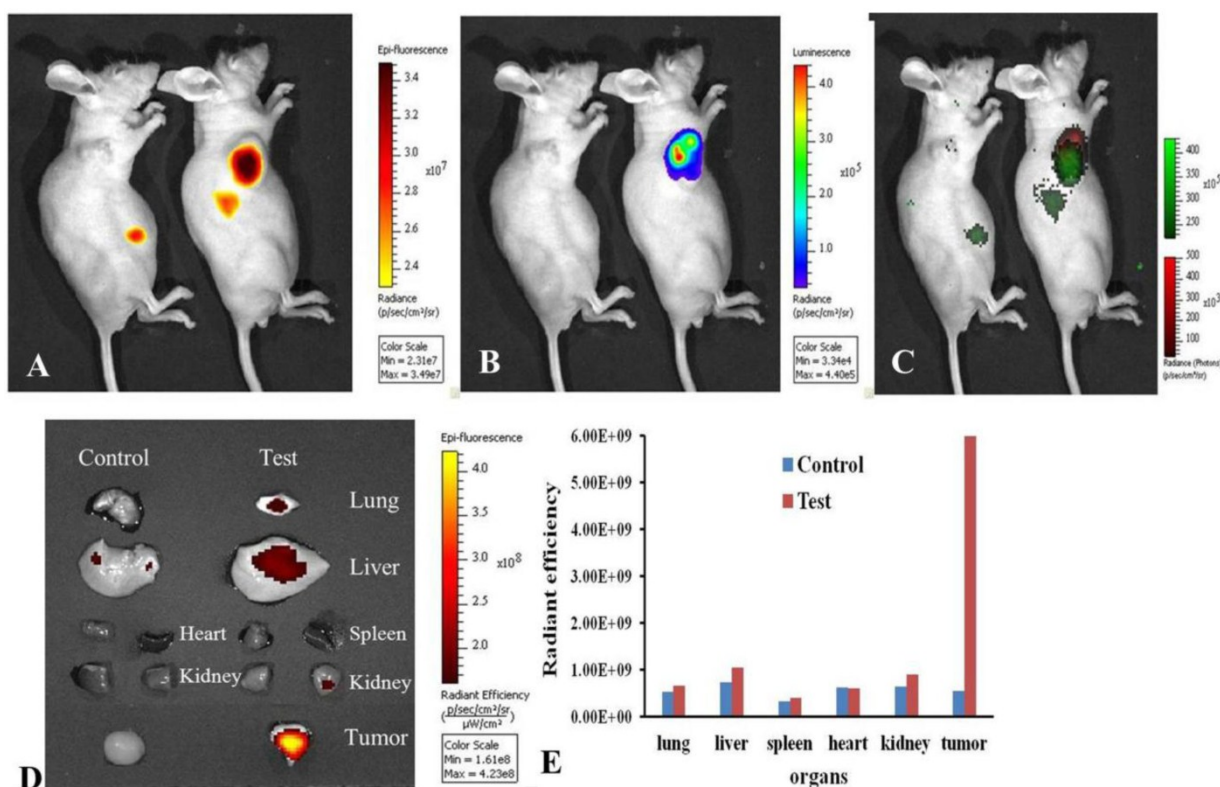
### Pathological analysis of tumor tissues and important organs

Pathological evaluation of excised major tissues including the liver, lung, spleen, kidney, heart, and tumor was performed. SSEA-1 is a specific, highly expressed surface antigen of mES cells, so the tumor tissues must express SSEA-1 once the mES cells migrated into the tumor tissues. As shown in Fig.5A, GFP and SSEA-1 fluorescent signals were significant

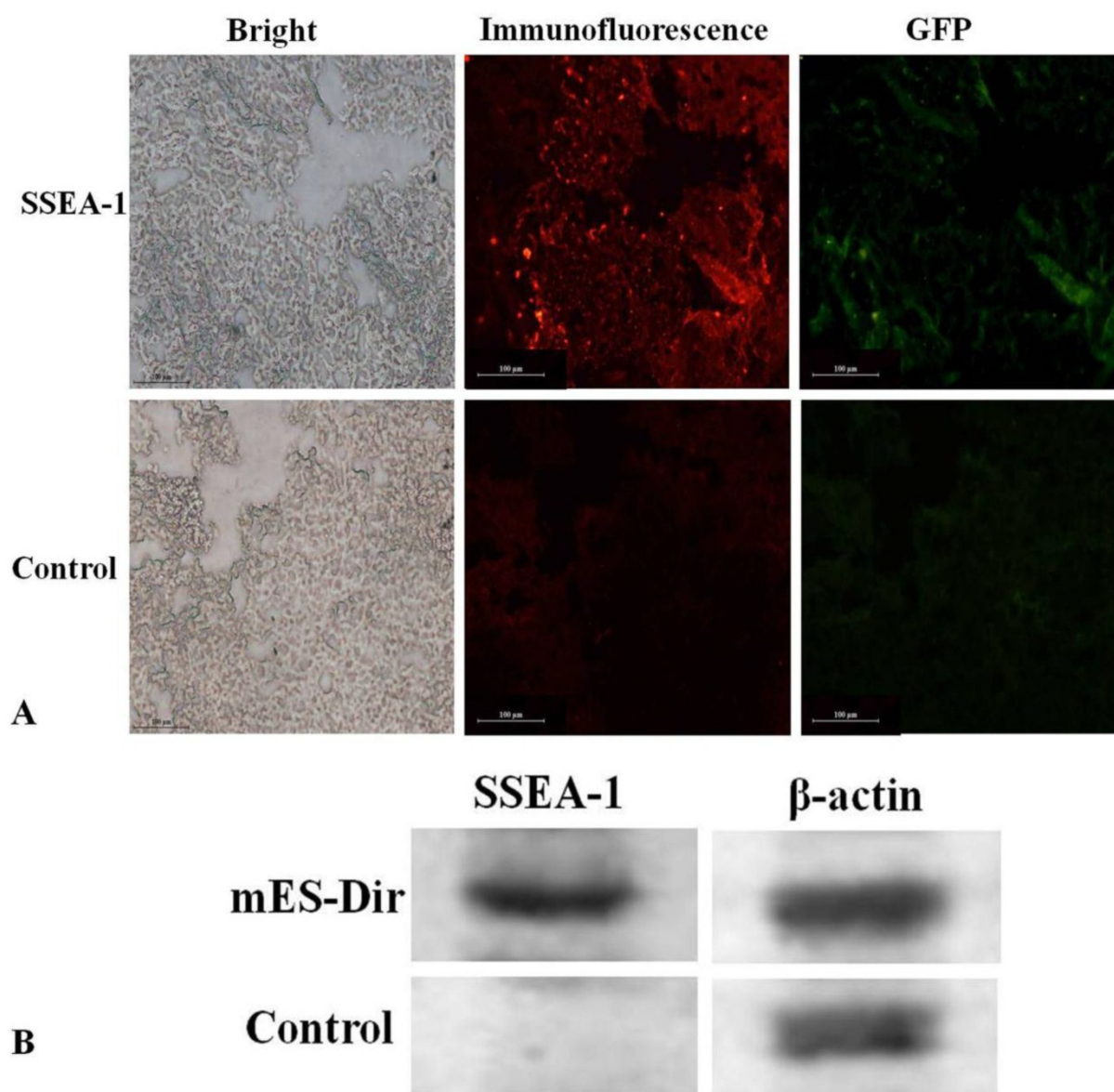
observed from the ultra-thin tumor tissue slices of the test group after performed immunofluorescence staining, and green fluorescence signals were consistent with fluorescent signals in the tumor tissues of the test group, no GFP and immunofluorescent signals were found in the tumor tissues of the control group. Furthermore, as shown in Supplementary Material: Fig.S2, no fluorescent signal was observed in the liver, lung, spleen, heart and kidney tissues of the test group. These results fully suggest that mES cells could specifically target gastric cancer cells *in vivo*.

### Western blot analysis of gastric cancer tissues

The expression of SSEA-1 in excised tumor tissues was investigated using Western blotting. The  $\beta$ -actin protein was used as a reference. Fig.5B shows that SSEA-1 was highly expressed in the tumor tissues of the test group, while it did not express in the tumor tissues of the control group. This result suggests that mES cells mainly located in the tumor tissues *in vivo*.



**Fig.4** Optical imaging of DiR-mES cells *in vivo/ex vivo*. (A) Fluorescence imaging of gastric cancer mouse models at 6 h post-injection at lateral position; (B) Bioluminescence imaging of gastric cancer mouse models at 6 h post-injection at lateral position; (C) Combination image of fluorescence and bioluminescence imaging shows that DiR-mES cells could target the tumor tissues exactly. (D) *Ex vivo* fluorescent image of the dissected organs and tumors in the control group and test group at 24 h post-injection. (E) Radiant efficiency of the tissues in the test group and control group.



**Fig.5** (A) Immunofluorescence analysis of the tumor tissues. (B) Western blot analysis of the tumor tissues.

### Possible mechanism of the migration of mES cells to gastric cancer cells

Tumor-homing is a complex, multistep process in which many cells move from distant locations to a tumor site. Homing mES cells may be activated by specific signals, and they can travel through the circulation, extravasate from vessels, migrate, and undergo phenotypic changes when it finally reaches the tumor site. The mechanism might be similar to the metastatic cascade [28]. So far, the process of tumor-homing associated with mES cells is still not clarified. The term “tumor-homing” indicates any action in which cells travel from a distant location to a tumor site, indicating an active filopodia-based mo-

tion through a tumor or surrounding local microenvironment based on local chemoattractants might be involved [29–36]. For example, CXCR4-CXCL12 loop was in charge of metastasis of the breast cancer [37]. The interaction between CXCL12 and CXCR4 has been implicated in the bone metastasis in prostate cancer [38]. The CXCL12/CXCR4 axis has been shown to be involved in the metastasis of non-small-cell lung cancer cells [39]. Thus, we speculate that CXCL12-CXCR4 loop may be involved in the course of mES cells migrating to the gastric cancer cells *in vivo*.

As shown in Fig.6, flow cytometer analysis results show that the positive rate of CXCR4 expression was 9.46 % in mES cells. ELISA results show that

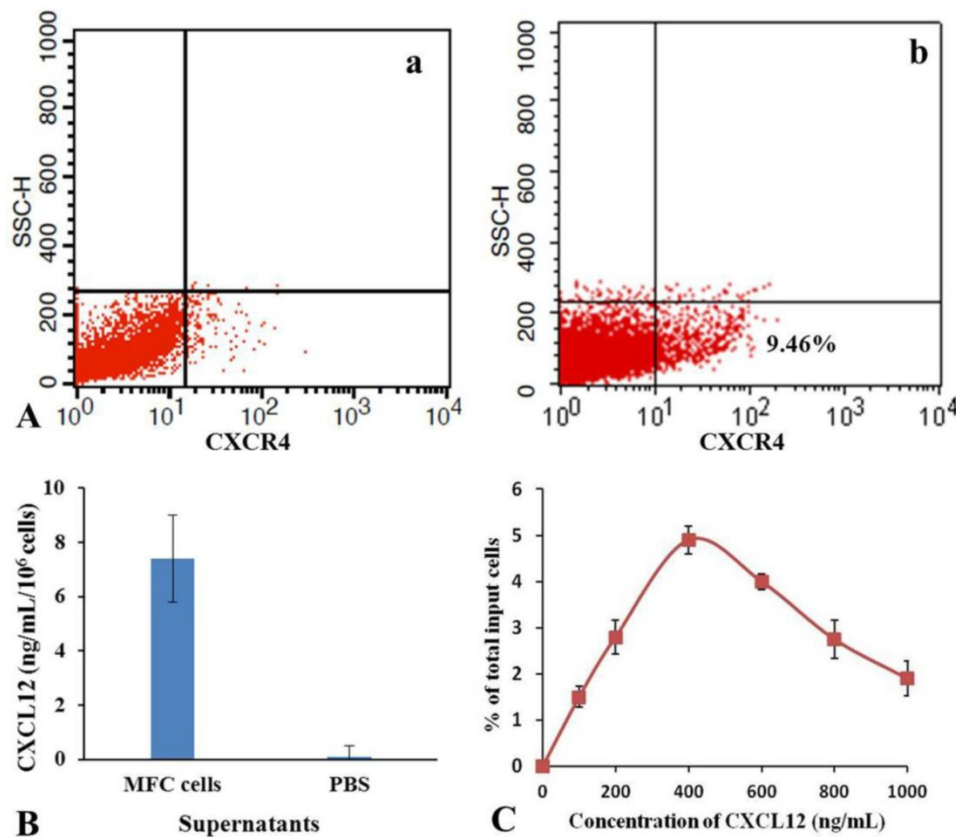


amounts of CXCL12 in the culture supernatant of MFC cells were  $7.2 \pm 0.7$  ng/mL. Cell and chemoattractant dose-response curves show that CXCL12 induced chemotaxis of mES cells in a dose-dependent, bell-shaped manner with maximum chemotactic response seen at 400 ng/mL of CXCL12. According to these results, we suggested a possible mechanism of mES cells migrating to gastric cancer cells *in vivo*. DiR-labeled mES cells enter the tail vein vessels, and then rapidly circulate into lung vessels. From there, they enter into artery vessels, circulating throughout the mouse body. Some mES cells enter the liver and are captured and endocytosed by the phagocyte, finally are trapped in the liver. High amounts of CXCL12 were found in the surrounding of the tumor tissue, providing a strong chemotactic signal. Since mES cells positive expressed CXCR4 receptor, they are very sensitive to CXCL12 factor secreted by gastric cancer tissue, which triggers the rapid mES cells migration toward gastric cancer tissues. In the course of migration of mES cells to gastric cancer tissues, it is possible that some other chemokine factors are also involved, therefore, a comprehensive study evaluat-

ing other potential mechanisms is under way.

Stem cell therapy is a promising method for cancer treatment [40-42]. Especially in 2004, there is one research reported that when the human embryonic stem (hES) cells were transfected with an anti-cancer molecular named "TRAIL", the transfected hES cells could migrate to the brain tumor site and significant prevent the tumor growth [43]. The targeting ability is the key for stem cell therapy of tumors. In this research, we clearly observed that mES cells have good ability of targeting gastric cancer. This discovery lays foundation for diagnosing early gastric cancer with embryonic stem cells.

In this study, our results indicate that DiR-labeled mES cells could target and recognize cancer cells *in vivo*. In addition to gastric cancer mouse model, nude mouse models with lung cancer, liver cancer, and prostate cancer were also prepared. It is found that DiR-labeled mES cells could target and recognize those different cancer cells *in vivo* as well (data not shown). However, the detailed molecular mechanism of tumor targeting still requires further investigation.



**Fig.6** Analysis of chemotaxis. (A) Expression of CXCR4 in isotype control(a) and mES cells (b) were determined by flow cytometry. (B) Secretion of CXCL12 in MFC cells was measured by enzyme-linked immunosorbent assay. (C) Cell and chemoattractant dose-response curves.

## Conclusions

In conclusion, this is the first report showing that DiR-labeled mES cells can selectively migrate to gastric cancer tissues *in vivo* and can be used for targeted NIR imaging of gastric cancer cells *in vivo*. The ligand-receptor combinations of CXCL12 and CXCR4 may be responsible for the migration of mES cells to the gastric cancer cells. Although the detailed mechanism still requires further investigation, we believe that this phenomenon may be general in other tumor models. Therefore, it has great potential in imaging detection of early and metastatic cancer cells in the near future.

## Supplementary Material

Fig.S1 – S2. <http://www.thno.org/v02p0618s1.pdf>

## Acknowledgements

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## Competing Interests

The authors have declared that no competing interest exists.

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