Remote actuation and on-demand activation of biomaterials pre-incorporated with

physical cues for bone repair

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Abstract

The high incidence of bone defect-related diseases caused by trauma, infection, and tumor

resection has greatly stimulated research in the field of bone regeneration. Generally, bone healing

is a long and complicated process wherein manipulating the biological activity of interventional

scaffolds to support long-term bone regeneration is significant for treating bone-related diseases.

It has been reported that some physical cues can act as growth factor substitutes to promote

osteogenesis through continuous activation of endogenous signaling pathways. This review

focuses on the latest progress in bone repair by remote actuation and on-demand activation of

biomaterials pre-incorporated with physical cues (heat, electricity, and magnetism). As an

alternative method to treat bone defects, physical cues show many advantages, including

effectiveness, noninvasiveness, and remote manipulation. First, we introduce the impact of

different physical cues on bone repair and potential internal regulatory mechanisms. Subsequently,

biomaterials that mediate various physical cues in bone repair and their respective characteristics

are summarized. Additionally, challenges are discussed, aiming to provide new insights and

suggestions for developing intelligent biomaterials to treat bone defects and promote clinical

translation.

Keywords: biomaterials, bone repair, on-demand activation, physical cues, remote actuation

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List of abbreviations

NIR	Near-infrared	PMMA	Polymethyl methacrylate
AMF	Alternating magnetic field	RGD	Arg-Gly-Asp
MNPs	Magnetic nanoparticles	COL	Collagen
MSCs	Mesenchymal stem cells	TGF-β	Transforming growth factor-β
HSPs	Heat shock proteins	BMP	Bone morphogenetic protein
ALP	Alkaline phosphatase	FAK	Focal adhesion kinase
PTT	Photothermal therapy	PLA	Poly-DL-lactide
BP	Black phosphorus	PPy	Polypyrrole
PDA	Polydopamine	PANI	Polyaniline
BMSCs	Bone mesenchymal stem cells	FPAT	Polyurethane
PCL	Poly(ε-caprolactone)	PVDF	Polyvinylidene fluoride
CDs	Carbon dots	P(VDF-TrFE)	Poly(vinylidene fluoridetrifluoroethylene)
GO	Graphene oxide	β-ТСР	β-Tricalcium phosphate
CNTs	Carbon nanotubes	PHB	Poly[(<i>R</i>)3-hydroxybutyrate]
НА	Hydroxyapatite	PHBV	Poly[3-hydroxybutyrate-co-3-hydroxyvalerate]
CS	Chitosan	SMF	Static magnetic field
ROS	Reactive oxygen species	cGMP	Cyclic guanosine monophosphate
PLGA	Poly(lactic-co-glycolic acid)	PKG	Protein kinase G
PCMs	Phase change materials	ERK	Extracellular signal-regulated kinase
MTT	Magnetothermal therapy		

1. Introduction

In recent decades, a large number of patients have suffered bone defects caused by bone tumors, osteoporosis, arthritis, and other related diseases, requiring orthopedic repair materials. Bone repair is a complicated, precise, and slow process that can be tuned by various factors. In addition to basic structural support, bone repairing materials are also desired to meet the specific functionalities performed by bioactive factors involved in bone healing at different repair stages. Accordingly, various synthetic scaffolds have been developed to provide biocompatible and bioactive platforms for bone tissue regeneration [1–4]. Physical cues such as heat, electricity, magnetism, ultrasound, light, and mechanical forces provide superior convenience with remote operation and have been extensively studied for their safety and efficacy in promoting bone regeneration by activating endogenous regulators [5]. However, how to well integrate physical cues with biomaterials to achieve precise and controlled modulation is challenging. This reveiw will discuss these physical cues pre-integrated into bone-repairing materials and activated by exogenous stimuli with high biosafety and strong tissue penetration, such as light, electricity, and magnetic field, enabling remote actuation and on-demand activation for bone regeneration.

As an emerging treatment modality, thermal-based therapy has made remarkable progresses in the treatment of tumors [6], osteoarthritis [7], bone infection [8], and other bone-related diseases [9]. Studies have shown that a mild thermal effect (40–43 °C) favors cell activity, proliferation and differentiation, thereby promoting bone defect repair. The highest osteogenic activity is observed with thermal stimulation at 42 °C twice a day, as prolonged exposure times at a certain temperature result in tissue anesthesia and cell death, whereas short exposure times have no effect [10,11]. However, the outcomes may be different for different disease models and laser parameters,

and extensive trials are needed to determine the optimal experimental conditions. In order to increase the utilization of thermal effect and meanwhile enable the safety of surrounding healthy tissues, exogenous stimuli that penetrate deep into the tissues, such as near-infrared (NIR) light [12] and magnetic field [13], are often used to activate implanted biomaterials *in situ* and generate local thermal energy for treatment. Accordingly, photosensitizers with strong NIR light absorption and high photothermal conversion efficiency have been developed for bone defect repair [14]. Besides light-induced thermal effect, resonance between the spin magnetic moment of the magnetic composite scaffold and an external alternating magnetic field (AMF) can also provide a local warm environment to promote bone repair [15].

Bioelectricity is an important physiological activity of living organisms, and bone is a natural piezoelectric material; therefore, electrical signals can be used as stimuli to promote bone growth and remodeling. Electrical stimulation can regulate many biological processes, ranging from cell cycle, migration, proliferation, restoring nerve conduction, muscle contraction, embryogenesis, and tissue regeneration [16]. Restoration of the natural bioelectric properties of healthy bone tissue using electroactive biomaterials with electrode-free, wireless, and self-charging capabilities, such as piezoelectricity, provides a favorable microenvironment for bone regeneration [17]. Commonly used electroactive materials in the biomedical field mainly include conductive materials (such as carbon-based nanomaterials, metallic materials, and conductive polymers, etc.), piezoelectric materials (like piezoelectric ceramics, piezoelectric polymers, etc.) [18] and other electroactive materials (magnetoelectric and photoelectric materials, etc.).

Low-frequency (below 1 MHz) homogeneous magnetic fields have received much attention for their ability to penetrate living tissues [19]. In addition, magnetic fields can improve blood circulation at bone defects, stimulate osteoblast and chondrocyte generation, inhibit the activity of

osteoclasts, and accelerate bone calcification, which helps combat osteoporosis and bone defects [20–22]. Magnetic nanoparticles (MNPs) have been widely used in drug delivery, magnetic resonance imaging, and bone repair due to their excellent biocompatibility and magnetism [23,24]. Under an exogenous magnetic field, the MNPs are rapidly magnetized, exerting a sustained weak magnetic force on the cells, thus making the superparamagnetic scaffolds better for bone regeneration than the magnetic field or MNPs alone. The positive effect of magnetic scaffolds, remotely coupled with magnetic fields, was confirmed on cell differentiation [25]. Although the exact mechanism is still being investigated, this non-/less invasive bone repair strategy is still of great interest.

Remote control of material properties through exogenous stimuli and on-demand activation of physical cues to guide the behavior of cells and organisms is important for developing bone tissue engineering. This review summarizes the generation of biomaterial-mediated physical cues (thermal effect, electrical and magnetic signals) and their impacts on bone repair processes, such as cell adhesion, migration, and differentiation (Figure 1). Material classification and corresponding outcomes of each physical cue are illustrated by *in vitro* and *in vivo* experimental results. This review will guide the development and clinical translation of intelligent stimulation-responsive materials for better bone regeneration outcomes.

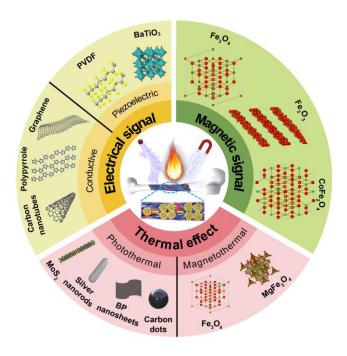


Figure 1. Schematic illustration of biomaterial-mediated physical cues (thermal effect, electrical and magnetic signals) for bone repair by remote actuation and on-demand activation.

2. Biomaterials-mediated mild thermal effect

Mild thermal therapy is flourishing as a promising method for bone regeneration. Exogenous stimuli can activate bone repairing scaffolds on demand to generate a mild thermal effect, accelerating the healing of bone and surrounding tissues and achieving targeted therapy and precise regulation. NIR light and magnetic fields are ideal energy sources due to their tissue penetration, remote operability, and spatiotemporal accuracy. With the rapid development of nanotechnology, a series of biomaterials (photosensitizers and magnetic materials) that can respond to NIR light and magnetic fields to generate efficient thermal response and achieve favorable thermotherapeutic effects have emerged. After implantation of these biomaterials, exogenous laser irradiation (photothermal therapy, PTT) and magnetic field stimulation (magnetothermal therapy, MTT) can effectively promote osteocyte proliferation, tissue regeneration, and mineralization (Table 1). This

section will discusses the impacts of the thermal effect on bone repair, followed by applications of photothermal and magnetothermal materials to treat bone defects.

Table 1. Summary of typical photothermal and magnetothermal materials for bone repair.

Classification	Materials ^{Ref}	Exogenous stimuli (in vitro)	Osteogenic impact (in vitro)	Animal models	In vivo results
		(iii viiro)	(iii viii 0)		
Photothermal	Photothermal eP@GNRs ^[26]		Promote osteogenic	Femur fracture in	Promote
metallic		cm ⁻² , 15 min	differentiation (BMSCs)	mice	fracture
materials					healing
	pAuPds ^[27]	808 nm, 2 W	Promote proliferation	Cranial defect in rat	Partial
		cm ⁻² , 3 min	(MC3T3-E1)	(Ø 8 mm)	bridging
Photothermal	P and Si	808 nm, 0.25 W	Promote adhesion, proliferation,	Femoral defect in rat	Partial
metal oxides	co-doped TiO ₂ ^[28]	cm ⁻² , 3 min	and differentiation (MC3T3-E1)	(Ø 1.6 mm)	bridging
	PPCL/Nd@WH ^[29]	808 nm, 0.5 W	Promote osteogenic	Cranial defect in rat	Partial
		cm ⁻² , 5 min	differentiation (BMSCs)	(Ø 4 mm)	bridging
Photothermal	PCL/MoS ₂ ^[30]	808 nm, 0.5 W	Promote osteogenic	Tibia defect in rat	Bridging
metal sulfides		cm^{-2} , 40.5 ± 0.5 °C for 60 s	differentiation (BMSCs)	(Ø 2.5 mm × 3 mm)	
	PTEB (MoS ₂) ^[31]	808 nm, 1.5 W	Promote proliferation and	Cranial defect in rat	Bridging
		cm ⁻² , 41 °C for 10 min	differentiation (BMSCs)	(Ø 5 mm)	
	D-CuS-PEG-PCL	1064 nm, 1.0 W	Promote osteogenic	Tibia defect in rat	Partial
	scaffold ^[32]	cm ⁻² , 10 min	differentiation (BMSCs)	$(\emptyset 3 \text{ mm} \times 3 \text{ mm})$	bridging
Photothermal	AMAD/MP hydrogel	808 nm, 1.0 W	Promote osteogenic	Cranial defect in rat	Partial
carbon materials	(MXene) ^[33]	cm^{-2} , 42 ± 0.5 °C for 120 s	differentiation (MC3T3-E1)	(Ø 5 mm)	bridging
	nHA/GO/CS scaffold ^[34]	808 nm, 1.0 W	Promote osteogenesis	Cranial defect in rat	Partial
		cm^{-2} , $42 \pm 0.5 ^{\circ}C$ for 60 s	(human BMSCs)	(Ø 5 mm)	bridging
	CNT-alginate gel ^[35]	NIR apparatus,	Promote mineral deposition	Cranial defect in rat	Partial
		4.0 W cm ⁻² , 42 °C for 15 min	(MG63 and DP cells)	(Ø 8 mm)	bridging

Other photothermal compounds	BMP-2@blood clot ^[36]	808 nm, 0.4 W cm ⁻² , 10 min	Promote osteoblast proliferation and differentiation (MC3T3-E1)	Cranial defect in mice (Ø 4 mm)	Partial bridging
	GelMA/PMMA/PDA hydrogel ^[37]	808 nm, 0.99 W cm ⁻² , 40 ± 0.5 °C for 60 s	Promote osteogenic differentiation (BMSCs)	Cranial defect in rat (Ø 5 mm)	Partial bridging
	CS/PCL/BP/PDA@Ag scaffold ^[38]	808 nm, 0.45 W cm ⁻² , 41 \pm 0.5 °C for 60 s	Promote osteogenic differentiation (BMSCs)	Femoral defect in rat (Ø 2.5 mm × 3 mm)	Partial bridging
	BPs@PLGA ^[39]	808 nm, 1.0 W cm ⁻² , 40.5 \pm 0.5 °C, 60 s	Promote osteogenesis (human BMSCs)	Tibia defect in rat (Ø 2.5 mm × 3 mm)	Partial bridging
	Apt19S-PCL-BP scaffold ^[40]	808 nm, 0.8 W cm ⁻² , 2 min	Promote osteogenic differentiation (MSCs)	Cranial defect in rat (Ø 5 mm)	Partial bridging
Magnetothermal materials	MCPC/GM/HMFNs/CP FX/Van (ferrite nanoparticles) ^[15]	AMF, 2.5 kW, 200 kHz, 18 A, 5 min	Promote osteogenic differentiation and mineralization (BMSCs)	Femoral defect in rat (Ø 3 mm × 3 mm)	Partial bridging
	$MION\text{-}RGD/agarose$ $(CoFe_2O_4@MnFe_2O_4)^{[41]}$	AMF, 1.35 kA m ⁻¹ , 5 min	Promote osteogenesis and biomineralization (MC3T3-E1)	Cranial defect in rat (Ø 5 mm)	Partial bridging

2.1. Impact of mild thermal effect

Temperature control is important for hyperthermia-based therapies because it regulates physiological changes through elevated temperature. High temperatures (>45 °C) trigger DNA and protein denaturation or internal stress response, leading to apoptosis or necrosis, which are suitable for antibacterial and anti-tumor treatments. Mild thermal effect resulted by NIR light and magnetic fields can promote osteogenic differentiation and new bone formation. A low-intensity laser can stimulate the proliferation and osteogenesis of osteoblasts at the fracture site, promote cell migration and differentiation, reduce inflammatory response, and relieve pain through photobiomodulation therapy (Figure 2A) [42,43]. Under NIR laser irradiation, the photosensitizers

enriched in the bone defect site produce a mild thermal effect, which in turn promotes the differentiation of mesenchymal stem cells (MSCs), and the maturation and mineralization of osteoblasts (Figure 2B) [42]. NIR light-mediated heat stress activates the expression of heat shock proteins (HSPs), subsequently influencing bone metabolic pathways and angiogenesis. This molecular response plays a crucial role in regulating the proliferation and differentiation of osteoblast-related cells (Figure 2C) [14]. Mild hyperthermia induced by magnetic composites stimulated under a magnetic field with strong tissue penetration can effectively restore critical-sized bone defect (Figure 2D), because the thermal effect significantly promotes osteogenic differentiation and biomineralization of preosteoblasts via the HSP90-activated PI3K/Akt pathway (Figure 2E) [41].

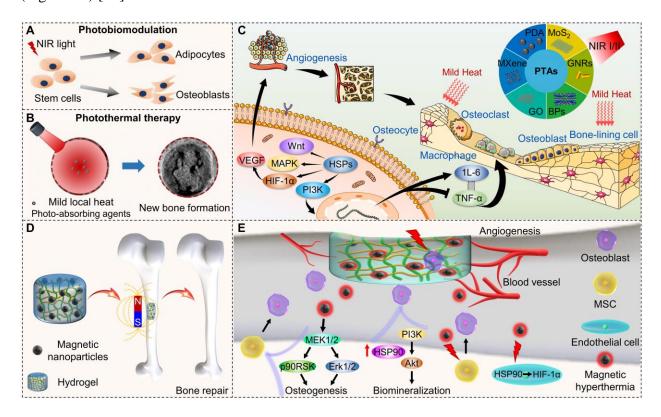


Figure 2. (A) Schematic of photobiomodulation under NIR light irradiation to regulate the attachment, proliferation, and differentiation of stem cells. Reproduced with permission from [42]. Copyright 2020, Ivyspring International. (B) New bone formation by mild thermal effect of NIR

light. Reproduced with permission from [42]. Copyright 2020, Ivyspring International. (C) Possible mechanisms of mild thermal effect promoting bone regeneration and angiogenesis; commonly used photosensitizers for bone regeneration. Reproduced with permission from [14]. Copyright 2023, Elsevier. (D) Schematic of magnetothermal effect promotes new bone formation. (E) Potential mechanisms of osteogenesis, biomineralization, and angiogenesis upon under magnetothermal effect.

In summary, the potential mechanisms of thermal effect on bone repair are listed as follows: (1) Warm stimulation helps to dilate blood vessels, improve local blood circulation, increase the supply of oxygen and necessary nutrients, and is beneficial to the healing of defects and surrounding soft tissues. (2) Heat stress induces the expression of osteogenesis-related proteins, such as alkaline phosphatase (ALP) and HSPs (especially HSP27, HSP47, HSP60, HSP70, and HSP90), which further promote immune remodeling, vascularization and osteogenesis. HSPs regulate the expression and conformation of osteogenic factors to adapt to a variety of physical and chemical stimuli such as hyperthermia, oxidative stress and ionizing radiation. When cells are exposed to high temperatures, the expression of HSPs is significantly increased, preventing protein denaturation and maintaining protein activity. Also, HSPs are involved in bone metabolism stages, such as bone resorption and bone regeneration, and play an important role in fracture healing [44]. For example, HSP60 can promote the formation of osteoclasts via p38 MAPK and NF-κB pathways and affect the status and proliferation of osteoblasts [44]. HSP27 is involved in specific regional and temporal expressions during tooth development [45]. The upregulation of HSP90 by hyperthermia activates the downstream PI3K/Akt pathway and promotes bone regeneration [41]. (3) Periodic mild photothermal stimulation can stimulate the immune system and related inflammatory processes, such as promoting macrophages to the anti-inflammatory M2 phenotype

and increasing the expression of anti-inflammatory cytokines [46]. (4) Thermal effect enhances angiogenesis through improving the expression of vascular endothelial growth factor and platelet-endothelial cell adhesion molecule (CD31) [47].

2.2. Photothermal materials

The photothermal phenomenon refers to the absorption of photon energy by the photosensitizers accumulated in the lesion under laser irradiation. Then electrons are activated from the ground state singlet to the excited singlet state. Subsequently, the electron returns to the lower vibrational level after multiple collisions and releases heat during this vibrational relaxation [48,49]. As a non-/less invasive treatment, PTT uses photosensitizers to convert light energy into heat for treatment and repair, demonstrating the characteristics of high spatiotemporal selectivity, slight side effects, and strong controllability [50,51]. The hyperthermia generated during PTT is often used to ablate tumors and combat bacterial infections, while mild PTT shows potential for bone repair. The laser adopted here is mainly the NIR light located in the transparency window of biological tissues. Due to the limited depth of tissue penetration, NIR light-induced PTT is more suitable for the repair of superficial tissue damage. With the rapid development of nanotechnology, the laser has evolved from NIR-I (750-1000 nm) to NIR-II (1000-1350 nm) region for deeper tissue penetration, higher biosafety, and larger maximum allowable exposure [52-54]. Photosensitizers with different absorption wavelengths have also been extensively studied. An ideal photosensitizer presents high photothermal conversion efficiency, good photothermal stability and biocompatibility. According to the type of materials, the commonly used photosensitizers in bone defect repair mainly include metals, metal oxides or sulfides, carbon materials, and other types of materials (such as black phosphorus (BP), polydopamine (PDA), etc).

2.2.1. Metallic photosensitizers

Metallic photosensitizers represented by gold nanomaterials show great potential in the biomedical field due to their easy synthesis, good biocompatibility, favorable stability, and outstanding photothermal effect. Notably, a study of the clinical pilot device using the local thermal effect of gold nanoparticles for prostate cancer treatment has demonstrated feasibility and safety in patients with localized low- or intermediate-risk prostate cancer, showing potential in clinical translation [55]. In addition to eradicating tumors, the mild thermal effect of metallic materials can also promote bone tissue regeneration. Gold nanorods modified with endogenous proteins collected from autologous blood can improve biocompatibility and reduce immune inflammation and rejection. Under laser irradiation, the mild photothermal effect of gold nanorods promotes osteogenic differentiation of bone mesenchymal stem cells (BMSCs) through MAPK, Akt, Smad, and β-catenin pathways [26]. Bismuth-doped glasses also show good photothermal performance. Under laser irradiation, these glasses improve biological activity and promote osteogenic cell proliferation, differentiation, and mineralization via photoinduced hyperthermia [56]. Porous AuPd alloy nanoparticles, as thermotherapeutic agents, significantly accelerate cell proliferation and bone regeneration through PTT [27]. Other metals (such as magnesium, iron, and manganese) and their alloys have attracted much attention in orthopedic implants due to their superior mechanical properties, biodegradability, and biological activity. Aside from stimulating the osteogenic differentiation of bone-forming cells by ionic products generated from metal degradation, the mild thermal effect caused by NIR light responsiveness also shows potential in bone regeneration [57,58].

2.2.2. Metal oxides or sulfides

Metal oxides or sulfides display improved osteogenic properties due to their mild photothermal stimulation. As a commonly used material in titanium implants, titanium dioxide

(TiO₂) is facile to prepare and has good biocompatibility. Although the absorption of TiO₂ is mainly located in the ultraviolet band, its absorption and photothermal performance in the NIR region can be effectively improved by doping ions (such as Si and P) to form black TiO₂ [59]. The mild photothermal effect of black TiO₂ and the release of doped ions triggered by NIR light can elevate the expression of osteogenic genes, regulate the behavior of osteoblasts, and also control inflammation, which can be used for peri-implantitis treatment [28,60,61]. A photothermal double-layer biomimetic periosteum based on Nd₂O₃ presents excellent bone tissue regeneration function [29].

MoS₂ nanosheets are a new class of layered two-dimensional metal sulfides. Due to the good biocompatibility and photothermal conversion efficiency, MoS₂ nanosheets show promising prospects in bone tissue engineering. MoS₂ and poly (ε-caprolactone) (PCL) nanofiber membranes obtained by electrospinning show favorable cell growth and osteogenesis properties, and the incorporation of MoS₂ improves the mechanical properties of nanofiber membranes without being affected by degradation. Under the mild hyperthermia effect induced by NIR light, these nanofiber membranes promoted the growth of BMSCs *in vitro* and helped to repair the rat tibial bone defect [30]. In another report, BMSCs were seeded into photothermal MoS₂-biotin-sucralose gelatin scaffolds to improve osteogenic activity. Subsequently, an osteoinductive extracellular matrix was used to cover these scaffolds to simulate the biomimetic microenvironment. The obtained scaffolds significantly induced bone regeneration in rats with bone defects [31].

Notably, the photothermal properties of some metal composites (such as SrCuSi₄O₁₀ [62], CuS [32], and WS₂ [63]) can be extended to the more prominent NIR-II region, which is beneficial for repairing deep tissue defects. CuS nanoparticle-PEG soft hydrogel-coated 3D hard PCL scaffolds demonstrate excellent photothermal properties. Under 1064 nm laser irradiation, the mild

thermal effect of 42 ± 0.5 °C effectively promoted the osteogenic differentiation of BMSCs and improved bone regeneration of rat tibial defect [32]. Cu²⁺ and PO₄³⁻ have been reported to have excellent tissue regeneration and osteogenesis abilities, and CuP-based bone-targeting nanosystems also exhibit NIR-II photo-responsiveness. Under 1064 nm laser irradiation, the resulting thermal effect effectively inhibits bone resorption caused by massive osteoclast differentiation, providing a promising treatment for osteolytic bone metastasis [64]. The CD/WS₂ heterojunctions, formed by the electrostatic assembly of CDs with WS₂, present enhanced NIR-II absorption and photothermal conversion efficiency and exhibit good photothermal effects under $1064 \text{ nm} (0.6 \text{ W cm}^{-2})$ laser irradiation after penetrating 10 mm-thick tissue. Periodic low-intensity laser irradiation and warm environment stimulate the expression of bone-related genes such as HSPs and accelerate osteoblast differentiation [63].

2.2.3. Carbon materials

Carbon materials (like MXene, carbon dots (CDs), graphene oxide (GO), carbon nanotubes (CNTs), etc.) are promising in the field of bone repair owing to their excellent biocompatibility and photothermal properties. The MXene-based material can attenuate local immune responses while promoting new bone formation by mild thermal stimulation [33,65,66]. CDs doped in drugloaded bone repairing materials can synergistically stimulate angiogenesis, promote osteoblast differentiation, and accelerate bone repair through NIR laser-triggered drug release and mild thermal effect [67]. CD-based composites can also generate photothermal effects in the NIR-II biowindow for osteosarcoma therapy and bone tissue regeneration [63]. Yang *et al.* developed a temperature-controlled multifunctional nano-hydroxyapatite (HA)/GO/chitosan (CS) scaffold for tissue repair after osteosarcoma resection. Under 808 nm laser irradiation, the hyperthermia generated by the scaffold (48 °C) could effectively kill osteosarcoma cells while promoting

osteogenesis of human BMSCs through a mild thermal effect (42 ± 0.5 °C) [34]. Another study reported that the mild thermal effect triggered by CNT and NIR laser irradiation (42 °C, 15 min per day) significantly up-regulated the expression of osteoinductive genes (ALP, osterix, and osteocalcin), thereby promoting mineral deposition and bone repair [35]. Thermosensitive hydrogels combined with carbon particles were injected into irregular bone defects and underwent sol-gel transition by body temperature, which could be used for conformal therapy. Results of the 8-week treatment of skull bone defect in rats with these hydrogels showed that the bone volume/total volume ratio reached 76.2%, significantly higher than 23.9% in the control group, demonstrating the effectiveness of this strategy [68].

2.2.4. Other photothermal materials

PDA, BP, and indocyanine green are widely used photothermal agents in bone repair, among which PDA could also reduce inflammation caused by reactive oxygen species (ROS) [69]. BP is a two-dimensional elemental material possessing a black metallic luster, a unique folded layered structure, and extraordinary optical and electronic properties desired by biomedical application [70,71]. BP exhibits good biocompatibility, high photothermal conversion performance, and suitable degradation ability. Under NIR laser irradiation, BP can induce a mild photothermal effect and promote bone regeneration by activating HSP-mediated signaling pathways [72]. Furthermore, BP is prone to decompose into phosphate ions in the presence of oxygen and/or water to extract Ca²⁺ from the physiological environment, thus promoting mineralization [38,73]. It has been reported that poly(lactic-*co*-glycolic acid) (PLGA) containing only 0.2 wt% BP (BPs@PLGA) shows an efficient NIR photothermal response, even when covered with 7 mm thick biological tissue [39]. The mild thermal effect and the release of elements required for bone formation endow the BP-based nanocomposite scaffolds with good osteogenic activity. Moreover, introducing other

functional units, such as antibacterial agents and exosomes, can further enrich the functions of this system.

As shown in Figure 3A, to selectively recruit MSCs to the injured site, the surface of electrospun PCL nanofibers doped with BP nanosheets is modified with the nucleic acid aptamer Apt19S, which can specifically recognize MSCs. Subsequently, the microparticles of phase change materials (PCMs) loaded with antibacterial drugs are deposited on the surface of these scaffolds. Under laser irradiation, the photothermal effect of BP activates the solid-liquid phase transition of these scaffolds (T > 39 °C), triggering drug release, thereby killing bacteria and preventing infection. The mild thermal effect-mediated upregulation of HSPs and biodegradation of BP synergistically promote osteogenic differentiation and biomineralization of MSCs. Micro-CT 3D reconstruction of bone tissue showed that new tissue formed at the edge and center of the defect in the Apt-PCL/BP group combined with NIR irradiation after 8 weeks of scaffold implantation, verifying the efficacy for bone regeneration *in vivo* (Figure 3B). This strategy provides valuable information for the rational design of biomimetic scaffolds [40].

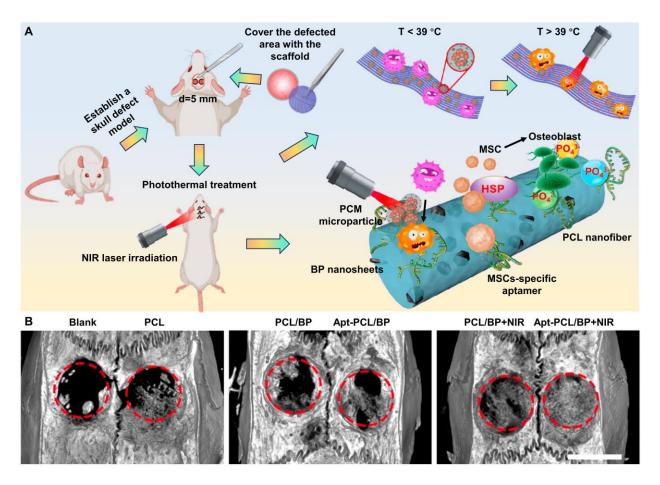


Figure 3. (A) Schematic illustration of a multifunctional scaffold for bone regeneration that kills bacteria, recruits endogenous MSCs, and maintains a mild photothermal effect. (B) Micro-CT 3D reconstruction of the repaired bone tissues at 8 weeks after implanted with different types of scaffolds in rat calvaria defect model. Scale bar = 5 mm. Reproduced with permission from [40]. Copyright 2023, American Chemical Society.

2.3. Magnetothermal materials

The magnetothermal effect refers to the local thermal effect caused by the resonance between the spin magnetic moment of the magnetic material gathered in the lesion and the external AMF. MTT has advantages such as high flexibility, non-/minimal invasiveness, and easy remote operation [74,75]. The depth and intensity of treatment can be controlled by adjusting the strength and frequency of AMF. Therefore, MTT is more beneficial for treating bone defect-related

diseases because the tissue penetration depth of the magnetic field is deeper than that of NIR light [76].

The properties of exogenous AMF and MNPs to promote MSC proliferation and osteogenic differentiation through targeted thermal effect have attracted much attention [77–79]. For instance, a US Food and Drug Administration (FDA)-approved polymethyl methacrylate (PMMA) material contains Fe₃O₄ nanoparticles to produce liquid phase PMMA-Fe₃O₄ bone cement, which is then injected into bone tumors to fill bone defects. The cured cement provides mechanical support for bone defects and alleviates pain, and the warm effect generated by the external AMF can repair bone defects [80]. Low melting point metals exhibit a strong magnetothermal effect upon AMF stimulation due to their high electrical and thermal conductivity. Bismuth alloy has the advantages of low melting point, good biocompatibility, strong bone affinity, and stability in the bone defect site for up to 210 days. Combined with the magnetothermal effect generated by AMF stimulation, the alloy can be used for long-term bone defect repair and analgesia [81]. Furthermore, the MNPs heat the scaffold and significantly accelerate its degradation with AMF, suggesting the benefits of the development of magnetically controlled degradation implants [82].

Superparamagnetic nano Mn-Zn-Cu-Gd ferrites with low Curie temperature (65 °C) were incorporated into bone cement matrix to avoid heat damage to normal tissues, achieve self-controlled hyperthermia, and promote the mineralization of osteoblasts [83]. The optimized osteoinductive nanoparticles-hydrogel composite prepared by embedding the Arg-Gly-Asp (RGD)-coated magnetic iron oxide nanoparticles (CoFe₂O₄@MnFe₂O₄) in agarose had a significant magnetothermal effect. The combination of the composite with the highly tissue-penetrating AMF produced a temperature of 41–42 °C, significantly promoting the osteogenic differentiation and biomineralization of preosteoblasts via the PI3K/Akt pathway activated by

HSP90 [41]. However, besides the adverse effects of magnetic fields on the body, the potential thermal damage to normal tissues and the long-term toxicity of materials are also important challenges in the clinical translation of MTT.

3. Biomaterials-mediated electrical signals

Bioelectricity is a key physiological activity of living organisms. Endogenous electric fields exist in several organs, such as skin, heart, nerve, and bone (Figure 4A), essential for maintaining normal physiological activity [84]. When bone tissue is damaged, the potential of the defect site decreases. Restoration of the local bioelectrical microenvironment by transmitting electrical signals through the damaged tissue takes time, and it is difficult for osteoblasts to migrate from the edge of the defect area to the center [85]. Under such circumstances, electroactive biomaterials serve as scaffolds for cell adhesion and structural support and also help to reconstruct the electrophysiological microenvironment of bone tissue, enhance local electrical stimulation, and regulate cell/tissue behavior and function. The ideal electroactive material must mimic the layered structure of natural bone, generate and/or transmit electrical signals, and regulate stem cell fate while being fully compatible and degradable in the physiological environment. Current research on electroactive materials mainly focuses on conductive materials, piezoelectric materials, and other electrically responsive biomaterials (such as magnetoelectric and photoelectric materials). Among them, conductive and piezoelectric materials are widely used in bone repair. Some representative data for conductive and piezoelectric materials are summarized clearly in Table 2 to facilitate comparison.

Table 2. Summary of typical conductive and piezoelectric materials for bone repair.

Classification	Materials ^{Ref}	Exogenous stimuli	Osteogenic impact	Animal models	In vivo
		(in vitro)	(in vitro)		results

Conductive	PCL/MWCNTs	10 μA, 5 min, three	Promote angiogenesis and	Cranial defect in rat	Partial
carbon	nanofibers ^[86]	times a week	mineralization (UMR-106)	(Ø 5 mm× 2.5 mm)	bridging
materials					
	PCL/graphene	N/A	Promote proliferation (MC3T3	Cranial defect in rat	Bridging
	scaffolds ^[87]		and THP-1)	(Ø 5 mm)	
			,	,	
	BP-CNTpega-gel ^[88]	100 mV mm ⁻¹ , 20	Promote osteogenic	Femoral and spinal	N/A
		Hz, 2 h per day	differentiation (MC3T3)	defects in rabbit	
Conductive	GelMA-PdMGSMW	4 V, 1 Hz, 10 ms for	Regulate adhesion and	N/A	N/A
metals	gels (Pd) ^[89]	2 continuous days	differentiation (C2C12)		
Conductive	FPAT membranes	N/A	Promote osteogenic	Cranial defect in rat	Partial
polymers	(aniline trimer) ^[90]		differentiation (MSCs)	(Ø 5 mm)	bridging
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Piezoelectric	$K_{0.5}Na_{0.5}NbO_3^{[91]}$	5 kV cm ⁻¹ , 20 min	Promote osteogenic	Femoral defect in rabbit	Partial
cements			differentiation (BMSCs)	$(5 \text{ mm} \times 3 \text{ mm})$	bridging
	BaTiO ₃ /Ti6Al4V	11.5 kV, 30 min	Promote osteogenic	Vertebral defect in sheep	Partial
	scaffolds ^[92]		differentiation (MSCs)	(12 mm × 6 mm)	bridging
	PLLA/Ca/Mn co-	6 kV cm ⁻¹ , 30 min	Promote osteogenic	Cranial defect in rat	Partial
	doped BaTiO ₃ ^[93]		differentiation (BMSCs)	(Ø 5 mm)	bridging
	BaTiO ₃ /P(VDF-TrFE)	13 kV, 30 min	Promote osteogenic	Cranial defect in rat	Partial
	membranes ^[94]		differentiation (BMSCs)	(Ø 5 mm)	bridging
	BaTiO ₃ /PLA ^[95]	6 kV, 10 min	Promote adhesion and	Cranial defect in rat	Partial
			proliferative (MC3T3-E1)	(Ø 5 mm)	bridging
Piezoelectric	MXene/PVDF	13 kV	Duamata astes asnis	Cranial defect in rat	Partial
	membranes ^[96]	15 KV	Promote osteogenic		
polymers	membranes		differentiation (MC3T3-E1)	(Ø 5 mm)	bridging
	PVFT-BGM	13 kV, 60 °C, 60	Promote osteogenic	Cranial defect in rat	Partial
	scaffolds ^[97]	min	differentiation (BMSCs)	(Ø 5 mm)	bridging
	P(VDF-TrFE)	13 kV, 60 °C, 60	Promote osteogenic	Cranial defect in rat	Partial
	membranes ^[98]	min	differentiation (BMSCs)	(Ø 5 mm)	bridging

PLLA nanofiber 0.08 MPa, 20 min Promote chondrogenic Femoral articular Bridging scaffold^[99] per day differentiation (ADSCs) cartilage defect in rabbit $(\emptyset \text{ 4 mm} \times 2 \text{ mm})$

3.1. Impact of electrical signals

Bone is considered a natural piezoelectric composite material, which generates piezoelectric phenomena and physiological potentials due to the electronic displacement of local electric fields caused by the mechanical deformation of collagen (COL) structures [100,101]. The piezoelectric effect in natural bone was first observed by Japanese scientists Fukada and Yasuda in 1957 [102] and later confirmed at the molecular level by infrared spectroscopy by Lipieca et al. in 2012 [103]. It has been reported that the piezoelectric constant (d₃₃) of bone tissues is about 0.7–2.3 pC N⁻¹ and the physiological potential ranges from –60 to –100 mV [104]. According to Wolf's Law, bone can constantly adjust its shape, strength, and density to resist external forces, and a persistent lack of force will cause bone resorption, resulting in decreased bone density [105]. With the change of mechanical stress, the potential gradient along COL fibers provides local stimulation to bone regeneration cells such as osteoblasts and osteocytes, which help to maintain the normal metabolism and physiological activities of bone tissues.

As an inducing factor, electrical stimulation plays an important role in bone maturation, remodeling, and reconstruction. The piezoelectric potential of bone generates negative charges under physiological loading, further stimulating the proliferation and differentiation of osteoblasts and thereby promoting bone regeneration [106]. The higher pressure load generates a higher piezoelectric potential in a specific range. Figure 4B displays the underlying mechanisms of electrical stimulation of osteogenesis, which may include activation of cell membrane ion channels, focal adhesion-associated mechanotransduction signaling axis, increase of local blood flow, and

regulation of cell signal transduction pathways [16,84]. Specifically, ion channels are hydrophilic microchannels that allow ions to pass through the cell membrane selectively. Upon stimulation of electrical signals generated by mechanical stress, Ca^{2+} channels on the surface of the cell membrane open, and Ca^{2+} influx leads to increased intracellular Ca^{2+} concentration, which, in turn, activates calcineurin and calmodulin. Calcineurin reacts with the phosphorylated nuclear factor of activated cells to generate the dephosphorylated nuclear factor NF-AT, which then translocates into the nucleus. NF-AT cooperates with other transcription factors to regulate gene transcription, promotes the synthesis of transforming growth factor- β (TGF- β) and bone morphogenetic protein (BMP), and regulates the production of extracellular matrix and cell metabolism [107]. In addition, Ca^{2+} can activate the blocking activity of gelsolin, release actin, and promote cell migration [108].

On the other hand, electroactive biomaterials enhance integrin receptor binding and aggregation by altering the conformation of adsorbed fibronectin through surface charges, accelerating the formation of focal adhesion complexes. Focal adhesion kinase (FAK) is activated by focal adhesion aggregation, which triggers the mechanotransduction signaling axis and promotes osteogenic differentiation through YAP/TAZ transcriptional coactivation of key genes involved in osteogenesis [109,110]. Electrical stimulation can promote vasodilation, increase the permeability of the vessel wall, and deliver white blood cells and oxygen to the wound to accelerate bone healing. Also, electrical stimulation promotes bone regeneration by activating signaling pathway-mediated osteogenic effects, such as enhanced expression and release of growth factors.

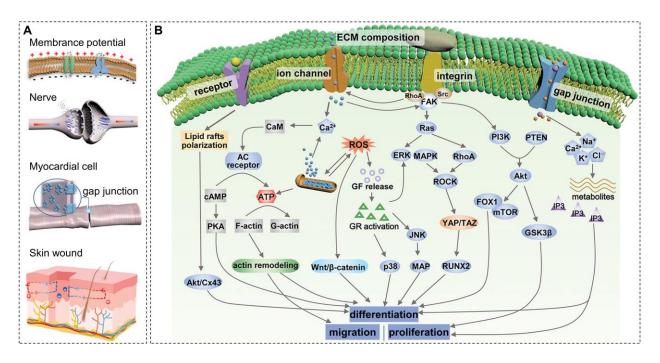


Figure 4. Schematic of (A) endogenous bioelectricity and (B) possible pathways involved in the biological response to electrostimulation. Reproduced with permission from [84]. Copyright 2021, Wiley-VCH.

3.2. Conductive materials

Carbon-based nanomaterials, metallic materials, and conductive polymers are widely used conductive materials for bone repair. Conductive materials amplify and deliver endogenous electrical signals to cells/tissues upon external electrical stimulation. This electrical signal can guide the osteogenic differentiation of bone-related cells and also change the surface charge density of the conductive matrix to promote mineral nucleation and growth, thus accelerating cell proliferation and differentiation.

3.2.1. Carbon-based nanomaterials

Carbon materials such as CNTs and graphene are ideal conductive biomaterials studied in bone regeneration, and CDs, nanodiamonds, and fullerenes have shown great potential [111]. Although these nanomaterials are mainly composed of the same carbon element, the differences

in structure, surface functionality, and size endow them with different characteristics. Besides the good mechanical properties, biological activity, biocompatibility, and chemical stability, carbon-based nanomaterials also demonstrate a large specific surface area and high electrical conductivity, which can be used to improve the mechanical properties and electrical conductivity of bone repairing material and stimulate bone regeneration. Incorporating CNTs into the polymer has been reported to effectively improve the tensile and compressive modulus of the composite scaffolds [112].

The similar surface roughness of CNTs to native COL enables them to positively affect cell adhesion, proliferation, and differentiation [113,114]. CNTs have a small diameter, a high specific surface area, and an ordered arrangement of internal carbon atoms through strong sp2 bonds. Electrons move freely in the tube to form one-dimensional conductive channels with a conductivity as high as 10^4 S cm⁻², which is ten thousand times that of copper [115]. Under direct current stimulation of $100~\mu\text{A}$, biodegradable poly-DL-lactide (PLA) nanofibers embedded with multiwalled CNTs fabricated by electrospinning can guide cellular elongation and proliferation, showing great potential in bone regeneration and fracture healing [116]. Exogenous electrical stimulation combined with conductive scaffolds can promote osteoclast formation and the expression of nuclear factor $\kappa\beta$ ligand RANKL, thus significantly promoting angiogenesis and mineralization and dominating the process of bone remodeling [86]. Furthermore, CNT-containing scaffolds exhibit sensitive electrochemical response signals to osteogenic differentiation and tissue mineralization at the cellular and animal levels, which can be used for long-term noninvasive and continuous monitoring of osteogenic differentiation levels [117].

Graphene is one of the strongest materials known. The formation of conjugated large π bonds by carbon atoms in the structure endow it with high electrical conductivity. Homogeneous doping

of PDA-reduced GO in the network enhances the mechanical properties of the network while providing a conductive pathway that allows it to act as an electroactive matrix to transmit electrical signals and regulate the proliferation and morphology of C2C12 cells [118]. BP is a graphene-like material with high electrical conductivity and topology due to its anisotropy, which is also an ideal conductive material for bone regeneration [119,120]. In particular, the large specific surface area of carbon-based biomaterials can be loaded with a variety of bioactive substances (growth factors, drugs, etc.) to synergize with the electrical activity to promote osteogenesis. Nonetheless, carbon materials are difficult to disperse uniformly, and agglomeration may reduce mechanical properties. The long-term safety of nondegradable carbon nanomaterials *in vivo* also requires further investigation.

3.2.2. Metallic materials

Gold and silver are commonly used conductive biomaterials in bone regeneration. Gold nanoparticle-doped hybrid hydrogel scaffolds could increase connexin 43 expression in neonatal rat cardiomyocytes, indicating that electroelastic scaffolds can potentially improve cardiomyocyte function [121]. Furthermore, silver nanoparticles could work as a conductive phase to enhance the electrical activity of other materials to promote bone repair [122]. The moderate dissociation in the physiological microenvironment is advantageous for metallic bone repairing materials because some metal ions are structural components of osteogenic enzymes and proteins. Introducing Fe³⁺ into hydrogels could significantly improve its conductivity as strain sensors in tissue repair [123]. However, the uncontrolled release of metal ions caused by wear or corrosion *in vivo* is prone to trigger inflammatory cascades. In constrast, metallic glass has higher wear and corrosion resistance. Upon electrical stimulation, the electrical signal amplified through gelatin methacryloyl gel,

containing conductive palladium-based metallic glass submicron wires, significantly promotes the formation, contractility, and metabolic activity of mouse myoblast C2C12 myotubes [89].

3.2.3. Conductive polymers

Conductive polymers, mainly polypyrrole (PPy), polyaniline (PANI), polythiophene, and poly(acetylene), are a new class of intelligent materials with long-range π electronic backbone structures formed by alternating arrangement of localized carbon-carbon single bonds and less localized double bonds. Under the stimulation of an external electric field, the internal charge carriers move along the conjugated π bonds, thus achieving the directional transfer of electrons, manifested as electrical conductivity. In the biomedical field, conductive polymers have attracted much attention due to their strong structural flexibility and redox reversibility compared with conductive carbon nanomaterials and metallic materials. Furthermore, according to the type and amount of dopant used, the conductivity of PPy and PANI can usually reach $10-10^3$ S cm⁻¹ and 30-200 S cm⁻¹, which is close to that in human tissues [124]. There are various methods for preparing and modifying conductive PANI, and selecting appropriate stabilizers helps improve integrative performance. The PANI-based films prepared with sodium dodecyl sulfate as a stabilizer were reported to have high electrical conductivity and good cytocompatibility, as well as non-irritant to the skin [125].

As shown in Figure 5A, fibrous aniline trimer (AT)-based polyurethane (FPAT) membranes prepared by electrospinning were used for calvarial defect repair. The FPAT electrospinning membranes with good electrical activity could increase Ca²⁺ concentration in MSCs, up-regulate the Ca²⁺ signaling pathway, promote osteogenic differentiation, and accelerate bone regeneration. In addition, the redox property of FPAT membranes allow them to remove ROS from the organism, which, in turn, promotes bone repair by inducing macrophage polarization toward the M2

phenotype (Figure 5B). Fluorescence staining indicates that electroactive membranes can stimulate the accumulation of intracellular Ca²⁺ and remove excessive intracellular ROS. The bone regeneration areas after FPAT membrane treatment are significantly larger than in other groups, indicating the superior bone regeneration ability with FPAT membranes *in vivo* [90]. In order to grasp the progress of bone repair, the combination of conductive materials with flexible electronic devices may be a potential approach [126]. However, there are few *in vivo* applications of conductive polymers, which may be due to their poor degradation, easy loss of conductivity, and physiological toxicity caused by small molecule residues. Dependence on external power supply, complex operation, and low efficiency are also major obstacles to practical applications of conductive materials.

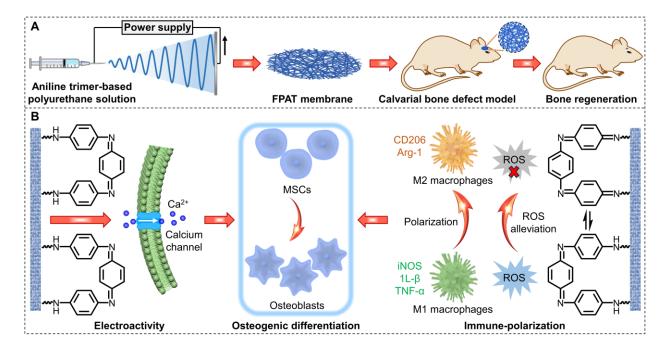


Figure 5. (A) Schematic of FPAT membranes prepared by electrospinning for calvarial defect repair. (B) The electroactive FPAT membranes restore the electrophysiological microenvironment, regulate site-specific cellular behaviors and promote osteogenic differention.

3.3. Piezoelectric materials

The piezoelectric effect refers to the physical phenomenon of generating potential difference between the two ends of a dielectric material under a mechanical load or deformation. The surface charge arises from the rearrangement of dipoles triggered by an external force, and the material can maintain a charged state for a period after polarization without continuous stimulation. Therefore, compared with conductive materials, piezoelectric materials are more suitable for clinical use. Due to the outstanding performance of bioelectric signals in repairing damaged tissues and restoring cellular functions, piezoelectric materials have attracted much attention in orthopedic applications [127,128]. According to the material type, the commonly used piezoelectric materials in bone repair mainly include ceramics and polymers, among which piezoelectric polymers include natural and synthetic types.

3.3.1. Piezoelectric ceramics

Barium titanate (BaTiO₃) [129], lithium niobate [130], and potassium sodium niobate [91] are well-known biological piezoelectric ceramics. The BaTiO₃ with tetragonal perovskite structure presents a high piezoelectric constant (d₃₃ > 190 pC N⁻¹), which generates polarization under an external force or a strong direct current electric field, separating the internal electrons from the holes and generating a voltage [131]. Although BaTiO₃ is not degradable *in vivo*, it still shows good biocompatibility and osteogenic ability and is an excellent piezoelectric material for hard tissue repair. BaTiO₃ is usually combined with nondegradable polymers (such as polyvinylidene fluoride (PVDF), poly(vinylidene fluoridetrifluoroethylene) (P(VDF-TrFE)), and polyamide 12) [94,132,133], degradable polymers (PLA, for instance) [95] and other inorganic materials (HA and Ti6Al4V, etc.) [104,134] because of its poor processing property, mechanical strength, and toughness.

The piezoelectric effect generated by polarized BaTiO₃/Ti6Al4V scaffold has been demonstrated to promote macrophage M2 polarization and immunomodulatory osteogenesis in MC-3T3 osteoblasts [134]. The piezoelectric BTCP ceramics prepared by two-step sintering of BaTiO₃ and β-tricalcium phosphate (β-TCP) form different types of charges through high-voltage polarization, thereby synergistically regulating cellular immunity and osteogenic function to guide bone healing. The negatively charged, polarized BTCP (BTCP–) promotes protein adsorption and extracellular Ca²⁺ influx of BMSCs, increases integrin α5β1, P-FAK, and P-ERK protein expression, and enhances osteogenic differentiation through FAK/ERK and BMP/Smad signaling pathways. The polarized BTCP ceramics with a positive surface charge (BTCP+) significantly inhibit the polarization of pro-inflammatory (M1) macrophages, alleviate the local inflammatory response, and form an immune microenvironment conducive to osteogenesis [135].

It has been reported that polarized BaTiO₃ enhanced calcium phosphate deposition [136], and the introduction of the piezoelectric phase promoted the adhesion and proliferation of mouse fibroblast L929 and human osteoblast SaOS2 cells [137]. BaTiO₃ possesses a superior intrinsic ability to maintain charged surfaces [95,138]. The original piezoelectric coefficient (d₃₃) of the nanocomposite membranes, containing BaTiO₃ nanoparticles embedded within a P(VDF-TrFE) matrix and immersed in the culture medium for 4 weeks, was maintained more than 90% [129]. After corona polarization, the surface potential of the flexible nanocomposite membrane prepared by BaTiO₃ nanoparticles, PDA, and PVDF reached -76.8 mV, consistent with the endogenous biological potential level. Moreover, it could maintain over half of the original surface potential at the bone defect site for 12 weeks [94].

When BaTiO₃ was combined with conductive silver and shape memory polymer of acrylate epoxidized soybean oil, the scaffolds demonstrated high accuracy, customizability, shape memory,

and piezoelectric properties, which could effectively promote bone regeneration [139]. HA, one of the main components of bone tissue, has also been shown to have piezoelectric activity due to its non-centrosymmetric monoclinic crystal structure [140,141]. However, the polarization conditions of HA-based ceramics are relatively harsh, usually requiring high direct current intensity (approximately 1–2 kV cm⁻¹) and high temperature (about 300–500 °C), which is an important reason to limit the development of piezoelectric HA to practical applications [142,143].

3.3.2. Piezoelectric polymers

Biopiezoelectric polymer-based composites have promising applications in regenerative medicine and tissue repair. Currently, the commonly used piezoelectric polymers for bone repair include synthetic (PVDF, PLLA, poly[(R)3-hydroxybutyrate] (PHB), etc.) and natural polymers (like COL, cellulose, and chitin). PVDF is a semi-crystalline polymer composed of five known crystalline phases, α , β , γ , δ , and ε , and is also one of the most widely studied piezoelectric polymers [144]. Among the crystalline structures, the polar β phase of PVDF shows the strongest piezoelectric activity due to the high density of dipoles in the same direction. β -phase PVDF has excellent biocompatibility, piezoelectric properties, and a large specific surface area, which can fit the COL fiber structure of the natural extracellular matrix and has been demonstrated to stimulate osteogenic differentiation. The better piezoelectric performance of PVDF with a higher content of β -phase has been shown [145].

Currently, there are two commonly used strategies to improve the β -phase content of PVDF. On one hand, introducing nanoparticles into the system could induce β phase nucleation [146]. The uniform distribution of the PDA-modified BaTiO₃ nanoparticles in the PVDF matrix increased the β -phase content from 46% to 59%, raising output voltage by 356% [132]. The β -phase content could also be enhanced by adding TiO₂ powder to the PVDF electrospinning solution with a

stretching effect on the PVDF chain [145]. On the other hand, the polarization and stretching effect of the high voltage electric field during electrospinning could promote the transformation of the non-polar α phase to the polar β phase of PVDF, and the content and crystallinity of the β phase could be effectively improved by adjusting the processing parameters [96].

PVDF is usually combined with other bioactive substances, such as hydrogel, nanocrystals, and GO to enhance piezoelectric effect and achieve excellent electrical signal stability and biological regulation. Piezoelectric PVDF composites could significantly promote cell adhesion, proliferation, and differentiation through surface electrical signals and enhance angiogenesis and osteogenic activity [146-149]. By modifying ZIF-8 nanocrystals in hierarchically architected PVDF foam, the resulting PVDF/ZIF-8 piezoelectric foams can realize a stable voltage output of up to 10 V without polarization. Piezoelectric PVDF affects the molecular trajectory of Zn²⁺ released from ZIF-8, up-regulates oxidative phosphorylation and ATP-coupled cation transmembrane transportation in vascular endothelial cells, thus enhancing the uptake of Zn²⁺ by cells, and guiding vascularized bone regeneration through micro-current stimulation and Zn²⁺ enrichment. PVDF and PVDF/ZIF-8 foam particles were mixed with commercially available bone grafts in equal proportions and filled in the box-shaped defect site of the mouse femur. The presence of PVDF foam-based particles significantly improved the electrical properties of the femur. After 4 weeks, the bone healing outcome of PVDF/ZIF-8-M combined with the piezoelectric condition was obvious compared to that of the non-piezoelectric PVDF/ZIF-8-M group, which was slightly worse. According to micro-CT results, PVDF/ZIF-8-M foam significantly promoted bone repair and avoided fractures under piezoelectric conditions [148]. Besides, piezoelectric PVDF-based copolymer (P(VDF-TrFE)) could recruit stem cells and

promote osteogenic differentiation by mimicking periosteal structure/function and physiological loading conditions [97,98].

However, it should be noted that BaTiO₃ and PVDF-based polymers are difficult to degrade *in vivo* and usually require secondary surgical removal after tissue repair is completed. Therefore, it is necessary to use biodegradable piezoelectric biomaterials. PLLA, PHB, and poly[3-hydroxybutyrate-co-3-hydroxyvalerate] (PHBV) polymers meet all requirements. PLLA is a semicrystalline polymer containing a C=O dipole, and its thermodynamically stable conformation is an α-crystalline phase with helical polymer chains, while the piezoelectric properties stem from the β-type structure formed by C=O rearrangement after polarization [150]. As a widely used biodegradable polymer, PLLA scaffold can generate a weak but stable electric field to promote cell migration/recruitment under external force stimulation. The experimental results of rabbit knee joint injury treated with PLLA nanofiber scaffold showed that piezoelectric effect could effectively stimulate cartilage regeneration [99]. However, the piezoelectric properties of PLLA are weak and their relation with bone growth promotion remains unclear and requires further investigation.

PHB and PHBV are aliphatic polyesters widely distributed in the cytoplasm of prokaryotic cells, especially bacteria, which can completely degrade into non-toxic products *in vivo* [151]. The existence of asymmetric carbon atoms associated with the polar C=O group in the structure endows PHB and PHBV with piezoelectric properties comparable to those of biological bone. Therefore, PHB- and PHBV-based piezoelectric scaffolds can fill bone tissue defects and stimulate new bone formation under dynamic mechanical conditions [152]. Nevertheless, the insufficient mechanical strength and uncontrollable degradation rate of PHB and PHBV limit their use.

Natural polymers such as polysaccharides (CS, sodium alginate, and hyaluronic acid, for instance) and proteins (COL, gelatin, fibrin, etc.) are also widely used in bone repair due to their good material accessibility and biocompatibility. Especially, natural polymeric materials with hierarchical structures such as COL, cellulose, and chitin possess piezoelectric properties with a piezoelectric constant d_{14} of approximately 0.2-2 pC N^{-1} , 0.1-0.2 pC N^{-1} , and 0.2-1.5 pC N^{-1} , respectively [153]. Natural polymers have superior biological affinity due to their properties similar to the extracellular matrix. The piezoelectric properties can be amplified by molecular engineering for applications in various fields such as bone regeneration. In 2009, Jolandan and Yu isolated a single COL-I fiber from a bovine Achilles tendon and found that these fibers mainly behave as shear piezoelectric materials with a piezoelectric constant d_{15} of about 1 pC N^{-1} [101]. The highly oriented and patterned structure of COL-I in bone and its ability to respond to mechanical loading confer piezoelectricity on bone tissue. Under the action of shear force, the COL fibers slide against each other, generating a piezoelectric charge, which in turn stimulates osteoblasts, induces mineralization and promotes bone formation [154]. The deposition of HA is an important step in bone repair. It has been reported that the piezoelectric dipole generated by deformed COL can promote HA deposition through electrochemical methods without catalyst [100].

3.3. Other electroactive materials

In recent years, the application of other electroactive materials such as magnetoelectric and optoelectronic materials in bone regeneration has attracted much attention. Magnetoelectric materials are composed of piezoelectric materials and magnetostrictive components. Electrical properties of these materials can be adjusted by external magnetic fields. Magnetostrictive materials move and rotate between magnetic domains in response to an external magnetic field,

thus stretching and deforming the material in length and volume, which in turn stimulates piezoelectric materials to generate electrical signals [155]. Liu *et al.* fabricated polarized flexible CoFe₂O₄@BaTiO₃/P(VDF-TrFE) core-shell particle-incorporated composite membranes with high magnetoelectric conversion efficiency. Under the stimulation of magnetic field, the magnetic driving force on the CoFe₂O₄ magnetic core transferred to the BaTiO₃ shell and increased its charge density, and enhanced the β phase transition of P(VDF-TrFE) matrix, leading to increased surface potential and osteogenic activation. Cranial defect experiments in male rats showed that repeated magnetic field applied to the composite membranes enhanced bone defect repair even under coexisting inflammatory conditions [156].

Photoelectric materials can absorb light energy and undergo photoelectron conversion reactions to produce electric energy. Three-dimensional biomimetic scaffold integrated with thin-film silicon-microstructures are able to regulate the membrane potential and intracellular calcium dynamics of stem cells through infrared light-induced electrical signals, and effectively promote cell proliferation, differentiation, and osteogenesis [157]. Nanogenerator has the characteristics of converting mechanical energy into electrical energy, but it is mainly used in energy harvesting, energy storage, health monitoring and other fields, and its application in bone repair needs to be further developed [158,159].

4. Biomaterials-mediated magnetic signals

As a non-/less invasive, safe, and convenient physical stimulation, the magnetic signal has significantly enhanced wound healing, bone defect repair, and osteoarthritis relief. Magnetic signals promote osseointegration and bone regeneration by activating cell surface receptors, stimulating the expression of bone-related genes, promoting cell adhesion, proliferation, migration, and differentiation, and inhibiting osteoclast formation [160]. However, magnetic stimulation of

cells by remote modulation of external magnetic field alone is insufficient. The combination of magnetic field and magnetic implants plays a special multifunctional synergistic role in clinical bone repair. The following section reviews the recent progress of magnetic signals mediated by biomaterials in bone tissue engineering. First, we review the impacts of magnetic signal on osteogenesis, and then we introduce the cutting-edge research on magnetic strategies based on MNPs and scaffolds to improve cell and tissue repair functions (Table 3).

Table 3. Summary of typical magnetic biomaterials for bone repair.

Materials ^{Ref}	Туре	Intensity	Osteogenic impact	Animal models	In vivo results
		(in vitro)	(in vitro)		
CS/PDA@MS (Fe ₃ O ₄) ^[161]	SMF	N/A	Promote adhesion and proliferation	Humerus defect in rat (Ø	Bridging
			(BMSCs)	$2 \text{ mm} \times 2 \text{ mm}$)	
RSF/TA/Fe ₃ O ₄ hydrogel ^[162]	SMF	125 mT	Promote osteoblast differentiation	Cranial defect in rat	Partial bridging
			(BMSCs)	(Ø 5 mm)	
PLLA/MgO/Fe ₃ O ₄	SMF	70–80 mT	Promote osteogenesis and	Cranial defect in rat	Partial bridging
scaffolds ^[163]			mineralization (BMSCs)	(Ø 8 mm)	
Fe ₃ O ₄ /PDA coating ^[164]	SMF	15 mT	Promote proliferation and osteoblast	Femoral defect in rabbit	Partial bridging
			differentiation (human BMSCs)	$(\emptyset 5 \text{ mm} \times 10 \text{ mm})$	

4.1. Impact of magnetic signals

Magnetic fields transmit the magnetic force between objects around a magnet or an electric current and are mainly divided into static magnetic field (SMF) [165], electromagnetic field [166], AMF [167], and rotating magnetic field [168]. Cell proliferation depends on various factors, such as the type, intensity, frequency, and duration of magnetic fields, so it is essential to determine the magnetic field range in which biological systems respond significantly [160]. Among them, SMF, as a magnetic field with constant strength and fixed directions, is the subject of research in bone

regeneration. SMF can be divided into weak (<1 mT, such as geomagnetic field), moderate (1 mT-1 T, such as common permanent magnets) and high (>1 T, like superconducting magnets) categories according to the magnetic field strength [169]. Magnetic fields of moderate intensity are most widely used, while strong SMF of 5-10 T has also been reported to have the ability to regulate the orientation of matrix proteins and cells [170,171]. As shown in Figure 6A, upon magnetic field stimulation, diamagnetic cell membranes alter membrane flux, activate intracellular ion channels, accelerate cell metabolism, and promote cell adhesion, proliferation and differentiation [172]. Long-term local SMF stimulation can enhance the binding of the implant to the host tissue, prevent the loss of bone density caused by surgical invasion, promote mineralization, and accelerate bone defect healing [173]. High magnetic flux inhibits osteoclast formation and differentiation by reducing tartrate-resistant acid phosphatase activity, leading to osteoclast apoptosis and necrosis [174]. SMF regulates the orientation of diamagnetic osteoblasts and extracellular matrix proteins, which helps to accelerate the growth process of osteoblasts [170,171].

There are several potential physical mechanisms about SMF interacts with bone tissue [175,176]: (1) Magneto-mechanical interaction. Bone is a tissue with minimal diamagnetism. In a homogeneous magnetic field with a sufficiently large magnetic gradient, material with anisotropic susceptibility rotates under magnetic torque drive until it reaches a stable orientation. In this case, phospholipid molecules will rotate and orient in the direction of the magnetic field, which expands the ion channels in the cell membrane and allows ions to pass through, increasing electrical conductivity and generating a strong current, followed by a series of biological effects that promote bone formation. (2) Electrodynamic interaction (also known as Hall effect). The COL of the bone matrix is negatively charged, and cations are adsorbed to the surface by electrostatic interactions.

At the same time, there is a voltage difference between the inside and outside of the cell membrane, which is usually positively charged on the outside and negatively charged on the inside. The charged ions move between the bone matrix and the cell membrane are subjected to the Lorentzian force in the magnetic field, forming a Hall voltage to induce further ion migration and improve the permeability of the cell membrane, promoting the extracellular ions to pass through the cell membrane and enhance cell activity. (3) Radical pair effect. SMF of moderate or weak strength affects the free radical or electron spin state of some biochemical reaction intermediates, altering the rate, yield, or product distribution of the reaction. (4) Activation of the cyclic adenosine phosphate system. Magnetic stimulation can activate the cyclic adenosine monophosphate system, which in turn activates various enzyme systems to induce special physiological functions of osteocytes and accelerate bone growth. Magnetism activates multiple signaling pathways within cells that synergistically promote osteogenesis, mineralization, and angiogenesis. These pathways include:

- A) Activation of the BMP-2/Smad pathway to significantly up-regulate the expression of osteogenesis-related genes and vascular endothelial growth factor [177].
- B) Activation of cyclic guanosine monophosphate (cGMP)/protein kinase G (PKG)/extracellular signal-regulated kinase (ERK) [162], TGFβ-Smads [164], and YAP/β-catenin signaling pathways [178].
- C) Up-regulation of the expression of bone-related genes (Runx2 and Osterix) and ALP activity. Activation of integrin signaling pathways, such as FAK, paxillin, RhoA, mitogen-activated protein kinase, and nuclear factor-kappaB, and up-regulation of BMP-2 and phosphorylation of Smad1/5/8 (Figure 6B) [24,172].

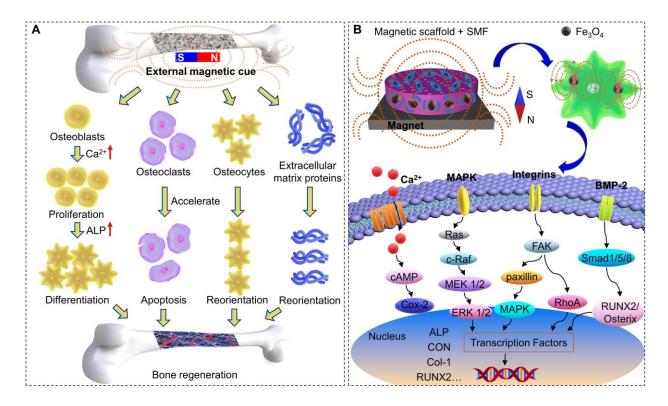


Figure 6. (A) Schematic of SMF promoting bone regeneration. (B) Schematic representation of magnetic scaffolds combined with SMF synergistically promoting osteogenic differentiation.

4.2. Magnetic composite scaffolds

MNPs, such as γ -Fe₂O₃ and Fe₃O₄, have shown great potential in bone tissue engineering due to their favorable chemical stability, biocompatibility, and high magnetic torque. In particular, low-dose MNPs possess a high safety profile *in vivo* as they can be absorbed as iron ions through ionization and participate in iron homeostasis. Importantly, the US FDA approved iron oxide nanoparticles for treating iron deficiency anemia [179]. In some studies, γ -Fe₂O₃ and Fe₃O₄ have been found to generate shear stress at the cellular level, promoting bone angiogenesis regardless of SMF [180,181]. Iron oxide nanoparticles and SMF alone could promote osteogenic differentiation and inhibit osteoclast activity, while the combination had a stronger effect [160]. In conjunction with 0.2 T AMF, α -Fe₂O₃/ γ -Fe₂O₃ nanocomposites significantly reduced the expression of TNF-a, an osteoinductive cytokine, which induces osteoclast formation and leads to

osteoporosis, increases Ca/P ratio and the expression of key bone formation markers OPN, col-1, OCN, DMP-1, and BMP-2, thereby promoting osteogenic differentiation [182]. Despite the obvious advantages, the toxicity and transient residence of MNPs exposed in large quantities to bone defects limit their practical applications. The construction of magnetic composites/scaffolds helps to overcome these shortcomings and promote clinical translation.

Magnetic composite scaffolds based on γ-Fe₂O₃, Fe₃O₄, and CoFe₂O₄ present special magnetic field responsiveness and can be used for tissue engineering, such as orthopedic implantation and intervention [25,183,184], especially for early bone fixation and repair [185]. Superparamagnetic scaffold combined with SMF demonstrates a stronger stimulating effect on cell proliferation and differentiation, and synergistically promotes the formation, integration, and remodeling of new bone [186]. Under an exogenous magnetic field, PLGA microspheres encapsulated with superparamagnetic iron oxide nanoparticles can significantly promote bone repair. When the actual feeding amount of iron oxide nanoparticles was 1.38%, the microspheres were labeled as PFe-II. In the rat femoral defect experiment, after 6 weeks of treatment with PFe-II combined with magnetic field, bone mineral density (263.97 ± 25.99 mg/cm³), trabecular thickness (0.58 \pm 0.08 mm), and bone volume/total volume (78.28 \pm 5.01%) were significantly higher than those of the PFe-II group (bone mineral density, 194.34 ± 26.71 mg/cm³; trabecular thickness, 0.41 ± 0.07 mm; bone volume/total volume, $50.49 \pm 6.41\%$). Moreover, the expressions of ALP, COL-I, OPN, and OCN in the repairing bone were significantly higher in the combination group than the PFe-II group, further clarifying the synergistic effect of the magnetic composites and the SMF on promoting bone regeneration [187].

MgO and magnetic Fe₃O₄ nanoparticles were loaded into the PLLA matrix to fabricate the biomimetic porous PLLA/MgO/Fe₃O₄ (PMF) scaffolds via selective laser sintering (Figure 7A).

With SMF, the magnetic torque effect of PLLA/MgO/Fe₃O₄ scaffolds (via integrin α V β 3/actin) enhanced the activity of Mg²⁺ channel proteins, thus facilitating the capture of Mg²⁺ by BMSCs in the microenvironment and inducing osteogenesis (Figure 7B). Coordinated with SMF, magnetic scaffolds increased the expression of osteogenesis-related genes (ALP, Runx2, OCN, and OPN) and mineralization *in vitro*. After 12 weeks of scaffold implantation, 3D reconstruction images showed that PM (PLLA/MgO), PMF, and PMF+SMF groups all promoted new bone formation to a certain extent. Among them, the PMF+SMF group exhibited the most obvious trend of new bone infiltration from the edge of the scaffold to the internal pores, further confirming its ability to promote bone differentiation and regeneration (Figure 7C) [163].

Exosomes derived from BMSCs pretreated with magnetic fields or low-dose MNPs also showed enhanced osteogenic and angiogenesis effects during bone regeneration [188]. Taking advantage of the responsiveness of MNPs to the SMF, dynamic stiffness of these scaffolds could be realized by changing the arrangement of magnetic particles to achieve an ideal biomimetic dynamic mechanical microenvironment and promote bone regeneration [189]. Hydrogels based on superparamagnetic HA nanorod (MagHA) with continuous biophysical and biochemical gradients generated incremental HA, mechanical and electromagnetic signals under external SMF stimulation to repair osteochondral units with perfect heterogeneity [190]. In addition to the dynamic structure generated by magnetic field responsiveness, MNPs could also optimize the interaction between cells and scaffolds, allowing cells to grow under natural biomimetic conditions. The magnetic scaffolds prepared by layer-by-layer assembly of superparamagnetic γ -Fe₂O₃ nanoparticles on the surface of PLGA/PCL electrospun scaffold could effectively promote the osteogenic differentiation of adipose-derived stem cells. Surface modification with γ -Fe₂O₃ improved interface hydropathy, elasticity, affinity for stem cells, and osteogenic differentiation

properties, providing a novel and convenient strategy for scaffolds with active stimulation and remote control functions [191].

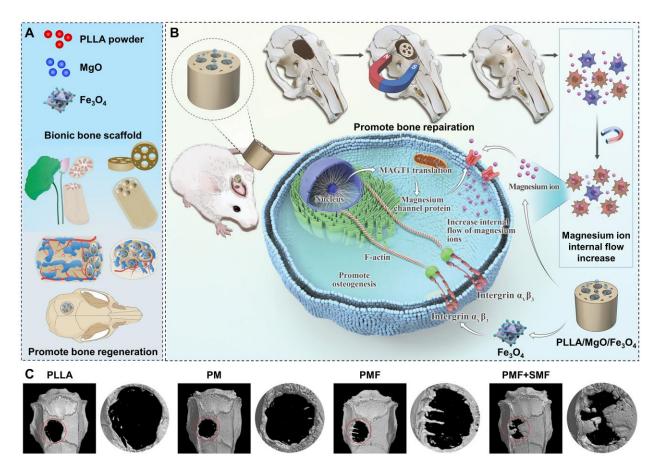


Figure 7. Schematics of (A) a biomimetic bone repair scaffold with a lotus seedpod-structured porous structure and (B) remote and accurate regulation of channel protein activity on the membrane of rBMSCs by SMF to promote Mg²⁺ influx and osteogenesis. (C) Micro-CT reconstruction on the horizontal plane and mid-sagittal plane of bone defect region 12 weeks after implantation, The red circle indicates the defect area. Reproduced with permission from [163]. Copyright 2023, Wiley-VCH.

5. Conclusions and perspectives

The ideal scaffolds to treat bone defect should provide structural support and promote specific biological activity required during bone healing. Despite growth factors are often introduced into

traditional bone repairing scaffolds to stimulate new bone formation, they have many disadvantages such as severe adverse effects, short half-life and high cost. In recent years, a variety of intelligent stimulation-responsive bone-repairing materials have been reported. These materials can respond to exogenous stimuli and promote bone repair. During bone healing, multiple signaling pathways are activated to accelerate bone regeneration through biomaterials-mediated mild thermal effect, electrical and magnetic signals. With advantages of intelligent responsiveness, strong controllability, low invasiveness, and high safety, such technology provides a promising strategy for treatment of bone defect.

Besides applications in anti-tumor strategies, eliminating infection, and promoting wound healing, thermal effect also shows excellent potential in the treatment of orthopedic diseases. The mild thermal effect promotes osteocyte proliferation by enhancing metabolism at the injured site, improving expression of osteogenesis-related proteins (such as ALP and HSPs), regulating inflammatory processes, and stimulating angiogenesis. Bone repairing materials loaded with photosensitizers or paramagnetic nanomaterials generate mild thermal effects when stimulated by NIR light or magnetic field, thereby regulating endogenous signaling pathways and accelerating bone regeneration. Various biomaterials responsive to NIR light and magnetic fields have been developed and incorporated into bone repairing scaffolds, demonstrating their efficacy to promote bone regeneration. NIR laser is considered a safe and effective means of external stimulation due to its unique spatio-temporal selectivity and few side effects. NIR light triggered PTT is commonly used for superficial tissue regeneration. In contrast, the magnetic field presents better tissue penetration, and MTT is more conducive to repairing deep tissue defects. In order to avoide damage to normal tissues, it is worth noting that three-dimensional, real-time, and accurate temperature monitoring is desired to realize therapeutic effects at the most extent.

Bioelectricity is one of the common physiological signals. Implantation of electroactive biomaterials helps to restore the bone surface potential at the site of bone defects wherein endogenous bioelectricity is subnormal and promotes bone healing. Electrical stimulation generally instigates bone regeneration by activating cellular voltage-gated Ca²⁺ channels, promoting the expression of integrins and bioactive factors, increasing local blood flow, and regulating cellular signal transduction pathways. Conductive materials are capable of charge transport at the cell membrane interface and regulate cell contact with cells, tissues and biological interfaces. However, conductive materials usually need to be connected to an external power supply, which is not convenience to clinical operation. Piezoelectric materials are characterized by spontaneous electricity. Under the stimulation of mechanical stress or strain, electric polarization occurs inside the piezoelectric material, and a charge with the opposite sign is generated and accumulated on the surface, thus promoting bone structure remodeling and regeneration. Piezoelectric biomaterials often require direct contact with damaged tissue to obtain local lowlevel electric fields induced by body motion, but it is challenging to achieve efficient and consistent piezoelectric output. In addition, magnetoelectric materials and photoelectric materials exhibit unique capacity for tuning electrical properties with magnetic field or light modulation, and can be activated in situ for bone defect repair. However, the microenvironment of different bone tissues varies greatly, bringing challenges to electroactive materials for clinical translation.

Despite MNPs or magnetic signals alone can significantly promote osteogenic differentiation, their combination can further improve the efficiency of bone repair. During bone healing, magnetic signals promote bone regeneration mainly through magnetic-mechanical and electric-dynamic interactions, free radical pair effect, and activation of cyclic adenosine monophosphate. Currently, the frontier technology of magnetic-assisted bone tissue engineering mainly focuses on bone

repairing scaffolds incorporated with MNPs, whereas single MNPs are rarely used. This is because the distribution of MNPs within the composite can lead to differences in magnetic moment gradients, resulting in positive impacts on cell activity, and the composite helps to prolong the residence time of the MNPs *in vivo*. Driven by an exogenous magnetic field, MNPs are always in dynamic motion. Therefore, it is critical to carry out non-invasive tracking technique and explore the degradation and distribution of MNPs during bone healing to better regulating their performance.

From the above introduction, it can be seen that for biomaterials that can generate thermal effect, electrical and magnetic signals to treat bone defect under external stimulation, conductive materials require the most complicated in vivo operation. When it comes to controllability, photothermal materials and conductive materials are more likely to mediate controllable physical cues in bone defect site, because photothermal materials combined with laser irradiation can bring relatively stable thermal effect, and conductive materials can also generate stable potential. The resulting temperature or potential can be conveniently regulated by adjusting the external stimulus. For piezoelectric materials, the surface potential may decrease due to electrostatic adsorption of proteins with prolonged implantation time. The arrangement of MNPs in bone defects is affected by the magnetic field. Uneven distribution of MNPs leads to inconsistent magnetic moment gradients, which affect cell behavior and bone regeneration. From the perspective of therapeutic outcomes, photothermal materials to treat deep tissue defects is less efficient than that of electroactive and magnetic materials due to the limited tissue penetration depth of NIR light. Although NIR optical fibers can be implanted into deep tissue defects to irradiate photothermal materials with improved efficacy, this invasive procedure usually causes additional damage to patients. The same dilemma also exists in conductive materials due to the requirement of metal

wire to connect external electric fields. In contrast, magnetic and piezoelectric materials have advantages in implementing patient-friendly, non-/less invasive treatments.

It is still challenging to employ biomaterial-mediated *in situ* physical cues to promote osteogenesis. First, although the effectiveness of biomaterial-mediated physical cues (mild thermal effect, electrical and magnetic signals) in bone repair has been extensively demonstrated and the underlying mechanisms have been proposed, bone repair is a complex process involving multiple physiological activities, and current research is still limited. Extensive collaboration and clinical trials are needed to further investigate the exact mechanisms.

Second, biomaterials-mediated physical cues are less effective than growth factors in promoting bone regeneration. Therefore, there is an urgent need to develop intelligent bone repairing materials with better performance and greater sensitivity to external stimuli. In addition, other physical cues can also significantly promote bone repair, such as ultrasound, which can accelerate bone regeneration processes [192,193]. Under pulsed ultrasound irradiation, the acoustically responsive scaffold generates enhanced acoustic trapping force to recruit stem cells, promoting bone tissue regeneration [194]. Ultrasound-trigged micro/nanorobots also demonstrate potential applications in multidisciplinary fields [195]. Scaffolds combining multiple stimulus-responsive bone repairing materials with improved functionalities might be a more effective strategy to treat bone defect.

Third, in existing reports, conditions, such as time, frequency, and intensity of applied external stimuli, vary significantly among different biomaterials and experimental models, making it challenging for follow-up studies. It is necessary to carry out multidisciplinary cooperation to further investigate the impacts of different stimulation parameters on the physiological microenvironment of different bone tissues.

Fourth, the strength of physical cues mediated by biomaterials depends primarily on the strength of the received external stimulus and homogeneous distribution of stimulus-response components within scaffolds. Heterogeneous output of physical cues may lead to inconsistent rates of new bone growth. Therefore, achieving efficient and consistent signal output and monitoring signal changes in realtime are desired.

Fifth, most of the literature has primarily verified the biosafety of biomaterials associated with physical cues. However, there is still a lack of systematic evaluation of the long-term physiological safety, metabolic mechanisms, and metabolites generated by degradation.

Sixth, most of the current studies on physical cues promoting bone repair are conducted in animal models. Due to the significant differences in species, the experimental results cannot be directly extrapolated to humans, especially to the elderly group with weaker vitality.

Finally, the currently widely studied synthetic bone scaffolds mainly include metallic materials, inorganic materials (bioceramics and bone cements, etc.), organic polymers (natural polymers and synthetic polymers), and composite materials. Different materials vary greatly in rigidity, porosity, hydrophilicity and degradability, so it is necessary to select appropriate single or composite materials according to the actual application scenario. In addition, the high specific surface area, porous structure, and appropriate degree of disordered nanostructures of bone repairing scaffolds also play great influences on bone regeneration. Therefore, a variety of factors need to be comprehensively considered to design scaffolds for better bone regeneration.

In summary, underlying mechanisms and the effects of physical cues (mild thermal effect and electrical and magnetic signals) elicited by various biomaterials for bone regenration with advantages of remote actuation and on-demand activation are elucidated, which show great potential in clinical translation.

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Competing Interests

The authors have declared that no competing interest exists.

References

- [1] Li C, Cui W. 3D bioprinting of cell-laden constructs for regenerative medicine. Eng. Regen. 2021; 2: 195–205.
- [2] Wang Y, Zhang H, Hu Y, Jing Y, Geng Z, Su J. Bone repair biomaterials: a perspective from immunomodulation. Adv. Funct. Mater. 2022; 32: 2208639.
- [3] Liu C, Xu X, Cui W, Zhang H. Metal-organic framework (MOF)-based biomaterials in bone tissue engineering. Eng. Regen. 2021; 2: 105–108.
- [4] Chen Z, Cai Z, Zhuang P, Li F, Cui W, Li Z. Living probiotic biomaterials for osteoporosis therapy. Biomed. Technol. 2023; 1: 52–64.
- [5] Liu S, Zhang L, Li Z, Gao F, Zhang Q, Bianco A, et al. Materials-mediated in situ physical cues for bone regeneration. Adv. Funct. Mater. 2023; 34: 2306534.
- [6] Zhou W, Ma X, Wang J, Xu X, Koivisto O, Feng J, et al. Co-delivery CPT and PTX prodrug with a photo/thermo-responsive nanoplatform for triple-negative breast cancer therapy. Smart Med. 2022; 1: e20220036.

- [7] Xue S, Zhou X, Sang W, Wang C, Lu H, Xu Y, et al. Cartilage-targeting peptide-modified dual-drug delivery nanoplatform with NIR laser response for osteoarthritis therapy. Bioact. Mater. 2021; 6: 2372–2389.
- [8] Li J, Liu X, Tan L, Cui Z, Yang X, Liang Y, et al. Zinc-doped Prussian blue enhances photothermal clearance of Staphylococcus aureus and promotes tissue repair in infected wounds. Nat. Commun. 2019; 10: 4490.
- [9] Yang C, Ma H, Wang Z, Younis M R, Liu C, Wu C, et al. 3D printed Wesselsite nanosheets functionalized scaffold facilitates NIR-II photothermal therapy and vascularized bone regeneration. Adv. Sci. 2021; 8: 2100894.
- [10] Chen S, Guo R, Liang Q, Xiao X. Multifunctional modified polylactic acid nanofibrous scaffold incorporating sodium alginate microspheres decorated with strontium and black phosphorus for bone tissue engineering. J. Biomater. Sci. Polym. Ed. 2021; 32: 1598–1617.
- [11] Sayed S, Faruq O, Hossain M, Im S-B, Kim Y-S, Lee B-T. Thermal cycling effect on osteogenic differentiation of MC3T3-E1 cells loaded on 3D-porous Biphasic Calcium Phosphate (BCP) scaffolds for early osteogenesis. Mater. Sci. Eng. C. 2019; 105: 110027.
- [12] Li Y, He J, Zhou J, Li Z, Liu L, Hu S, et al. A conductive photothermal non-swelling nanocomposite hydrogel patch accelerating bone defect repair. Biomater. Sci. 2022; 10: 1326–1341.
- [13] Lei Q, Chen Y, Gao S, Li J, Xiao L, Huang H, et al. Enhanced magnetothermal effect of high porous bioglass for both bone repair and antitumor therapy. Mater. Des. 2023; 227: 111754.

- [14] Yu Z, Wang H, Ying B, Mei X, Zeng D, Liu S, et al. Mild photothermal therapy assist in promoting bone repair: related mechanism and materials. Mater. Today Bio. 2023; 23: 100834.
- [15] Zhao Y, Hu L, Dai H, Zhuang P, Huang K, Hou W, et al. Smart trap-capture-kill antibacterial system for infected microenvironment improvement and vascularized bone regeneration via magnetic thermotherapy. Adv. Funct. Mater. 2023; 33: 2214734.
- [16] Heng B C, Bai Y, Li X, Lim L W, Li W, Ge Z, et al. Electroactive biomaterials for facilitating bone defect repair under pathological conditions. Adv. Sci. 2023; 10: 2204502.
- [17] Alvarez-Lorenzo C, Zarur M, Seijo-Rabina A, Blanco-Fernandez B, Rodríguez-Moldes I, Concheiro A. Physical stimuli-emitting scaffolds: the role of piezoelectricity in tissue regeneration. Mater. Today Bio 2023; 22: 100740.
- [18] Zheng T, Huang Y, Zhang X, Cai Q, Deng X, Yang X. Mimicking the electrophysiological microenvironment of bone tissue using electroactive materials to promote its regeneration. J. Mater. Chem. B. 2020; 8: 10221–10256.
- [19] Sapir-Lekhovitser Y, Rotenberg M Y, Jopp J, Friedman G, Polyak B, Cohen S. Magnetically actuated tissue engineered scaffold: insights into mechanism of physical stimulation. Nanoscale. 2016; 8: 3386–3399.
- [20] Sun Y, Fang Y, Li X, Li J, Liu D, Wei M, et al. A static magnetic field enhances the repair of osteoarthritic cartilage by promoting the migration of stem cells and chondrogenesis. J. Orthop. Translat. 2023; 39: 43–54.
- [21] Li X, Zou Q, Man Y, Li W. Synergistic effects of novel superparamagnetic/upconversion HA material and Ti/magnet implant on biological performance and long-term in vivo tracking. Small. 2019; 15: 1901617.

- [22] Marycz K, Kornicka K, Röcken M. Static magnetic field (SMF) as a regulator of stem cell fate—new perspectives in regenerative medicine arising from an underestimated tool. Stem Cell Rev. Rep. 2018; 14: 785–792.
- [23] Zhao S, Yu X, Qian Y, Chen W, Shen J. Multifunctional magnetic iron oxide nanoparticles: an advanced platform for cancer theranostics. Theranostics. 2020; 10: 6278–6309.
- [24] Yun H-M, Ahn S-J, Park K-R, Kim M-J, Kim J-J, Jin G-Z, et al. Magnetic nanocomposite scaffolds combined with static magnetic field in the stimulation of osteoblastic differentiation and bone formation. Biomaterials. 2016; 85: 88–98.
- [25] Sadeghzadeh H, Dianat-Moghadam H, Del Bakhshayesh A R, Mohammadnejad D, Mehdipour A. A review on the effect of nanocomposite scaffolds reinforced with magnetic nanoparticles in osteogenesis and healing of bone injuries. Stem Cell Res. Ther. 2023; 14: 194.
- [26] Shan H, Zhou X, Tian B, Zhou C, Gao X, Bai C, et al. Gold nanorods modified by endogenous protein with light-irradiation enhance bone repair via multiple osteogenic signal pathways. Biomaterials. 2022; 284: 121482.
- [27] Zhang X, Cheng G, Xing X, Liu J, Cheng Y, Ye T, et al. Near-infrared light-triggered porous AuPd alloy nanoparticles to produce mild localized heat to accelerate bone regeneration. J. Phys. Chem. Lett. 2019; 10: 4185–4191.
- [28] Xue Y, Zhang L, Liu F, Zhao Y, Zhou J, Hou Y, et al. Surface bandgap engineering of nanostructured implants for rapid photothermal ion therapy of bone defects. Adv. Healthcare Mater. 2022; 11: 2200998.

- [29] Li Q, Liu W, Hou W, Wu X, Wei W, Liu J, et al. Micropatterned photothermal double-layer periosteum with angiogenesis-neurogenesis coupling effect for bone regeneration. Mater. Today Bio. 2023; 18: 100536.
- [30] Ma K, Liao C, Huang L, Liang R, Zhao J, Zheng L, et al. Electrospun PCL/MoS₂ nanofiber membranes combined with NIR-triggered photothermal therapy to accelerate bone regeneration. Small. 2021; 17: 2104747.
- [31] Zhang J, Zhang B, Zheng Z, Cai Q, Wang J, Shu Q, et al. Tissue-engineered bone functionalized with MoS₂ nanosheets for enhanced repair of critical-size bone defect in rats. Adv. Funct. Mater. 2021; 32: 2109882.
- [32] Xue X, Zhang H, Liu H, Wang S, Li J, Zhou Q, et al. Rational design of multifunctional CuS nanoparticle-PEG composite soft hydrogel-coated 3D hard polycaprolactone scaffolds for efficient bone regeneration. Adv. Funct. Mater. 2022; 32: 2202470.
- [33] Wu M, Liu H, Zhu Y, Chen F, Chen Z, Guo L, et al. Mild photothermal-stimulation based on injectable and photocurable hydrogels orchestrates immunomodulation and osteogenesis for high-performance bone regeneration. Small. 2023; 19: 2300111.
- [34] Ma L, Feng X, Liang H, Wang K, Song Y, Tan L, et al. A novel photothermally controlled multifunctional scaffold for clinical treatment of osteosarcoma and tissue regeneration.

 Mater. Today. 2020; 36: 48–62.
- [35] Yanagi T, Kajiya H, Kawaguchi M, Kido H, Fukushima T. Photothermal stress triggered by near infrared-irradiated carbon nanotubes promotes bone deposition in rat calvarial defects. J. Biomater. Appl. 2015; 29: 1109–1118.

- [36] Fan Q, Bai J, Shan H, Fei Z, Chen H, Xu J, et al. Implantable blood clot loaded with BMP-2 for regulation of osteoimmunology and enhancement of bone repair. Bioact. Mater. 2021; 6: 4014–4026.
- [37] Wu Y, Zhang X, Tan B, Shan Y, Zhao X, Liao J. Near-infrared light control of GelMA/PMMA/PDA hydrogel with mild photothermal therapy for skull regeneration. Biomater. Adv. 2022; 133: 112641.
- [38] Zhao Y, Peng X, Wang D, Zhang H, Xin Q, Wu M, et al. Chloroplast-inspired scaffold for infected bone defect therapy: towards stable photothermal properties and self-defensive functionality. Adv. Sci. 2022; 9: 2204535.
- [39] Tong L, Liao Q, Zhao Y, Huang H, Gao A, Zhang W, et al. Near-infrared light control of bone regeneration with biodegradable photothermal osteoimplant. Biomaterials. 2019; 193: 1–11.
- [40] Zhang X, Li Q, Li L, Ouyang J, Wang T, Chen J, et al. Bioinspired mild photothermal effect-reinforced multifunctional fiber scaffolds promote bone regeneration. ACS Nano. 2023; 17: 6466–6479.
- [41] Wang L, Hu P, Jiang H, Zhao J, Tang J, Jiang D, et al. Mild hyperthermia-mediated osteogenesis and angiogenesis play a critical role in magnetothermal composite-induced bone regeneration. Nano Today. 2022; 43: 101401.
- [42] Wan Z, Zhang P, Lv L, Zhou Y. NIR light-assisted phototherapies for bone-related diseases and bone tissue regeneration: a systematic review. Theranostics. 2020; 10: 11837–11861.
- [43] Ateş G B, Ak A, Garipcan B, Gülsoy M. Methylene blue mediated photobiomodulation on human osteoblast cells. Lasers Med. Sci. 2017; 32: 1847–1855.

- [44] Hang K, Ye C, Chen E, Zhang W, Xue D, Pan Z. Role of the heat shock protein family in bone metabolism. Cell Stress and Chaperones. 2018; 23: 1153–1164.
- [45] Leonardi R, Barbato E, Paganelli C, Lo Muzio L. Immunolocalization of heat shock protein 27 in developing jaw bones and tooth germs of human fetuses. Calcified Tissue International. 2004; 75: 509–516.
- [46] Koh J-M, Lee Y-S, Kim Y S, Park S-H, Lee S H, Kim H-H, et al. Heat shock protein 60 causes osteoclastic bone resorption via toll-like receptor-2 in estrogen deficiency. Bone. 2009; 45: 650–660.
- [47] Qu X, Guo Y, Xie C, Li S, Liu Z, Lei B. Photoactivated MXene nanosheets for integrated bone–soft tissue therapy: effect and potential mechanism. ACS Nano. 2023; 17: 7229–7240.
- [48] Kong X, Wan G, Li B, Wu L. Recent advances of polyoxometalates in multi-functional imaging and photothermal therapy. J. Mater. Chem. B. 2020; 8: 8189–8206.
- [49] Jablonski A. Efficiency of anti-stokes fluorescence in dyes. Nature. 1933; 131: 839–840.
- [50] Wang H, Chang J, Shi M, Pan W, Li N, Tang B. A dual-targeted organic photothermal agent for enhanced photothermal therapy. Angew. Chem. Int. Ed. 2018; 58: 1057–1061.
- [51] Jung H S, Verwilst P, Sharma A, Shin J, Sessler J L, Kim J S. Organic molecule-based photothermal agents: an expanding photothermal therapy universe. Chem. Soc. Rev. 2018; 47: 2280–2297.
- [52] Bashkatov A N, Genina E A, Kochubey V I, Tuchin V V. Optical properties of human skin, subcutaneous and mucous tissues in the wavelength range from 400 to 2000 nm. J. Phys. D: Appl. Phys. 2005; 38: 2543–2555.
- [53] Smith A M, Mancini M C, Nie S. Second window for in vivo imaging. Nat. Nanotechnol. 2009; 4: 710–711.

- [54] Lin H, Gao S, Dai C, Chen Y, Shi J. A two-dimensional biodegradable niobium carbide (MXene) for photothermal tumor eradication in NIR-I and NIR-II biowindows. J. Am. Chem. Soc. 2017; 139: 16235–16247.
- [55] Rastinehad A R, Anastos H, Wajswol E, Winoker J S, Sfakianos J P, Doppalapudi S K, et al. Gold nanoshell-localized photothermal ablation of prostate tumors in a clinical pilot device study. Proc. Natl. Acad. Sci. U.S.A. 2019; 116: 18590–18596.
- [56] Wang L, Long N J, Li L, Lu Y, Li M, Cao J, et al. Multi-functional bismuth-doped bioglasses: combining bioactivity and photothermal response for bone tumor treatment and tissue repair. Light: Sci. Appl. 2018; 7: 1.
- [57] Liu Y, Li T, Ma H, Zhai D, Deng C, Wang J, et al. 3D-printed scaffolds with bioactive elements-induced photothermal effect for bone tumor therapy. Acta Biomater. 2018; 73: 531–546.
- [58] Long J, Zhang W, Chen Y, Teng B, Liu B, Li H, et al. Multifunctional magnesium incorporated scaffolds by 3D-Printing for comprehensive postsurgical management of osteosarcoma. Biomaterials. 2021; 275: 120950.
- [59] Ullattil S G, Narendranath S B, Pillai S C, Periyat P. Black TiO₂ nanomaterials: a review of recent advances. Chem. Eng. J. 2018; 343: 708–736.
- [60] Xu B, Li Z, Ye Q, Li G, Xu M, Li Y, et al. Mild photothermal effect of titania nanotubes array as a promising solution for peri-implantitis. Mater. Des. 2022; 217: 110641.
- [61] Xue Y, Zhang L, Liu F, Kong L, Ma D, Han Y. Fluoride releasing photothermal responsive TiO₂ matrices for antibiosis, biosealing and bone regeneration. J. Control. Release. 2023; 363: 657–669.

- [62] Zhang X, Wei H, Dong C, Wang J, Zhang T, Huang L, et al. 3D printed hydrogel/bioceramics core/shell scaffold with NIR-II triggered drug release for chemophotothermal therapy of bone tumors and enhanced bone repair. Chem. Eng. J. 2023; 461: 141855.
- [63] Geng B, Qin H, Shen W, Li P, Fang F, Li X, et al. Carbon dot/WS₂ heterojunctions for NIR-II enhanced photothermal therapy of osteosarcoma and bone regeneration. Chem. Eng. J. 2020; 383: 123102.
- [64] Zeng Y, Pan Z, Yuan J, Song Y, Feng Z, Chen Z, et al. Inhibiting osteolytic breast cancer bone metastasis by bone-targeted nanoagent via remodeling the bone tumor microenvironment combined with NIR-II photothermal therapy. Small. 2023; 19: 2301003.
- [65] Zhang L, Zhang H, Zhou H, Tan Y, Zhang Z, Yang W, et al. A Ti₃C₂ MXene-integrated near-infrared-responsive multifunctional porous scaffold for infected bone defect repair. J. Mater. Chem. B. 2024; 12: 79–96.
- [66] Zhao Y, Kang H, Xia Y, Sun L, Li F, Dai H. 3D printed photothermal scaffold sandwiching bacteria inside and outside improves the infected microenvironment and repairs bone defects. Adv. Healthcare Mater. 2023; 13: 2302879.
- [67] Liu S, Han Z, Hao J-N, Zhang D, Li X, Cao Y, et al. Engineering of a NIR-activable hydrogel-coated mesoporous bioactive glass scaffold with dual-mode parathyroid hormone derivative release property for angiogenesis and bone regeneration. Bioact. Mater. 2023; 26: 1–13.
- [68] Wei C, Jin X, Wu C, Zhang W. Injectable composite hydrogel based on carbon particles for photothermal therapy of bone tumor and bone regeneration. J. Mater. Sci. Technol. 2022; 118: 64–72.

- [69] Luo Z-Y, Liu Z-H, Yu H-D, Chen A-J, Du Z, Cai Y-R, et al. Multifunctional mesoporous polydopamine near-infrared photothermal controlled release of kartogenin for cartilage repair. Mater. Des. 2023; 231: 112007.
- [70] Cheng L, Cai Z, Zhao J, Wang F, Lu M, Deng L, et al. Black phosphorus-based 2D materials for bone therapy. Bioact. Mater. 2020; 5: 1026–1043.
- [71] Tan L, Li M, Luo Z, Cai K, Hu Y. Black phosphorus biomaterials for photo-controlled bone tissue engineering. Compos. Part B Eng. 2022; 246: 110245.
- [72] Tan L, Hu Y, Li M, Zhang Y, Xue C, Chen M, et al. Remotely-activatable extracellular matrix-mimetic hydrogel promotes physiological bone mineralization for enhanced cranial defect healing. Chem. Eng. J. 2022; 431: 133382.
- [73] Shao J, Ruan C, Xie H, Chu P K, Yu X-F. Photochemical activity of black phosphorus for near-infrared light controlled in situ biomineralization. Adv. Sci. 2020; 7: 2000439.
- [74] Pan J, Xu Y, Wu Q, Hu P, Shi J. Mild magnetic hyperthermia-activated innate immunity for liver cancer therapy. J. Am. Chem. Soc. 2021; 143: 8116–8128.
- [75] Afzalipour R, Khoei S, Khoee S, Shirvalilou S, Raoufi N J, Motevalian M, et al. Thermosensitive magnetic nanoparticles exposed to alternating magnetic field and heat-mediated chemotherapy for an effective dual therapy in rat glioma model. Nanomedicine. 2021; 31: 102319.
- [76] Lee H, Thirunavukkarasu G K, Kim S, Lee J Y. Remote induction of in situ hydrogelation in a deep tissue, using an alternating magnetic field and superparamagnetic nanoparticles.

 Nano Res. 2018; 11: 5997–6009.

- [77] Zhuang H, Lin R, Liu Y, Zhang M, Zhai D, Huan Z, et al. Three-dimensional-printed bioceramic scaffolds with osteogenic activity for simultaneous photo/magnetothermal therapy of bone tumors. ACS Biomater. Sci. Eng. 2019; 5: 6725–6734.
- [78] Zhao Y, Fan T, Chen J, Su J, Zhi X, Pan P, et al. Magnetic bioinspired micro/nanostructured composite scaffold for bone regeneration. Colloids Surf. B. 2019; 174: 70–79.
- [79] Li Y, Huang L, Tai G, Yan F, Cai L, Xin C, et al. Graphene oxide-loaded magnetic nanoparticles within 3D hydrogel form high-performance scaffolds for bone regeneration and tumour treatment. Compos. Part A Appl. Sci. Manuf. 2022; 152: 106672.
- [80] Yu K, Liang B, Zheng Y, Exner A, Kolios M, Xu T, et al. PMMA-Fe₃O₄ for internal mechanical support and magnetic thermal ablation of bone tumors. Theranostics. 2019; 9: 4192–4207.
- [81] He Y, Zhao Y, Fan L, Wang X, Duan M, Wang H, et al. Injectable affinity and remote magnetothermal effects of Bi-based alloy for long-term bone defect repair and analgesia. Adv. Sci. 2021; 8: 2100719.
- [82] Hao L, Li J, Wang P, Wang Z, Wu Z, Wang Y, et al. Spatiotemporal magnetocaloric microenvironment for guiding the fate of biodegradable polymer implants. Adv. Funct. Mater. 2021; 31: 2009661.
- [83] Zhao S, Zhang K, Li G, Zhang Z, Li X, Cai B, et al. Novel degradable super-paramagnetic bone cement with self-controlled hyperthermia ability. Mater. Des. 2022; 218: 110676.
- [84] Liu Z, Wan X, Wang Z L, Li L. Electroactive biomaterials and systems for cell fate determination and tissue regeneration: design and applications. Adv. Mater. 2021; 33: 2007429.

- [85] Nauth A, Schemitsch E, Norris B, Nollin Z, Watson J T. Critical-size bone defects: is there a consensus for diagnosis and treatment? J. Orthop. Trauma. 2018; 32: S7–S11.
- [86] e Silva E P, Huang B, Helaehil J V, Nalesso P R L, Bagne L, de Oliveira M A, et al. In vivo study of conductive 3D printed PCL/MWCNTs scaffolds with electrical stimulation for bone tissue engineering. Bio-des. Manuf. 2021; 4: 190–202.
- [87] Wang W, Junior J R P, Nalesso P R L, Musson D, Cornish J, Mendonça F, et al. Engineered 3D printed poly(ε-caprolactone)/graphene scaffolds for bone tissue engineering. Mater. Sci. Eng. C. 2019; 100: 759–770.
- [88] Liu X, George M N, Li L, Gamble D, Miller A L, Gaihre B, et al. Injectable electrical conductive and phosphate releasing gel with two-dimensional black phosphorus and carbon nanotubes for bone tissue engineering. ACS Biomater. Sci. Eng. 2020; 6: 4653–4665.
- [89] Ahadian S, Banan Sadeghian R, Yaginuma S, Ramón-Azcón J, Nashimoto Y, Liang X, et al. Hydrogels containing metallic glass sub-micron wires for regulating skeletal muscle cell behaviour. Biomater. Sci. 2015; 3: 1449–1458.
- [90] Tang J, Zhang Y, Fang W, Man Y, Zhang J, Zhao Q, et al. Electrophysiological microenvironment and site-specific cell behaviors regulated by fibrous aniline trimer-based polyurethanes in bone progressive regeneration. Chem. Eng. J. 2023; 460: 141630.
- [91] Yu P, Ning C, Zhang Y, Tan G, Lin Z, Liu S, et al. Bone-inspired spatially specific piezoelectricity induces bone regeneration. Theranostics. 2017; 7: 3387–3397.
- [92] Liu W, Li X, Jiao Y, Wu C, Guo S, Xiao X, et al. Biological effects of a three-dimensionally printed Ti6Al4V scaffold coated with piezoelectric BaTiO₃ nanoparticles on bone formation. ACS Appl. Mater. Interfaces. 2020; 12: 51885–51903.

- [93] Zheng T, Yu Y, Pang Y, Zhang D, Wang Y, Zhao H, et al. Improving bone regeneration with composites consisting of piezoelectric poly(L-lactide) and piezoelectric calcium/manganese co-doped barium titanate nanofibers. Compos. B Eng. 2022; 234: 109734.
- [94] Zhang X, Zhang C, Lin Y, Hu P, Shen Y, Wang K, et al. Nanocomposite membranes enhance bone regeneration through restoring physiological electric microenvironment. ACS Nano. 2016; 10: 7279–7286.
- [95] Dai X, Yao X, Zhang W, Cui H, Ren Y, Deng J, et al. The osteogenic role of barium titanate/polylactic acid piezoelectric composite membranes as guiding membranes for bone tissue regeneration. Int. J. Nanomed. 2022; 17: 4339–4353.
- [96] Fu Y, Huang S, Feng Z, Huang L, Zhang X, Lin H, et al. MXene-functionalized ferroelectric nanocomposite membranes with modulating surface potential enhance bone regeneration. ACS Biomater. Sci. Eng. 2023; 9: 900–917.
- [97] Zhao F, Zhang C, Liu J, Liu L, Cao X, Chen X, et al. Periosteum structure/function-mimicking bioactive scaffolds with piezoelectric/chem/nano signals for critical-sized bone regeneration. Chem. Eng. J 2020; 402: 126203.
- [98] Zhang C, Liu W, Cao C, Zhang F, Tang Q, Ma S, et al. Modulating surface potential by controlling the β phase content in poly(vinylidene fluoridetrifluoroethylene) membranes enhances bone regeneration. Adv. Healthcare Mater. 2018; 7: 1701466.
- [99] Liu Y, Dzidotor G, Le T T, Vinikoor T, Morgan K, Curry E J, et al. Exercise-induced piezoelectric stimulation for cartilage regeneration in rabbits. Sci. Transl. Med. 2022; 14: eabi7282.

- [100] Noris-Suárez K, Lira-Olivares J, Ferreira A M, Graterol A, Feijoo J L, Lee S W. Electrochemical influence of collagen piezoelectric effect in bone healing. Mater. Sci. Forum. 2007; 544–545: 981–984.
- [101] Minary-Jolandan M, Yu M-F. Nanoscale characterization of isolated individual type I collagen fibrils: polarization and piezoelectricity. Nanotechnology. 2009; 20: 085706.
- [102] Fukada E, Yasuda I. On the piezoelectric effect of bone. J. Phys. Soc. Jpn. 1957; 12: 1158–1162.
- [103] Lipiec E, Kowalska J, Wiecheć A, Zieliński P M, Kwiatek W M, Iwaniec M. Infrared spectroscopy in molecular study of the piezoelectric effect in pig's shin bone. Acta Phys. Pol. 2012; 121: 539–542.
- [104] Tang Y, Wu C, Wu Z, Hu L, Zhang W, Zhao K. Fabrication and in vitro biological properties of piezoelectric bioceramics for bone regeneration. Sci. Rep. 2017; 7: 43360.
- [105] Ahn A C, Grodzinsky A J. Relevance of collagen piezoelectricity to "Wolff's Law": a critical review. Med. Eng. Phys. 2009; 31: 733–741.
- [106] Khare D, Basu B, Kumar Dubey A. Electrical stimulation and piezoelectric biomaterials for bone tissue engineering applications. Biomaterials. 2020; 258: 120280.
- [107] Balint R, Cassidy N J, Cartmell S H. Electrical stimulation: a novel tool for tissue engineering. Tissue Eng., Part B. 2013; 19: 48–57.
- [108] Li X, Kolega J. Effects of direct current electric fields on cell migration and actin filament distribution in bovine vascular endothelial cells. J. Vasc. Res. 2002; 39: 391–404.
- [109] Shen S, He X, Chen X, Dong L, Cheng K, Weng W. Enhanced osteogenic differentiation of mesenchymal stem cells on P(VDF-TrFE) layer coated microelectrodes. J. Biomed. Mater. Res. B. 2021; 109: 2227–2236.

- [110] Raic A, Friedrich F, Kratzer D, Bieback K, Lahann J, Lee-Thedieck C. Potential of electrospun cationic BSA fibers to guide osteogenic MSC differentiation via surface charge and fibrous topography. Sci. Rep. 2019; 9: 20003.
- [111] Liu H, Chen J, Qiao S, Zhang W. Carbon-based nanomaterials for bone and cartilage regeneration: a review. ACS Biomater. Sci. Eng. 2021; 7: 4718–4735.
- [112] Qian D, Dickey E C, Andrews R, Rantell T. Load transfer and deformation mechanisms in carbon nanotube-polystyrene composites. Appl. Phys. Lett. 2000; 76: 2868–2870.
- [113] Ross A M, Jiang Z, Bastmeyer M, Lahann J. Physical aspects of cell culture substrates: topography, roughness, and elasticity. Small. 2012; 8: 336–355.
- [114] Lu Y-L, Cheng C-M, Leduc P R, Ho M-S. Controlling the mechanics and nanotopography of biocompatible scaffolds through dielectrophoresis with carbon nanotubes. Electrophoresis. 2008; 29: 3123–3127.
- [115] Hopley E L, Salmasi S, Kalaskar D M, Seifalian A M. Carbon nanotubes leading the way forward in new generation 3D tissue engineering. Biotechnol. Adv. 2014; 32: 1000–1014.
- [116] Shao S, Zhou S, Li L, Li J, Luo C, Wang J, et al. Osteoblast function on electrically conductive electrospun PLA/MWCNTs nanofibers. Biomaterials. 2011; 32: 2821–2833.
- [117] Huang Y, Zhang L, Ji Y, Deng H, Long M, Ge S, et al. A non-invasive smart scaffold for bone repair and monitoring. Bioact. Mater. 2023; 19: 499–510.
- [118] Tang P, Han L, Li P, Jia Z, Wang K, Zhang H, et al. Mussel-inspired electroactive and antioxidative scaffolds with incorporation of polydopamine-reduced graphene oxide for enhancing skin wound healing. ACS Appl. Mater. Interfaces. 2019; 11: 7703–7714.

- [119] Su Y, Zeng L, Deng R, Ye B, Tang S, Xiong Z, et al. Endogenous electric field-coupled PD@BP biomimetic periosteum promotes bone regeneration through sensory nerve via Fanconi anemia signaling pathway. Adv. Healthcare Mater. 2023; 12: 2203027.
- [120] Jing X, Xu C, Su W, Ding Q, Ye B, Su Y, et al. Photosensitive and conductive hydrogel induced innerved bone regeneration for infected bone defect repair. Adv. Healthcare Mater. 2023; 12: 2201349.
- [121] You J-O, Rafat M, Ye G J C, Auguste D T. Nanoengineering the heart: conductive scaffolds enhance Connexin 43 expression. Nano Lett. 2011; 11: 3643–3648.
- [122] Shuai C, Liu G, Yang Y, Qi F, Peng S, Yang W, et al. A strawberry-like Ag-decorated barium titanate enhances piezoelectric and antibacterial activities of polymer scaffold. Nano Energy. 2020; 74: 104825.
- [123] Chai C, Guo Y, Huang Z, Zhang Z, Yang S, Li W, et al. Antiswelling and durable adhesion biodegradable hydrogels for tissue repairs and strain sensors. Langmuir. 2020; 36: 10448–10459.
- [124] Kaur G, Adhikari R, Cass P, Bown M, Gunatillake P. Electrically conductive polymers and composites for biomedical applications. RSC Adv. 2015; 5: 37553–37567.
- [125] Kašpárková V, Humpolíček P, Capáková Z, Bober P, Stejskal J, Trchová M, et al. Cell-compatible conducting polyaniline films prepared in colloidal dispersion mode. Colloids Surf. B. 2017; 157: 309–316.
- [126] Wang Y, Guo J, Xu D, Gu Z, Zhao Y. Micro-/nano-structured flexible electronics for biomedical applications. Biomed. Technol. 2023; 2: 1–14.

- [127] Nain A, Chakraborty S, Barman S R, Gavit P, Indrakumar S, Agrawal A, et al. Progress in the development of piezoelectric biomaterials for tissue remodeling. Biomaterials. 2024; 307: 122528.
- [128] Khan A, Joshi R, Sharma M K, Huang C-J, Yu J-H, Wang Y-L, et al. The potential of organic piezoelectric materials for next-generation implantable biomedical devices. Nano Trends. 2024; 6: 100032.
- [129] Bai Y, Dai X, Yin Y, Wang J, Sun X, Liang W, et al. Biomimetic piezoelectric nanocomposite membranes synergistically enhance osteogenesis of deproteinized bovine bone grafts. Int. J. Nanomed. 2019; 14: 3015–3026.
- [130] Li J, Mou X, Qiu J, Wang S, Wang D, Sun D, et al. Surface charge regulation of osteogenic differentiation of mesenchymal stem cell on polarized ferroelectric crystal substrate. Adv. Healthcare Mater. 2015; 4: 998–1003.
- [131] Park J B, Kelly B J, Kenner G H, Von Recum A F, Grether M F, Coffeen W W. Piezoelectric ceramic implants: in vivo results. J. Biomed. Mater. Res. 1981; 15: 103–110.
- [132] Shuai C, Liu G, Yang Y, Yang W, He C, Wang G, et al. Functionalized BaTiO₃ enhances piezoelectric effect towards cell response of bone scaffold. Colloids Surf. B. 2020; 185: 110587.
- [133] Shuai C, Liu G, Yang Y, Qi F, Peng S, Yang W, et al. Construction of an electric microenvironment in piezoelectric scaffolds fabricated by selective laser sintering. Ceram. Int. 2019; 45: 20234–20242.
- [134] Wu H, Dong H, Tang Z, Chen Y, Liu Y, Wang M, et al. Electrical stimulation of piezoelectric BaTiO₃ coated Ti6Al4V scaffolds promotes anti-inflammatory polarization

- of macrophages and bone repair via MAPK/JNK inhibition and OXPHOS activation. Biomaterials. 2023; 293: 121990.
- [135] Mao L, Bai L, Wang X, Chen X, Zhang D, Chen F, et al. Enhanced cell osteogenesis and osteoimmunology regulated by piezoelectric biomaterials with controllable surface potential and charges. ACS Appl. Mater. Interfaces. 2022; 14: 44111–44124.
- [136] Park Y-J, Hwang K-S, Song J-E, Ong J L, Ralph Rawls H. Growth of calcium phosphate on poling treated ferroelectric BaTiO₃ ceramics. Biomaterials. 2002; 23: 3859–3864.
- [137] Dubey A K, Basu B, Balani K, Guo R, Bhalla A S. Multifunctionality of perovskites BaTiO₃ and CaTiO₃ in a composite with hydroxyapatite as orthopedic implant materials. Integr. Ferroelectr. 2011; 131: 119–126.
- [138] Tandon B, Blaker J J, Cartmell S H. Piezoelectric materials as stimulatory biomedical materials and scaffolds for bone repair. Acta Biomater. 2018; 73: 1–20.
- [139] Li G, Li Z, Min Y, Chen S, Han R, Zhao Z. 3D-printed piezoelectric scaffolds with shape memory polymer for bone regeneration. Small. 2023; 19: 2302927.
- [140] Itoh S, Nakamura S, Kobayashi T, Shinomiya K, Yamashita K, Itoh S. Effect of electrical polarization of hydroxyapatite ceramics on new bone formation. Calcif. Tissue Int. 2006; 78: 133–142.
- [141] Gandhi A A, Wojtas M, Lang S B, Kholkin A L, Tofail S A M, Bandyopadhyay A. Piezoelectricity in poled hydroxyapatite ceramics. J. Am. Ceram. Soc. 2014; 97: 2867–2872.
- [142] Itoh S, Nakamura S, Nakamura M, Shinomiya K, Yamashita K. Enhanced bone regeneration by electrical polarization of hydroxyapatite. Artif. Organs. 2006; 30: 863–869.

- [143] Itoh S, Nakamura S, Nakamura M, Shinomiya K, Yamashita K. Enhanced bone ingrowth into hydroxyapatite with interconnected pores by electrical polarization. Biomaterials. 2006; 27: 5572–5579.
- [144] Lu L, Ding W, Liu J, Yang B. Flexible PVDF based piezoelectric nanogenerators. Nano Energy. 2020; 78: 105251.
- [145] Liu J, Cheng Y, Wang H, Yang D, Liu C, Dou W, et al. Regulation of TiO₂@PVDF piezoelectric nanofiber membranes on osteogenic differentiation of mesenchymal stem cells. Nano Energy. 2023; 115: 108742.
- [146] Fernandes M M, Correia D M, Ribeiro C, Castro N, Correia V, Lanceros-Mendez S. Bioinspired three-dimensional magnetoactive scaffolds for bone tissue engineering. ACS Appl. Mater. Interfaces. 2019; 11: 45265–45275.
- [147] Wu J, Chen T, Wang Y, Bai J, Lao C, Luo M, et al. Piezoelectric effect of antibacterial biomimetic hydrogel promotes osteochondral defect repair. Biomedicines. 2022; 10: 1165.
- [148] Chen J, Song L, Qi F, Qin S, Yang X, Xie W, et al. Enhanced bone regeneration via ZIF-8 decorated hierarchical polyvinylidene fluoride piezoelectric foam nanogenerator: coupling of bioelectricity, angiogenesis, and osteogenesis. Nano Energy. 2023; 106: 108076.
- [149] Abzan N, Kharaziha M, Labbaf S. Development of three-dimensional piezoelectric polyvinylidene fluoride-graphene oxide scaffold by non-solvent induced phase separation method for nerve tissue engineering. Mater. Des. 2019; 167: 107636.
- [150] Tajitsu Y. Fundamental study on improvement of piezoelectricity of poly(L-lactic acid) and its application to film actuators. IEEE Trans. Ultrason. Ferroelectr. Freq. Control. 2013; 60: 1625–1629.

- [151] Dwivedi R, Pandey R, Kumar S, Mehrotra D. Poly hydroxyalkanoates (PHA): role in bone scaffolds. J. Oral. Biol. Craniofac. Res. 2020; 10: 389–392.
- [152] Chernozem R V, Surmeneva M A, Shkarina S N, Loza K, Epple M, Ulbricht M, et al. Piezoelectric 3-D fibrous poly(3-hydroxybutyrate)-based scaffolds ultrasound-mineralized with calcium carbonate for bone tissue engineering: inorganic phase formation, osteoblast cell adhesion, and proliferation. ACS Appl. Mater. Interfaces. 2019; 11: 19522–19533.
- [153] Fukada E. History and recent progress in piezoelectric polymers. IEEE Trans. Ultrason. Ferroelectr. Freq. Control. 2000; 47: 1277–1290.
- [154] Carter A, Popowski K, Cheng K, Greenbaum A, Ligler F S, Moatti A. Enhancement of bone regeneration through the converse piezoelectric effect, a novel approach for applying mechanical stimulation. Bioelectricity. 2021; 3: 255–271.
- [155] Li C, Zhu A, Yang L, Wang X, Guo Z. Advances in magnetoelectric composites for promoting bone regeneration: a review. J. Mater. Chem. B. 2024; 12: 4361–4374.
- [156] Liu W, Zhao H, Zhang C, Xu S, Zhang F, Wei L, et al. In situ activation of flexible magnetoelectric membrane enhances bone defect repair. Nat. Commun. 2023; 14: 4091.
- [157] Wang H, Tian J, Jiang Y, Liu S, Zheng J, Li N, et al. A 3D biomimetic optoelectronic scaffold repairs cranial defects. Sci. Adv. 2023; 9: eabq7750.
- [158] Khan A, Joshi R, Sharma M K, Ganguly A, Parashar P, Wang T-W, et al. Piezoelectric and triboelectric nanogenerators: promising technologies for self-powered implantable biomedical devices. Nano Energy. 2024; 119: 109051.
- [159] Lai Y-H, Chen Y-H, Pal A, Chou S-H, Chang S-J, Huang E W, et al. Regulation of cell differentiation via synergistic self-powered stimulation and degradation behavior of a

- biodegradable composite piezoelectric scaffold for cartilage tissue. Nano Energy. 2021; 90: 106545.
- [160] Xia Y, Sun J, Zhao L, Zhang F, Liang X-J, Guo Y, et al. Magnetic field and nano-scaffolds with stem cells to enhance bone regeneration. Biomaterials. 2018; 183: 151–170.
- [161] Ma M, Zou F, Abudureheman B, Han F, Xu G, Xie Y, et al. Magnetic microcarriers with accurate localization and proliferation of mesenchymal stem cell for cartilage defects repairing. ACS Nano. 2023; 17: 6373–6386.
- [162] Zou Y-P, Liang H-F, Wang B, Zhang Q-C, Su D-H, Lu S-Y, et al. Precipitation-based silk fibroin fast gelling, highly adhesive, and magnetic nanocomposite hydrogel for repair of irregular bone defects. Adv. Funct. Mater. 2023; 33: 2302442.
- [163] Yan Z, Sun T, Tan W, Wang Z, Yan J, Miao J, et al. Magnetic field boosts the transmembrane transport efficiency of magnesium ions from PLLA bone scaffold. Small. 2023; 19: 2301426.
- [164] Huang Z, He Y, Chang X, Liu J, Yu L, Wu Y, et al. A magnetic iron oxide/polydopamine coating can improve osteogenesis of 3D-printed porous titanium scaffolds with a static magnetic field by upregulating the TGFβ-Smads pathway. Adv. Healthcare Mater. 2020; 9: 2000318.
- [165] Wu D, Kang L, Tian J, Wu Y, Liu J, Li Z, et al. Exosomes derived from bone mesenchymal stem cells with the stimulation of Fe₃O₄ nanoparticles and static magnetic field enhance wound healing through upregulated miR-21-5p. Int. J. Nanomed. 2020; 15: 7979–7993.
- [166] Yan Z, Wang D, Cai J, Shen L, Jiang M, Liu X, et al. High-specificity protection against radiation-induced bone loss by a pulsed electromagnetic field. Sci. Adv. 2022; 8: eabq0222.

- [167] Kesse X, Adam A, Begin-Colin S, Mertz D, Larquet E, Gacoin T, et al. Elaboration of superparamagnetic and bioactive multicore–shell nanoparticles (γ-Fe₂O₃@SiO₂-CaO): a promising material for bone cancer treatment. ACS Appl. Mater. Interfaces. 2020; 12: 47820–47830.
- [168] Han Y, Yang H, Hua Z, Nie S, Xu S, Zhou C, et al. Rotating magnetic field mitigates ankylosing spondylitis targeting osteocytes and chondrocytes via ameliorating immune dysfunctions. Cells. 2023; 12: 972.
- [169] Zhang J, Ding C, Ren L, Zhou Y, Shang P. The effects of static magnetic fields on bone. Prog. Biophys. Mol. Biol. 2014; 114: 146–152.
- [170] Torbet J, Ronzière M-C. Magnetic alignment of collagen during self-assembly. Biochem. J. 1984; 219: 1057–1059.
- [171]Kotani H, Iwasaka M, Ueno S, Curtis A. Magnetic orientation of collagen and bone mixture.

 J. Appl. Phys. 2000; 87: 6191–6193.
- [172] Shuai C, Cheng Y, Yang W, Feng P, Yang Y, He C, et al. Magnetically actuated bone scaffold: microstructure, cell response and osteogenesis. Compos. B: Eng. 2020; 192: 107986.
- [173]Yan Q C, Tomita N, Ikada Y. Effects of static magnetic field on bone formation of rat femurs. Med. Eng. Phys. 1998; 20: 397–402.
- [174] Di S, Tian Z, Qian A, Li J, Wu J, Wang Z, et al. Large gradient high magnetic field affects FLG29.1 cells differentiation to form osteoclast-like cells. Int. J. Radiat. Biol. 2012; 88: 806–813.
- [175] Zhang J, Ding C, Ren L, Zhou Y, Shang P. The effects of static magnetic fields on bone. Prog. Biophys. Mol. Biol. 2014; 114: 146–152.

- [176] Shuai C, Yang W, Peng S, Gao C, Guo W, Lai Y, et al. Physical stimulations and their osteogenesis-inducing mechanisms. Int. J. Bioprint. 2018; 4: 138.
- [177] Tang Y-Q, Wang Q-Y, Ke Q-F, Zhang C-Q, Guan J-J, Guo Y-P. Mineralization of ytterbium-doped hydroxyapatite nanorod arrays in magnetic chitosan scaffolds improves osteogenic and angiogenic abilities for bone defect healing. Chem. Eng. J. 2020; 387: 124166.
- [178] Yu C, Yang W, Yang L, Ye L, Sun R, Gu T, et al. Synergistic effect of magneto-mechanical bioengineered stem cells and magnetic field to alleviate osteoporosis. ACS Appl. Mater. Interfaces 2023; 15: 19976–19988.
- [179] Trujillo-Alonso V, Pratt E C, Zong H, Lara-Martinez A, Kaittanis C, Rabie M O, et al. FDA-approved ferumoxytol displays anti-leukaemia efficacy against cells with low ferroportin levels. Nat. Nanotechnol. 2019; 14: 616–622.
- [180] Meng J, Xiao B, Zhang Y, Liu J, Xue H, Lei J, et al. Super-paramagnetic responsive nanofibrous scaffolds under static magnetic field enhance osteogenesis for bone repair in vivo. Sci. Rep. 2013; 3: 2655.
- [181] Fan D, Wang Q, Zhu T, Wang H, Liu B, Wang Y, et al. Recent advances of magnetic nanomaterials in bone tissue repair. Front. Chem. 2020; 8: 745.
- [182] Marycz K, Sobierajska P, Roecken M, Kornicka-Garbowska K, Kępska M, Idczak R, et al. Iron oxides nanoparticles (IOs) exposed to magnetic field promote expression of osteogenic markers in osteoblasts through integrin alpha-3 (INTa-3) activation, inhibits osteoclasts activity and exerts anti-inflammatory action. J. Nanobiotechnol. 2020; 18: 33.
- [183] Xu H-Y, Gu N. Magnetic responsive scaffolds and magnetic fields in bone repair and regeneration. Front. Mater. Sci. 2014; 8: 20–31.

- [184] Tang B, Shen X, Yang Y, Xu Z, Yi J, Yao Y, et al. Enhanced cellular osteogenic differentiation on CoFe₂O₄/P(VDF-TrFE) nanocomposite coatings under static magnetic field. Colloids Surf. B. 2021; 198: 111473.
- [185] He J, Hu H, Zeng X, Lan F, Wu F, Wu Y. A magnetic hydroxyapatite composite scaffold-based magnetic therapy for bone repair: an experimental study in canis lupus familiaris. Regen. Biomater. 2017; 4: 97–103.
- [186] Zeng X B, Hu H, Xie L Q, Lan F, Jiang W, Wu Y, et al. Magnetic responsive hydroxyapatite composite scaffolds construction for bone defect reparation. Int. J. Nanomed. 2012; 7: 3365–3378.
- [187] Zhao Y-Z, Chen R, Xue P-P, Luo L-Z, Zhong B, Tong M-Q, et al. Magnetic PLGA microspheres loaded with SPIONs promoted the reconstruction of bone defects through regulating the bone mesenchymal stem cells under an external magnetic field. Mater. Sci. Eng. C. 2021; 122: 111877.
- [188] Wu D, Chang X, Tian J, Kang L, Wu Y, Liu J, et al. Bone mesenchymal stem cells stimulation by magnetic nanoparticles and a static magnetic field: release of exosomal miR-1260a improves osteogenesis and angiogenesis. J. Nanobiotechnol. 2021; 19: 209.
- [189] Li S, Dong C, Lv Y. Magnetic liquid metal scaffold with dynamically tunable stiffness for bone tissue engineering. Biomater. Adv. 2022; 139: 212975.
- [190] Zhang L, Dai W, Gao C, Wei W, Huang R, Zhang X, et al. Multileveled hierarchical hydrogel with continuous biophysical and biochemical gradients for enhanced repair of full-thickness osteochondral defect. Adv. Mater. 2023; 35: 2209565.

- [191] Chen H, Sun J, Wang Z, Zhou Y, Lou Z, Chen B, et al. Magnetic cell–scaffold interface constructed by superparamagnetic IONP enhanced osteogenesis of adipose-derived stem cells. ACS Appl. Mater. Interfaces. 2018; 10: 44279–44289.
- [192] Padilla F, Puts R, Vico L, Raum K. Stimulation of bone repair with ultrasound: A review of the possible mechanic effects. Ultrasonics. 2014; 54: 1125–1145.
- [193] Assanah F, Grassie K, Anderson H, Xin X, Rowe D, Khan Y. Ultrasound-derived mechanical stimulation of cell-laden collagen hydrogels for bone repair. J. Biomed. Mater. Res. Part A. 2023; 111: 1200–1215.
- [194] He Y, Li F, Jiang P, Cai F, Lin Q, Zhou M, et al. Remote control of the recruitment and capture of endogenous stem cells by ultrasound for in situ repair of bone defects. Bioact. Mater. 2023; 21: 223–238.
- [195] Huang D, Cai L, Li N, Zhao Y. Ultrasound-trigged micro/nanorobots for biomedical applications. Smart Med. 2023; 2: e20230003.