

Review

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# Prostate-specific membrane antigen-targeted surgery in prostate cancer: Accurate identification, real-time diagnosis, and precise resection

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# Abstract

Radical prostatectomy (RP) combined with pelvic lymph node dissection (PLND) is the first step in multimodal treatment of prostate cancer (PCa) without distant metastases. For a long time, the surgical resection range has been highly dependent on the surgeon's visualization and experience with preoperative imaging. With the rapid development of prostate-specific membrane antigen positron emission tomography and single-photon emission computed tomography (PSMA-PET and PSMA-SPECT), PSMA-targeted surgery has been introduced for a more accurate pathological diagnosis and complete resection of positive surgical margins (PSMs) and micro-lymph node metastases (LNMs). We reviewed PSMA-targeted surgeries, including PSMA-PET-guided prostatic biopsy (PSMA-TB), PSMA-targeted radio-guided surgery (PSMA-RGS), PSMA-targeted fluorescence-guided surgery (PSMA-FGS), and multi-modality/multi-targeted PSMA-targeted surgery. We also discuss the strengths and challenges of PSMA-targeted surgery, and propose that PSMA-targeted surgery could be a great addition to existing surgery protocols, thereby improving the accuracy and convenience of surgery for primary and recurrent PCa in the near future.

Keywords: prostate cancer; prostate-specific membrane antigen (PSMA); molecular imaging; intraoperative; image-guided surgery

# 1. Introduction

As the most frequently diagnosed cancer, prostate cancer (PCa) is the second leading cause of death in men worldwide [1]. Radical prostatectomy (RP) with pelvic lymph node dissection (PLND) is the optimal choice for patients with intermediate-to high-risk PCa with or without regional lymph node

metastases (LNMs) [2, 3]. Accurate and complete resection of PCa lesions and positive LNMs while preserving healthy tissues as much as possible is essential but difficult, depending on the surgeon's experience in the interpretation of preoperative imaging and anatomical knowledge. Prostate-specific membrane antigen (PSMA)-targeted surgery has revolutionized traditional surgery and is a new trend in PCa surgery.

Prostate-specific membrane antigen (PSMA), also known as FOLH1 or GCP II, is a folate gamma glutamyl carboxypeptidase that is highly expressed in more than 90% of PCa lesions-including primary lesions, LNMs, and bone metastases-with 100-1000-fold higher expression level than that in normal prostate (NP) tissues [4, 5]. The significantly higher PSMA expression in PCa makes it an excellent target for PSMA-targeted molecular imaging and therapies [6, 7]. PSMA-targeted surgery involves using PSMA as a molecular marker to enhance the precision of PCa for an intuitive, fast and reliable way to discriminate PCa tumors from NP tissue, making the outcome of surgery less dependent on the surgeon's own experience and preoperative imaging, visual findings, or frozen sections during surgery.

PSMA-targeted surgery can be categorized as PSMA-PET-guided targeted prostatic biopsy (PSMA-TB), PSMA-targeted radio-guided surgery (PSMA-RGS), or PSMA-targeted fluorescence-guided surgery (PSMA-FGS), according to the aim and type of imaging agent.

PSMA-TB uses the precision of prebiopsy PSMA-PET to improve the accuracy of image-guided PCa biopsy [8, 9]. For this purpose, PSMA-PET/MRI is the preferred imaging modality because of the better spatial resolution of MRI; PSMA-PET/CTguided biopsy is also reliable [10, 11]. For PSMA-RGS, the agent contains a PSMA-targeted ligand, linker, gamma-emitting radionuclide, and such <sup>111</sup>In-PSMA imaging and therapy (<sup>111</sup>In-PSMA-I&T), <sup>99m</sup>Tc-PSMA imaging and surgery (<sup>99m</sup>Tc-PSMA-I&S) etc. [12, 13]. For PSMA-FGS, the agent is comprised of a PSMA-targeting ligand, linker, and fluorescent probe, such as OTL78 and IS-002 etc. [14-16].

PSMA-RGS has been clinically applied in hospitals in Europe and Australia, and the first Phase IIa study of PSMA-FGS agents has been completed in 18 PCa patients [15]. Although PSMA-FGS began its clinical trials later than PSMA-RGS, it is considered by many to be more convenient than PSMA-TB and PSMA-RGS, due to its radiation-free nature, making it popular among patients and surgeons.

PSMA-targeted surgery has the potential to replace traditional guided surgery methods for PCa. In this review, we aim to provide an overview of the status of PSMA-targeted surgery for PCa, highlighting its efficiency in detecting positive LNMs and positive surgical margins (PSMs). We also focus on patient selection before PSMA-targeted surgery and follow-up after surgery, and the several clinical questions that urgently need to be addressed.

# 2. Selecting proper candidates for PSMA-targeted surgery

As shown in Table 1, the inclusion criteria for PSMA-targeted surgeries remain unclear. In most studies on recurrent PCa, PSMA-PET is necessary, whereas the lymph node invasion nomogram is mostly used for patients with primary PCa. In fact, PSMA-PET should be conducted for each patient before prostate biopsy to perform PSMA-TB to improve the diagnostic accuracy of PSMA-PET for clinically significant prostate cancer (csPCa) [17].

Table 1. Inclusion Criteria of patients for PSMA-RGS.

Year	Country	Inclusion Criteria
Primary PCa		
2015[13]	Germany	Primary PCa and suspicion of recurrent LNMs on preoperative PSMA PET/CT
2018[38], 2021[49]	Germany	Suspicion of LNMs on preoperative PSMA PET/CT
2022[50]	Italy	Intermediate- or high-risk cN0cM0 PCa at conventional imaging with a risk of LNI of >5% (Briganti nomogram risk)
2022[51]	Australia	Primary high-risk PCa (≥cT3a, international society of urological pathology (ISUP) Grade Group ≥3 or PSA of ≥ 15 ng/mL) with potential LNMs (Briganti nomogram risk >10% or on preoperative imaging)
Recurrent PCa		
2016[37]	Germany	Rising PSA level to ≥0.2 ng/mL after primary curative therapy; <sup>68</sup> Ga-PSMA-11 PET-positive soft tissue lesions
2018[45]	Germany	Recurrent PCa on preoperative PSMA PET/CT
2022[42]	Netherlands	Up to three pelvic PCa recurrences (nodal or local) on preoperative PSMA PET/CT
2022[54]	Germany	Recurrent PCa after initial RP with positive LNMs on preoperative PSMA PET/CT
2023[42]	Germany	Patients with positive lesion detected on PSMA PET imaging in the pelvis or retroperitoneum suspicious for lymph node metastases (LNMs) or local recurrence.
2023[69]	Germany	Conforming initial treatment with either radical prostatectomy (RPE) or curatively intended external beam radiation, prostate-specific antigen (PSA) ≥0.5 ng/mL, positive 6%Ga-PSMA I&T PET/CT scan with no more than 2 intense PSMA-positive lymph nodes (maximum uptake at least 6 kBq/mL), and no signs of local recurrence or distant metastases, as well as informed consent to participate in this study.

Compared with the traditional viewpoint of using PSMA-PET as a staging tool after pathologically confirmed PCa, PSMA-TB may further improve the accuracy of biopsy and select suitable patients with higher uptake for PSMA-targeted surgery (Figure 1). Before PSMA-targeted surgery, patients with csPCa must be correctly diagnosed, and the presence of distant metastases (M1 stage) must be excluded. Even in patients with previously negative biopsy results, PSMA-TB can achieve a sensitivity of 100.0%, a specificity of 68.4%, a positive predictive value (PPV) of 66.7%, a negative predictive value (NPV) of 100.0%, and an accuracy of 80.6% for the detection of csPCa [18, 19]. Furthermore, PSMA-PET and multiparametric magnetic resonance imaging (mpMRI) should



both be used as first-line imaging modalities to diagnose and stage primary PCa before prostate biopsy (PB) [6, 7].

PSMA-PET clearly illuminates the borders of primary PCa. In comparison between PSMA-PET and robot-assisted radical prostatectomy (RARP) results, the PSMA-TB recommendation correlates better with the final histopathological results of poly(ADP-ribose) polymerase (PARP) tests. PSMA-PET can accurately detect PCa in 93.3% (28/30) and cover the highest Gleason score (GS) in 86.7% (26/30) of patients with intermediate-to-high-risk Pca [20].

Compared with mpMRI, PSMA-PET/CT has similar sensitivity (83.1% vs. 90.1%; p = 0.267). The missed lesions by PSMA-PET are smaller than those missed by mpMRI ( $0.72 \text{ cm}^3 vs. 1.66 \text{ cm}^3$ ; p = 0.034) in Pca patients with similar serum prostate-specific antigen (PSA) levels, prostate volumes, and PSA density (PSAD) [10]. In patients undergoing PI-RADs v2 mpMRI imaging, PSMA-PET can additionally identify 81.8% (9/11) of patients (GS  $\geq$  3+4) with csPCa [10]. PSMA-PET/CT can also be used to predict pathological upgrading of the GS from mpMRItargeted PB to RP, with SUV<sub>max</sub> (OR: 1.223, 95% CI: 1.068–1.399, P = 0.003) as the most significant factor [21]. A higher SUV<sub>max</sub> (high avidity) can be used to improve the diagnosis of csPCa using PSMA-TB [22, 23]

Furthermore, compared with traditional systematic PB, PSMA-PET/CT targeted PB may improve the detection rate of clinically significant PCa with GS  $\geq$  7 (3+4) [11, 24]. The sensitivity (100.0% *vs.* 90.9%), specificity (80.3% *vs.* 78.9%), PPV (87.9% *vs.* 78.9%), NPV (100% *vs.* 84.9%), and accuracy (92.0% *vs.* 86.2%) of PSMA-TB were higher than those of mpMRI-TB when an SUV<sub>max</sub> cutoff value of 8 was used [25]. In another study, PSMA-TB was performed

for repeat PB in 31 patients with previously negative PB results, but persistently elevated serum PSA levels [19]. PCa was found in 48.4% (15/31) of patients, 38.7% (12/31) of whom had csPCa [19]. The sensitivity, specificity, PPV, NPVs, and accuracy of PSMA-PET to detect csPCa were 100.0%, 68.4%, 66.7%, 100.0%, and 80.6%, respectively [19].

In addition to PSMA-PET/CT, PSMA-PET/MRI can also be used for guided PB. In a prospective study, PSMA-PET/MRI-guided targeted PB successfully detected csPCa in 61.9% (26/42) of patients, achieving patient-based sensitivity, specificity, PPV, NPV, and accuracy of 96%, 81%, 93%, 89%, and 90%, respectively [8]. In 55 patients with positive PSMA-PET results, 82.0% (146/178) and 92.7% (51/55) had PCa, and 85.5% (47/55) had csPCa [26]. In a single-center study of 120 patients with elevated serum PSA levels (>4 ng/mL), transgluteal PSMA-TB detected more PCa and csPCa lesions than template PB (csPCa: 20/25 [80.0%] vs. 15/60 [25.0%], P < 0.01; PCa: 21/25 [84.0%] vs. 19/60 [31.6%], P < 0.01) [11]. Similarly, in 56 patients with positive PSMA-PET-avid lesions, transgluteal PSMA-TB confirmed PCa lesions in 96.4% (54/56), of which csPCa lesions were found in 44.4% (24/54) of patients [17]. PSMA-TB detected more csPCa lesions than systemic PB (81.7% vs. 76.7%, P < 0.001), whereas systemic PB can still have additional diagnostic benefits in 6.6% (4/60) patients [27].

For patients with regional LNMs, RP±PLND should be performed. PSMA-PET/CT can be used to exclude patients with distal LNMs (M1a stage), bone metastases (M1b stage), or visceral metastases (M1c stage) from planned curative radical prostatectomy [28]. For LNMs with a diameter ≥2.3 mm, PSMA-PET can achieve a detection rate of 50.0% [29]. The detection rate goes down significantly for LNMs with

a diameter < 2.3 mm; small LNMs can be difficult to discover by preoperative PSMA-PET.

Therefore, PSMA-targeted surgery is needed to illuminate smaller LNMs intraoperatively and provide a chance to completely remove all primary lesions and regional LNMs. Based on the agent, PSMA-targeted surgery can be further divided into PSMA-RGS, PSMA-FGS, and multi-modality PSMA-targeted surgery.

# 3. PSMA-RGS

A series of agents have been developed for PSMA-RGS [30]. For example, PSMA-I&T can be labeled with 68Ga for PET imaging, 177Lu for targeted radionuclide therapy, and 111In for RGS or single-photon emission computed tomography/ computed tomography (SPECT/CT) [31]. Mease et al. first established PSMA-targeted PET agent <sup>18</sup>F-DCFBC and its first in-patient imaging study in five patients with metastatic PCa [32, 33]. With the advent of 68Ga-PSMA PET for PCa, PSMA-targeted RGS was introduced to intraoperatively detect PSMA-avid LNMs with atypical localization and inconspicuous morphology. To the best of our knowledge, PSMA-targeted RGS systems, such as PSMA-I&S and PSMA-I&T, are currently the most mature methods for intraoperative navigation in clinical practice (Figure 2A-K). For theranostic purposes, PSMA ligands could be labeled with short half-life imaging

isotopes, such as <sup>68</sup>Ga (T<sub>1/2</sub>: 68 minutes), <sup>18</sup>F (T<sub>1/2</sub>: 110 minutes), <sup>111</sup>In (T<sub>1/2</sub>: 2.8 days), <sup>99m</sup>Tc (T<sub>1/2</sub>: 6 hours), and for therapeutic isotopes, <sup>177</sup>Lu (T<sub>1/2</sub>: 6.7 days], <sup>225</sup>Ac (T<sub>1/2</sub>: 10.0 days), etc. Documented theranostic isotopes and their characteristics are shown in Table 2. We, aimed to further summarize the recent updates of PSMA-RGS according to agent type.

### 3.1 IIIn-labeled PSMA-RGS

Although the detection rate of <sup>111</sup>In SPECT/CT is lower than that of <sup>68</sup>Ga-PSMA-11 PET/CT, the longer half-time of <sup>111</sup>In enables <sup>111</sup>In-PSMA-I&T to be appropriate for PSMA-RGS [34]. Currently, most PSMA-targeted RGS systems are still highly dependent on γ-detecting probes and γ-emitting radiotracers, such as <sup>111</sup>In and <sup>99m</sup>Tc [35].

<sup>111</sup>In-PSMA is the most widely used agent for PSMA-RGS, successfully identifying 59% (29/49) PSMA-avid lesions by  $\gamma$ -probe. Among them, 97% of lymph nodes were confirmed as LNMs with a mean size of 7.9 mm (range, 0.8-20.0 nm) [36]. The effectiveness of <sup>111</sup>In-PSMA RGS is moderate due to interfering tracer accumulation in surrounding organs, leading to a more limited tumor-tobackground ratio (TBR) *in vivo* compared with *ex vivo*. This issue might be mitigated by extending the time between the administration of the tracer and the PSMA-RGS procedure.

 Table 2. Isotopes and relevant characteristics for PSMA-targeted radionuclides.

Isotopes	Agents	Imaging/therapeutic properties	Half-lives	Stage	Strengths	Limitations			
Commonly used									
<sup>111</sup> In	<sup>111</sup> In-PSMA-I&T <sup>111</sup> In-PSMA-617	Imaging-PSMA-RGS	2.83 d	Clinical trial	Earliest used agents	Suboptimal nuclear properties, high cost, and limited availability of <sup>111</sup> InCl <sub>3</sub>			
99mTc	<sup>111</sup> In-PSMA-I&S	Imaging-PSMA-RGS	6.02 h	Clinical trial	Optimal nuclear properties, lower cost, and easily available	Inferior ability in pre-operative imaging, compared with PET/CT			
<sup>177</sup> Lu	<sup>177</sup> Lu-PSMA-617; <sup>177</sup> Lu-PSMA-I&T	Beta-therapy	6.65 d	Clinical trial	Most widely used agents for therapy of CRPC	Heterogeneous PSMA expression after ADT			
<sup>225</sup> Ac	<sup>225</sup> Ac-PSMA-617	Alpha-therapy	10.00 d	Clinical trial	Higher level of radiobiologic effectiveness when compared with Beta-therapy	Heterogeneous PSMA expression after ADT			
<sup>18</sup> F	18F-DCFPyL	PET/CT	110 min	FDA approved 2021	High resolution	Relatively limited availability			
<sup>68</sup> Ga	<sup>68</sup> Ga-PSMA-I&T <sup>68</sup> Ga-PSMA-11	PET/CT	6.7 h	FDA approved 2020 (PSMA-11)	High resolution	Relatively limited availability			
Uncomm	only used								
<sup>44</sup> Sc	<sup>44</sup> Sc-PSMA-617	PET/CT	3.93 h	Clinical trial	More appropriate for delayed imaging				
<sup>64</sup> Cu	<sup>64</sup> Cu-CA003; <sup>64</sup> Cu-CA003	PET/CT	12.7 h	Preclinical		Instability of 64Cu complexes in vivo			
<sup>89</sup> Zr	<sup>89</sup> Zr-PSMA-DFO	PET/CT	78.4 h	Clinical trial	Localize lesions with weak PSMA expression				
131I	131I-MIP-1095	Beta-therapy	8.02 d	Clinical trial	A potential alternative to 177Lu-PSMA				
<sup>161</sup> Tb	<sup>161</sup> Tb-PSMA-617	Alpha-therapy	7.90 d	Preclinical	A potential alternative to 177Lu-PSMA				
<sup>165</sup> Er	<sup>165</sup> Er-PSMA-617	Alpha-therapy	10.40 h						
<sup>212</sup> Pb	<sup>212</sup> Pb-CA012	Alpha-therapy	10.64 h	Preclinical	A potential alternative to <sup>225</sup> Ac-PSMA				
<sup>213</sup> Bi	<sup>213</sup> Bi-PSMA-617; <sup>213</sup> Bi-PSMA I&T <sup>213</sup> Bi-JVZ-008	Alpha-therapy	45.59 min	Clinical trial	A potential alternative to <sup>225</sup> Ac-PSMA				
<sup>227</sup> Th	<sup>227</sup> Th-PSMA-TTC (BAY 2315497)	Alpha-therapy	18.70 d	Preclinical	Synergistic antitumor efficacy in combination with darolutamide				



Figure 2. Chemical structures of agents in PSMA-targeted RGS/FGS. (A-K) PSMA ligands for PSMA-RGS; (L-Y) PSMA ligands for PSMA-FGS. Blue, yellow, green, red, orange areas separately highlight the PSMA binding motif, chelated area for the labeled radioisotope, albumin binder, linker, and labeled near-infrared fluorescent probe.

Table 3. PSMA-RGS in PCa patients.

Year	Country	Retro/ pros	Patient number	Patient type	Preoperative staging	Gamma probe	Agents for PSMA-RGS	Age	tPSA (ng/ml)	GS	sensitivit y	specificit y	PPV	NPV	Accuracy
2015[13]	Germany	R	5	Primary PCa; Recurrent PCa	68Ga-PSMA-1 1 PET/CT	Crystal Probe CXS-SG60 3	111In-PSMA-I& T	75 (IQR 64-75)	2.45 (IQR 0.46- 4.36).		100.0%	100.0%	100.0 %	100.0 %	100.0%
2016[37]	Germany	R	31	Recurrent Pca	68Ga-PSMA-1 1 PET/CT	Crystal Probe CXS-SG60 3	111In-PSMA-I& T	68.2 (Range 52-76)	1.3 (IQR 0.57-2.53)	7 (≤6-10)	92.3%	93.5%	88.9%	95.6 %	93.1%
2018[38]	Germany	R	6	Primary Pca	68Ga-PSMA-1 1 PET/CT	Ex situ gamma probe	111In-PSMA-617	71 (IQR 66.5-75.5)	7.03 (IQR 1.31- 12.02)	7 (7-9)	92.1%	98.9%	94.6%	98.4 %	97.7%
2018[45]	Germany	R	31	Recurrent PCa	68Ga-PSMA-1 1 PET/CT	Crystal Probe CXS-SG60 3	99mTc-PSMA-I &S	66.7(IQR 60.5-73.5)	1.13 (IQR: 0.71- 2.35)	7 (5-9)	83.6%	100.0%	100.0 %	89.2 %	93.0%
2021[49]	Germany	R	6	Primary PCa; Recurrent PCa	68Ga-PSMA-1 1 PET/CT; 18F-PSMA-10 07	In situ/Ex situ gamma probe	99mTc-PSMA-I &S	64(IQR 54.7-67.3)	33.3 (IQR: 4.6-64.4)	8 (7b-9)	76.6%	94.4%	89.4%	86.9 %	
2022[42]	Netherla nds	Р	20	Recurrent Pca	68Ga-PSMA-1 1; 18F-DCFPyl PET/CT	DROP-IN Gamma Probe	99mTc-PSMA-I &S	68 (IQR 66–72)	1.02 (IQR 0.46- 2.43).		86.0%	100.0%	100.0 %	95.0 %	80.0%
2022[50]	Italy	Р	12	Primary PCa	68Ga-PSMA-1 1 PET/MRI	DROP-IN Gamma Probe	99mTc-PSMA-I &S	70 (IQR 66-71)	8.7 (IQR: 4.8-15.5)	8 (7a-9)	63.0%	99.0%	83.0%	96.0 %	
2022[51]	Australia	Р	12	Primary PCa	68Ga-PSMA-1 1 PET/CT	DROP-IN gamma probe	99mTc-PSMA-I &S	68 (IQR 57–69)	9.15 (IQR: 6.0 - 21.2)	9 (8-9)	76.0%	69.0%	50.0%	88%,	
2022[54]	Germany	R	364	Recurrent Pca	PSMA PET/CT	Crystal Probe CXS-SG60 3	111In-PSMA-I& T or 99mTc-PSMA-I &S	67 (IQR 62-71)	1.0 (IQR: 0.5 - 1.9)	7a (6-≥9)					
2023[42]	Germany	Р	85	Recurrent Pca	PSMA PET/CT	Crystal Probe CXS-SG60 3	99mTc-PSMA-I &S	63 (IQR 60-69) Open; 64 (IQR60-6 7) Robot	0.58 (0.38, 0.87) Open; 0.44 (0.31, 0.71) Robot						
2023[69]	Germany	Р	6	Recurrent Pca	68Ga-PSMA I&T PET/CT	Crystal Probe CXS-SG60 3	67Ga-PSMA I&T	69.5 (IQR 63–73)	2.7 (IQR: 1.125 - 3)	7a (6-9)				<u> </u>	
2024[36]	Netherla nds	р	20	Primary PCa	18F-PSMA PET/CT	DROP-IN gamma probe	1111n-PSMA-I& T	69 (Range 57–79)	22.2 (2.9–117)		66.7%	99.8%	96.8%	97.0 %	

In 2015, Maurer et al. first traced LNMs intraoperatively with a gamma probe in five patients receiving injections of 111In-PSMA-I&T 24 h before surgery (Table 3) [13]. Although the preliminary case report included only five patients, they successfully proved that PSMA-targeted RGS was feasible and safe [13]. Furthermore, Rauscher et al. performed <sup>111</sup>In-PSMA-I&T RGS in 31 patients with recurrent PCa, with a sensitivity of 92.3% and specificity of 93.5% [37] (Table 3). Recently, the DETECT Trial reported that 111In-PSMA-I&T facilitated the resection of PSMA-avid lesions, of which 97% were confirmed as LNMs in 20 newly diagnosed PCa patients [36]. In addition to PSMA-I&T, another theranostic ligand, <sup>111</sup>In-PSMA-617, was used for PET/CT imaging and PSMA-RGS in six primary PCa patients with potential LNMs [38] (Table 3). The sensitivity, specificity, PPV, NPV, and accuracy using cut-off SUVs normalized to lean body mass (SUL) were 92.1%, 98.9%, 94.6%, 98.4%, and 97.7%, respectively [38]. In these studies, PSMA-targeted RGS could not discriminate LNMs intraoperatively within the surgery, and a  $\gamma$ -probe was used to confirm LNMs by counts per second (count/s, CPS) after ex vivo removal of the tissue

samples [37, 38].

Using drop-in y probes (DIGPs) can overcome limited maneuverability traditional the of laparoscopic gamma probes to facilitate RGS. A DIGP was designed with a 135° grip for optimal performance during robotic-assisted radical prostatectomy (RARP), while a 45° grip DIGP has been shown to be more appropriate for *ex vivo* clinical setups [39]. In one study using a DIGP with nontargeted tracers (99mTc-nanocolloid + ICG), 10 high-risk patients with PCa underwent sentinel pelvic lymph node dissection (PLND) with no side effects [40]. In another study, with a median uptake time from injection to surgery of 5.4 h, and ex vivo examinations were also performed with a gamma probe and near-infrared fluorescence (NIRF) imaging [41]. All LNMs were identified using DIGP and 91.0% were identified by in vivo NIRF imaging [41]. Although sentinel PLND can detect micro-LNMs, its non-targeting nature may limit its further use, especially in patients with recurrent PCa whose lymphatic drainage has been altered by previous PLNDs. Recently, Maurer et al. performed a systematic comparative analysis between robotassisted and open salvage PSMA-RGS, proving the safety and efficiency of robot-assisted PSMA-RGS with a longer operating time and equal estimated blood loss [42]. Complete biochemical response (cBR, PSA < 0.2 ng/mL 2–16 weeks after PSMA-RGS) was 57% (34/61) in the open group and 61% (14/24) in the robotic group [42]. The same research group further studied the reasons for negative histopathology from PSMA-RGS in 302 patients with recurrent PCa, showing that false positive lesions on PSMA-PET in most cases had negative histopathology and that only a few true-positive lymph nodes were missed by surgery [43]. All true-positive lesions were adjacent to the internal iliac arteries or the prostatic fossa [43].

# 3.2 <sup>99m</sup>Tc-labeled PSMA-RGS

The production and wider applications of PSMA-RGS with 111In-PSMA-I&T still face some limitations, such as suboptimal nuclear properties, high cost, and limited availability of <sup>111</sup>InCl<sub>3</sub> [12]. To overcome the limitations of 111In-PSMA-I&T in PSMA-RGS, Robu et al. successfully used mercaptoacetyl triserine (MAS<sub>3</sub>) to replace the DOTAGA chelator in PSMA-I&T for 99mTc labeling and D-Tyr-D-2-Nal to replace the 3-iodo-D-Tyr-D-Phe sequence to obtain a stronger linker [12]. Compared to <sup>111</sup>In, <sup>99m</sup>Tc-labeled PSMA-I&T can better meet clinical needs as a more cost-effective and readily available agent. Therefore, 99mTc-labeled agents were also introduced into the PSMA-RGS. The biodistributions of <sup>111</sup>In labeled and <sup>99m</sup>Tc labeled agents are similar, but the absorbed radiation doses can vary greatly; the total body exposure of <sup>111</sup>In is 20-fold higher than that of <sup>99m</sup>Tc (0.1 mGy/MBq vs. 0.0047 mGy/MBq) [44]. Considering the total body exposure and availability of the two isotopes, technetium is the preferred choice.

Compared with <sup>111</sup>In, <sup>99m</sup>Tc-PSMA-RGS has been evaluated in more clinical trials, and its resolution is highly dependent on the γ-probe. The commonly used Crystal Probe CXS-SG603 has a resolution of 14 mm and is, therefore, optimally suited for identifying LNMs around 14 mm. The median diameter of correctly identified LNMs by <sup>99m</sup>Tc-PSMA-RGS is 12 mm (range: 3-25 mm) [43]. Compared with PSMA PET/CT, <sup>99m</sup>Tc-PSMA-I&S SPECT/CT can detect only 56.8% (25/44) lesions but <sup>99m</sup>Tc-PSMA-RGS can detect LNMs as small as 3 mm [45]. Compared with false positive LNMs in <sup>99m</sup>Tc-PSMA-RGS, true positive lesions had higher SUVmax on preoperative <sup>68</sup>Ga-PSMA-11 PET [8.9 (IQR 3.8-13.8) *vs.* 2.5 (IQR 2.2– 5)] [43].

In 31 patients with PCa, <sup>99m</sup>Tc-PSMA-RGS yielded a sensitivity of 83.6%, specificity of 100.0%, and accuracy of 93.0%. The minimum diameter of detected LNMs was 3 mm [45] (Table 3). <sup>99m</sup>Tc-

PSMA-RGS has also been introduced in robot-assisted PSMA-guided salvage PLND on the robotic da Vinci® platform [46]. After careful measurement of occupational radiation exposure in all personnel, 99mTc-PSMA-RGS was subsequently introduced in additional clinical trials [47]. During the PSMA-targeted surgery with 99mTc-PSMA-I&S, the measured effective dose ranged from 0 to 5 µSv, including all the personnel during labeling, quality control, syringe preparation, patient administration, patient imaging, and PSMA-targeted surgery. The mean effective doses for anesthesiology technician, scrub nurse, surgical nurse, and surgeon were  $0.6 \pm$  $0.5, 3.2 \pm 1.3, 0.2 \pm 0.4$ , and  $1.0 \pm 0.0 \mu$ Sv, respectively. The total effective dose would be 320 µSv/year if as many as 100 procedures were performed [47]; the maximum effective dose was safe for all personnel.

*In vivo* and *ex vivo* gamma probes (Crystal Probe CXS-SG603; Crystal Photonics, Berlin, Germany) were simultaneously used to remove all metastatic lesions depicted on <sup>68</sup>Ga-PSMA-11 PET [45]. The cutoff value to discriminate positive specimens from the background was 4 CPS [45] (Table 3). <sup>99m</sup>Tc-PSMA-I&S was also validated as a safe and effective imaging technique for SPECT/CT in four healthy volunteers and 10 patients with primary PCa for initial staging [48]. The injected activities of 562–828 MBq can be translated into effective doses of 3.33–4.42 mSv [48]. In an *ex situ* analysis, <sup>99m</sup>Tc-PSMA-I&S RGS discriminated LNMs with a sensitivity of 76.6%, specificity of 94.4%, PPV of 89.4%, and NPV of 86.9% [49] (Table 3).

In a phase 2 trial including 12 cN0cM0 primary PCa patients undergoing RARP with extended pelvic nodal dissection (ePLND), <sup>99m</sup>Tc-PSMA-I&S-RGS was used to intraoperatively identify LNMs [50] (Table 3). The per-patient sensitivity, specificity, PPV, and NPV of <sup>99m</sup>Tc-PSMA-I and S-RGS were 67.0%, 100.0%, 100.0%, and 90.0%, respectively [50]. The per-region sensitivity, specificity, PPV, and NPV of PSMA-RGS were 63.0%, 99.0%, 83.0%, and 96.0%, respectively [50].

The main challenges in <sup>99m</sup>Tc-PSMA-I&S RGS are micro-LNMs with diameters of <3 mm [50]. A similar study was conducted in Australia [51] (Table 3), wherein Gondoputro *et al.* evaluated the performance of <sup>99m</sup>Tc-PSMA-I&S in 12 patients with primary PCa who underwent RARP with ePLND [51]. The sensitivity, specificity, PPV, and NPV were 76.0%, 69.0%, 50.0%, and 88.0%, respectively. Additionally, >90% of missed LNMs were micrometastases with diameters ≤3 mm [51]. In 2022, a phase III clinical trial was performed using a DIGP with <sup>99m</sup>Tc-PSMA-I&S in robot-assisted PSMA-RGS, including 20 patients with recurrent PCa [52] (Figure 6). The sensitivity and specificity of robot-assisted PSMA-RGS for the detection of LNMs were 86.0% and 100.0 %, respectively [52]. In addition to PSMA-I&S, another PSMA-I&T-based analog named PSMA-I&F was also successfully conjugated to <sup>68</sup>Ga/<sup>177</sup>Lu for targeted imaging and therapy with high affinity [53].

In summary, <sup>99m</sup>Tc-PSMA-RGS has been translated into clinical use, facilitating better detection of positive LNMs than conventional ePLND in patients with PCa. The limitations of <sup>99m</sup>Tc-PSMA agents include its inability to detect micro-LNMs and lesions with low to intermediate levels of PSMA. Therefore, it is important to develop new strategies to improve the imaging accuracy for better PSMAtargeted surgery. Current strategies include a combination of near-infrared dyes and dual-target methods.

# 3.3 Considerations regarding PSMA-RGS

Regarding PSMA-RGS, several key areas need further investigation to optimize patient care and outcomes. The follow-up of PCa patients receiving PSMA-RGS needs to be standardized and include an evaluation of whether positive LNMs may be missed in PSMA-RGS and their impact on prognosis. Requirements and patient selection criteria need to be established for PSMA-targeted RGS candidates, to be verified before the injection of radiotracers. Lastly, it is also critical to assess the prescribed imaging strategy, including the first preoperative imaging (<sup>68</sup>Ga-PSMA-PET/CT) and the subsequent second preoperative imaging (PSMA SPECT/CT) after administering the radiotracer for PSMA-RGS.

To investigate the oncological outcomes of patients with salvage PLND via PSMA-RGS, Knipper et al. showed that, in 94% (343/364) of patients receiving PSMA-RGS, 48.1% (165/343) can reach a PSA level of <0.2 ng/mL at 2-16 weeks after PSMA-RGS, with a 2-year biochemical recurrence-free survival of 32.0% and therapy-free survival of 58.0% [54] (Table 3). Adjuvant systemic therapy should be added to further improve the overall survival in patients with recurrent PCa undergoing PSMA-RGS [55]. For PSMA-targeted RGS, both robot-assisted and open surgeries are appropriate for patients with recurrent PCa. Ambrosini et al. compared 24 patients (28%) undergoing robot-assisted RGS with 61 patients (72%) receiving open PSMA-RGS and found that they had comparable oncological outcomes with no safety concerns [42].

PSMA-PET imaging and SPECT/CT imaging should both be performed preoperatively with different purposes. PSMA-PET imaging before or after biopsy provides accurate staging, while SPECT/CT confirms the presence of PSMA-avid lesions and tracer uptake. PSMA-PET/CT can detect nearly double the number of LNMs compared with <sup>99m</sup>Tc-PSMA-I&S SPECT/CT during PSMA RGS preoperative imaging [45, 56] (Figure 1). As <sup>99m</sup>Tc-PSMA-I&S SPECT/CT clearly has an inferior resolution, it should only be considered for preoperative imaging when PSMA PET is unavailable [57].

As shown in Figure 1, patients with suspected PCa are recommended to undergo serum PSA testing and PSMA-PET before biopsy for PSMA-TB and accurate staging. PSMA SPECT/CT is performed after radiotracer injection to confirm specific uptake by PCa lesions. After 3 months, postoperative PSMA-PET is performed to evaluate the surgical outcome. The performance of PSMA-targeted surgery can be improved using similar approaches to those applied to PSMA-targeted theranostic agents, such as 177Lu-PSMA-617. Both PSMA-targeted surgery and radioligand therapy require a high tumor uptake and TBR, as well as a relatively long circulation half-life. For this purpose, albumin binders have been conjugated with PSMA-targeted radioligands to prolong blood retention time with minimal adverse effects [58]. Albumin, as one of most abundant proteins in human serum, exerts an important role as a carrier protein of hormones or fatty acids [58]. Its strengths, including nontoxicity, prolonged half-time, biodegradability and absence of immunogenicity enabled it as a promising candidate for theranostic agents [58]. The albumin binding Evans blue (EB) moiety has proven to be a safe and effective way to improve the pharmacokinetics of PSMA-617. Compared with 177Lu-PSMA-617, 177Lu-EB-PSMA-617 accumulates significantly more in PSMA+ PCa tumors, reduces the needed dose of radioactivity [59, 60], is safe, and shows improved pharmacokinetics and therapeutic efficacy in 76 patients with progressive metastatic castration-resistant prostate cancer (mCRPC) [60-63]. Another EB modified PSMA radioligand 177Lu-LNC1003 can further improve therapeutic efficacy by greatly enhancing uptake and retention, even in PCa with moderate level of PSMA expression [64, 65]. Another series of dual PSMA and albumin binding ligands were also reported, such as PSMA+albumin (RPS-025, RPS-063, RPS-067, RPS-072) [66-68]. For example, the dual-targeted dye RPS-027 showed higher tumor uptake and increased tumor-to-tissue distribution than MIP-1095 in PSMA-targeted a-therapy [66]. Then, a series of ligands (RPS-061, RPS-063, RPS-066, RPS-067, RPS-068, and RPS-069) based RPS-027 was developed for higher tumor uptake and lower kidney uptake, but just in its preclinical models [67]. Among all albumin-binding agents, LNC1003 has just finished its Phase 1 trial and showed best potential in clinical translation.

Table 4	PSMA-FGS	in PCa	patients.
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Year	Stage	Country	Institution	Excitation/emission	NCT#	Agents	Patient number	Patient Type	FGS type
2023[15]	Phase 2a	Netherlands	Leiden University Medical Center, Leiden University, Leiden	774/— nm	European Trial Database, 2019-002393-31;	OTL78	18	Primary PCa with ISUP≥7	RP±ePLND (da Vinci surgical system)
2023[16]	Phase I	USA	University of California, San Francisco	774/793 nm	NCT04574401	IS-002	24	High-risk primary prostate cancer	RP+ePLND (da Vinci surgical system)

In addition to <sup>111</sup>In and <sup>99m</sup>Tc, <sup>67</sup>Ga with three low gamma emission energy levels at 93, 185, and 300 keV has also been introduced to synthesize <sup>67</sup>Ga-PSMA I&T, with the aim of overcoming the potential disadvantages of currently used radionuclides such as <sup>111</sup>In, <sup>99m</sup>Tc, and <sup>68</sup>Ga [69] (Table 3). With a longer half-life, <sup>67</sup>Ga-PSMA RGS can be performed 1 day after injection of <sup>67</sup>Ga-PSMA I&T, with the option of a further delay of 1 or 2 more days [69].

To improve the diagnostic efficiency of PSMAtargeted agents, a series of glutamate-urea-glutamate or glutamate-urea-lysine pharmacophores incorporating second-generation single amino acid chelators were developed for 99mTc-labeled PSMA SPECT imaging, such as MIP-1404, MIP-1405, MIP 1428, and MIP-1427 [70, 71]. Among them, MIP-1404 achieved the best binding affinity by PCa tumors  $(1.07 \pm 0.89)$ nM) with rapid clearance from the kidneys and non-tumor tissues [70]. Compared with MIP-1405, MIP-1404 showed high-contrast imaging and lower urinary activity (7% vs. 26%) in six healthy men and six patients with metastatic PCa [72]. 99mTc-MIP-1404 has shown good accuracy in the detection of PCa lesions and LNMs; however, it has not been used in PSMA-RGS. 99mTc-MIP-1404 was first tested in four clinical trials involving 39 patients [73]. In a multicenter phase 2 clinical trial including 105 intermediate- to high-risk PCa patients, 99mTc-MIP-1404 detected 93.3% (98/105) of primary PCa patients, with a sensitivity of 50.0% and a specificity of 87.0% for the detection of LNMs [74]. Similar results were reported in another study in 2018, wherein Schmidkonz et al. performed <sup>99m</sup>Tc-MIP-1404 SPECT/CT in 93 patients with pathologically confirmed primary PCa, and detected 96.7% (90/93) of LNMs [75]. TU<sub>prostate</sub> was significantly higher in patients with a GS > 7 or PSA > 10 ng/ml [75]. In 60 biochemically recurrent PCa patients, 99mTc-MIP-1404 SPECT/CT could detect 91.4% of PCa patients with  $PSA \ge 2 \text{ ng/ml}$  and 40.0% of PCa patients with PSA <2 ng/ml [73]. In a larger study including 225 PCa patients with biochemical recurrence, detection rates were 90.0% in patients with PSA  $\geq$  2 ng/mL and 54.0% in patients with PSA < 2 ng/mL, leading to treatment management in 74.0% of the patients [76]. In biochemically recurrent PCa patients with PSA 0.2-0.5 ng/mL, the detection rate of 99mTc-MIP-1404 SPECT/CT was 44.0% [77].

Although 99mTc-MIP-1404 has not been used in PSMA RGS, it has potential to become the next <sup>99m</sup>Tc-labeled agent for PSMA RGS. Compared with other SPECT/CT tracers, 99mTc-MIP-1404 has better imaging performance, achieving similar а lesion-to-liver ratio to 68Ga-PSMA-11 PET in patients with mCRPC (median, 18.2 vs. 17.3; P = 0.93) [78]. With favorable radiation dosimetry, compared to 68Ga-PSMA-11 (0.0088 mSv/MBq vs. 0.022 mSv/MBq), 99mTc-MIP-1404 has shown imrpoved efficacy in patients with intermediate-to-high-risk PCa [74, 78]. 99mTc-MIP-1404 could be used as a secondary alternative when PSMA-PET/CT is unavailable.

PSMA PET/CT has also been used for intraoperative imaging [79]. Intraoperative PET/CT imaging (AURA10; XEOS Medical, Gent, Belgium) was first used to detect specimens from RP and lymphadenectomy ex vivo in a prospective two-center feasibility study [80, 81]. This technique can provide a quicker method to detect PSMs in resected specimens than frozen section analysis; however, this type of imaging also has obvious shortcomings. The ex vivo specimen PET/CT imaging cannot determine whether all positive lesions (prostatic lesions and LNMs) have been totally removed by surgery because no postoperative PSMA PET/CT is performed, whether the estimated dose per procedure is safe for personnel, and whether the ex vivo imaging can correctly match the final pathological results of the whole prostate. Therefore, we believe that ex vivo PET/CT imaging of specimens should not be the preferred method for PSMA-RGS.

# 4. PSMA-FGS

# 4.1 PSMA-FGS in clinical stage

Compared to PSMA-RGS, PSMA-FGS is a more promising and attractive strategy for intraoperative navigation because it is more intuitive. It is dynamically fitted for the da Vinci surgical system, equipped with a fluorescent imaging system. Currently, OTL78 and IS-002 are two agents for PSMA-FGS that have entered clinical trials [14-16] (Table 4). The median size of correctly detected LNMs by PSMA-FGS-using OLT 78 was 1.9 mm (IQR 1.8-2.8), much smaller than that of PSMA-RGS. The exact resolution of IS-002 is yet to be determined. The sensitivity of *ex vivo* imaging with OLT78 was higher than its *in vivo* imaging, because the limited range of motion of a laparoscope and limited range of fluorescence may cause difficulties in the in vivo visualization of the apex of the prostate, which is located in the deep region of pelvis [15]. Compared with the laparoscope, robotic surgery systems may greatly facilitate deep pelvic visualization and can be better integrated into a robot-assisted fluorescenceguided surgery workflow.

In a single-arm, phase 2a, feasibility trial, Stibbe et al. showed that OTL78 was well tolerated and had the potential to achieve complete oncological resection of PSMs and positive LNMs, with the optimal dose of 30 µg/kg at 24 h preoperatively (Figures 4-5) [15]. Similarly, Nguyen et al. conducted a phase 1, single-center, dose-escalation study to evaluate another near-infrared agent, IS-002, in 24 men with high-risk PCa by RP+ePLND [16]. The PSMA-FGS with IS-002 achieved a PPV of 97% and NPV of 45% for LNMs, and PPV of 100% and NPV of 80% for locoregional/residual disease detection with an injection dose of  $25 \,\mu g/kg$  (Figures 4–5) [16]. These studies using OTL78 or IS-002 demonstrate the excellent performance of PSMA-FGS in the detection of PCa lesions and LNMs, as clinicians prefer real-time imaging and radiation-free agents.

# 4.2 PSMA FGS in the preclinical stage

In addition to OTL78 and IS-002, other agents for PSMA FGS are still in their respective preclinical stages (Figure 2L-W). Based on the binding motif, PSMA-targeted agents in FGS can be divided into antibody-conjugated NIRF dyes, small-moleculeconjugated NIRF dyes, and activable NIRF dyes.

# 4.2.1 Antibody-conjugated NIRF agents

In 2011, a humanized anti-PSMA antibody, huJ591, was conjugated with indocyanine green (ICG); however, huJ591-ICG was guenched until uptake by PSMA<sup>+</sup> PCa tumors [82] (Table 5). The quenching ability of huJ591-ICG was mainly due to the unique property of ICG, which loses its fluorescence when bound to the antibody, but regains fluorescence when it is separated from the antibody after internalization and degradation of the ICG conjugate in the lysosome [82]. Despite the advantages of ICG, such as documented safety and deep tissue penetration, the fluorescence quenching could hinder the visibility and identification of target areas. To maintain fluorescence and bypass the quenching ability of ICG conjugates, a series of IRDyes were developed and conjugated with huJ591. The IRDye800CW is a hydrophilic dye that can be easily synthesized and maintained in vitro and in vivo.

IRDye800CW was introduced to replace ICG to bind huJ591 as ProsaFluor (huJ591-ICG-IRDye800CW) [83] (Table 5). Unlike IRDye800CW, IRDye700DX is another dye that could provide simultaneous NIRF imaging and NIR-infrared photoimmunotherapy, where IRDye700DX, upon activation by NIR light of a wavelength that correlates to the absorption spectrum of the agent, produces reactive oxygen species (ROS) and thus induces localized ROS-mediated cell death in targeted cells. IRDye700DX is a hydrophilic dye that differentiates it from previous hydrophobic dyes, associated advantages including with easier distribution throughout the body without the need for complex solubilizing agents, no aggregation in aqueous solutions leading to less non-specific binding to various tissues, facilitated conjugation with targeting molecules, and therefore overall better biocompatibility. Both IRDye800CW and IRDye700DX have been approved by the US Food and Drug Administration (FDA) to start clinical trials, indicating their safety for use in FGS.

Nagaya et al. incubated a fully human anti-PSMA antibody with IRDye700DX to develop anti-PSMA-IR700, showing the potential of PSMAtargeted FGS and near-infrared photoimmunotherapy (NIR-PIT) [84] (Table 5). Chen et al. synthesized a PSMA-targeted theranostic photosensitizer, YC-9, by conjugating IRDye700DX to a PSMA-targeted Lys-Glu urea-based molecule for NIR-PIT [85]. Furthermore, they developed a long-circulating PSMA-targeted bacteriochlorophyll-based photosensitizer (BPP) for NIRF imaging within surgery and NIR-PIT [86]. To further improve its clinical use, another fully human anti-PSMA antibody, named PSMAb, based on a PSMA-specific single-chain variable fragment (scFv) named gy1 [87, 88] (Table 5), was used. Both PSMAb and gv1 have been labeled with IRDye800CW to illuminate PSMA+ PCa tumors [87, 88]. ScFvD2B, another high-affinity anti-PSMA scFv, was conjugated to X770 (an NRIF probe) [89] (Table 5) with IRDye700DX, DTPA, and <sup>111</sup>In to develop the multi-modality agent, DTPA-D2B-IRDye700DX [90] (Table 5).

Antibodies or antibody fragment conjugates can be used for the optical imaging of PSMA-positive tumors. The advantages of antibody conjugates include a long circulation half-life, providing a long time window for FGS, and reliable safety because most of the antibodies have been approved by the FDA for treatment in clinical trials. However, due to the nonspecific uptake of antibodies by the liver and spleen, only a small portion of the conjugates are available to bind the PSMA-positive tumor. Therefore, for PSMA-FGS, a focus has instead been placed on small-molecule conjugated NIRF agents.

Year	Country	Institution	Type of ligands	Agents	Aim	In-vivo experiment
2011[82]	USA	National Cancer Institute	Humanized Antibody	ICG-huJ591	FGS	PC3±PSMA
2016[83]	USA	Johns Hopkins University School of Medicine	Humanized Antibody	J591-IR800	FGS	PC3±PSMA; LMD±PSMA
2017[84]	USA	National Cancer Institute	Fully human antibody	Anti-PSMA-IR700	FGS/PI T	PC3±PSMA
2019[87]	China	Fourth Military Medical University	Fully human antibody	PSMAb-IRDye800CW	FGS	PC3±PSMA
2016[88]	China	Fourth Military Medical University	scFv	gy1-IRDye800CW	FGS	PC3±PSMA
2016[89]	Italy	Verona University	scFv	X770-scFvD2B	FGS	LNCaP
2019[90]	Netherlan ds	Radboud university medical center	scFv	111In-DTPA-D2B-IRDye700 DX	FGS/PI T	LS174T±PSMA
2010[91]	USA	Washington State University	PSMA inhibitor CTT-54.2	Cy5.5-CTT-54.2	FGS	LNCaP; PC3
2009[92]	USA	Johns Hopkins Medical Institutions	PSMA inhibitor YC-273	YC-27 3-IRDye800CW	FGS	PC3±PSMA
2014[93]	USA	Case Western Reserve University	PSMA inhibitor PSMA-1	PSMA-1-IRDye800CW; PSMA-1-Cy5.5	FGS	PC3±PSMA
2017[85]	USA	Johns Hopkins Medical Institutions	IRDye700DX	YC-9	FGS	PC3±PSMA
2020[86]	Canada	University Health Network	PSMA inhibitor	BPP	FGS	PC3±PSMA
2019[14]	USA	On Target Laboratories	PSMA inhibitor	OTL78	FGS	LNCaP;22Rv1; PC3; A549
2017[94]	Japan	Yoshida Shimoadachi-cho	PSMA inhibitor	800CW-SCE	FGS	LNCaP; PC3
2017[95]	USA	Massachusetts General Hospital and Harvard Medical School	PSMA inhibitor KUE	ZW800-1	FGS	LNCaP; PC3
2018[112 ]	Canada	University Health Network	PSMA inhibitor	64Cu-LC-Pyro	PET/C T FGS	PC3±PSMA
2022[96]	USA	Johns Hopkins Medical Institutions	PSMA inhibitor	DyLight800-10	FGS	PC3±PSMA
2022[99]	Germany	German Cancer Research Center (DKFZ)	PSMA inhibitor PSMA-617	PSMA-927	FGS	LNCaP; PC3
2022[102 ]	Netherlan ds	Radboud University Medical Center	PSMA inhibitor PSMA-N064	PSMA-N064inc	FGS/PI T	LS174T±PSMA
2022[103 ]	Netherlan ds	Leiden University Medical Center	PSMA inhibitor EuK	99mTc-EuK-(SO3)Cy5-mas3	FGS	Porcine model (15 non-tumor-bearing pigs)
2023[105 ]	China	Peking Union Medical College and Chinese Academy of Medical Sciences	PSMA inhibitor KUE	Cy7	FGS	C4-2;22RV1; PC3
2022[100 ]	UK	King's College London	PSMA inhibitor urea-based Glu-Lys	Cy7.5	FGS	LNCaP; PC3; DU145; DU145
2023[97]	China	Northwestern Polytechnical University	PSMA inhibitor Lys-urea-Glu	iridium(III) complex	FGS	LNCaP; 22Rv1; PC3
2022[101 ]	Italy	University of Torino	PSMA-617	IRDye700DX	FGS	LNCaP; PC3
2022[104 ]	Japan	The University of Tokyo	Activatable	5GluAF-2MeTG	FGS	LNCaP; PC3

#### 4.2.2 Small molecule-conjugated NIRF agents

Small-molecule inhibitors include CTT-54.2, YC273, PSMA-1, SCE, KUE, and OTL78 [14, 91-95] (Table 5). Cy5.5-CTT-54.2 displayed high targeting potency for PSMA, but in vivo studies are lacking [91]. YC-273-IRDye800CW, Similarly, PSMA-1-IRDye800CW, PSMA-1-Cy5.5, and SCE-IRDye800CW have been established and preliminarily validated in PSMA<sup>+</sup> PCa tumor-bearing mice [92-94]. In addition to polymethine fluorophores, Bao et al. reported a zwitterionic (ZW) fluorophore conjugated directly with a urea-containing PSMA ligand, lysine-ureaglutamate (KUE), ZW800-1, for the NIRF imaging of PSMA<sup>+</sup> PCa tumors [95]. Kularatne et al. designed and synthesized OTL78, which has a circulation half-life of 17 min and excellent TBR (5:1) for PSMA-targeted NIRF surgery [14]. Chen et al. reported that PSMA-targeted NIRF dyes for PSMA FGS and DyLight800-10 achieved the highest uptake in PCa tumors [96] (Table 5). Furthermore, Yang et al. conjugated an iridium(III) complex with the inhibitor Lys-urea-Glu to create a PSMA-targeted iridium(III) probe [97].

<sup>68</sup>Ga-PSMA PET/CT and Preoperative ICG-guided PLND were combined in early studies, achieving a sensitivity of 92.8% and specificity of 39.1% [98]. However, due to ICG by itself not being tumors-specific, the specificity of PLND was low. To develop PSMA-targeted NIRF dyes for guided surgery, several studies have attempted to develop PSMA-targeted ligands using PSMA-PET/CT. Eder et al. also developed a PSMA-targeted NIRF dye, Glu-urea-Lys-2-Nal-Chx-Lys (IRDye800CW)-DOTA (PSMA-927), based on the theranostic agent PSMA-617 [99] (Table 5). Moreover, PSMA-Cy7.5-2 outperformed PSMA-IRDye800CW in vitro and in vivo [100].

To evaluate the performance of the PSMAtargeted agents, PSMA-negative PC3 cells were transfected with a PSMA-expressing lentivirus to obtain PC3-PSMA<sup>+</sup> cells, which were used to evaluate the efficacy of PSMA-targeted NIRF dyes. Similarly, IRDye700DX was also conjugated to PSMA-617 as IRDye700DX-PSMA for preliminary validation in PC3-PSMA<sup>+</sup> tumor mice [101]. Derks *et al.* modified the PSMA peptide linker from glutamic acid to a lvsine residue and synthesized three dualmodality DOTA(GA)-IRDye700DX-PSMA ligands (PSMA0N01, PSMA-N02, and PSMA-N03) based on the existing PSMA backbone structures (PSMA-1007/PSMA-617) [102] (Table 5). Dell'Oglio et al. developed and validated the performance of intraoperative PSMA-mediated FGS using EuK-(SO<sub>3</sub>)Cy5mas<sub>3</sub> in a porcine model, demonstrating its clinical translational potential [103] (Table 5). However, while the porcine model enables the study of robot-assisted radical RP within a large animal model, unlike humans or mice, it does not have PSMA-positive tumors; therefore, targeted agents still need further evaluation in mouse models and clinical trials.

# 4.2.3 Activatable NIRF dyes

PSMA also has glutamate carboxypeptidase (CP) activity. Activatable dyes, namely "turn on/turn off" agents, can highlight tumors with a lower background (Figure 3A-B). Kawatani et al. developed 5GluAF-2MeTG as the first in-class activatable NIRF dye for FGS patients with PCa [104](Table 5). 5GluAF-2MeTG was designed based on previous findings that aryl glutamate conjugates with an azoformyl linker can be recognized by PSMA and have a sufficiently low lowest unoccupied molecular orbital energy level to quench the dve via photoinduced electron transfer [104]. After incubation with recombinant human PSMA, 5GluAF-Fl showed significantly increased (>200 fold) fluorescence, which was mainly converted to highly fluorescent fluorescein [104]. Xing et al. developed a self-quenching NIRF dye, Cy-KUE-OA, with dual PCa-membrane affinity. The targeted dye was anchored to the phospholipids of the cell membrane of PCa cells, dequenched, and showed strong Cy7 fluorescent light. In addition to in vitro and in vivo imaging in PSMA+ PCa tumor-bearing mice, the study further confirmed the performance of Cy-KUE-OA on surgically resected specimens of PCa tissues, LNMs, and healthy tissues from patients [105] (Table 5). Compared to previous preclinical studies, the validation of resected PCa specimens in this study was solid for clinical translation, providing a new method to validate the performance of dyes. The resected specimen validation in this study is an excellent method to validate the performance of dyes, bridging the gap between preclinical studies and clinical trials.

# 5. Multi-modality/multi-targeted PSMA-targeted surgery

With regard to the future development of PSMA-guided surgery, fluorescent and radioactive tracers are complementary and used in combination

with multi-modality-guided surgery [106]. Because PSMA-RGS and PSMA-FGS have their own strengths and shortcomings, multi-modality PSMA-targeted dyes with radiotracers and fluorescent dyes in a hybrid tracer were simultaneously developed for PSMA-RGS and PSMA-FGS (Figure 3 C-F). Integrating radioactive and fluorescent signatures into a single dye can enable surgeons to improve their intraoperative performance. The PSMA-11-based multi-modality agent 68Ga-Glu-urea-Lys-HBED-CC-IRDye800CW (68Ga-PSMA-11-IRDye800CW) also achieved accurate intraoperative detection of PSMA+ PCa tumors in tumor-bearing mice and healthy pigs [107]. To overcome the nonspecific accumulation of PSMA-11 in non-malignant tissues, a histidine- (H) and glutamic acid (E)-containing linker ((HE)<sub>3</sub>-linker) was introduced between the chelator and the PSMA-binding motif [108]. Compared to 68Ga-PSMA-11-IRDye800CW, 68Ga-Glu-urea-Lys-(HE)3-HBED-CC-IRDye800CW (68Ga-PSMA-914) can be used for preoperative PET imaging and PSMA-guided PARP with faster excretion and a lower background signal [109].

Another agent, NYM016, was designed using the NIRF dye Cy7 and NOTA chelator for <sup>68</sup>Ga labeling. <sup>68</sup>Ga-NYM016 was evaluated in a patient with recurrent PCa with metastatic lesions, showing an SUV<sub>max</sub> of 18.93 in recurrent lesions [110]. Furthermore, Aras *et al.* synthesized a multimodality agent, <sup>18</sup>F-BF3-Cy3-ACUPA, and determined its safety and feasibility in 10 PCa patients by preoperative PET and intraoperative FGS [111]. Harmatys *et al.* designed <sup>64</sup>Cu-LC-Pyro, a low molecular weight PSMA-targeted agent with long plasma circulation time and high tumor-to-background ratio (10:1) for PET/CT, NIRF imaging and NIR-PIT at the same time [112].

Cordonnier et al. designed an oleic acid-coated spherical NaYF4:Yb,Tm@NaYF4 core/shell upconversion nanoparticle (UCNP) labeled with <sup>125</sup>I for PSMA-targeted FGS/RGS in LNCaP-Luc-bearing mice [113]. Cheng et al. developed a theranostic nanotexaphyrin for PSMA targeted SPECT/CT imaging and PIT through the chelation of metal isotopes (111In/175Lu), achieving a TBR of 2.7 [114]. Although the multi-modality method combines the strengths of PSMA-RGS and PSMA-FGS, challenges have also been exemplified. For instance, the diagnostic standard for PCa lesions and LNMs in multi-modality PSMA-targeted surgery, decisions regarding contradictory results of PSMA-RGS and PSMA-FGS, and the hybrid modality weakening the performance of each imaging modality remain to be elucidated. Radiation and contradictory results from multi-modality imaging may limit its further development and clinical translation.





# Preoperative



Intraoperative



**Postoperative** 



Figure 4. Robot-assisted <sup>99m</sup>Tc-PSMA radio-guided surgery procedure. Preoperative (A) CT, (B) PSMA PET/CT, and (C) <sup>99m</sup>Tc-PSMA-l&S SPECT/CT imaging demonstrate a prevesical lesion (indicated with an arrow) in a patient with biochemical recurrence after primary treatment for prostate cancer. (D) During the surgery, the DROP-IN probe is inserted in the abdomen through a trocar (E) and is autonomously maneuvered by the surgeon using a Da Vinci surgical console. (F, G) *In vivo* radioactivity measurement of a suspected prostate cancer recurrence. (H) *Ex vivo* radioactivity measurement of the resected specimen. Histological analysis revealed a lymph node metastasis on (I) hematoxylin and eosin staining and (J) PSMA immunohistochemistry. CT = computed tomography; PET = positron emission tomography; PSMa = prostate-specific membrane antiger; SPECT = single-photon emission CT. The figures were adapted with permission from [52], copyright 2022 European Urology © Elsevier, Inc.



Figure 5. Fluorescence workflow for surgical margin in apical prostate resection of OTL78(A-C) and IS-002 (D-F). (A) The entire prostate is imaged directly after resection *ex vivo* on the backtable. Increased fluorescence signal is visible at the apex [15]. (B) Subsequent assessment of the resection bed shows residual fluorescence signal (indicated with an arrow) where the apex used to be [15]. (C) A fluorescent, PSMA-expressing positive surgical margin is seen on *ex vivo* gross macroscopy [15]. (D) White light and sensitive Firefly image of the excised prostate 24 h after IS-002 injection (100 lg/kg) showing a strong, focal fluorescent signal in the primary tumor [16]. (E) White light and sensitive Firefly images of an area where residual disease was identified in the anterior apical prostate resection bed near the urethra. Sensitive Firefly pinpointed (Fig. 2E) highly specific IS-002 fluorescent signal in the resection ded not deemed suspicious in white light endoscopy that was subsequently biopsied [16]. (F) White light and sensitive Firefly images after biopsy. Insert in Figure IF shows a strong fluorescent signal or transected focal tumor extension. Gold arrows indicate biopsy location; gold stars mark transected tumor extension on prostate surface in Figures 2D and 2F. Dotted white lines (Fig. 2E) indicate instrument and prostate boundaries [16]. The figures(A-C) were adapted with permission from [15, 52], copyright 2022 Lancet Oncology © Elsevier, Inc. The figures (D-F) were adapted with permission from [16], copyright 2023 Eur Urol Oncol © Elsevier, Inc.



Figure 6. Fluorescent imaging of a lymph node metastasis in PSMA-FGS of OTL78(A-B) and IS-002 (C-D). (A) In-vivo bright-field, near-infrared and fluorescence color overlay imaging of a lymph node metastasis within a lymph node cluster using the VisionSense imaging system [15]. (B) Corresponding haematoxylin and eosin tumor delineation, PSMA immunohistochemistry, and fluorescence imaging show a PSMA-positive metastatic lymph node with OTL78 uptake (OTL78 30 µg/kg). (C) White light endoscopy and sensitive Firefly image of tumor, positive lymph node for patient 2 (IS-002 25 µg/kg) [16]. (D) Corresponding tumor-negative contralateral lymph node packages for patient 2 (IS-002 25 µg/kg) [16]. The figures(A-B) were adapted with permission from [15, 52], copyright 2022 Lancet Oncology © Elsevier, Inc. The figures (C-D) were adapted with permission from [16], copyright 2023 Eur Urol Oncol © Elsevier, Inc.



It is well known that PSMA expression is heterogeneous and can vary between patients, lesions in the same patient, or even within the same lesion. To better recognize all LNMs during PSMA RGS, multi-targeting methods have also been attempted (Figure 3G-K). Integrin  $\alpha v\beta 3$  is a reasonable co-target with PSMA for neo-vasculature which incorporates the Arg-Gly-Asp (RGD) binding motif. EUKLcRGDfK-IRDye800 could be used to illuminate either the PSMA<sup>+</sup> or integrin  $\alpha \nu \beta 3^+$  xenografts in mice [115]. Dual-targeting agents with PSMA-targeting ability and complementary molecules have also been introduced [116]. PSMA and gastrin-releasing peptide receptor (GRPR) are complementarily expressed in PCa tumors, as has been validated in prostatectomy samples and patients [117-119]. For targeting GRPR, RM-26 (GRPR antagonist) and its dual-targeting radiotracers (RGD-RM26-03, denoted as LNC1015) have showed favorable pharmacokinetics and high clinical feasibility for PET/CT imaging of cancer [120-124]. Similarly, Schollhammer *et al.* also retrospectively studied 20 frozen samples with various GS group using <sup>111</sup>In-PSMA-617 and GRPR <sup>111</sup>In-RM2[117]. They antagonist found that <sup>111</sup>In-PSMA-617 had a higher uptake in the higher GS group, whereas <sup>111</sup>In-RM2 had higher binding in the lower GS group [117]. The dual-tracer method was also validated in recent clinical studies. Qiu et al. also showed that dual-tracer (NOTA-P2-RM26 and PSMA-617) PET/CT-targeted biopsy (TB) achieved a

higher PCa detection rate (69.77%) than systematic biopsy SB (29.29%) or mpMRI-TB (36.14%) [125]. GRPR/PSMA TB is better than GRPR TB or PSMA TB alone: the csPCa omission diagnostic rate of dual-tracer PET/CT-TB (15.38%) was significantly lower than 68Ga-GRPR PET/CT-TB (48.72%) or 68Ga-PSMA PET/CT-TB (28.21%) [125]. Recently, a PSMA/GRPR-targeting radioligand named 68Ga-BQ7812 was also developed but just in preclinical stage [126]. This compound holds promise for increasing the chance of specific tumor binding, thereby improving detection accuracy, especially in patients with heterogeneous expression of either target. Fibroblast activation protein alpha (FAP) is a transmembrane protease highly expressed in the tumor microenvironment of PCa [127]. Boinapally et al. developed small-molecule PSMA- and FAPtargeting moieties with 64Cu, named 64Cu-FP-L1 and <sup>64</sup>Cu-FP-L2, respectively [116], with the potential to be used for PSMA-FAP dual-targeted guided PCa surgery. These PSMA/FAP dual-targeted radioligands enable imaging of lesions expressing FAP, PSMA, or both on the tumor cell surface or within the tumor microenvironment. These multi-targeted dyes were developed for patients with varying levels of PSMA expression in PCa tumors, including those with high, low, or no PSMA expression. While as nearly all treatment-naïve patients receiving PSMA-RGS had PSMA-positive PCa tumors expressing 10-100-fold more PSMA than surrounding benign

prostatic tissues [5] and loss of PSMA expression is commonly observed in patients with CRPC who do not usually undergo further surgery, a well-performing PSMA single-targeted dye may suffice for most clinical needs in PSMA-RGS or PSMA-FGS, multi-targeted conjugates offer promise for patients with PCa tumors displaying low or reduced PSMA expression. These conjugates provide additional advantages for managing heterogeneous tumors.

Thus, we believe that establishment and clinical validation of a well-performing PSMA-targeted dye (such as LNC1003) with reliable safety and a long operation time window can meet the greatest clinical need for PSMA-targeted surgery. For a better understanding of the development of PSMA-targeted surgery, a chronological timeline highlighting major events and significant agents for PSMA-targeted guided surgery was shown in Figure 7.

# 6. Conclusions

PSMA-targeted surgery can lead to significant changes in traditional surgical procedures for PCa because of its high sensitivity in the detection of PSMs and micro-LNMs compared to any preoperative imaging or visualization. PSMA-targeted surgery may benefit PCa patients with PSMA-TB because of a more accurate pathological diagnosis by PSMA-RGS or PSMA-FGS for a more accurate resection of all prostatic and metastatic lesions. PSMA-TB, PSMA-RGS, and PSMA-FGS should be used as overall strategies for patients with primary csPCa or recurrent PCa, thereby greatly facilitating clinical diagnosis and surgical treatment. Among the above strategies, PSMA-RGS has entered clinical use in many trials, but we believe that PSMA-FGS will be a developing trend in the near future, as it is radiation-free and intuitively used in real-time within surgery. The multi-targeted method may be more suitable for CRPC radiotherapy than PSMA-targeted surgery.

Multi-modality methods have strengths, but the shortcomings of both agents and contradictory results may limit their use. Importantly, the close correlation among preoperative imaging, PSMA-TB, PSMA-FGS, and follow-up evaluations after surgery requires further investigation. However, a well-performing clinical standard to select the most suitable candidates for PSMA-RGS and PSMA-FGS is still lacking. More attention should be paid to potential PSMA-FGS dyes, and their efficacy should be validated in more clinical trials in the near future. With the rapid development of PSMA-targeted dyes, PSMA-targeted surgery, and its related standard flow would completely change conventional surgical methods and improve the level of precision surgery for all patients with PCa.

# Abbreviations

RP: Radical prostatectomy;

PLND: pelvic lymph node dissection;

PCa: prostate cancer;

PSMA PET: Prostate specific membrane antigen positron emission tomography;

PSMA: prostate-specific membrane antigen;

PSMs: positive surgical margins;

LNMs: lymph node metastases;

PSMA-TB: PSMA PET guided prostatic biopsy;

PSMA-RGS: PSMA targeted radio-guided surgery;

PSMA-FGS: PSMA targeted fluorescence-guided surgery;

FOLH1: Folate hydrolase 1;

GCP II: Glutamate carboxypeptidase II;

NP: normal prostate;

111In-PSMA-I&T: 111Indium-PSMA imaging and therapy;

99mTc-PSMA- I&S: 99mTechnetium-PSMA imaging and surgery;

csPCa: clinically significant prostate cancer;

mpMRI: multiparametric magnetic resonance imaging;

PB: prostate biopsy;

RARP: robot-assisted radical prostatectomy;

PSA: prostate-specific antigen;

PPV: positive predictive value;

NPV: negative predictive value;

OT: operating time;

EBL: equal estimated blood loss;

cBR: Complete biochemical response;

MAS3: mercaptoacetyl triserine;

ePLND: extended pelvic nodal dissection;

TFS: therapy-free;

SPECT/CT: Single-Photo Emission Computed

Tomography/Computer Tomography;

CP: carboxypeptidase;

GRPR: gastrin-releasing peptide receptor.

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#### Ethical approval

This article does not contain any studies with human participants or animals performed by any of the authors.

# **Competing Interests**

The authors have declared that no competing interest exists.

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