

1 **Supplementary Data**

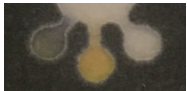

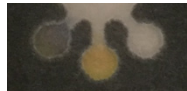
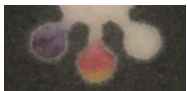

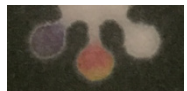






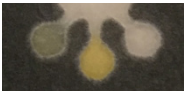



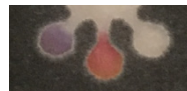










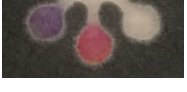








2 Table S1. Viscosity Measurement

% SMC	Shear rate,1/s	True Visc, mPa-s			Mean	SD
0.25%	7000	1.53	1.538	1.541	1.54	0.0035
0.50%	6000	1.871	1.866	1.872	1.87	0.0032
0.75%	4000	2.174	2.181	2.178	2.18	0.0035
1.00%	4000	2.721	2.734	2.721	2.73	0.0075
1.25%	3000	4.181	4.169	4.165	4.17	0.0083
1.50%	2000	5.036	5.199	5.054	5.10	0.0894

3 SMC: sodium carboxymethylcellulose, Visc: Viscosity

4
5
6
7
8
9
10
11
12
13
14
15
16
17

1 Table S2. The pH color change on proposed PADs

No.	%SCMC	Color change on PADs			pH on PADs	pH meter
		1	2	3		
1	0.25				6.6-6.8	6.401 ± 0.018
2	0.25				8.0	8.003 ± 0.002
3	0.50				7.2-7.4	7.284 ± 0.037
4	0.50				6.8-7.0	6.858 ± 0.002
5	0.75				5.8	5.700 ± 0.060
6	0.75				7.8 – 8.0	7.826 ± 0.045
7	1.00				7.8 – 8.0	8.252 ± 0.001
8	1.00				7.0	7.199 ± 0.051
9	1.25				7.0	7.107 ± 0.019
10	1.25				8.5	8.908 ± 0.049
11	1.50				6.6	6.552 ± 0.021
12	1.50				7.8	7.896 ± 0.010

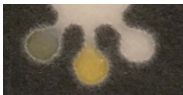

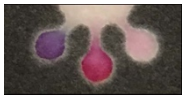


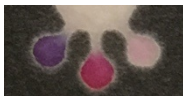
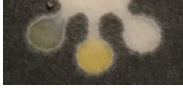
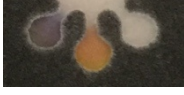
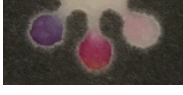

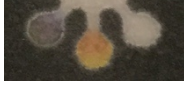
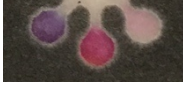

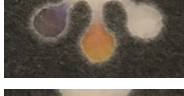
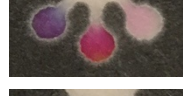
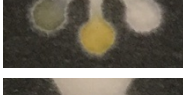
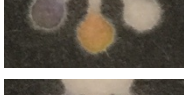
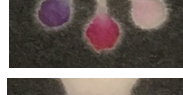










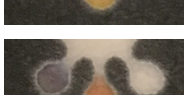
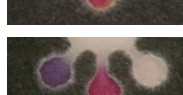
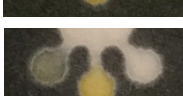



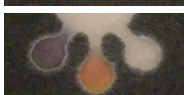
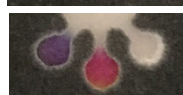

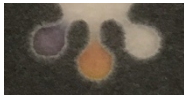

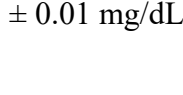


2

3 SCMC: sodium carboxymethylcellulose, PADs: paper-based analytical devices

4

5

1 Table S3. Reagent stability

Days	Spiked concentration (mg/dL)	Condition	PADs (mg/dL) (n=6)	pH on PADs		
				Low (5.651)	Normal (7.132)	High (10.186)
2	1.00	RT	0.94 ± 0.23			
		4 °C	0.90 ± 0.13			
4	1.00	RT	0.81 ± 0.12			
		4 °C	0.96 ± 0.19			
7	1.00	RT	0.99 ± 0.09			
		4 °C	1.02 ± 0.10			
10	1.00	RT	0.97 ± 0.13			
		4 °C	1.07 ± 0.17			
14	1.00	RT	0.68 ± 0.15			
		4 °C	1.07 ± 0.13			
21	1.00	RT	0.72 ± 0.18			
		4 °C	1.03 ± 0.31			
30	1.00	RT	0.92 ± 0.20			
		4 °C	1.04 ± 0.08			

2 Day 30 Spectrophotometer Nitrite concentration 1.01 ± 0.01 mg/dL

1 1. pH color index preparation

2 For preparation the pH color scale, The PADs design with diameter 4 mm of detection
3 area was fabricated using wax printing technique and 0.3 μL of each indicator was deposited
4 in to pH sensing area. Phenol red indicator was required 0.6 μL or 2 drops of solution. The
5 PADs were allowed to dry at room temperature for 5 min. The scale for reading of pH was
6 conducted as follows: 0.5 μL of buffer solution or saliva sample containing a various of pH
7 in range 5 to 10 were added on top of sensing area. The change of color on PADs was
8 recorded within 1-2 min via smartphone.

9

10 2. Optimization the optimum types and pH of washing buffer

11 To investigate the effect of pH on the formation of azo dye complex, citrate buffer
12 with pH 5, Na-phosphate buffer with pH 7.2, borate buffer with pH 10 and distilled water
13 were prepared and tested on proposed device. In this study, 0.7 μL of Griess reagent was
14 immobilized into detection areas and control area and left to dry in the dark plate for 10 min.
15 Then, 13 μL of saliva specimen containing 0.5 or 1.0 mg/dL of nitrite was added into sample
16 area. The fluid can be directly absorbed into sample area and flowed through the channel.
17 Finally, each buffer was applied into sample area and waiting until saliva fluid was spread
18 out into detection areas, approximately 5-7 min. Figure S1 represents the color signal of each
19 buffer and the photographs of PADs after adding Na-phosphate buffer.

20

21

22

23

24

25

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25

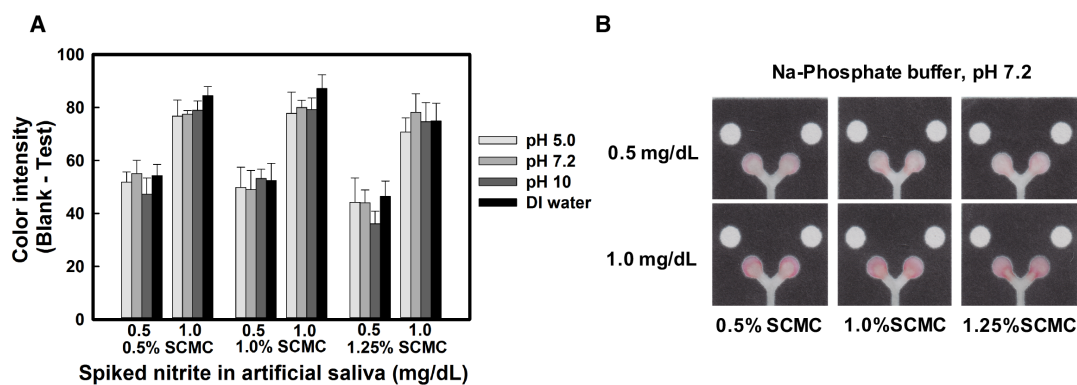


Figure S1. (A) Represent the color intensity of azo dye complex after adding with citrate buffer, Na-phosphate buffer, borate buffer and distilled water (n=4) (B) Photographs of PADs with nitrite concentration 0.5 mg/dL and 1 mg/dL in various viscosity of saliva specimen

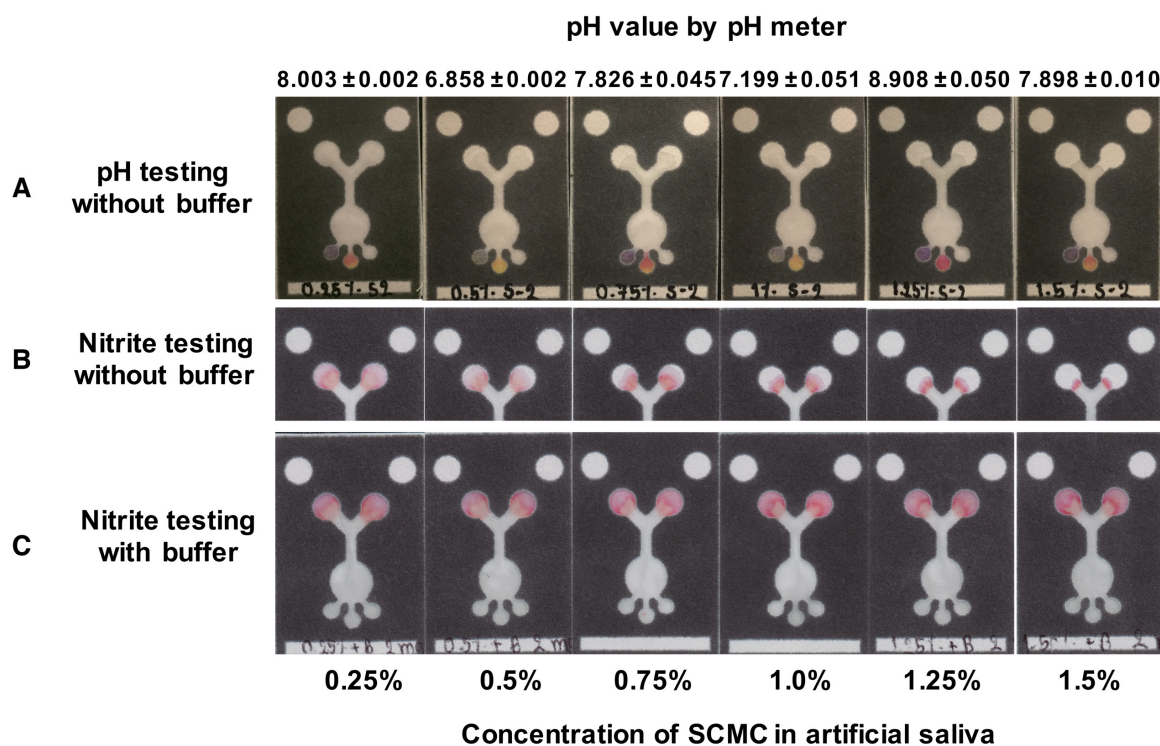
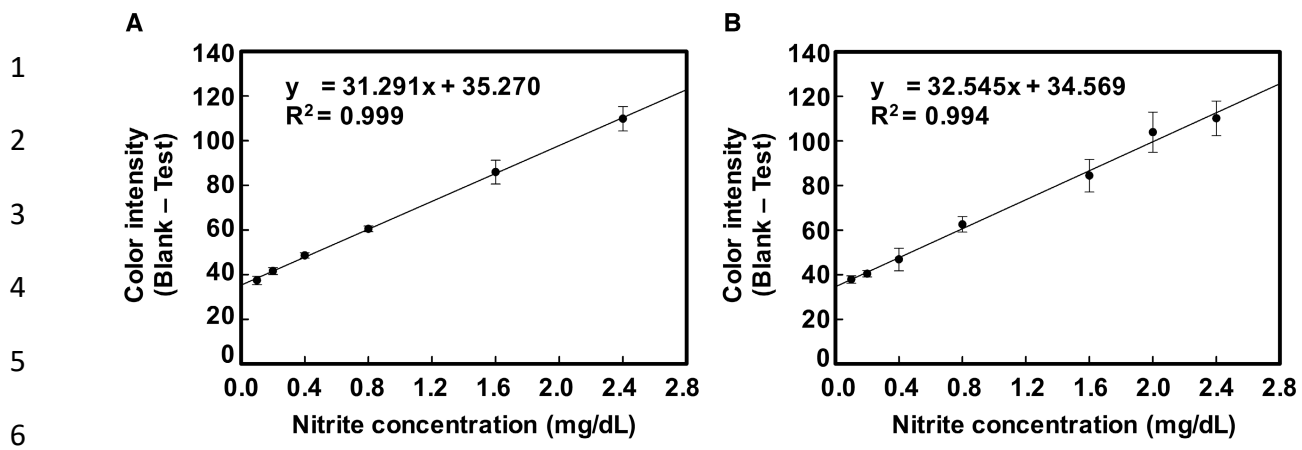
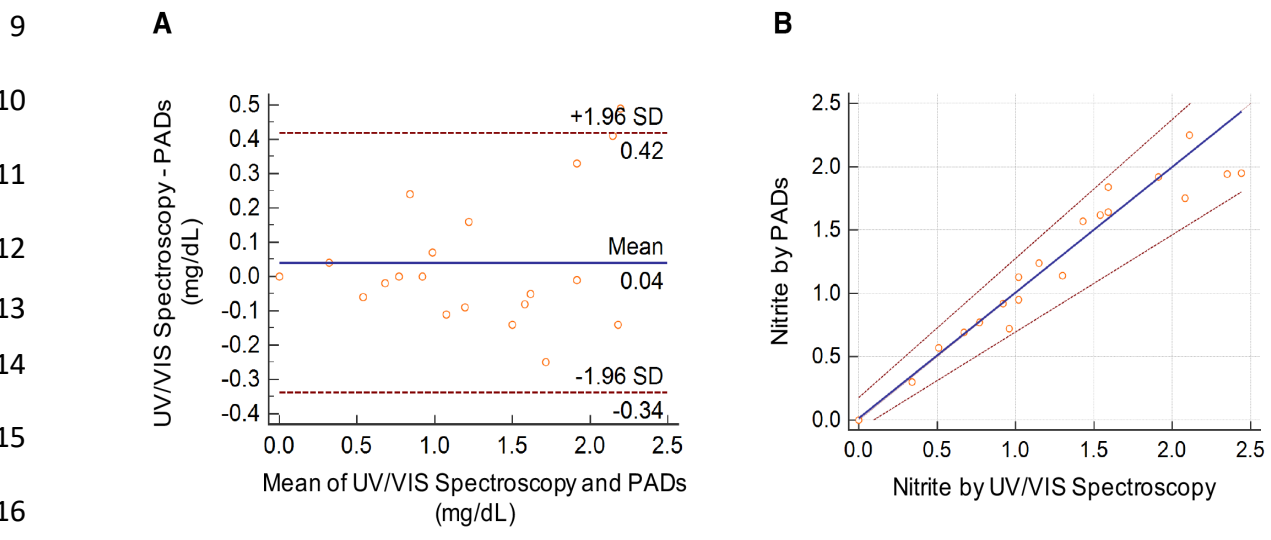


Figure S2. (A) The pH testing on proposed PADs was not affected by the viscosity of saliva. (B-C) Nitrite measurement was effected by the viscosity of solution and the accuracy for nitrite quantification was improved by added the washing buffer to spread out saliva solution.



7 Figure S3. (A) Show analytical curve for nitrite measurement from artificial saliva containing
 8 0.50% SCMC (n=6) (B) Show data from 1.50% SCMC in artificial saliva (n=6)



17 Figure S4. Comparison of nitrite levels obtained for the proposed-PADs and spectrophotometric
 18 assay. The results are shown as (A) a Bland-Altman plot and (B) a Passing-Bablok regression
 19 analysis.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17

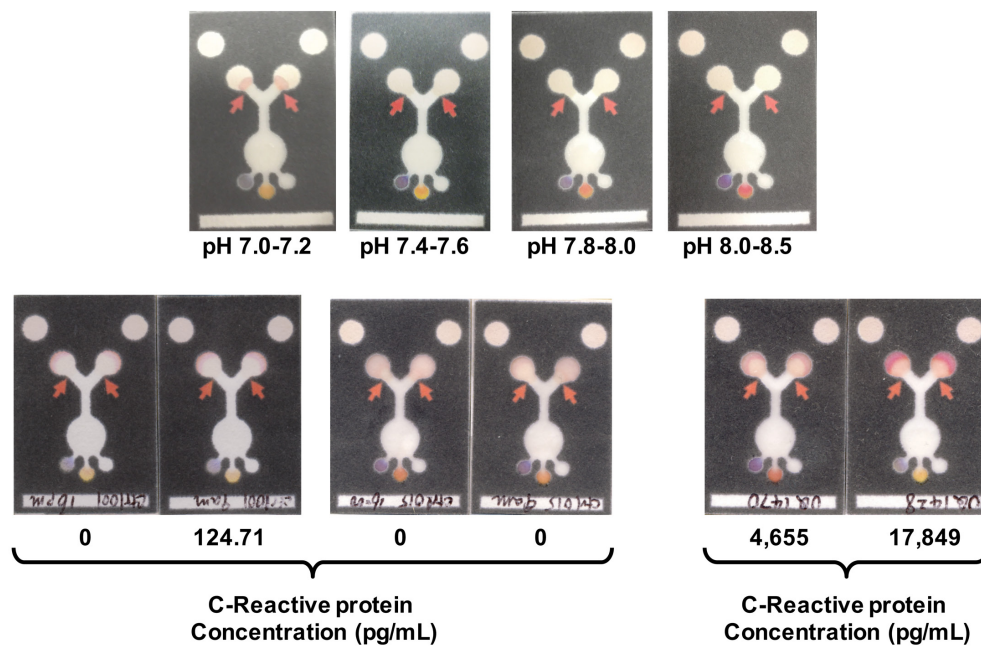


Figure S5. Real sample analysis (A) pH testing on PADs (B) Saliva contain low concentration of C-reactive protein (CRP) (C) Saliva contain high concentration of C-reactive protein (CRP)