

## Supplemental Information

### Supplemental Materials and Methods

#### RNA extraction and quantitative real-time PCR (qRT-PCR)

Total RNA was extracted from cultured cells using TRIzol reagent (Invitrogen, Cat No. 15596018) according to the manufacturer's instructions. cDNA was synthesized from total RNA using random primers. The qRT-PCR primers for *TFAP4*, *CD133*, *NANOG*, *SOX2*, *ABCG2*, *BM11*, *CD44*, *EpCAM*, *ALDH1A1*, *JUN*, *CCND1*, *TCF1*, *TCF4*, *DVL1*, *LEF1* and glyceraldehyde-3-phosphate dehydrogenase (*GAPDH*) were designed using Primer Express version 5.0 software (Applied Biosystems). *TFAP4* forward primer: 5'-GCTGAGTCTCG GGGGTTAGT-3'; *TFAP4* reverse primer: 5'-GTGCCCTCTTTGCAACATTT-3'; *CD133* forward primer: 5'-TTTTGGATTCATA TGCCTTCTGT-3'; *CD133* reverse primer: 5'-ACCC ATTGGCATTCTCTTTG-3'; *NANOG* forward primer: 5'-AT GGAGGAGGGAAGAGGAGA -3'; *NANOG* reverse primer: 5'-GATTTGTGGGCCTGAAGAAA-3'; *SOX2* forward primer: 5'-GCTTAGCCTCGTCGATGAAC-3'; *SOX2* reverse primer: 5'-AACCCCAAGATGCACA ACTC-3'; *ABCG2* forward primer: 5'-AAGCCATTGGTGTTCCT TG -3'; *ABCG2* reverse primer: 5'-CTGGATCCTGAGCCTTTGG-3'; *BM11* forward primer: 5'-CAGGTGGGGATTA GCTCAG-3'; *BM11* reverse primer: 5'-CTTTCATTGTCTTTTCCGCC-3'; *CD44* forward primer: 5'-CGTGAATACACCTGCAAAG -3'; *CD44* reverse primer: 5'-CGGACA CC ATGGACAAGTTT-3'; *EpCAM* forward primer: 5'-GCTGGTGTGTGAACACTGCT-3'; *EpCAM* reverse primer: 5'-ACGCGTTGTGATCTCCTTCT-3'; *ALDH1A1* forward primer: 5'-CCACTCACTGAATCATGCA -3'; *ALDH1A1* reverse primer : 5'-TGAGCCAGTCACCTGT GTTC-3'; *c-Jun* forward primer: 5'-GTCCTTCTTCTCTTGCGTGG-3'; *c-Jun* reverse primer: 5'-GGAGACAAGTGGCAGAGTCC-3'; *CCND1* forward primer: 5'-AGTTGTTGGGGCTC CTCAG-3'; *CCND1* reverse primer: 5'-AG ACCTTCGTTGCCCTCTGT-3'; *TCF1* forward primer: 5'-GTGCTGCTGCAGGTAGGACT-3'; *TCF1* reverse primer: 5'-CCATCCTCAAAG

AGCTGGAG-3'; *TCF4* forward primer: 5'-GCCTCTCATCAC TACAGCA-3'; and *TCF4* reverse primer, 5'-GGATGGGGGATTTGTCCTAC 3' *DVLI* forward primer: 5'-TGCCCCAC CCATCAGTACCCTG-3'; *DVLI* reverse primer: 5'-CCCCTGCTCTTGCTCCCTTCA CT-3'; *LEF1* forward primer: 5'-GCAGCTATCAACCAGATCC-3'; *LEF1* reverse primer: 5'-GATGTAGGCAGCTGTCATTC-3'; *GAPDH* forward prime: 5'-ACATCCCCTCACCAAT ACAAC-3'; *GAPDH* reverse primer: 5'-TAGCCAAATCATACTGCTCGTC-3'. Expression data were normalized to the geometric mean of housekeeping gene *GAPDH* to control the variability in expression levels and calculated as  $2^{-[(Ct \text{ of gene}) - (Ct \text{ of } GAPDH)]}$ , where Ct represents the threshold cycle for each transcript.

### **Plasmids, retroviral infection, and transfection**

The primers used to amplify the cDNA are: TFAP4 primer up : 5'-GCCGAATTCACCATG GACTAC AAGGACGACGATGACAAGGAGTATTTTCATGGTGCCAC-3'; TFAP4 primer down : 5'- GCCGCTAGCTCAGGGAAGCTCCCCGTCCCCCG-3'. The amplified PCR products was further digested by BamH1 and EcoR1 restriction enzymes and subcloned into pSin-EF2 lentiviral vector. The sequences of RNAi#1 and RNAi#2 are GCCTTGCCAACAT TCCACTAA and TTAGTGGAATGTTGGCAAGGC, respectively. The reporter plasmids and siRNA were transfected into HCC cells using Lipofectamine 2000 (Life Technologies), according to the manufacturer's instructions. The sense strand sequences of siRNAs, which were designed to target human cells, were as follows: si*DVLI*, 5'- GCAGAGUGAAGGGAGC AAATT-3'; si*LEF1*, 5'-UCAGAUGUCAACUCCAAACA A-3'. The reporter plasmids containing wild-type (CCTTTGATC; TOP flash) or mutated (CCTTTGGCC; FOP flash) TCF/LEF DNA binding sites were purchased from Upstate Biotechnology Company.

### **Primer for Chromatin immunoprecipitation-qPCR (ChIP-qPCR)**

LEF1 forward 1: 5'- AAACCTTAACAGATGCGTTCAGCAG -3'; LEF1 reverse 1: 5'- CCA CTAATTGGCTAATAAAAACAGA -3'; LEF1 forward 2: 5'-ACAGTGCAA AAGGATCGTT

TTTTTT -3'; LEF1 reverse 2: 5'- TGTCATTATTTCAAACCTTTCCAGA -3'; LEF1 forward 3: 5'- TTCGTCTACTGCAAGAGCCAAGTTC -3'; LEF1 reverse 3: 5'- CTGGGAAGTGCA CGCAGATATG -3'; LEF1 forward 4: 5'- ATAGGGCAATTCATTTTAGCCATCC -3'; LEF1 reverse 4: 5'- AAGGAACAAATGAAATGCTATTAAT -3'; LEF1 forward 5: 5'- CTCTGGG CTGTGAGTGTGAGCGGAC -3'; LEF1 reverse 5: 5'- TTTGGTGTTTTTGGTCCACTGTG GG - 3'; TCF4 forward 1: 5'- ACGTGTGTCAACTATCATGGAAGGA-3'; TCF4 reverse 1: 5'-CTTTACATTTCTACAGTGCAATGTT-3'; TCF4 forward 2: 5'-AGGATTAATAATTGA AGGA ATATA-3'; TCF4 reverse 2: 5'-GGTGCAAAAGGTTAAGGCTGACTTG-3'; TCF4 forward 3: 5'- ACAAACCCAGCTTGGTCAAGGCCAT-3'; TCF4 reverse 3: 5'-GCAAAAT GAAATGAA ACAAGTGTC-3'; TCF4 forward 4: 5'- AGTTTGTTACTTTATTTTACTATTT -3'; TCF4 reverse 4: 5'- AATTTCCCCGCTGCGACCTACAAC -3'; TCF4 forward 5: 5'-AA TGGTGAAAATATATGAACTGTGG -3'; TCF4 reverse 5: 5'-CTTTATAACAAAAAGCCAG GCACTG - 3'; DVL1 forward 1: 5'- TCTGAAAGTACGTGGAGGACGGGAC -3'; DVL1 reverse 1: 5'- CCCATCTCCCCAAGACCTCCCTCCC -3'; DVL1 forward 2: 5'- CACCTGT GTCTGAGCAGCCGTGTTG -3'; DVL1 reverse 2: 5'- AGGAGGTGGGGGTCAGCCGAG AGCC -3'; DVL1 forward 3: 5'- GGCTCCCCCGCCCCACCCACGAC -3'; DVL1 reverse 3: 5'- CTGGAAGGACTGGCGGCTGCCTGTC -3'; DVL1 forward 4: 5'-CCGGCCCTGTCC TCGCGCCTGCATG -3'; DVL1 reverse 4: 5'- GATGGGGAAGGAGCCTGTCAGCACC -3'; DVL1 forward 5: 5'- GACCTTGGGCCGGTAAGCCAGGGTC -3'; DVL1 reverse 5: 5'- CAG GACCCTGGCCGACGGATGACTC -3'. The primers used to generate DVL1 and LEF1 luciferase reporters are: DVL1 primer up: 5'- TCTGAAAGTACGTGGAGGACGGGAC-3'; DVL1 primer down: 5'- CCCATCTCCCCAAGACCTCCCTCCC-3'; LEF1 primer up: 5'- AA ACCTTAACAGATGCGTTCAGCAG -3'; LEF1 primer down: 5'- CCACTAATTGGCTAAT AAAAACAGA -3'.

## Supplemental Figure legends

**Supplemental Figure 1. Elevated TFAP4 expression is associated with overall survival in human HCC.** Kaplan-Meier analysis of the association between TFAP4 expression and overall survival in the TCGA dataset (A) and 197 primary HCC specimens (B). HCC: hepatocellular carcinoma; TCGA: the cancer genome atlas; TFAP4: transcription factor AP-4.

**Supplemental Figure 2. TFAP4 overexpression activates the Wnt/ $\beta$ -catenin signaling.** (A) GSEA analysis indicating a significant correlation between TFAP4 mRNA expression and Wnt-suppressed gene signatures (LABBE\_WNT3A\_TARGETS\_DN, LABBE\_TARGETS\_OF\_TGFB1\_AND\_WNT3A\_DN). (B) Prediction and validation of the interaction between TFAP4 and the *TCF4* promoter. (C) Real-time PCR analysis of the mRNA expression of *TCF4*, *LEF1* and *DVL1* genes. Bars in B and C indicate the mean  $\pm$ SD of three independent experiments; \*  $P < 0.05$ . *DVL1*: dishevelled segment polarity protein 1; GSEA: gene set enrichment analysis; *LEF1*: lymphoid enhancer binding factor 1; TFAP4: transcription factor AP-4; *TCF4*: transcription factor 4.

**Supplemental Figure 3. TFAP4 overexpression enhances the TIC-like phenotype through activation of Wnt/ $\beta$ -catenin pathway in human HCC.** (A) Real-time PCR analysis of stemness-related markers in the indicated cells. (B) Real-time PCR analysis of the expression of Wnt/ $\beta$ -catenin signaling pathway downstream genes in the indicated cells. Bars represent the mean  $\pm$ SD of three independent experiments. \* $P < 0.05$ . HCC: hepatocellular carcinoma; TIC: tumor-initiating cell; TFAP4: transcription factor AP-4.

## Supplemental Tables

**Supplemental Table 1. Clinicopathological Characteristics of Clinical Samples and Expression of TFAP4 in Liver Cancer**

<b>Characteristics</b>	<b>No. patients</b>	<b>(%)</b>
<b>Age (years)</b>		
≤ 60	104	(52.8)
> 60	93	(47.2)
<b>Gender</b>		
Male	109	(55.3)
Female	88	(44.7)
<b>Clinical stage</b>		
I-II	63	(32.0)
III-IV	134	(68.0)
<b>Histological differentiation</b>		
Well/Moderate	127	(64.5)
Poor	70	(35.5)
<b>HBsAg</b>		
positive	175	(88.8)
negative	22	(11.2)
<b>AFP</b>		
> 400 ng/ml	171	(86.8)
≤ 400 ng/ml	26	(13.2)
<b>Tumor size</b>		
> 5 cm	121	(61.4)

≤ 5 cm	76	(38.6)
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**Vital status (at follow-up)**

Alive	131	(66.5)
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Death due to liver cancer cause	66	(33.5)
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**Expression of TFAP4**

Low expression	87	(44.2)
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High expression	110	(55.8)
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**Relapse**

Yes	63	(32.0)
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No	134	(68.0)
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**Supplemental Table 2. Correlation between TFAP4 Expression and Clinicopathologic Characteristics of Liver Cancer Patient**

Characteristic		TFAP4 Protein level		Chi-square test
		Low(87)	High(110)	P value
<b>Age (years)</b>	≤ 60	46	58	<i>0.984</i>
	> 60	41	52	
<b>Gender</b>	Male	53	56	<i>0.160</i>
	Female	34	54	
<b>Clinical stage</b>	I-II	49	14	<i>&lt;0.001</i>
	III-IV	38	96	
<b>Histological differentiation</b>	Well/Moderate	63	64	<i>0.038</i>
	Poor	24	46	
<b>Tumor size (cm)</b>	> 5	42	79	<i>0.001</i>
	≤ 5	45	31	
<b>AFP (ng/ml)</b>	> 400	71	100	<i>0.055</i>
	≤ 400	16	10	
<b>Vital status</b>	Alive	65	66	<i>0.030</i>
	Death	22	44	
<b>HBsAg</b>	Positive	77	98	<i>0.897</i>
	Negative	10	12	
<b>Relapse</b>	Yes	19	44	<i>0.007</i>
	No	68	66	

**Supplemental Table 3. Spearman Analysis of Correlation between TFAP4 and  
Clinicopathological Characteristics**

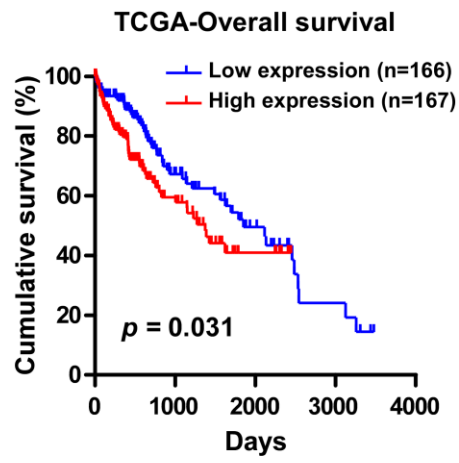
<b>Variables</b>	<b>TFAP4 expression level</b>	
	<b>Spearman Correlation</b>	<b>P Value</b>
<b>Survival time</b>	-0.305	<i>&lt;0.001</i>
<b>Vital status</b>	0.155	<i>0.030</i>
<b>HBsAg</b>	0.009	<i>0.898</i>
<b>Age</b>	0.001	<i>0.984</i>
<b>Clinical stage</b>	0.403	<i>&lt;0.001</i>
<b>Histological differentiation</b>	0.148	<i>0.038</i>
<b>AFP</b>	0.136	<i>0.056</i>
<b>gender</b>	0.100	<i>0.162</i>
<b>Relapse</b>	0.193	<i>0.006</i>
<b>Tumor size</b>	0.240	<i>0.001</i>



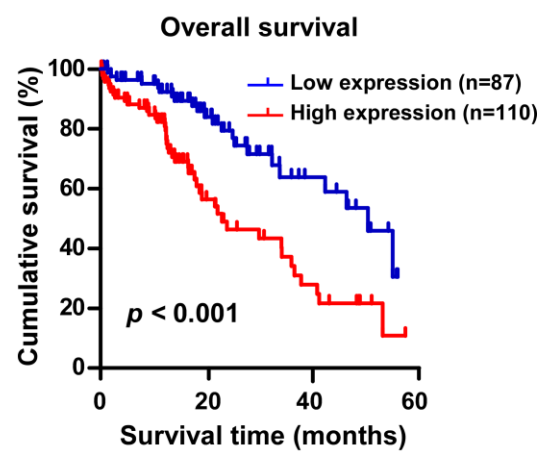
**Supplemental Table 4. Univariate and Multivariate Analyses of Various Prognostic Parameters in Liver Cancer Patients by Cox-regression Analysis**

	Univariate analysis			Multivariate analysis		
	Relative risk	95% confidence interval	<i>P</i>	Relative risk	95% confidence interval	<i>P</i>
<b>Stage</b>	0.125	1.127-1.838	<i>0.003</i>	1.360	1.033-1.792	<i>0.029</i>
<b>Recurrence</b>	0.250	1.260-3.360	<i>0.004</i>	1.703	1.022-2.839	<i>0.041</i>
<b>TFAP4</b>	0.263	1.585-4.451	<i>&lt;0.001</i>	1.825	1.027-3.244	<i>0.040</i>
<b>Histological differentiation</b>	0.252	1.070-2.873	<i>0.026</i>	1.792	1.080-2.974	<i>0.024</i>

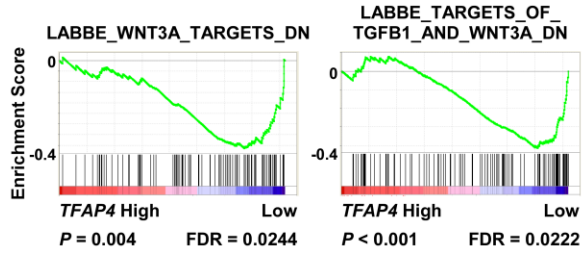
**A**



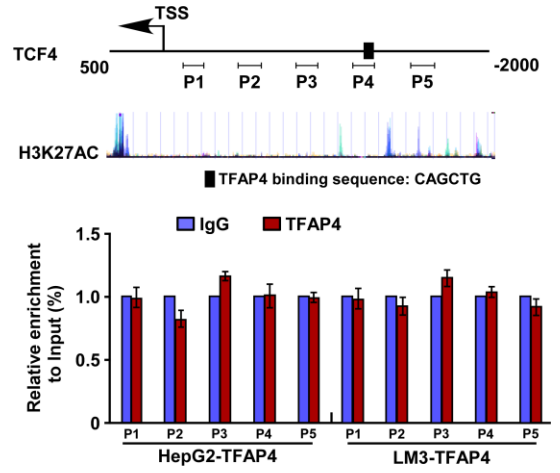
**B**



**A**



**B**



**C**

