

Supplementary Material

Direct Administration of Nerve-Specific Contrast to Improve Nerve Sparing Radical Prostatectomy

Authors: Connor W. Barth¹ and Summer L. Gibbs^{1,2,3*}

Affiliations:

¹Biomedical Engineering Department, Oregon Health & Science University, Portland, OR 97201.

²Knight Cancer Institute, Oregon Health & Science University, Portland, OR 97201.

³OHSU Center for Spatial Systems Biomedicine, Oregon Health & Science University, Portland, OR 97201.

*To whom correspondence should be addressed: Summer L. Gibbs, Ph.D. Oregon Health & Science University, Collaborative Life Sciences Building, 2730 SW Moody Ave, Mail Code: CL3SG, Portland, OR 97201. Email: gibbs@ohsu.edu, Phone: 503-494-8940

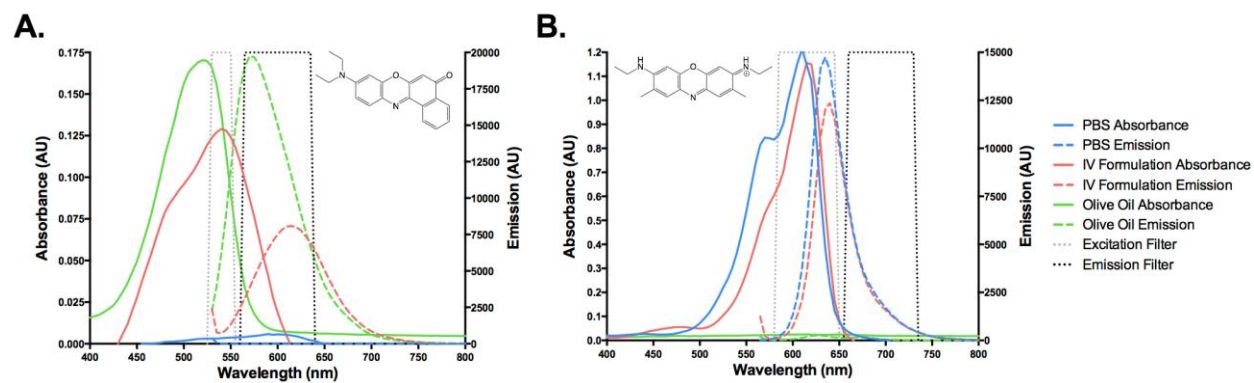


Figure S1: Fluorescence spectra and structures of Oxazine 4 and Nile Red fluorophores. (A) Absorbance and emission spectra for 10 μ M (A) Nile Red and (B) Oxazine 4 in PBS, co-solvent formulation, or olive oil with overlaid excitation and emission filter spectra used for *in vivo* imaging. The fluorophores' chemical structures are inlaid into each graph.

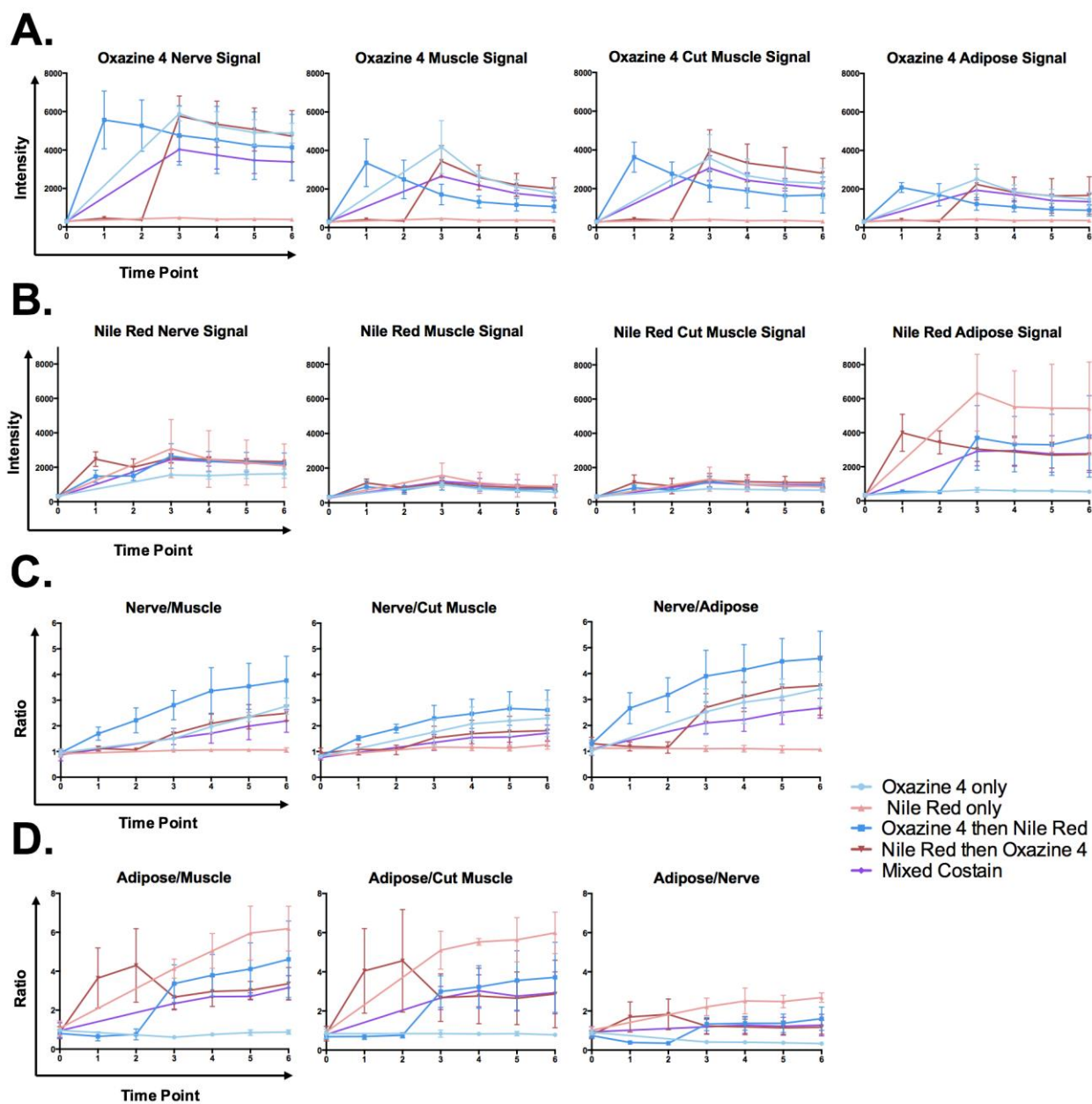
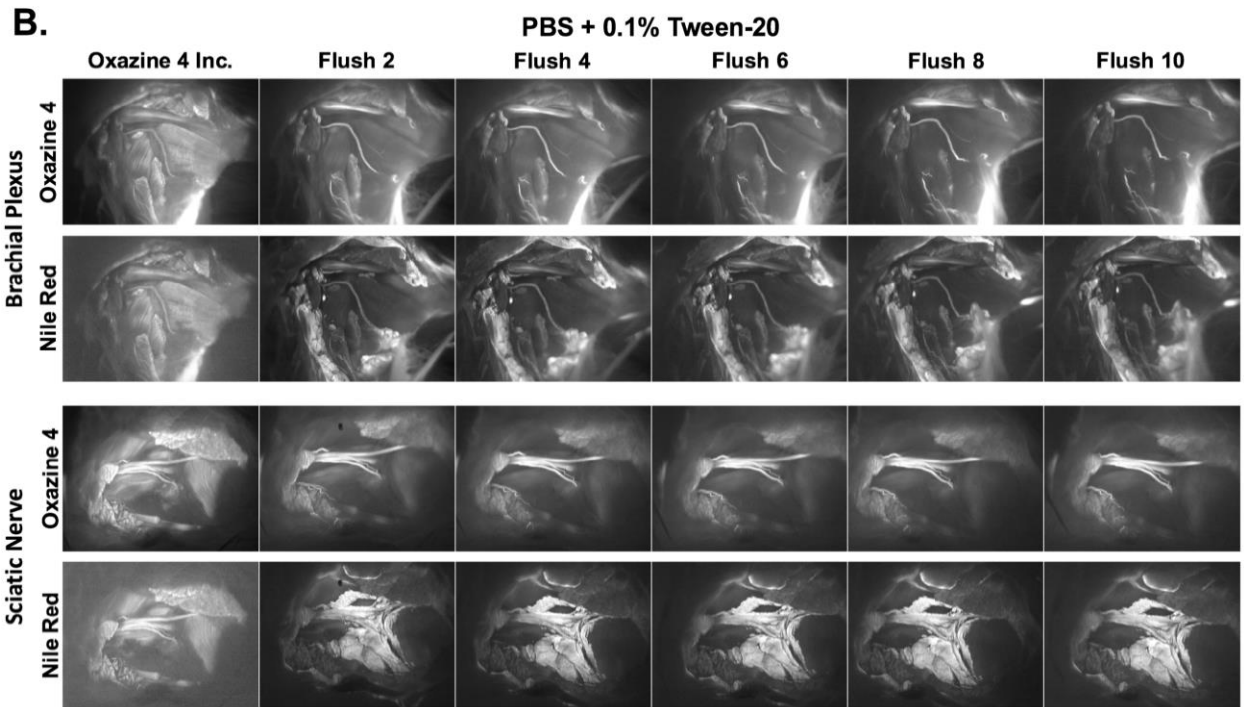
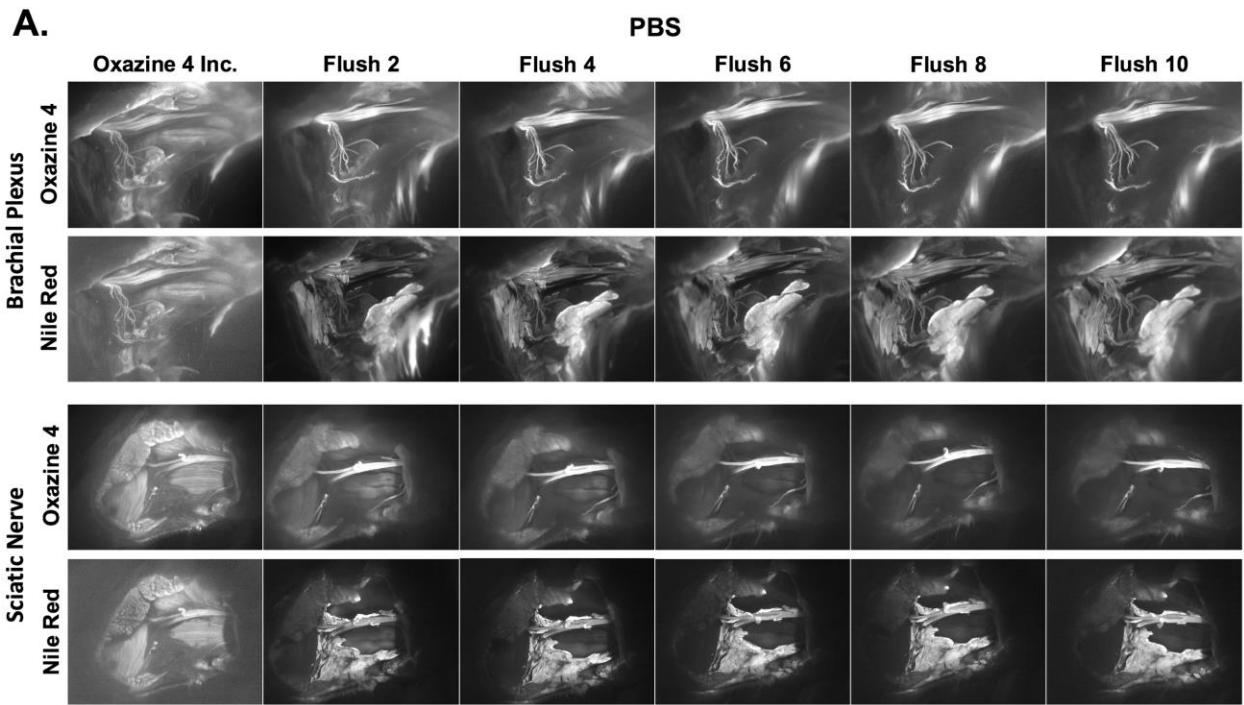


Figure S2: Fluorophore cross-talk between Oxazine 4 and Nile Red. Fluorescence signal intensities for each major visible tissue type and nerve to background tissue ratios were determined using region of interest analysis on images collected in the (A) Oxazine 4 fluorescence channel or (B) Nile Red fluorescence channel. The (C) nerve or (D) adipose to background tissue ratios were calculated from the intensity values determined through region of interest analysis. All data was determined from images collected for n=3 nerve sites at several time points: 0 = prior to the initial Oxazine 4 incubation, 1 = following the Oxazine 4 incubation, 2 = following the first blank co-solvent formulation wash, 3 = following the Nile Red incubation, 4 = following the second blank co-solvent formulation wash, 5 = following the PBS + 0.1% Triton X-100 wash, 6 = following the PBS + 0.1% Tween-20 wash. Time point 0 was considered the baseline fluorescence intensity for each tissue type. The data was determined from images collected at either 1 s exposure time for the time 0 point images or 2.5 ms exposure time for all other time points. All data in presented as mean +/- standard deviation.



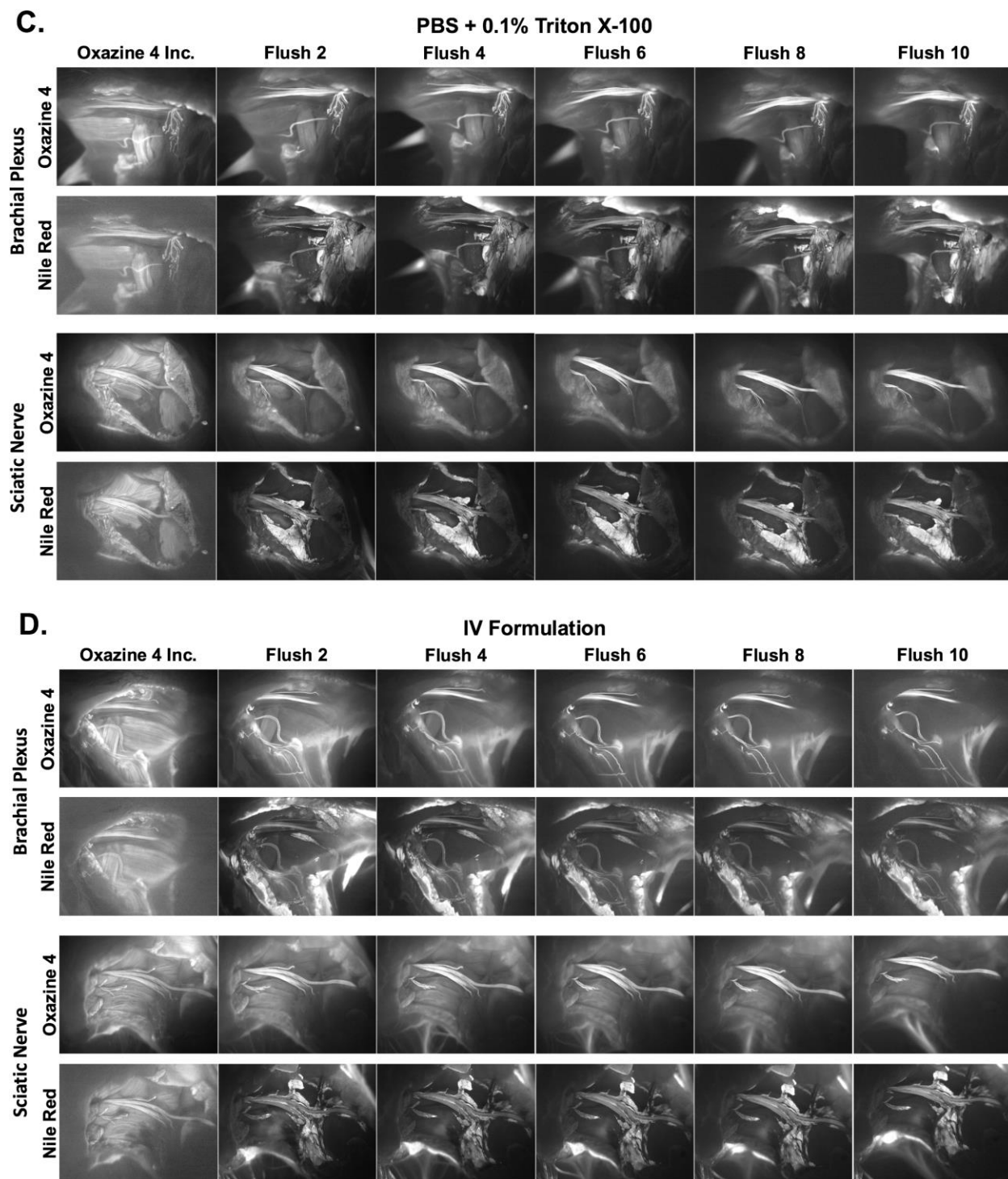


Figure S3: *Washing solution composition comparison.* Representative fluorescence images for nerve sites washed with (A) PBS, (B) PBS + 0.1% Tween-20, (C) PBS + 0.1% Triton X-100, or (D) blank co-solvent formulation in a series of 10 flushes are shown. All images are representative of data collected for n=3 nerve sites following completion of each fluorophore incubation and flush step. All images were collected at 2.5 ms exposure time.

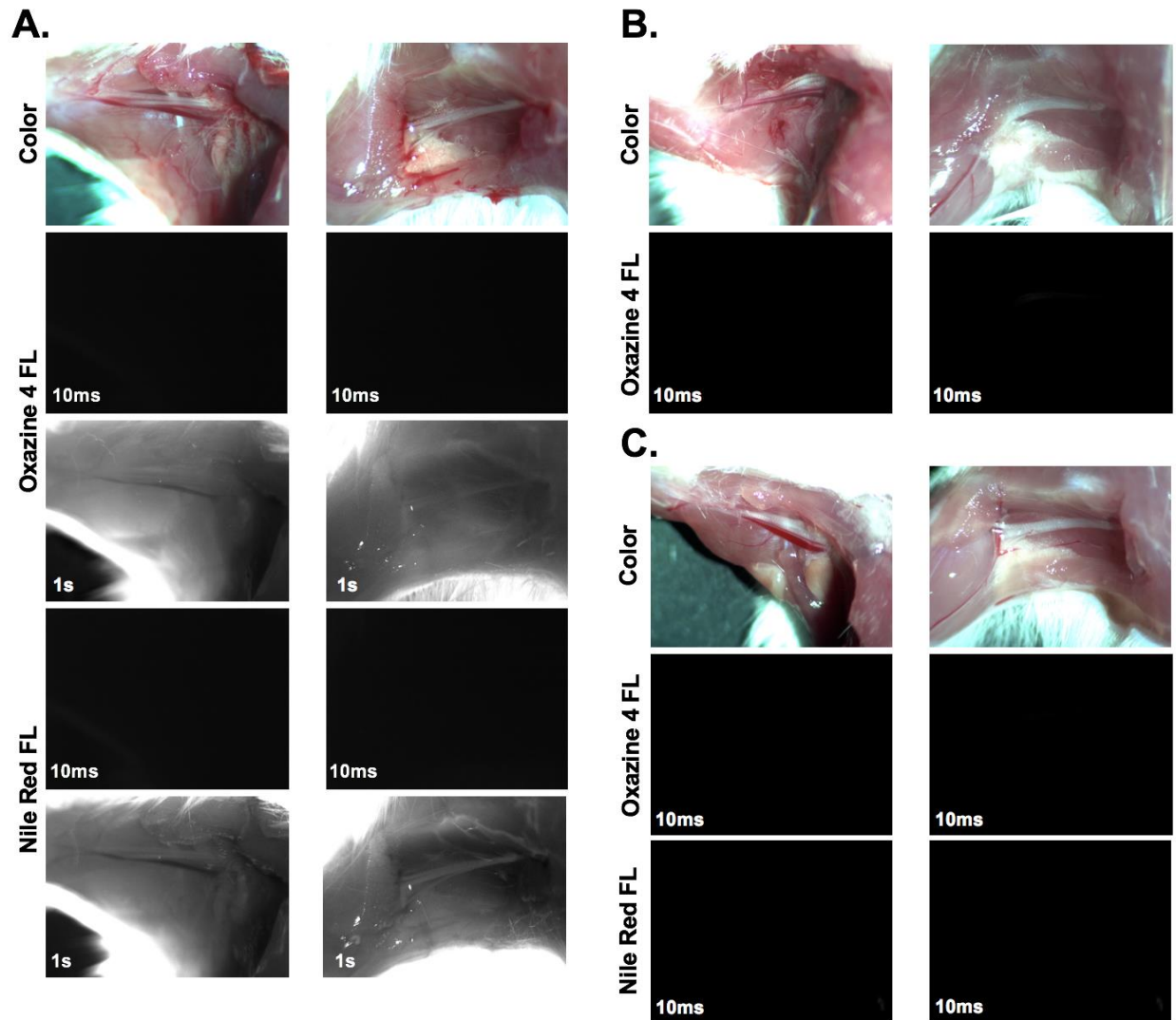


Figure S4: *Murine control images and video rate systemic administration images.* Representative fluorescence images for (A) unstained nerve sites used to calculate control nerve to background tissue ratios, (B) nerve sites stained by systemic administration of Oxazine 4 alone taken at 10 ms exposure time, and (C) nerve sites stained by systemic administration of Oxazine 4 and Nile Red taken at 10 ms exposure times. All images are representative of data collected for n=3 nerve sites following completion of the staining procedure.

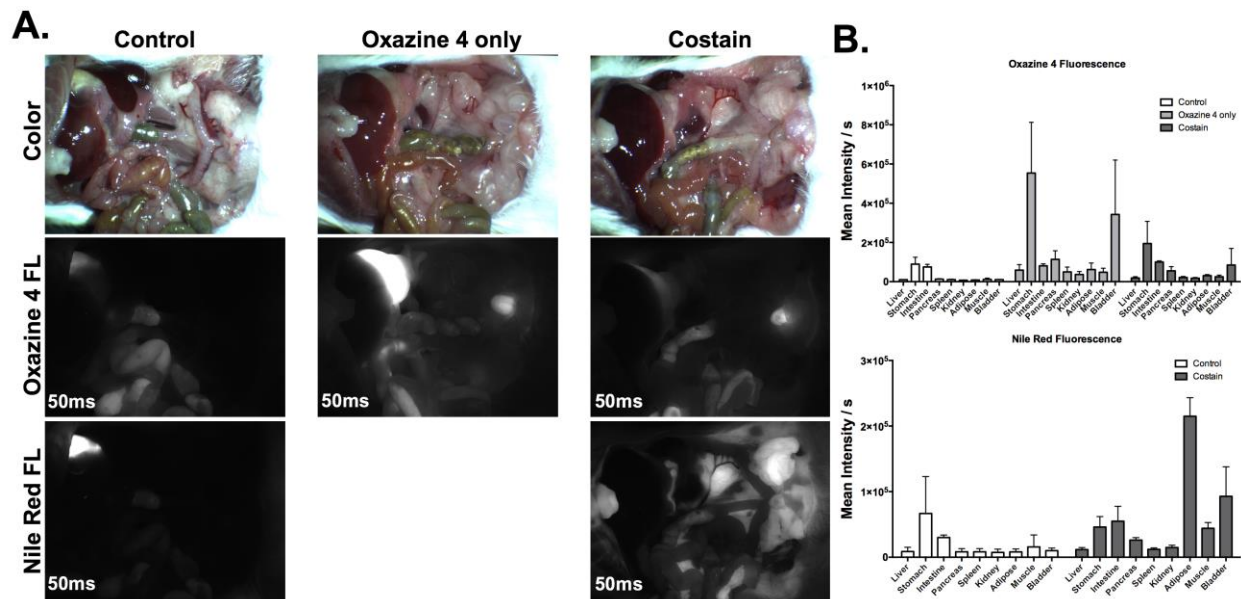


Figure S5: Fluorophore peritoneal biodistribution following systemic administration. **(A)** Representative color and fluorescence images for control blank co-solvent, Oxazine 4 only, and Oxazine 4 with Nile Red co-stain systemically administered mice. **(B)** Fluorophore biodistribution was quantified using region of interest analysis. The mean intensities normalized to exposure time (Mean Intensity/second (s)) as shown. All images were collected at 50 ms exposure time and are displayed with equal contrast and brightness. The data is presented as mean +/- standard deviation.