## **Combination Therapy for Ulcerative Colitis: Orally Targeted Nanoparticles Prevent Mucosal Damage and Relieve Inflammation**

Bo Xiao<sup>1,2,⊠</sup>, Zhan Zhang<sup>2</sup>, Emilie Viennois<sup>2,3</sup>, Yuejun Kang<sup>1</sup>, Mingzhen Zhang<sup>2</sup>, Moon Kwon Han<sup>2</sup>, Jiucun Chen<sup>1</sup>, Didier Merlin<sup>2,3</sup>

1. Institute for Clean Energy and Advanced Materials, Faculty for Materials and Energy, Southwest University, Chongqing, 400715, P. R. China

2. Institute for Biomedical Sciences, Center for Diagnostics and Therapeutics, Georgia State University, Atlanta, 30302, USA

3. Atlanta Veterans Affairs Medical Center, Decatur, 30033, USA

Corresponding authors: Bo Xiao, Ph.D., Institute for Clean Energy and Advanced Materials, Faculty of Materials and Energy, Southwest University, Chongqing, 400715, P. R. China. Email: hustboxiao@gmail.com or bxiao@gsu.edu; Tel: +86-23-6825-4762; Fax: +86-23-6825-4969



Figure S1. (a) Agarose gel electrophoresis of siRNA and (b) corresponding photo of agarose gel. Lane 1, control siRNA; Lane 2, siRNA extracted from HA-siCD98-NPs.



Figure S2. In vitro cytotoxicity assays of HA-siCD98-NPs, HA-CUR-NPs and HAsiCD98/CUR-NPs against (a) Colon-26 cells for 24 h, (b) Raw 264.7 macrophages for 24 h, (c) Colon-26 cells for 48 h and (d) Raw 264.7 macrophages for 48 h. Triton X-100 was used as the positive control to produce a maximum cell death rate (100%). Cell culture medium was used as a negative control (death rate defined as 0%). Toxicity is given as the percentage of viable cells remaining after treatment for 24 h. Each point represents the mean  $\pm$  S.E.M. (n=5). Statistical significance was assessed using Student's *t*-test (\**P*<0.05 and \*\**P*<0.01).



Figure S3. *In vitro* cytotoxicity assays of HA-CUR-NPs against (a) Colon-26 cells and (b) Raw 264.7 macrophages for 5 h. Triton X-100 was used as the positive control to produce a maximum cell death rate (100%). Cell culture medium was used as a negative control (death rate defined as 0%). Toxicity is given as the percentage of viable cells remaining after treatment for 24 h. Each point represents the mean  $\pm$  S.E.M. (n=5). Statistical significance was assessed using Student's *t*-test (\**P*<0.05 and \*\**P*<0.01).



Figure S4. *In vitro* down-regulation capability of CD98 mRNA expression by various NPs. (a) CD98 mRNA expression levels in Colon-26 cells and Raw 264.7 macrophages treated by blank HA-NPs for 24 h or 48 h. (b) CD98 mRNA expression levels in Colon-26 cells and Raw 264.7 macrophages treated by HA-scrambled siRNA-NPs for 24 h. (c) CD98 mRNA expression levels in Colon-26 cells and Raw 264.7 macrophages treated by HA-scrambled siRNA-NPs for 24 h. (c) CD98 mRNA expression levels in Colon-26 cells and Raw 264.7 macrophages treated by HA-siCD98/CUR-NPs for 72 h or 96 h. Each point represents the mean  $\pm$  S.E.M. (n=3).



Figure S5. Typical images of stomach, small intestine and caecum imaging showing the distribution of orally HA-functionalized NPs embedded in hydrogel at four different time points (0, 8, 16 and 24 h).



Figure S6. Histological scores determined from H&E-stained colons. Each point represents the mean  $\pm$  S.E.M. (n = 3).

## Table S1

Primers used in this study.

Primer name	Sequence	Description
CD98-F	5'-GAGGACAGGCTTTTGATTGC-3'	CD98 gene RT-PCR forward primer
CD98-R	5'-ATTCAGTACGCTCCCCAGTG-3'	CD98 gene RT-PCR reverse primer
TNF-α-F	5'-AGGCTGCCCCGACTACGT-3'	Tumor necrosis factor gene RT-PCR forward primer
TNF-α-R	5'-GACTTTCTCCTGGTATGAGATAGCAAA-3'	Tumor necrosis factor gene RT-PCR reverse primer
36B4-F	5'-TCCAGGCTTTGGGCATCA-3'	36B4 gene RT-PCR forward primer
36B4-R	5'-CTTTATCAGCTGCACATCACTCAGA-3'	36B4 gene RT-PCR reverse primer