

Supplementary data:

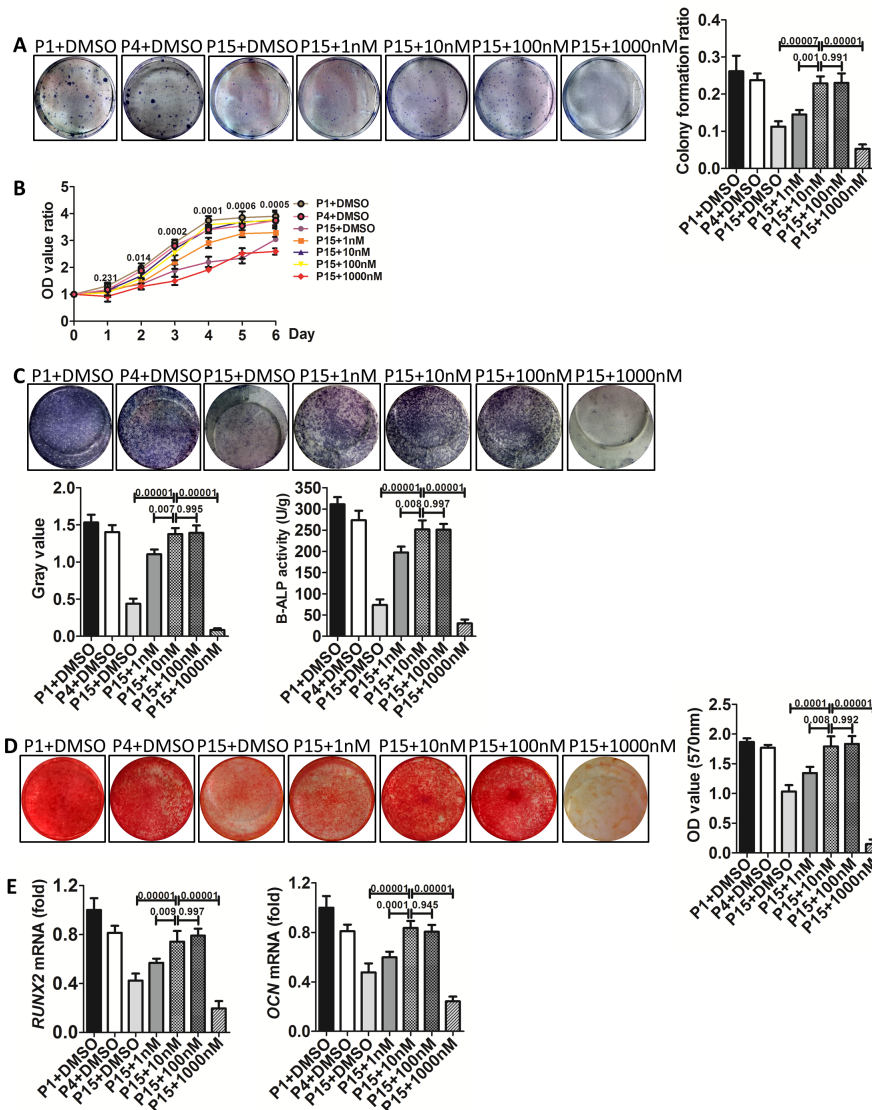


Fig.S1

Fig. S1 Optimal concentration of melatonin for stemness preservation of long-term passaged BMMSCs.

Rat BMMSCs cultured with DMSO for 1 passage (P1), 4 passages (P4), and 15 passages (P15) and with different concentration of melatonin for 15 passages were used in the following analysis. (A) 5×10^2 BMMSCs were seeded in 5-cm dishes for 10 days. CFU-F was analyzed by toluidine blue staining and the colony ratio was counted (n=3). (B) 1×10^3 /well BMMSCs were seeded in 96-well plates. Proliferation of BMMSCs was detected from day 0 to day 6 (n=3). (C) 1×10^5 /well BMMSCs seeded in 12-well plates were induced with osteogenic medium for 7 days. Activity of ALP was detected by ALP staining and quantified with Image-Pro Plus 6.0 software (n=3). Activity of B-ALP was detected by ELISA and the values were normalized to corresponding total protein concentration

(n=3). **(D)** BMMSCs were induced with osteogenic medium for 28 days. Mineralized nodules were detected by alizarin red staining and quantified with a spectrophotometer after dissolving with isopropanol (n=3). **(E)** *Runx2* and *Ocn* mRNA levels of BMMSCs were analyzed by Real-time RT-PCR. *β -actin* was used as the loading control for quantification (n=3). Data are shown as mean \pm SD. *P* value is presented in each graph.

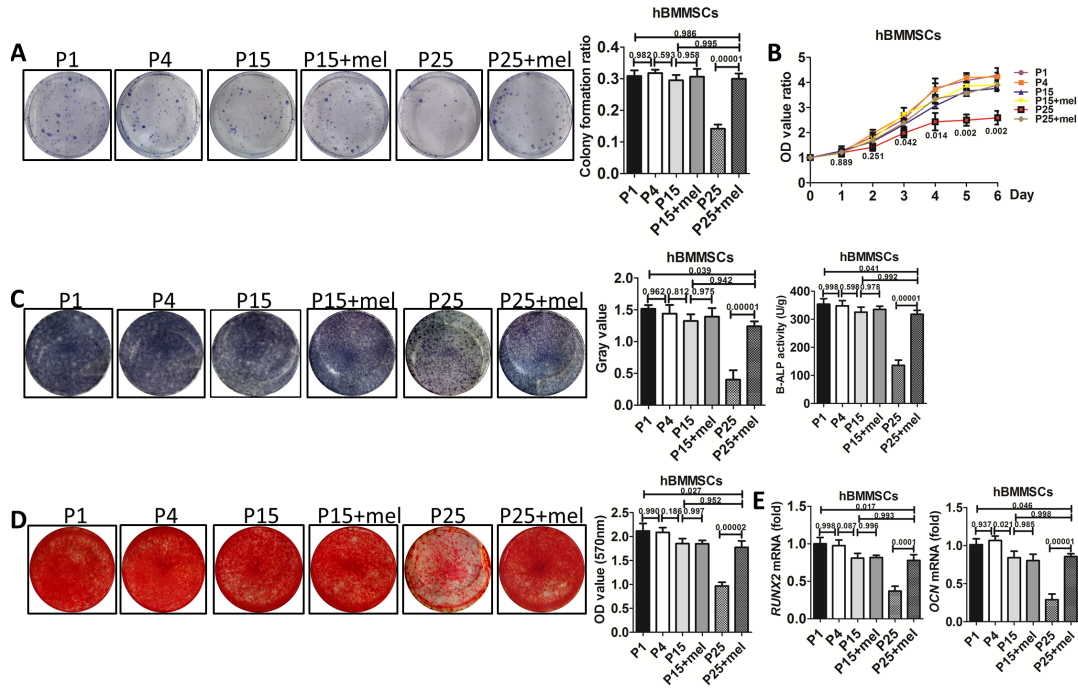


Fig.S2

Fig. S2 Melatonin treatment prevents the stemness loss of long-term passed human BMMSCs.

Human BMMSCs cultured with DMSO for 1 passage (P1), 4 passages (P4), 15 passages (P15) and 25 passages (P25), or with melatonin for 15 passages and 25 passages were used in the following analysis. **(A)** 5×10^2 BMMSCs were seeded in 5-cm dishes for 10 days. CFU-F was analyzed by toluidine blue staining and the colony ratio was counted ($n=3$). **(B)** 1×10^3 /well BMMSCs were seeded in 96-well plates. Proliferation of BMMSCs was detected from day 0 to day 6 ($n=3$). **(C)** 1×10^5 /well BMMSCs seeded in 12-well plates were induced with osteogenic medium for 7 days. Activity of ALP was detected by ALP staining and quantified with Image-Pro Plus 6.0 software ($n=3$). Activity of B-ALP was detected by ELISA and values were normalized to corresponding total protein concentration ($n=3$). **(D)** BMMSCs were induced with osteogenic medium for 28 days. Mineralized nodules were detected by alizarin red staining and quantified with a spectrophotometer after dissolving with isopropanol ($n=3$). **(E)** *Runx2* and *Ocn* mRNA levels of BMMSCs were analyzed by Real-time RT-PCR. *β -actin* was used as the loading control for quantification ($n=3$). Data are shown as mean \pm SD. *P* value is presented in each graph.

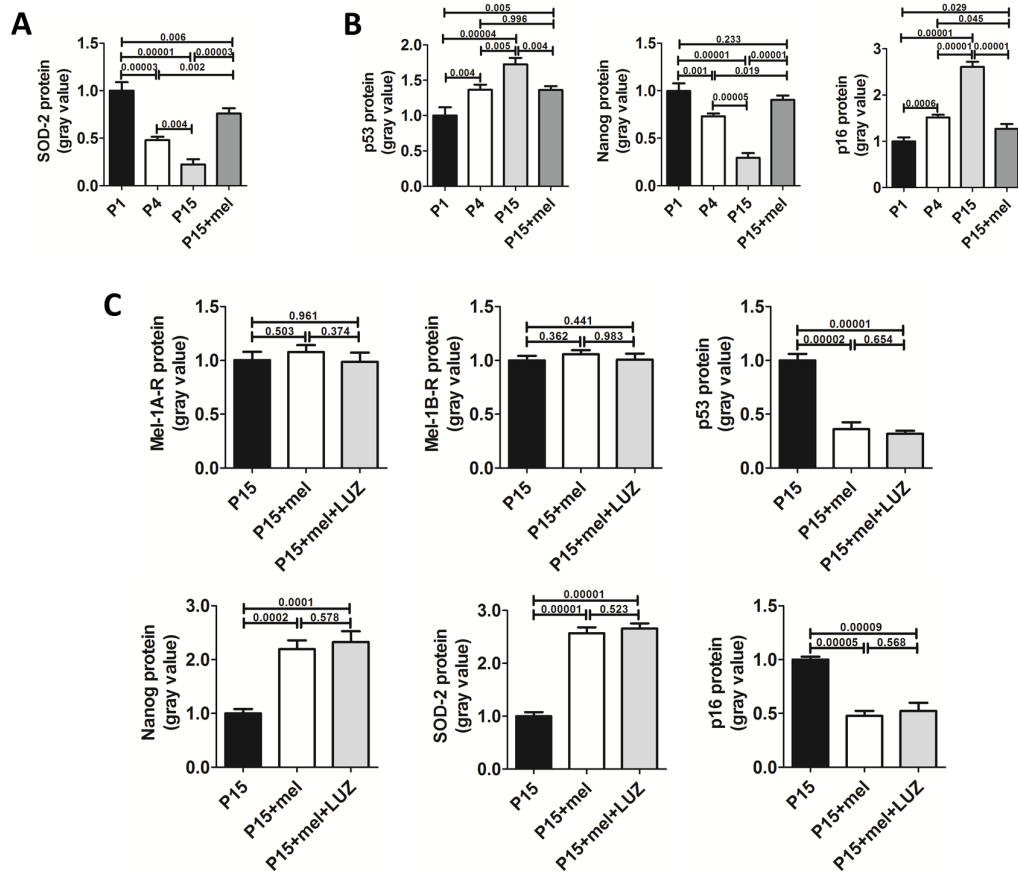


Fig.S4

Fig. S4 Quantification of western blots in Figure 8.

Expression of SOD2, p53, p16, NANOG, Mel-1A-R and Mel-1B-R protein was detected by western blot analysis. The gray values of specific protein bands were analyzed by Image J software. β -actin was used as the loading control (n=3). (A) Quantification of blots in Fig. 8C. (B) Quantification of blots in Fig. 8F. (C) Quantification of blots in Fig. 8P. Data are shown as mean \pm SD. *P* value is presented in each graph.

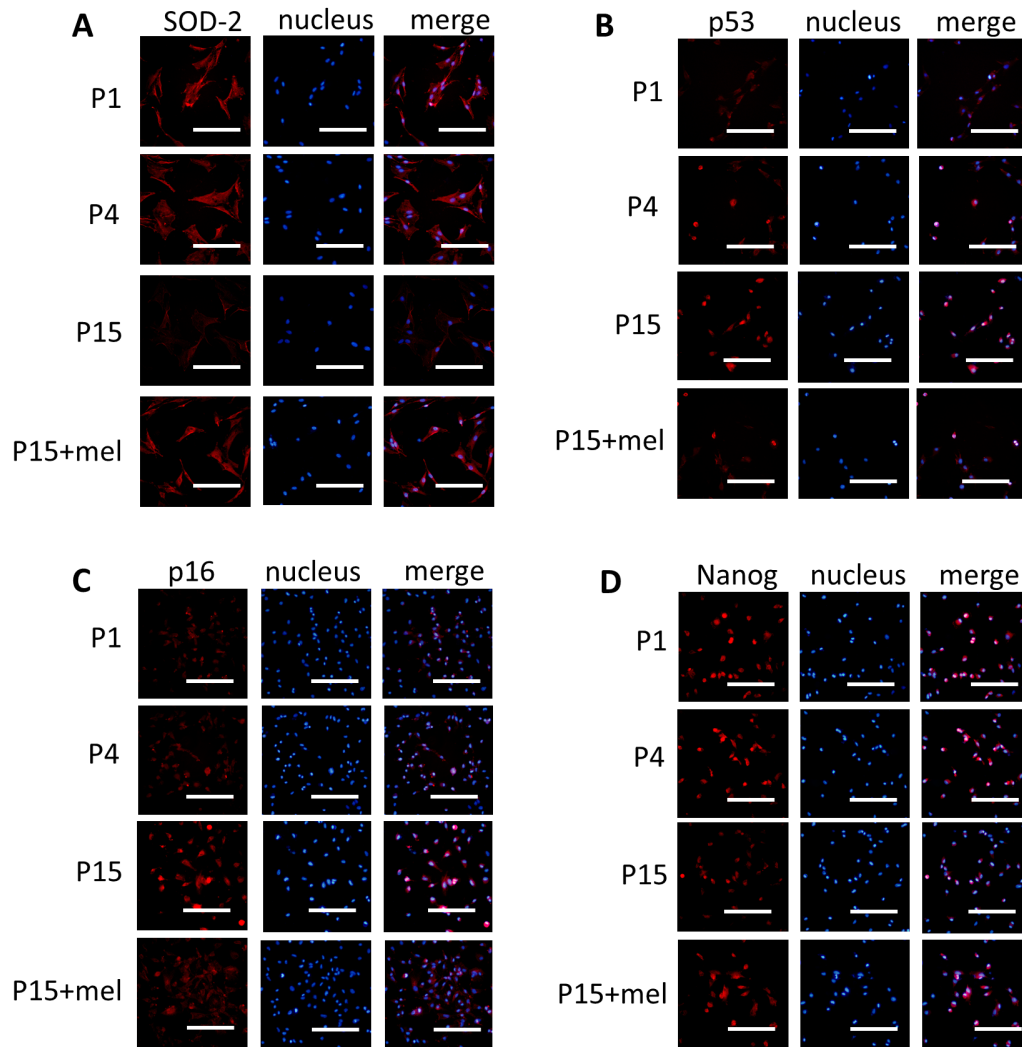


Fig.S5

Fig. S5 Melatonin treatment increases SOD2 levels, inhibits p53 pathway, and preserves NANOG expression in long-term passaged BMMSCs.

Rat BMMSCs were treated with DMSO for 1, 4 and 15 passages or with 10 nM melatonin for 15 passages. (A) SOD2, (B) p53, (C) p16 and (D) NANOG protein accumulations in rat BMMSCs were detected by immunofluorescence. Cell nuclei were counterstained by Hoechst 33342 (n=3). Scale bar, 200 μ m.

Supplementary Table:

Supplementary Table 1. Sequence of primers for realtime RT-PCR

mRNA/primer	Forward (5'-3')	Reverse (5'-3')
<i>r-β-actin</i>	GGAGATTACTGCCCTGGCTCCTA	GACTCATCGTACTCCTGCTTGCTG
<i>r-Runx2</i>	CATCGCCGGGAATGATGAG	TGTGAAGACCGTGATGGTCAAAGTG
<i>r-Ocn</i>	CCACCCGGGAGCAGTGT	GAGCTGCTGTGACATCCATACTTG
<i>r-Sod2</i>	CACATTAACGCGCAGATCATG	CCTTAGGGCTCAGGTTTGTC
<i>r-Nanog</i>	TTCTGAACCTGAGCTATAAGCAG	GAGAATAGCTGCAATGGATG
<i>r-p16</i>	TCCTCCGCTGGGAACGT	GGCGTGCTTGAGCAGAAGTT
<i>r-p53</i>	CCCATCCTTACCATCATCACG	TGCTGGTGGGCAGTGCTCTCT
<i>r-Mel-1A-R</i>	TTGTGGCGAGTTTAGCTGTG	TTTACCCTCCGTCTGACCTG
<i>r-Mel-1B-R</i>	TACATCAGCCTCATCTGGCTT	CACAAACTGCGAACATGGT
<i>h-β-actin</i>	TGGCACCCAGCACAATGAA	CTAAGTCATAGCCGCCTAGAAGCA
<i>h-Runx2</i>	AGATGATGACACTGCCACCTCTG	GGGATGAAATGCTTGGGAACT
<i>h-Ocn</i>	CCTCACACTCCTCGCCCTATT	CCCTCCTGCTTGACACAAA