Supporting Information for

Histone deacetylase inhibitors delivery using nanoparticles with intrinsic passive tumor targeting properties for tumor therapy.

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3. ¹H NMR of compound 4b.



5. HPLC of compound 4b.

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Time (min)	% ACN	%H ₂ 0+0.1% TFA	Flow (mL/min)						
0.0	85.0	15.0	0.250						
5.0	85.0	15.0	0.500						
10.0	85.0	15.0	1.000						
15.0	85.0	15.0	1.000						

Composition of eluting system for the determination of the half-life.



6. HPLC of compound 2.



7. Table S1. Release kinetic of compounds 2 from its prodrug 4b at pH 3, 4.3, 5 and 7.3 expressed in percent.

The quantity of released compound 2 versus its prodrug 4b is expressed in relative percentage pH 3

time (min.)	0	24	43	76	94	164	193	246	300	376	1260
2	0.00	6.75	6.53	21.35	29.92	42.02	47.76	63.82	73.03	74.77	87.52
4b	100.00	93.25	93.47	78.65	70.08	57.98	52.24	36.18	26.97	25.23	12.48

pH 4.3											
time (min)	0	17	34	51	71	97	113	188	286	415	1376
2	0,00	2,88	4,49	6,05	7,61	9,28	11,60	18,46	27,06	37,17	63,59
4b	100,00	97,12	95,51	93,95	92,39	90,72	88,40	81,54	72,94	62,83	36,41

pH 5.0

P11 010					
time (min)	0	30	122	1080	1370
2	0,00	7,10	10,00	38,56	52,42
4b	100,00	92,90	90,00	61,44	47,58

pH 7.3

P						
time(min)	0	271	1380	2760	4380	11520
2	0,00	1,22	14,36	26,60	48,31	77,57
4b	100,00	98,78	85,64	73,40	51,69	22,43

8. Figure S1. Example of HPLC monitoring of 4b at pH= 4.3



Measure made at t = 17 min

9. Figure S2. Transmission electron miscroscopy images of the synthesized nanoparticles 8.





TEM pictures were performed with a Hitachi H7650 microscope operating at an accelerating voltage of 120 kV. For the particle size and morphology observation, samples diluted about 100 times were deposited on a 200 mesh carbon film-coated copper grids surface ($3 \times 5 \mu$).

10. Figure S3. Groups description and schedule of the in vivo experimental protocol.





11. Figure S4. Quantification of NPs11 accumulation in different organs over time.



Quantification of the biodistribution of NPs 11 in different organs. C57Bl/6 mice bearing orthotopic AK7 tumors were injected with 60 μ g of NPs 11 per g of mice in the tail vein. The graphics represent the quantification of NPs 11 accumulation in dissected tumor, liver, spleen, kidneys, brain, ovaries and in blood over time. Values are means ± S.E.M. of results obtained on 5 mice.

12. Figure S5. Effect of compound 2 and NPs 8 on histone H3 acetylation in tumors.



Immunohistochemistry using anti-acetylated histone H3 antibody and HES staining of tumors. Tissues were fixed in 4% CH₂O in PBS, embedded in paraffin, cut into 5- μ m sections. Immunohistochemistry and histology were performed on tissues slices (paraffin-embedded) by Cellular and Tissue Imaging Core Facility of Nantes University (MicroPICell) using anti-acetylated histone H3 antibody (Active Motif, 1/100) or hematoxylin, eosin and safran staining (HES), respectively. Pictures were obtained using a NanoZoomer 2.0HT (Hamamatsu).

13. Figure S6. Effect of compound 2 and NPs 8 on histone H3 acetylation in liver and in kidneys.



Immunohistochemistry of liver and kidneys using anti-acetylated histone H3 antibody. Tissues were fixed in 4% CH2O in PBS, embedded in paraffin, cut into 5- μ m sections. Immunohistochemistry and histology were performed on tissues slices (paraffin-embedded) by Cellular and Tissue Imaging Core Facility of Nantes University (MicroPICell) using anti-acetylated histone H3 antibody (Active Motif, 1/100). Pictures were obtained using a NanoZoomer 2.0HT (Hamamatsu).