

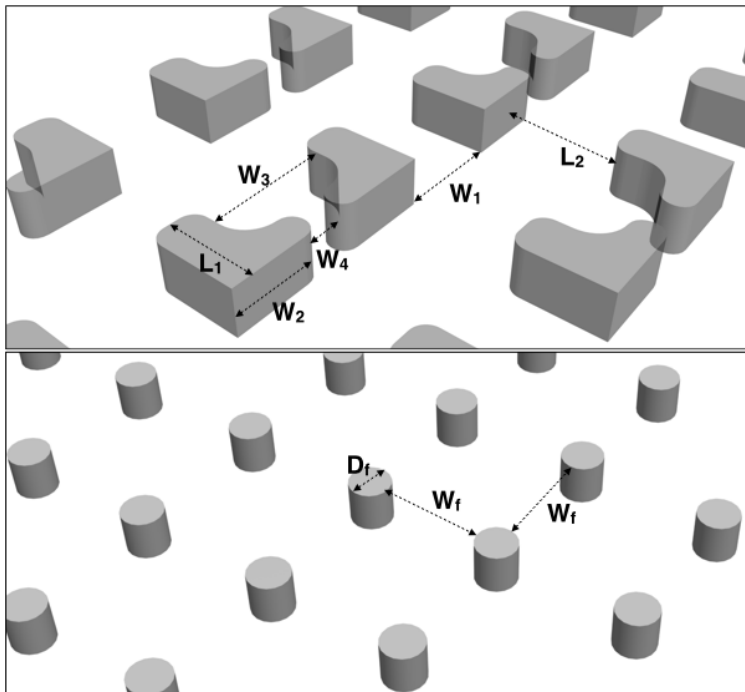
## Supplementary Information

### On Chip Analysis of CNS Lymphoma in Cerebrospinal Fluid

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*Design of microfluidic platform:* The fluidic system has a single-layer structure that is composed of a capture site region, a fluidic channel, and a debris filter at the inlet. Injected fluids (e.g. cells, buffers, antibodies) first pass through the microfilter array (200  $\mu\text{m}$  in diameter) in order to filter large aggregates and debris. The fluids then pass through the capture site region (12000  $\mu\text{m}$  in width; 5800  $\mu\text{m}$  in length). Figure S1 (top) shows the detailed dimensions of the single-cell capture sites, which were designed to capture lymphocytes  $\sim 10\mu\text{m}$  in diameter. There are two capture regions with different gap sizes ( $W_1 = 30\ \mu\text{m}$  and  $16\ \mu\text{m}$ ;  $L_2 = 40\ \mu\text{m}$  and  $25\ \mu\text{m}$ ) to enhance the capture rate. Figure S1(bottom) shows the schematic of a column filter at the sample inlet, that removes large debris and cell aggregates. The height of the fluidic channel is  $25\ \mu\text{m}$ .



**Figure S1.** Fluidic structures in microfluidic chip. (Top) Design parameters for the capture sites:  $W_1 = 30\ \mu\text{m}$  and  $16\ \mu\text{m}$ ;  $W_2 = 10\ \mu\text{m}$ ;  $W_3 = 14\ \mu\text{m}$ ;  $W_4 = 4\ \mu\text{m}$ ;  $L_1 = 15\ \mu\text{m}$ ;  $L_2 = 40\ \mu\text{m}$  and  $25\ \mu\text{m}$ . (Bottom) Structure of the on-chip column filter.  $D_f = 200\ \mu\text{m}$  and  $W_f = 800\ \mu\text{m}$ .

**Table S1.** Cell Counts in CSF

	Normal <sup>1</sup>	CNS lymphoma <sup>2,3</sup>	Inflammation <sup>4</sup>
B cells (per mL)	0-30	10-500,000	200-43,000
T cells (per mL)	150-2,000	250-180,000+ (max 97.2%)	9,000-460,000
Monocytes (per mL)	80-1,100	—	—
Granulocytes (per mL)	20-430	—	—
NK cells (per mL)	0-50	max 7.4%	1,500-50,000

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- 2 Quijano S, Lopez A, Manuel Sancho J, Panizo C, Deben G, Castilla C, Antonio Garcia-Vela J, Salar A, Alonso-Vence N, Gonzalez-Barca E, et al. Identification of Leptomeningeal Disease in Aggressive B-cell Non-hodgkin's Lymphoma: Improved Sensitivity of Flow Cytometry. *J Clin Oncol* **2009**; 27: 1462-1469.
- 3 Schroers R, Baraniskin A, Heute C, Vorgerd M, Brunn A, Kuhnenn J, Kowoll A, Alekseyev A, Schmiegel W, Schlegel U, et al. Diagnosis of Leptomeningeal Disease in Diffuse Large B-cell Lymphomas of the Central Nervous System By Flow Cytometry and Cytopathology. *Eur J Haematol* **2010**; 85: 520-528.
- 4 Kleine TO, Albrecht J, Zofel P. Flow Cytometry of Cerebrospinal Fluid (csf) Lymphocytes: Alterations of Blood/csf Ratios of Lymphocyte Subsets in Inflammation Disorders of Human Central Nervous System (cns). *Clin Chem Lab Med* **1999**; 37: 231-241.

**Table S2.** Antibodies used in the current report.

<b>Antigen</b>	<b>Clone</b>	<b>Provider</b>	<b>Fluorochrome (on chip)</b>
CD10	HI10a	BioLegend	—
CD19	HIB19	BioLegend	R-PE
CD20	2H7	BioLegend	R-PE
CD45	HI30	BioLegend	—
Ki-67	B56	BD Biosciences	Alexa Fluor 488
$\alpha$ light chain	MHK-49	BioLegend	Brilliant Violet 421
$\lambda$ light chain	JDC-12	BD Biosciences	Alexa Fluor 647